

DEVELOPMENT OF ROBUST ANALYTICAL METHODS USING DESIGN SPACE METHODOLOGY

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1. Introduction

Nowadays, the concept of Quality by Design and Design Space (DS) have become widespread in the field of pharmaceutical and analytical science. In the framework of method development, the DS computation can be carried out using design of experiment approach (DoE). The combination of these methodologies (DoE-DS) are applied to optimize the operating conditions governing a separation and to estimate the robustness of the developed method, simultaneously.

1.1. What is the Design Space?

ICH Q8(R2) guideline provides a harmonized guidance to improve the robustness and reliability of pharmaceutical development. In this guideline, DS is defined as "the multidimensional combination and interaction of input variables (e.g. material attributes) and process parameters that have been demonstrated to provide assurance of quality". In the method development framework, the DS defines a space wherein the separation is complete taking into account the uncertainty in models, measurements and process.

This guideline follows with "working within the design space is not considered as a change". Therefore, the DS can be considered as a zone of theoretical robustness as the method parameters modifications do not result in significant variations in terms of separation quality.

1.2. How to compute the Design Space?

First, DoE are used to model the retention factors against some chromatographic factor (e.g. pH, gradient time, temperature, etc.). The beginning, the apex and the end of each peak (i.e. respectively, t_B , t_R and t_E) are individually modelled by multiple linear regressions.

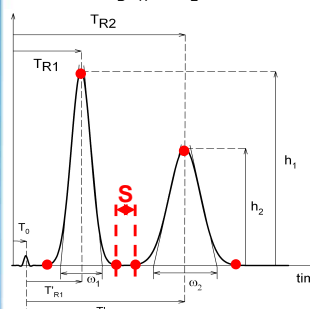


Fig. 1: Calculation of S for the critical pair complete ($S > 0$ min) and where the error is relatively low ($P(S > 0)$ is high) as shown by Eq. 1.

$$DS = \{x_0 \in \chi | E_{\theta} [P(S > \lambda) | \theta] \geq \pi\} \quad \text{Eq. 1.}$$

Where x_0 is a point in the experimental domain, χ , λ is the threshold on this criterion, π is the selected probability of acceptance. Symbols P and E respectively correspond to probability and mathematical expectation.

Thus, the robustness is evaluated by the quality criteria (π) and the DS shape. A high π and a large DS size means that the method is robust.

2. Results

2.1. Pharmaceutical formulation. The first example is the optimization of the separation of a common cold pharmaceutical formulation containing phenylephrine, chlorpheniramine maleate, paracetamol, E110 (sunset yellow) and PVP-K30. A full factorial design was used to model the retention times against the pH and the gradient time.

Factors	Levels				
pH	2.6	4.4	6.3	8.1	10.0
Gradient time (T_G , min)	10	20	30		

Table 1: Full factorial design

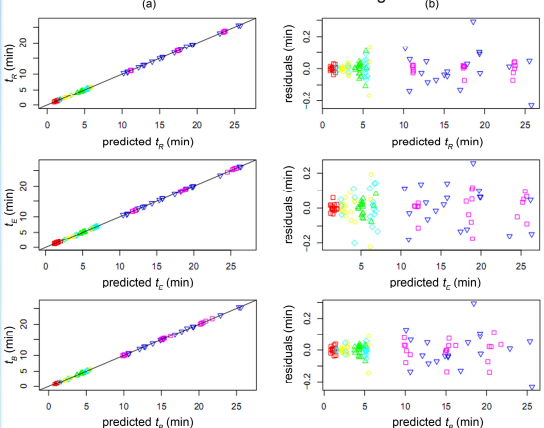


Fig. 2: (a) Predicted versus experimental values for t_R , t_E and t_B . (b) Corresponding residuals plots.

This design leads to 17 experiments. ($n = 15+2$ independent repetitions at the centre)

As depicted on Fig 2, the adequacy between the experimental retention times and the predicted one is very good ($R^2 > 0.99$). Furthermore, the corresponding residuals were normally distributed between -0.2 and 0.2 min.

Maleate
Phenylephrine
Saccharine
Paracetamol
Chlorpheniramine
PVP-K30

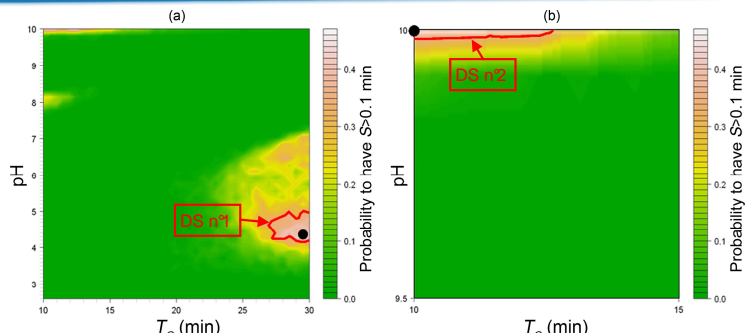


Fig. 3: (a) Probability surface for the separation criterion (acceptance limit $\lambda=0.1$ min) optimum n^1 is depicted by the black circle (pH 4.3 - $T_G=28.0$ min). (b) Magnification of the upper left zone, optimum n^2 is depicted by the black circle (pH 10.0 - $T_G=10$ min).

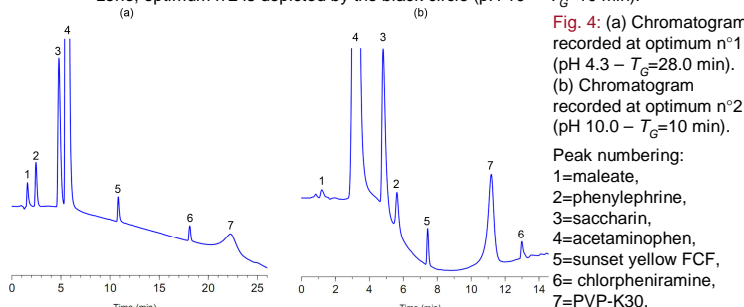


Fig. 4: (a) Chromatogram recorded at optimum n^1 (pH 4.3 - $T_G=28.0$ min). (b) Chromatogram recorded at optimum n^2 (pH 10.0 - $T_G=10$ min).
Peak numbering:
1=maleate,
2=phenylephrine,
3=saccharin,
4=acetaminophen,
5=sunset yellow FCF,
6= chlorpheniramine,
7=PVP-K30.

2.2. Test sample. The second example is the optimization of the separation of a system suitability test mixture. The same full factorial design (see Table 1) was used.

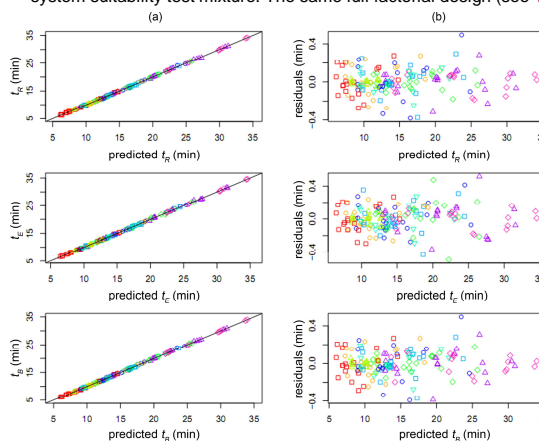


Fig. 5: (a) Predicted versus experimental values for t_R , t_E and t_B . (b) Corresponding residuals plots.

As depicted on Fig 5, the adequacy between the experimental retention times and the predicted one is also very good ($R^2 > 0.995$). Furthermore, the corresponding residuals were normally distributed between -0.4 and 0.4 min.

Peaks labels and numbering
 (1) Atenolol
 (2) Pindolol
 (3) Q290
 (4) Propranolol
 (5) Indoprofen
 (6) Naproxen
 (7) Warfarin
 (8) RA-Imp2
 (9) Retinoic Acid

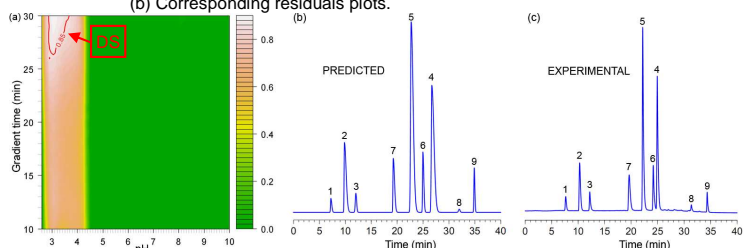


Fig. 6: (a) depicts the probability surface to obtain a separation (S) of at least 0 min. The quality level is set to 85%. (b) predicted chromatogram at pH 3.0 and with $T_G = 30$ min. (c) chromatogram recorded at this operating condition.

This DoE-DS methodology was also successfully applied on a cannabinoids mixture [B. De Backer et al., J. Chrom. B 877 (2009) 4115], on the separation of sulindac and its related impurities [F. Krier et al., J. Pharm. Biomed. Anal., In press], on the separation of 18 antimalarial drugs [R.D. Marini et al., Chemistry Today 28 (2010) 10-14] on extracts of *spirospERM penduliflorum* [M. Rafamantanana et al.] and on extracts of *strychnos usambarensis* [I. Nistor et al.].

3. Conclusions

Even if the number of experiments needed (n) might seem high regarding to conventional optimization approaches, the DoE-DS methodology is able to simultaneously model the chromatographic behaviour of each compound and provides optimal conditions in which the prediction error was analyzed in order to evaluate the method robustness.