STRYCHNOCHROMINE, AN UNUSUAL C₁₈ INDOLINE ALKALOID FROM STRYCHNOS GOSSWEILERI

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Abstract—Through a combination of spectroscopic techniques (UV, IR, MS, ¹H and ¹³C NMR) it has been possible to deduce the plane structure and the relative configuration of strychnochromine, a dextrorotatory alkaloid, isolated from the root bark of *Strychnos gossweileri*. The alkaloid is 4a-(2-hydroxyethyl)-4-oxo-1,2,2a,3,4,4a,5,10,11,12a-decahydropyrrolizino-[1,7-cd]-carbazol.

INTRODUCTION

Extensive study of the constituents of the African plant Strychnos gossweileri has resulted in the isolation of a number of alkaloids among which is the new and peculiar compound, strychnochromine [1-5]. The name strychnochromine, derived from $\chi\rho\omega\mu\alpha$ (Greek = colour), was chosen by one of us (C.C.) because this substance was the only alkaloid from ca 100 isolated at Liège from numerous Strychnos species, to present notable and stable colours with various TLC spray reagents (see Experimental) [3, 6]. As will be shown, its structure (1) is of interest because of its unusual skeleton.

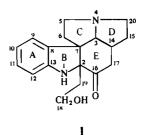
RESULTS AND DISCUSSION

The elemental composition $C_{18}H_{22}N_2O_2$ was established by high-resolution mass measurement of the [M]⁺ at m/z 298. Beside the ubiquitous m/z 130, 143 and 144 characteristic of the tryptamine moiety, the most important peaks were found at m/z 229, 200 (base peak), 187, 182 and 112.

The presence of a hydroxyl group, suspected by the peak at m/z 281 (loss of OH), was confirmed by acetylation of 1 (pyridine-Ac₂O) at room temperature. Comparison of the mass spectrum of 1 with that of the acetylated derivative revealed that the peaks at m/z 298, 229, 200 and 187 were shifted to 340 [M]⁺, 271 [M-C₄H₅O]⁺, 242 [M-C₅H₈NO]⁺ and 229 [M-C₆H₉NO]⁺, respectively, in the monoacetylated derivative.

The UV absorption spectrum, suggestive of an indoline, exhibited in addition a significant absorbance in the 330 nm region, which revealed the existence of a further chromophore. The UV spectrum is somewhat reminiscent of that of a 2-acylindole [7, 8]. [The lone-pair electrons of the amino function (N-1) attached to the benzene ring may interact with the π -system of the carbonyle chromophore]. The IR spectrum exhibited a band at 1700 cm⁻¹ assignable to a carbonyl, possibly a ketone, and a band at 745 cm⁻¹, attributable to an *ortho*-disubstituted benzene ring.

A more detailed understanding of the structure of the molecule was gained from its 360 MHz ¹H NMR spectrum (Table 1). In addition to the expected foursome of resonances from the aromatic nucleus, there are two mobile protons (NH/OH) and three mutually non-interacting spin systems, two -CH2CH2- units and one $-C(=O)-CH_2-CH(-CH)-CH_2CH_2$ system. One of the ethylene units is due to an hydroxyethyl (HOCH₂CH₂-) appendage, as is shown by the characteristic high-frequency shift of the $-OCH_2$ resonances in O-acetyl strychnochromine. The second ethylene unit probably arises from a tryptamine moiety; it features a noteworthy chemical shift difference of $\delta 1.31$ for the two diastereotopic hydrogen atoms of one of the methylenes. In the eight spin system -C(=O)-CH₂-CH (-CH)-CH₂CH₂the presence of a carbonyl next to the lone methylene group rests on two properties of the hydrogen atoms. Firstly the magnitude of the gem coupling constant (-17 Hz) and secondly their very easy exchange for deuterium on treatment of strychnochromine with de-



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	Chemical	Chemical		
н	shift	Н	shift	
3	2.97	15A	2.22	
5 A	3.18	15 B	1.66	
5B	2.95	17 A	2.98	
6A	3.27	17 B	2.51	
6 B	1.96	18A	3.94	
9	6.94	18 B	3.95	
10	6.62	19A	2.50	
11	7.02	19 B	2.18	
12	6.53	20 A	2.89	
14	2.41	20 B	2.50	

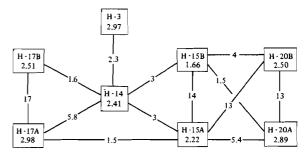
*Certain shift equivalences in CDCl₃ are raised in mixtures of CDCl₃-C₆H₆, sometimes at the expense of creating new ones. All couplings were verified by double irradiation experiments. Resolution enhancement was used in the extraction of the coupling constants. Coupling constants (Hz) $J_{6A,6B} = 12.8; J_{5A,5B} = 12.4; J_{6A,5A} = 6; J_{6A,5B} = 8.5;$ $J_{6B,5A} = 10.5; J_{6B,5B} = 4; J_{18A,18B} = 11.2; J_{19A,19B} =$ = 14.8.

uterium oxide. The chemical shift of the distal methylene and of the methine shows that they must be bonded to nitrogen.

On the basis of the above data, we propose that strychnochromine has structure 1. The formula explains why after reacting strychnochromine with an excess of methyl iodide/sodium hydride the FAB mass spectrum indicates $[M]^+$ at m/z 355, i.e. an increment of 57 mass units. There are indeed four methylation sites, OH, NH (indoline), N (quaternization) and the enol form of the ketone.

The ¹³C NMR chemical shifts of strychnochromine are presented in Table 2; they are fully supportive of structure 1. There is a ketonic carbonyl, four protonated and two non-protonated aromatic carbon atoms, one of which has a relatively small δ value, while the aliphatic region contains two non-protonated carbons, two methines and seven methylenes. Three of the methylenes, C-17 and those of the hydroxyethyl side-chain, were unambiguously assigned by reference to C-17 deuterated and to Oacetyl strychnochromine, respectively. The pair of methylenes at δ 55 and 44 has a larger sum of ¹J H, ¹³C coupling than the pair of methylenes at $ca \ \delta 28$, which shows that the former carbons are bonded to nitrogen and must be assigned to the pair (C-5, C-20). It follows that the pair (C-15, C-6) lies at $ca \delta 28$ and such a δ value for C-6 is rather small in comparison with numerous other indoline alkaloids [9].

There remains to consider the stereochemistry of strychnochromine, i.e. the relative configuration of the four asymmetric carbon atoms. The small ${}^{3}J$ H-3, H-14 (2.3 Hz) shows that the rings E and D are *cis*-fused. In ring D, the CH₂CH₂ fragment features for one pair of vicinal protons a very large coupling constant (13 Hz), and for the other pair a very small one (1.5 Hz) which constitutes evidence that this ring assumes a unique conformation with a large torsion angle C-14-C-15-C-20-N-4. But



Scheme 1. Chemical shifts and coupling constants of selected ¹H spin systems of 1.

because the opposite part of the D ring must be relatively unpuckered, rings C and D are cis-fused. Based on the following facts and reasoning, E and C are also cis-fused. H-14 is equatorial with respect to ring D, because ${}^{3}J$ to both C-15 hydrogens is small, but also with respect to ring E, and for the same reason. This peculiar orientation is confirmed by a long-range coupling of 1.5 Hz between the axial H-15 atom and one of the H-17 atoms, which implies that the torsion angles defined by the atoms H-C-17-C14-C-15-H under consideration are both anti (W pattern of long-range coupling). Thus, not only is C-14-C-17 axial with respect to D, but C-17-C-16 continues that path away from D, and then for geometric reasons E and C and cis-fused. As a corollary, a twist conformation may be ascribed to ring E-nothing exceptional for a cyclohexanone. The tricyclic moiety C, D, E is thus all-cis configured. As far as the B, E fusion is concerned, a number of chemical shift phenomena that we have drawn attention to earlier are rationalized by accepting a synperiplanar (eclipse) orientation of the C-7-C-6 and the C-2-C-19 bonds: the truly enormous chemical shift difference of H-6 A and H-6 B, the low frequency position of C-6 and perhaps even the chemical shift of C-8. By implication, the rings E and B are cis-fused.

Concerning the absolute stereochemistry of strychnochromine, we feel that it would be hazardous to deduce it from the presently reported chiroptical properties because adequate model or reference compounds are lacking. The absolute configuration can in principle also be obtained by Bijvoet X-ray diffraction on single crystals but so far we have failed to obtain strychnochromine in the crystalline state.

Structure 1 is notable for two reasons, the pyrrolizidine moiety (rings C and D) instead of the concomitant presence of pyrrolidine (ring C) and piperidine (ring D) and the hydroxyethyl side chain located at C-2 rather than at C-20. Strychnochromine presents some analogies but also striking differences from desethylibophyllidine and other alkaloids of the 21-nor-(+) pandolane type, previously isolated from the leaves of *Tabenanthe iboga*, *T.* subsessilis and *Tabernaemontana albiflora* [10, 11]. Therefore, 1 might arise biogenetically by oxidative fission of the 20-21 bond of a hypothetical precursor and loss of C-21 followed by contraction of the usual piperidine ring and migration of the sidechain (C-18–C-19) to C-2.

Table 2. ¹³C NMR spectral data of 1 (22.4 MHz, CDCl₃, TMS as internal standard)

С	Chemical shift	Multiplicity	С	Chemical shift	Multiplicity
2	58:4	S	12	112.0	d
3	67.3	d	13	143.6	S
5	44.7ª	t	14	40.5	d
6	27.5 ^b	t	15	28.0 ^b	t
7	53.8	S	16	206.7	\$
8	119.2	S	17	42.0	t
9	126.4	d	18	58.5	t
10	116.9	d	19	38.5	t
11	128.0	d	20	55.2ª	t

^{a,b} These values may be interchanged.

EXPERIMENTAL

Plant material. Root bark of *S. gossweileri* Exell. was collected at Matadi in the Bas-Zaïre region (province of Kinshasa) of Zaïre (Voucher specimen DUVIGNEAUD 193, Herbarium of the National Botanical Garden of Belgium at Meise).

Isolation. Extraction followed the usual procedure, which has been described elsewhere [1, 3]. Strychnochromine is a minor alkaloid present in the fraction of tertiary alkaloids containing mainly dolichantoside. The mixt. was purified by prep. TLC on silica gel (2 mm) in the system EtOAC-*iso*PrOH-NH₄OH (16:3:1) and elution with MeOH from which 1 is pptd by Et₂O. The free base is a white and amorphous powder, sol. in CHCl₃ and MeOH. NMR spectra were run in CDCl₃ (¹³C) or in CDCl₃-C₆D₆ (¹H). Strychnochromine must be stored in a cool, dry place, protected from light.

Detection of 1 by TLC. Colour reactions [12]: (i) H₂SO₄ gives a violet colour, stable for several years [6], (ii) Ce(SO₄)₂ in H₂SO₄ gives a red colour turning violet after some hr, (iii) FeCl₃ in HClO₄ also gives a red colour turning violet, (iv) Van Urk's reagent (*p*-dimethylaminobenzaldehyde-HCl) gives a colour varying from turquoise to violet after some hr, (v) Gibbs's reagent (dibromoquinonechlorimide) also gives a turquoise colour slow-ly turning violet.

Spectral analysis of 1. UV $\lambda_{max}^{MeOH.}$ nm (log ε) 208 (4.46), 248 (4.23), 330 (3.43). IR ν_{max}^{KBr} cm⁻¹ 3400, 2930, 1700 (>C=O), 1600, 1480, 1440, 1405, 1320, 1260, 1105, 1040, 745 (unsubstituted indoline ring) cm⁻¹. EIMS 70 eV, m/z (rel. int.) 298 [M]⁺ (C₁₈H₂₂N₂O₂ obs. 298.1681; calc. 298.1672), 281 (12), 253 (5), 229 (27), 200 (100), 187 (52), 182 (27), 172 (22), 169 (15), 156 (8), 154 (8), 144 (9), 130 (10), 112 (18). Peak matching measurements on acetylated 1: 340.1783 (C₂₀H₂₄N₂O₃ requires 340.1786), 281.1648 (C₁₈H₂₁N₂O requires 281.1653), 271.1443 (C₁₆H₁₉N₂O₂ requires 271.1446), 242.1175 (C₁₅H₁₆NO₂ requires 242.1181), 229.1104 (C₁₄H₁₅NO₂ requires 229.1103), 182.0969 (C₁₃H₁₂N requires 182.0970). CD: Δε(λ_{max}): +28.3 (330 nm), -4.3 (288 nm), -13 (265 nm), +8.6 (235 nm) in MeOH (c = 0.005). [α]_D²⁰: +600° (MeOH; c 0.05). ¹H and ¹³C NMR data: see Tables 1 and 2.

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REFERENCES

- 1. Coune, C. and Angenot, L. (1978) Planta Med. 34, 53.
- 2. Coune, C. and Angenot, L. (1978) Phytochemistry 17, 1447.
- 3. Coune, C. (1978) Pl. Med. Phyt. 12, 106.
- 4. Coune, C. and Angenot, L. (1980) Herba Hungarica 19, 189.
- 5. Coune, C., Tavernier, D., Caprasse, M. and Angenot, L.
- (1984) Planta Med. 50, 93.
 6. Denoël, A., Jaminet, F., Detilleux, G., Van Sumsen, M. and Merveille, L. (1953) Contribution à l'étude chimique des Strychnos du Congo Belge p. 155. Ministère des colonies, Bruxelles.
- Tits, M. Tavernier, D. and Angenot, L. (1980) *Phytochemistry* 19, 1531.
- 8. Verpoorte, R. (1986) J. Nat. Prod. 49, 1.
- Verpoorte, R., Van Beek, T. A., Riegman, R. L. M., Hylands, P. J. and Bisset, N. G. (1984) Org. Magn. Res. 22, 328 and 345.
- Khuong-Huu, F., Cesario, M., Guilhem, J. and Goutarel, R. (1976) Tetrahedron 32, 2539.
- 11. Kan, C. Husson, H. P., Jacquemin, H., Kan, S. K. and Lounasmaa, M. (1980) Tetrahedron Letters 55.
- Baerheim-Svendsen, A. and Verpoorte, R. (1983) J. Chromatogr. Library Vol 23A, Chromatography of Alkaloids part A TLC. Elsevier, Amsterdam.

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