

## EFFECTS OF INSECTICIDE TREATMENTS ON INSECT DENSITY AND DIVERSITY IN VEGETABLE OPEN FIELDS

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### ABSTRACT

Vegetable open field areas increased for 15 years in Wallonia, mainly in Hesbaye. To be in accordance with quality standards, especially in terms of agro-chemical residues (R.M.L.), biological pest control was developed and allowed to reduce the insecticide use, leading to have safer fresh products. Among cultivated species in Wallonia, leguminous crops represent more than 85% of the vegetable production. To assess the impact of insecticide treatment on both pests (mainly aphids) and beneficial insects (predators and parasitoids), broad bean (*Vicia faba* L.) fields were investigated during all the production duration. Twelve fields between Waremmes and Hannut were visited weakly from May to July. In each field, control untreated and treated plots were investigated. Insects were caught using yellow traps and determined until the family level. Approximately 90.000 insects belonging to 59 major families (99% of captures) and 64 minor families were identified. These results showed that biodiversity in terms of family numbers was significantly higher in unsprayed plots. In addition, biodiversity and biomass (insect density) increased gradually during the season. Evaluation of pest and beneficial diversity and density was discussed in relation to aphidic ( $\lambda$ -cyhalothrin, pirimicarb) treatments and the development of I.P.M. program in vegetable crops.

**Key-words:** broad bean, aphids, insecticides, yellow traps, biological control

### INTRODUCTION

For the middle of '80 vegetables areas increased in Wallonia (South of Belgium) because of a will to diversify agricultural productions. More than 30% of the Walloon vegetable production is produced in Hesbaye (APHW, 1997), the region we investigated. We choose broad bean according to the economic importance of Leguminous in this region and to crucial aphid control problems. Indeed, among cultivated species in Wallonia, leguminous crops represent more than 85% of the vegetables production (APHW, 1997) moreover *Fabaceae* species are such as pea, bean and broad bean are continuously present in agro-ecosystems as crop or as green fertiliser. Like this, alternative habitats for pests are always available. The main problem associated with such crops is aphid control. To avoid outbreaks, insecticides are widely sprayed, leading to problems as resistance to active substances, environment pollution and agro-chemical residues on food.

Natural enemies contribute greatly to reducing aphid populations and collateral damage in broad bean crops. But, the widespread and routine use of insecticides display negative effects on beneficial species populations.

Several studies such as LOWE *et al.* (1983) have shown that unselective insecticide sprayings lead to a worsening of aphid problems by suppressing beneficials. Future integrated pest management strategies in broad bean crops will be developed especially to minimise adverse effects of insecticide spraying on natural enemies. The conservation of beneficials to control aphid pest by using selective insecticides at lower concentrations is one of the potential ways to promote I.P.M. . The first step of this approach is to determine the effects of common insecticide treatments on broad bean field entomofauna, especially on aphidiphagous natural enemies. The results will be discussed in relation to the impact on aphid control.

## MATERIAL AND METHODS

Twelve broad bean fields were investigated in Hesbaye region between Hannut and Waremmé. Each field was divided in an untreated plot of 250 square meters which only herbicide sprayings received and a normally treated one constituted by the remaining area. In treated plots, required fungicidal and aphicidal treatments were applied at recommended doses. Active substances sprayed were  $\lambda$ -cyhalothrin, pirimicarb (aphicids) alone or in combination and vinclozolin and carbendazim (fungicides). An average of two insecticide and two fungicidal treatments were applied on each field for the ten weeks of cropping. Spraying period was variable for each field but more than nine applications on ten were made between 25<sup>th</sup> May and 28<sup>th</sup> June, with a maximum at sixth week. Eleven on the 12 fields received both pirimicarb and  $\lambda$ -cyhalothrin in association ("Okapi®") or in pure formulation ("Pirimor®" and "Karaté®") and ten on the 12 have been sprayed with both vinclozoline ("Ronilan®") and carbendazim ("Virolex®"). One field received  $\lambda$ -cyhalothrin only and two fields have been sprayed only with "Virolex". Insects were trapped using tree yellow traps per plot. Each trap was disposed on the corner of a one meter triangle and weekly emptied. Insects caught were determined until the family taxonomical level except for *Thysanoptera* (to order) and for some particular beneficials and pests which were determined until the species. Results of the latter work was presented elsewhere (FRANCIS *et al.*, 2001). Trapping last ten weeks (as long as the crop was present), from 2<sup>th</sup> Mai to 11<sup>th</sup> July. Statistical analysis were performed using the Minitab® software (12.2 version). ANOVA was realised on data after a log (x+1) transformation.

## RESULTS

### General diversity

Amount of 89579 insects belonging to nine orders and 123 families were caught and determined on the bean cultivation. More than 99% of them

belong to 59 families while 64 families were represented only by 325 individuals (Appendix 1). *Diptera* (57108 indiv.) was largely the most abundant order (Fig. 1). *Hemiptera* (9999 indiv.), *Hymenoptera* (8394 indiv.), *Coleoptera* (6855 indiv.) and *Thysanoptera* (6107 indiv.) represent each one from 11 to 7%. The four other orders (*Neuroptera*, *Mecoptera*, *Lepidoptera* and *Odonatoptera*) corresponded to only 1103 individuals.

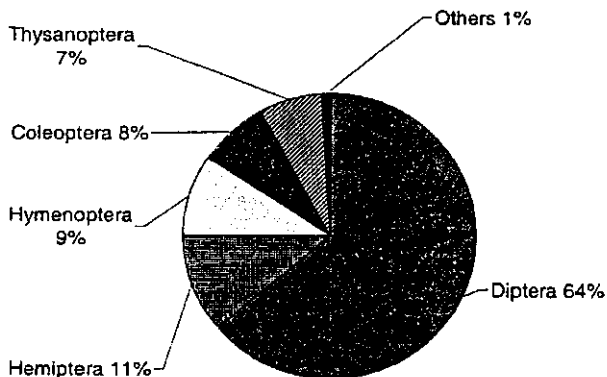


Figure 1 : Insects repartition according to taxonomical orders

### Effects of treatments on insect density

Insects density in traps remained relatively low and stable between 54.1 and 91.7 insects during weeks 2 to 5 (Fig. 2). Then captures became fewer on week 6 before to increase constantly the two following weeks. Insect density dropped on week 9 before to increasing again last week. On the whole spraying period (weeks 4 to 10), no significant was observed between the two plots in term of insects density difference ( $P = 0.367$  ;  $F = 0.82$ ). Considering each trapping week separately, significant difference between plots was observed only week 6 ( $P = 0.02$  ;  $F = 9.90$ ). At this time, insect density was higher in untreated plots traps (59.5 indiv./trap) than in treated ones (54.1 indiv./trap).

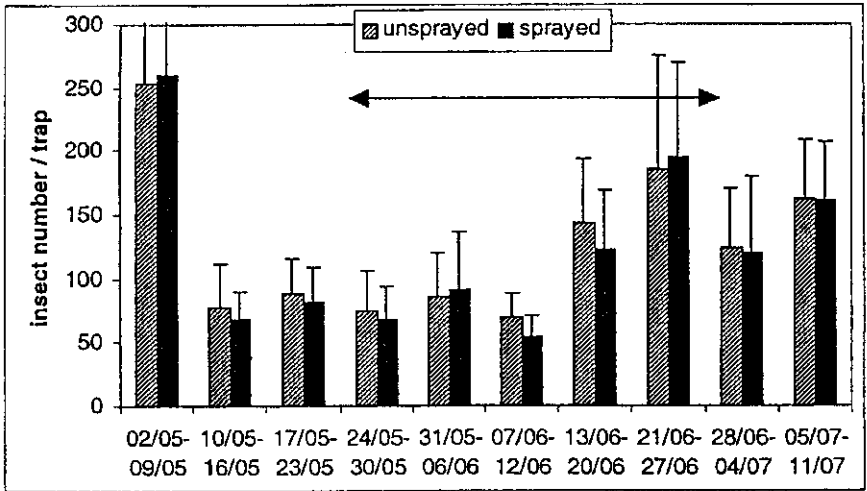


Figure 2: Mean insect number per yellow trap in relation to insecticide applications (arrow on the graph indicates treatments period)

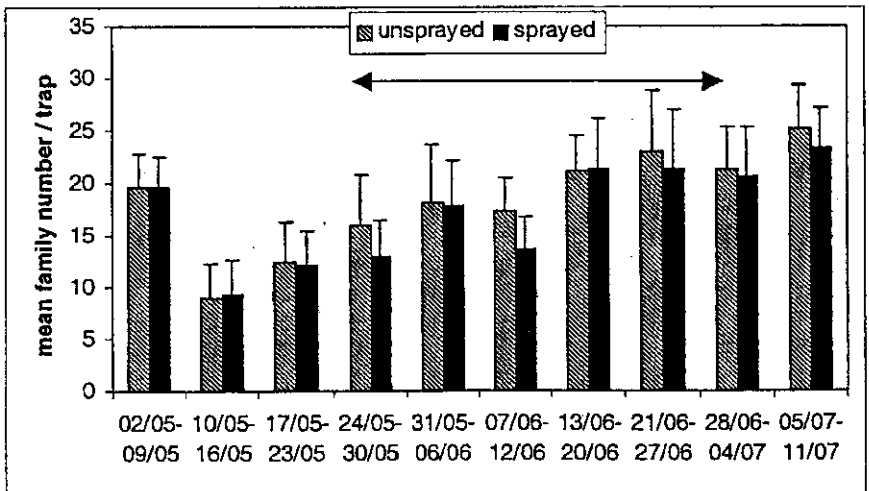


Figure 3: Mean insect number per yellow trap in relation to insecticide applications (arrow on graph indicates treatments period)

### Effects of treatments on insect diversity

Insect diversity in term of family number per trap showed a trend to increase through the time (Fig. 3). Untreated plot traps (20.3 families) were

significantly richer in families on the spraying period than treated plot ones (18.8 families) ( $P = 0.023$ ;  $F = 5.17$ ).

Obvious difference between treated and untreated plots, started at the beginning of spraying period (fourth week). Except at week seven, treated plots remained less diversified until crop end. Significant differences were observed week four and six ( $0.000 < P < 0.005$ ;  $8,58 < F < 9.90$ ).

### Effects of treatments on insect families

Fifty eight families were sufficiently numerous to allow statistical analysis. During insecticide spraying period, 37 families were more abundant in untreated plot traps (Fig. 4) whereas 22 families were caught in higher number in treated plot traps (Fig. 5). Marked differences are observed for height families more abundant in untreated plots (*Crysolmelidae*, *Sphecidae*, *Dolichopodidae*, *Elateridae*, *Pieridae*, *Panorpidae*, *Carabidae* and *Tachinidae*). Fifteen families including important pests or beneficials (*Carabidae*, *Elateridae*, *Tipulidae*, *Cantharidae*, *Coccinellidae*, *Aphididae*) in broad bean crop, were significantly more present in untreated plot traps ( $0.000 < P < 0.054$ ;  $3.18 < F < 22.18$ ). Only two families of micro-wasps were significantly ( $0.050 < P < 0.055$ ;  $3.18 < F < 3.87$ ) more numerous in treated plots.

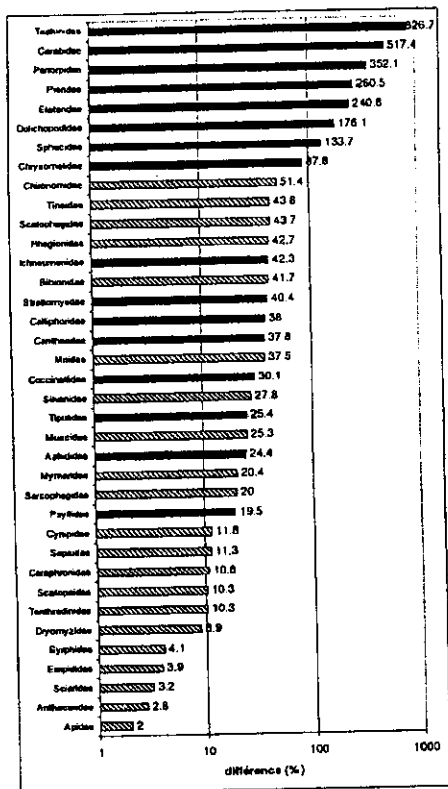


Figure 4: Differences of insect families (in %) between treated and untreated plots (families decreasing in untreated plots) Significant differences at  $P = 0.05$  are black colored

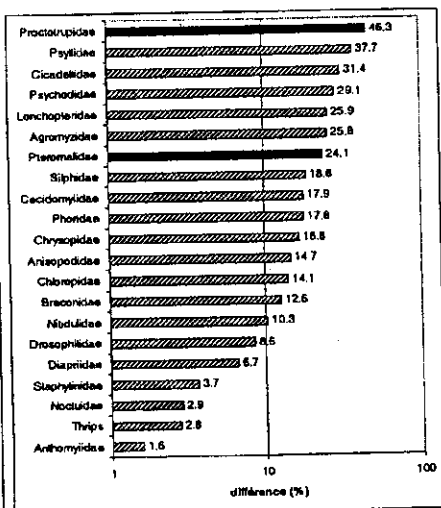


Figure 5: Differences of insect families (in %) between treated and untreated plots (families increasing in treated plots) Significant differences at  $P = 0.05$  are black colored

## DISCUSSION

Effects of treatments on insect density were unclear (may be due to the relatively low pressure of treatment in comparison on a "standard season"). During weeks 2, 3, 4, 6 and 9 mean insect density in untreated plots was higher than in treated ones. Inverse phenomenon was observed on weeks 5 and 8, in the middle of spraying period. Small differences (never more than 23%) and large standard deviations explained that significant differences were never observed, except on sixth week. The spring fly of *Cavariella aegopodii* Scop., an aphid species not damageable for broad bean, explained abnormal density in traps observed on first week. Treatment effects on broad bean insect fields diversity was more obvious. From the

beginning of insecticide sprayings, treated plots remained globally less diversified and effects on diversity continued after the end of sprayings. Information on insect density were disconcerting. This could be due to a rapid re-colonisation from surrounding environment which was frequently observed in small plots (Holland *et al.*, 2000). While we observed a decrease of aphid populations after insecticide treatments, negative effects on numerous beneficial insects families were also demonstrated. Among predator families, *Carabidae* and *Dolichopodidae* were highly affected by insecticide applications and were confirmed to be very sensible families (Vickerman *et al.*, 1987). Moreover, *Coccinellidae* were significantly less present in sprayed plots ( $P = 0.042$ ;  $F = 4.19$ ) and. This is more worrying concerning the aphid control. Beetle density in the field is influenced by insecticide applications but is also linked to the prey abundance (Hodek and Honek, 1996). Prey host scarcening could partially explain the reduction of *Coccinellidae* presence in treated plots. At the opposite, syrphid adults are extremely mobile and were present in traps not depending on treatments which not seemed affect syrphid abundance. Indeed, yellow traps are adapted to catch only insect mobile stages like syrphid adults. Then, only observations of nymphal larvae could provide accurate information on insecticide treatments effects and allow to conclude in term of aphid control. Paradoxically, treatments were not negative towards 21 families and demonstrate the presence of indirect and complex interactions between insect species. This phenomenon was frequently observed with phytophagous species (Evans *et al.*, 1996). Here, no serious pest family was clearly favoured by sprayings. At the opposite, treatments enhance two important families of parasitoid (*Pteromalidae* and *Proctotrupidae*). Nevertheless, beneficial effects of spraying were globally less marked than noxious ones. Finally, treatment didn't cause the disappearance of one of the 58 families which were statistically discussed. Observed disappearance of *Bibionidae* and *Cantharidae* seemed to be only linked to their own phenology. Indeed, they have disappeared from untreated plots at the same time.

## CONCLUSIONS

This study showed that there is a possibility that prophylactic and systematic treatments exacerbate the problems of aphids in broad bean by reducing important natural enemies populations, even if a part of treatments was mad using pirimicarb. Proposed solutions should be monitor insect populations in the field and reduce insecticide concentrations whose increase selectivity. These actions, linked to closed environment survey (Colignon *et al.*, 2000), should contribute to a future integrated pest management strategy in broad bean crops.

## ACKNOWLEDGEMENTS

The authors would like to thank the Walloon Region Agriculture Minister and particularly Mr Happart, for the funding of this work which is part of a research program entitled "Evaluation et utilisation de l'entomofaune utile en cultures maraîchères de plein champ".

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## Appendix 1: Insect total numbers according to families

Order	Family	Number	Order	Family	Number
THYSANOPTERA		6107	ODONATOPTERA	<i>Platycnemidae</i>	1
HEMIPTERA	<i>Aphididae</i>	8797	ORTHOPTERA	<i>Acridae</i>	5
	<i>Cicadellidae</i>	292		<i>Tetrigidae</i>	2
	<i>Psyllidae</i>	647	HEMIPTERA	<i>Coreidae</i>	5
	<i>Miridae</i>	221		<i>Lygaeidae</i>	2
	<i>Anthracoridae</i>	18		<i>Nabidae</i>	11
NEUROPTERA	<i>Chrysopidae</i>	316		<i>Pentatomidae</i>	5
MECOPTERA	<i>Panorpidae</i>	74		<i>Reduviidae</i>	1
LEPIDOPTERA				<i>Cercopidae</i>	6
	<i>Pieridae</i>	64	NEUROPTERA	<i>Hemerobidae</i>	1
	<i>Noctuidae</i>	576	LEPIDOPTERA	<i>Incurvaridae</i>	1
	<i>Theidae</i>	57		<i>Nymphalidae</i>	7
DIPTERA	<i>Syrphidae</i>	6464		<i>Dioscorophoridae</i>	1
	<i>Anthomyiidae</i>	20501		<i>Papilionidae</i>	4
	<i>Muscidae</i>	1176		<i>Phorophoridae</i>	1
	<i>Calliphoridae</i>	7987	DIPTERA	<i>Anthomyzidae</i>	1
	<i>Scatophagidae</i>	565		<i>Asilidae</i>	3
	<i>Sarcophagidae</i>	475		<i>Ceratopogonidae</i>	1
	<i>Tachinidae</i>	38		<i>Chamaemyiidae</i>	1
	<i>Agromyzidae</i>	192		<i>Coelopaedidae</i>	5
	<i>Anisopodidae</i>	74		<i>Conopidae</i>	5
	<i>Bibionidae</i>	997		<i>Culicidae</i>	5
	<i>Cecidomyiidae</i>	106		<i>Ephyrididae</i>	2
	<i>Chironomidae</i>	87		<i>Heleomyzidae</i>	11
	<i>Chloropidae</i>	1523		<i>Lauxaniidae</i>	9
	<i>Dolichopodidae</i>	581		<i>Lonchaeidae</i>	1
	<i>Drosophilidae</i>	4850		<i>Micropezidae</i>	6
	<i>Dryomyzidae</i>	133		<i>Opomyzidae</i>	6
	<i>Empididae</i>	4847		<i>Otilidae</i>	4
	<i>Lonchopteridae</i>	180		<i>Palaopteridae</i>	2
	<i>Mycetophoridae</i>	400		<i>Pipunculidae</i>	2
	<i>Phoridae</i>	224		<i>Pleystomatidae</i>	6
	<i>Psilidae</i>	117		<i>Scleromyzidae</i>	10
	<i>Psychodidae</i>	70		<i>Simuliidae</i>	8
	<i>Rhagionidae</i>	37		<i>Tephritidae</i>	16
	<i>Scalopsidae</i>	315		<i>Xylophagidae</i>	3
	<i>Sciariidae</i>	422	HYMENOPTERA	<i>Bethylidae</i>	3
	<i>Sepsidae</i>	1811		<i>Chrysididae</i>	8
	<i>Stratiomyidae</i>	892		<i>Encyrtidae</i>	2
	<i>Tipulidae</i>	1938		<i>Eucharitidae</i>	1
HYMENOPTERA	<i>Tenthredinidae</i>	1685		<i>Eulophidae</i>	2
	<i>Ichneumonidae</i>	1066		<i>Eumenidae</i>	2
	<i>Apidae</i>	2487		<i>Eupeimidae</i>	8
	<i>Braconidae</i>	629		<i>Eurytomidae</i>	3
	<i>Ceraphronidae</i>	106		<i>Evanidae</i>	1
	<i>Cynipidae</i>	1051		<i>Formicidae</i>	7
	<i>Diapriidae</i>	135		<i>Leucospidae</i>	1
	<i>Myrmecidae</i>	150		<i>Perilampidae</i>	17
	<i>Proctotrupidae</i>	170		<i>Pompilidae</i>	4
	<i>Pteromalidae</i>	809		<i>Trichogrammatidae</i>	3
	<i>Sphecidae</i>	27		<i>Torymidae</i>	10
COLEOPTERA	<i>Cantharidae</i>	560		<i>Vanhorniidae</i>	1
	<i>Cerambycidae</i>	34		<i>Vespididae</i>	6
	<i>Chrysomelidae</i>	1010	COLEOPTERA	<i>Buprestidae</i>	1
	<i>Curculionidae</i>	819		<i>Cerambycidae</i>	1
	<i>Elaeidae</i>	53		<i>Cetoniidae</i>	6
	<i>Nitidulidae</i>	1830		<i>Histeridae</i>	29
	<i>Staphinidae</i>	58		<i>Hydrophilidae</i>	16
	<i>Sitonaidae</i>	419		<i>Lucanidae</i>	1
	<i>Staphilinidae</i>	725		<i>Oedemeridae</i>	13
	<i>Coccinellidae</i>	1262		<i>Melyridae</i>	1
				<i>Mordellidae</i>	11
				<i>Scarabaeidae</i>	4
				<i>Tenebrionidae</i>	2