



Original Investigation | Surgery

# Effect of a Combined Drug Approach on the Severity of Ischemia-Reperfusion Injury During Liver Transplant

## A Randomized Clinical Trial

Nicolas Meurisse, MD; Markoen Mertens, MD; Steffen Fieuws, PhD; Nicholas Gilbo, MD, PhD; Ina Jochmans, MD, PhD; Jacques Pirenne, MD, PhD; Diethard Monbaliu, MD, PhD

### Abstract

**IMPORTANCE** In a porcine model of liver transplant, a combined drug approach that targeted the donor graft and graft recipient reduced ischemia-reperfusion injury, a major hurdle to the success of liver transplant.

**OBJECTIVE** To assess the effect of a clinical form of a perioperative combined drug approach delivered immediately before implantation to the procured liver and to the liver recipient on the degree of ischemia-reperfusion injury.

**DESIGN, SETTING, AND PARTICIPANTS** This unicentric, investigator-driven, open-label randomized clinical trial with 2 parallel arms was conducted in Belgium from September 2013 through February 2018, with 1-year follow-up. Adults wait-listed for a first solitary full-size liver transplant were screened for eligibility. Exclusion criteria were acute liver failure, kidney failure, contraindication to treatment, participation in another trial, refusal, technical issues, and death while awaiting transplant. Included patients were enrolled and randomized at the time of liver offer. Data were analyzed from May 20, 2019, to May 27, 2020.

**INTERVENTIONS** Participants were randomized to a combined drug approach with standard of care (static cold storage) or standard of care only (control group). In the combined drug approach group, following static cold preservation, donor livers were infused with epoprostenol (ex situ, portal vein); recipients were given oral  $\alpha$ -tocopherol and melatonin prior to anesthesia and intravenous antithrombin III, infliximab, apotransferrin, recombinant erythropoietin- $\beta$ , C1-inhibitor, and glutathione during the anhepatic and reperfusion phase.

**MAIN OUTCOMES AND MEASURES** The primary outcome was the posttransplant peak serum aspartate aminotransferase (AST) level within the first 72 hours. Secondary end points were the frequencies of postreperfusion syndrome, ischemia-reperfusion injury score, early allograft dysfunction, surgical complications, ischemic cholangiopathy, acute kidney injury, acute cellular rejection, and graft and patient survival.

**RESULTS** Of 93 randomized patients, 21 were excluded, resulting in 72 patients (36 per study arm) in the per protocol analysis (median recipient age, 60 years [IQR, 51.7-66.2 years]; 52 [72.2%] men). Peak AST serum levels were not different in the combined drug approach and control groups (geometric mean, 1262.9 U/L [95% CI, 946.3-1685.4 U/L] vs 1451.2 U/L [95% CI, 1087.4-1936.7 U/L]; geometric mean ratio, 0.87 [95% CI, 0.58-1.31];  $P = .49$ ) (to convert AST to  $\mu$ kat/L, multiply by 0.0167). There also were no significant differences in the secondary end points between the groups.

*(continued)*

### Key Points

**Question** Can a combined drug approach targeting the preimplantation liver graft and recipient attenuate the degree of ischemia-reperfusion injury?

**Findings** In this open-label randomized clinical trial of 72 liver transplant recipients allocated to either static cold storage (standard of care) with an add-on combined drug approach delivered to the preimplantation liver and recipient or to standard of care only, peak aspartate aminotransferase serum levels and other functional, laboratory, and survival outcomes were similar between groups.

**Meaning** The findings suggest that use of a downstream combined drug approach that targets the preimplantation liver graft and recipient is not clinically effective for decreasing ischemia-reperfusion injury.

+ [Visual Abstract](#)

+ [Supplemental content](#)

Author affiliations and article information are listed at the end of this article.

**Open Access.** This is an open access article distributed under the terms of the CC-BY License.

Abstract (continued)

**CONCLUSIONS AND RELEVANCE** In this randomized clinical trial, the combined drug approach targeting the post-cold storage graft and the recipient did not decrease ischemic-reperfusion injury. The findings suggest that in addition to a downstream strategy that targets the preimplantation liver graft and the graft recipient, a clinically effective combined drug approach may need to include an upstream strategy that targets the donor graft during preservation. Dynamic preservation strategies may provide an appropriate delivery platform.

**TRIAL REGISTRATION** ClinicalTrials.gov Identifier: [NCT02251041](https://clinicaltrials.gov/ct2/show/study/NCT02251041)

*JAMA Network Open.* 2023;6(2):e230819. doi:10.1001/jamanetworkopen.2023.0819

## Introduction

Liver transplant is a life-saving therapy for liver failure, but its success is limited by a marked shortage of organs and high mortality among individuals on the waiting list.<sup>1</sup> Attempts to improve this imbalance have led to less restrictive organ donation criteria and an expansion of the donor pool with so-called extended-criteria donors, including donors of more advanced age, donors with steatotic livers, donation after circulatory death (DCD), and donor livers exposed to prolonged static cold ischemia time (CIT).<sup>2</sup>

Livers from extended-criteria donors are more susceptible to ischemia-reperfusion injury (IRI),<sup>3</sup> the principal cause of liver graft dysfunction and primary nonfunction after liver transplant.<sup>4-6</sup> Ischemia-reperfusion injury is a self-amplifying process involving 2 interrelated phases of ischemic and reperfusion injury.<sup>7</sup> The ischemic insult is characterized by a dysfunction of the mitochondrial respiratory chain,<sup>8</sup> sinusoidal endothelial cell barrier impairment and Kupffer cell activation,<sup>9</sup> activation of cell death programs, and alterations in expression of genes, such as hypoxia-inducible factor.<sup>5,8</sup> At reperfusion, the mitochondrial burst of reactive oxygen species (ROS) overburdens the endogenous antioxidant capacity<sup>9,10</sup> and ATP depletion deepens, all contributing to cell death.<sup>8,11</sup> The subsequent expression of danger-associated molecule patterns activates innate immune responses that promote the release of inflammatory cytokines and chemokines and complement and coagulation factors.<sup>7,8,11</sup> Ischemia-reperfusion injury invariably results in local damage that may lead to graft dysfunction,<sup>5</sup> ischemic cholangiopathy (IC),<sup>12</sup> and remote organ dysfunction such as acute kidney injury (AKI).<sup>13,14</sup> Since IRI is associated with increased short- and long-term morbidity and mortality,<sup>15</sup> finding strategies to effectively mitigate IRI is considered a research priority in the field of liver transplantation.

Over the past decades, evidence from small- and large-animal models has indicated that pharmacological targeting of IRI pathways is a promising strategy to attenuate IRI,<sup>7,16</sup> for example, through reduction of ROS and ROS-induced effects, blockade of immune activation, or modulation of cytokine responses.<sup>3</sup> While, to our knowledge, only 3 randomized clinical trials<sup>17-19</sup> have shown clinical efficacy of single-drug treatment with inhaled nitric oxide, *N*-acetylcysteine, or recombinant P-selectin glycoprotein ligand IgG, none of these agents are used in clinical practice. Since IRI has multiple pathogenetic pathways, a multitarget pharmacological strategy that interferes with several steps in the IRI cascade is likely to be more effective.<sup>16,20,21</sup> Preclinical evidence to support this comes from work by some of us in a stringent porcine DCD model of liver transplant and severe IRI in which a combined drug approach (CDA) consisting of agents that target well-known pathogenic IRI mechanisms was used.<sup>9</sup> Streptokinase and epoprostenol were flushed through the liver prior to static cold storage, and the recipient received intravenous glycine,  $\alpha_1$ -acid glycoprotein, mitogen-activated protein kinase inhibitor,  $\alpha$ -tocopherol, and glutathione.<sup>10</sup> The CDA allowed for a steady and spontaneous liver function recovery, eliminated primary nonfunction, and improved recipient survival; it prevented Kupffer cell activation and subsequent tumor necrosis factor  $\alpha$  production, stabilized glutathione depletion, and reduced redox-active iron and bile salt toxicity.<sup>10</sup> In a

subsequent mechanistic study, the CDA was found to be associated with reduced expression of inflammation-regulating genes involved in pathways of cytokine activity and apoptosis.<sup>22</sup>

Although advocated over the past decade, to our knowledge, a CDA strategy has so far not been tested in the clinic.<sup>16,20,21</sup> We conducted a randomized clinical trial (RCT) that assessed the effect of a CDA modified to the clinical setting on the severity of IRI after liver transplant.

---

## Methods

### Study Design

This investigator-initiated, open-label RCT (Combined Drug Approach to Prevent Ischemia-Reperfusion Injury During Transplantation of Livers [CAPITL]) with 2 parallel arms was conducted between September 2013 and February 2018, with 1-year follow-up, at the University Hospitals Leuven, Belgium. Ethical approval was obtained from the Ethics Committee Research UZ / KU Leuven and the Federal Agency for Medicines and Health Products of Belgium. Before recruitment started, the trial was registered at ClinicalTrials.gov (NCT02251041) and approved by the Centre for Evidence in Transplantation. Written informed consent was obtained from study participants when they entered the transplant waiting list. The study was conducted in accordance with principles of the declaration of Helsinki<sup>23</sup> and followed the Consolidated Standards of Reporting Trials (CONSORT) reporting guideline. The study protocol is available in [Supplement 1](#).

### Study Population

Adults aged 18 years or older who were wait-listed for a first solitary full-size liver transplant were screened for eligibility at the time of liver offer. Recipients of whole livers from donations after brain death and/or circulatory death (Maastricht category 3<sup>24</sup>) were eligible. Exclusion criteria were acute liver failure as an indication for transplant, dialysis prior to transplant, hypersensitivity or a condition with a specific contraindication to any component of the CDA (**Table 1**), participation in another clinical trial, technical issues with administration of the CDA, and patient refusal.

### Randomization

At the time of organ offer, eligible patients were enrolled and randomized. A random allocation sequence was generated using computer-based random numbers with permuted block sizes of 2, 4, and 6. No stratification was performed on a priori-defined potential confounders such as CIT and priority on the waiting list through the Model for End-stage Liver Disease (MELD)<sup>48</sup> score. Data were recorded in an internet-secure electronic case report form (Eonix) at the University Hospitals Leuven.

### Intervention

As part of the standard of care, all livers were preserved by static cold storage following classic organ procurement. All liver grafts were flushed with 1 L of Institut Georges Lopez (IGL-1) preservation solution through the portal vein ex situ at the end of the back-table preparation at the recipient center. All transplants were done with caval replacement using venovenous bypass. After completion of caval and portal anastomosis, reperfusion of the liver was initiated. The standard of care further included an immunosuppressive regimen consisting of tacrolimus (trough levels between 5 and 8 µg/L), mycophenolate mofetil (500 mg, 2 times), and steroids (tapered to 3 months).<sup>14</sup>

In addition to standard of care, patients in the CDA group received a combination drug regimen that was adapted from the CDA regimen used in the porcine study.<sup>10</sup> Drugs and doses were selected for their proven IRI-protective effects without drug-related adverse events (Table 1).<sup>10,25-47</sup> Nine agents were administered in 3 distinct steps and with a specific sequence that, according to pharmacokinetic data, allowed for a peak plasma concentration during the reperfusion ([Supplement 1](#)). Recipients were given α-tocopherol (500 mg, 5 mL) and melatonin (6 mg) orally prior to transportation to the operating room. Epoprostenol (500 µg, 50 mL, 2 minutes) was added

to 1 L of IGL-1 and administered as part of the ex situ flush of the donor liver at the end of the back-table work before implantation. Last, the recipient was given intravenous antithrombin III (3000 IU, 60 mL, 2 minutes), infliximab (3 mg/kg, 7.5 mL/kg, 3 hours), apotransferrin (170 mg/kg, 3.4 mL/kg, 3 hours), recombinant erythropoietin-β (2 doses of 30 000 IU administered separately, 6-hour interval, 0.6 mL, 2 minutes), C1-inhibitor (1000 IU, 10 mL, 5 minutes), and glutathione (3 g, 20 mL, 2 minutes) during the anhepatic and the reperfusion phase.

**End Point Measures**

The primary end point was defined as the difference between the 2 study arms in peak serum aspartate aminotransferase (AST) levels within the first 72 hours after graft reperfusion, an accepted

**Table 1. Components, Doses, Mechanisms, and Relevant Experimental and Clinical Evidence of the Combined Drug Approach**

Component	Dose	Mechanisms	Experimental and clinical evidence
α-Tocopherol	500 mg	Antioxidant, ROS scavenger, increases the release of glutathione and prostacyclin	Improvement of IRI severity, elimination of primary nonfunction, reduction of inflammatory cytokines such as TNF-α, improvement in liver function, reduction of bile salt toxicity, and increase of survival in a stringent pig DCD model of LT <sup>10</sup> Improvement of hepatocellular injury, lipid peroxidation, and function in rat liver in situ reperfusion model <sup>25</sup> Reduction of AST levels and ICU LOS after human partial liver resection <sup>26</sup> Prevention of inflammation and parenchymal injury in isolated rat liver perfusion model <sup>27</sup>
Melatonin	6 mg	Synergistic antioxidant properties with α-tocopherol and glutathione, ROS scavenger	Decrease of TNF-α production and improvement of hepatocellular injury, function, and microcirculation in rat liver ex situ reperfusion model <sup>28,29</sup>
Epoprostenol	500 µg	Vasodilatation, antioxidant, inhibition of platelet aggregation, reduction of leukocyte activation and adhesion	Included in proof of concept; improvement in IRI severity, elimination of primary nonfunction, reduction of inflammatory cytokines such as TNF-α, improvement in liver function, reduction of bile salt toxicity, and increase in survival in a stringent pig DCD model of LT <sup>10</sup> Improvement of hepatocellular injury and survival rate in pig liver in situ reperfusion model <sup>30</sup> Decrease of serum aminotransferase and lactate and sinusoidal congestion in pig LT model <sup>31</sup> Improvement of hepatocellular injury <sup>32</sup> and biliary strictures <sup>33</sup> after administration in donor during human LT
Antithrombin III	3000 IU	Improvement of microcirculation (anticoagulation and vasodilatation), anti-inflammatory, antioxidant	Improvement in creatinine values, malondialdehyde levels, myeloperoxidase activity, and histological damage in rat kidney in situ reperfusion model <sup>34</sup> Reduction of neutrophil rolling and adhesion in a feline mesentery in situ reperfusion model <sup>35</sup> Increase in the release of prostacyclin, improvement of blood flow, and decrease of cytokine-induced neutrophil chemoattractant and myeloperoxidase in rat liver in situ reperfusion model <sup>36</sup> Reduction of graft thrombosis incidence in human simultaneous pancreas kidney transplant <sup>37</sup>
Infliximab	3 mg/kg	Anti-inflammatory blocking TNF-α	Prolonging of long-term survival after human intestinal transplant when used as a component of an immunomodulatory strategy <sup>38</sup> Improvement of hepatocellular injury and decrease of myeloperoxidase in rat liver in situ reperfusion model <sup>39</sup> Improvement of hepatocellular injury, apoptosis, and survival in mice liver in situ reperfusion model <sup>40</sup>
Apotransferrin	170 mg/kg	Redox-active iron chelator	Included in proof of concept; improvement of IRI severity, elimination of primary nonfunction, reduction of inflammatory cytokines such as TNF-α, improvement of liver function, reduction of bile salt toxicity, and increase in survival in a stringent pig DCD model of LT <sup>10</sup> Improvement of kidney reperfusion injury and acute kidney failure, decrease of complement activation and ROS formation in mice in situ reperfusion model <sup>41</sup>
EPO-β	2 Doses of 30 000 IU administered separately	Anti-inflammatory, antiapoptotic, antioxidant	Improvement of hepatocellular injury and apoptosis in pig liver in situ reperfusion model <sup>42</sup> Reduction of inflammatory cytokine (IL-6 and TNF-α) and hospital LOS after human partial liver resection and Pringle maneuver <sup>43</sup>
C1-inhibitor	1000 U	Inhibition of classic (very strong), lectine (strong), and alternative (strong) pathways of the complement activation, regulation of intrinsic and fibrinolytic pathways of the coagulation cascade, anti-inflammatory	Improvement of hepatocellular injury and function in rat liver in situ reperfusion model <sup>44</sup> Improvement of hepatocellular injury and decrease of inflammatory cell infiltration and histological damage in pig liver ex situ reperfusion model <sup>45</sup> Improvement of hepatocellular injury, liver regeneration, and survival in mice liver in situ reperfusion model <sup>46</sup> Improvement of myocardial injury, hemodynamic parameters, and hospital LOS in human coronary artery bypass grafting <sup>47</sup>

Abbreviations: AST, aspartate aminotransferase; DCD, donation after circulatory death; EPO-β, recombinant erythropoietin-β; ICU, intensive care unit; IL-6, interleukin 6; IRI, ischemia reperfusion injury; LOS, length of stay; LT, liver transplant; ROS, reactive oxygen species; TNF-α, tumor necrosis factor α.

clinical surrogate of liver IRI that correlates with patient and graft survival and postreperfusion liver histological assessment.<sup>49,50</sup> Serum AST levels were analyzed in the central laboratory of the University Hospitals Leuven by means of a colorimetric method with a 4-U/L detection limit (Hitachi-Roche Modular P, Roche Diagnostics).

Secondary end points included the frequencies of postreperfusion syndrome,<sup>51</sup> severity of histological IRI,<sup>10,52</sup> early allograft dysfunction (EAD),<sup>53</sup> severe postoperative surgical complications (Clavien-Dindo classification  $\geq 3b$ ) within the first 30 days after transplant,<sup>54</sup> IC,<sup>55</sup> AKI,<sup>14,56</sup> acute cellular rejection,<sup>57</sup> 1-year graft survival, and 1-year patient survival (detailed definitions are provided in Supplement 1).

## Safety

All adverse events were prospectively collected in both groups, categorized as adverse events or serious adverse events, and adjudicated for a possible causal relationship with the treatment.

Treatment-related adverse events were considered within the first 7 days following transplant.

## Statistical Analysis

A sample size of 58 patients was needed to detect a 50% reduction in peak AST, as determined with a 2-sided, 2-sample pooled *t* test of the mean ratio with log-normal data with  $\alpha$  equal to 5% and 80% power. The assumed coefficient of variation was 116%, as obtained from a series of 308 patients who underwent liver transplant at the University Hospitals Leuven between January 2007 and October 2011. Because we anticipated a dropout rate of 20% and aimed to have complete blocks in the randomization, it was planned to include a total of 72 patients (36 per group) in the study.

Results for the primary end point are reported for the full analysis set and a per protocol set. For secondary outcomes, the results for the per protocol set are given.

For the primary end point, a linear regression model was used on log-transformed values with treatment group as a factor, yielding a ratio of geometric means with 95% CIs. In a sensitivity analysis, MELD score and CIT were added as confounders. The primary end point analysis was repeated in the full analysis set with exclusion of patients who received a DCD liver. Odds ratios and 95% CIs are given for the comparison of frequencies of EAD, AKI, postreperfusion syndrome, and severe surgical complications. Mann-Whitney U tests with the Hodges-Lehmann estimator as effect size were performed to compare acute cellular rejection Banff<sup>58</sup> and IRI scores. For IC and clinically relevant graft loss, the hazard ratio from a Fine-Gray model treating death without the event of interest as a competing risk is reported as well as the cumulative incidence estimates after 1 year. Death-censored graft loss and recipient mortality were visualized using (the complement of) Kaplan-Meier estimates with (exact) log-rank test. All analyses were performed from May 20, 2019, to May 27, 2020, using SAS software, version 9.4 (SAS Institute Inc). Two-sided  $P < .05$  was considered to be statistically significant.

## Results

### Recruitment

Between September 2013 and February 2018, 310 consecutive patients were screened for the trial (Figure 1). Of these, 93 patients met the inclusion criteria and were randomized. Due to poor donor liver quality (as assessed by the procuring surgeon), 4 patients were excluded from the control arm and 11 from the CDA arm because livers were discarded and not transplanted. In the CDA arm, 2 patients were excluded because of a last-minute decision to perform a combined liver-kidney transplant (exclusion criterion) and 2 patients were regarded unfit for transplant. As such, the full analysis set included 74 patients (36 in the control arm and 38 in the CDA arm). Two patients did not receive CDA for logistical reasons; hence, the per protocol set contained 36 patients in each arm (median recipient age, 60 years [IQR, 51.7-66.2 years]; 52 men [72.2%] and 20 women [27.8%]). After 60 patients were randomized, it was noted that all postrandomization exclusions up until that

moment occurred in the CDA group (n = 8). To avoid a substantial imbalance in group size in the full analysis set, it was decided to create a new sequence of variable block sizes by increasing the probability to be allocated to the CDA group; this yielded a higher number of patients being randomized to the CDA group (53 in the CDA group vs 40 in the control group) but comparable numbers in the full analysis set (38 in the CDA group vs 36 in the control group).

**Donor, Preservation, and Recipient Demographics**

Overall, the 2 study arms were balanced with respect to donor and recipient demographics and baseline characteristics (Table 2). The cause of donor death included a higher rate of trauma in the control group than in the CDA group (11 of 36 [30.6%] vs 6 of 38 [15.8%]) and a higher rate of cerebrovascular accident in the CDA group than in the control group (20 of 38 [52.6%] vs 11 of 36 [30.6%]).

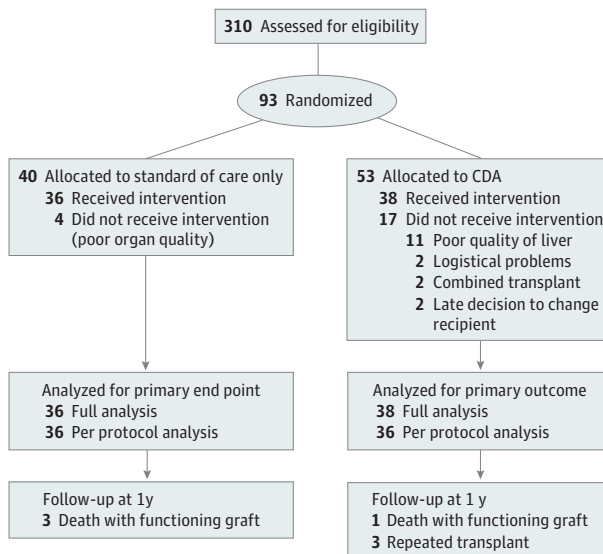
**Primary End Point**

In the per protocol analysis, the peak AST level within the first 72 hours following reperfusion was not different in the CDA group compared with the control group both without adjustment for CIT and MELD score (geometric mean, 1262.9 U/L [95% CI, 946.3-1685.4 U/L] vs 1451.2 U/L [95% CI, 1087.4-1936.7 U/L]; geometric mean ratio, 0.87 [95% CI, 0.58-1.31]; P = .49) and with adjustment for these factors (geometric mean, 1261.0 U/L [95% CI, 964.1-1649.5 U/L] vs 1453.3 U/L [95% CI, 1111.0-1901.0 U/L]; geometric mean ratio, 0.87 [95% CI, 0.59-1.27]; P = .46) (to convert AST to  $\mu$ kat/L, multiply by 0.0167). Furthermore, subgroup analysis in recipients of livers donated after brain death adjusted for CIT and MELD score did not show any difference in peak AST levels between the CDA and control groups (geometric mean, 1327.6 U/L [95% CI, 946.7-1861.8 U/L] vs 1710.3 U/L [95% CI, 1210.7-2416.1 U/L]; geometric mean ratio, 0.77 [95% CI, 0.48-1.26]; P = .29) (Table 3).

**Secondary End Points**

There was no difference in the IRI score or frequencies of postreperfusion syndrome, EAD, severe surgical complications, IC, AKI, and acute cellular rejection between arms (Table 3 and eTable 1 in Supplement 2). Overall, 1-year patient survival was not different between groups (Figure 2A). Four recipients died in the CDA group and 3 in the control group during 1-year follow-up. Three deaths in the CDA group but none in the control group were attributed to graft failure. Death-censored 1-year

Figure 1. Flow Diagram of Enrollment, Randomization, and Follow-up of Patients



CDA indicates combined drug approach.

graft survival was similar between the CDA and control groups (Figure 2B). Graft failure in the CDA group was due to 2 events of mycotic aneurysms and 1 event of IC requiring retransplantation.

### Safety and Adverse Events

A significant difference in the proportion of patients with at least 1 serious adverse event was not observed between groups (10 of 36 [27.8%] in the CDA group and 8 of 36 [22.2%] in the control group;  $P = .79$ ) (eTable 2 in Supplement 2). None of the events were considered to have a causal relationship to the intervention.

Table 2. Baseline Donor and Graft Preservation Characteristics and Recipient Characteristics

Characteristic	Participants <sup>a</sup>	
	CDA group (n = 38)	Control group (n = 36)
<b>Donor and graft preservation</b>		
Donor age, median (IQR), y	57 (49-70)	59 (48-68)
<b>Donor type</b>		
DBD	26 (68.4)	23 (63.9)
DCD	12 (31.5)	13 (36.1)
<b>Warm ischemia time if DCD, median (IQR), min<sup>b</sup></b>		
All phases	17 (16-21)	16 (12-25)
Agonal phase <sup>c</sup>	10 (6-13)	9 (2-13)
ICU LOS, median (IQR), d	3 (1-7)	4 (3-7)
<b>Donor cause of death</b>		
Trauma	6 (15.8)	11 (30.6)
CVA	20 (52.6)	11 (30.6)
Anoxia	0	2 (5.6)
Other	12 (31.6)	12 (33.3)
<b>Type of preservation solution</b>		
HTK	6 (15.8)	5 (13.9)
UW	7 (18.4)	5 (13.9)
IGL-1	25 (65.8)	25 (69.4)
Other	0	1 (2.8)
DBD duration, median (IQR), h <sup>d</sup>	13.3 (10.5-17.1)	14.3 (10.0-20.8)
Donor hepatectomy duration, median (IQR), min <sup>e</sup>	34.5 (25.0-46.0)	32.5 (26.0-42.5)
CIT, median (IQR), h <sup>f</sup>	5.8 (4.8-8.1)	5.8 (5.1-8.1)
<b>Recipient</b>		
Age, median (IQR), y	59 (53-66)	60 (51-68)
MELD score, median (IQR) <sup>g</sup>	13.2 (8.7-19.1)	13.8 (10.8-19.4)
<b>Indication for live transplant</b>		
Metabolic disease	4 (10.5)	3 (8.3)
HCC	18 (47.4)	12 (33.3)
Chronic liver disease	25 (65.8)	18 (50.0)
Ethyl	23 (60.5)	18 (50.0)
HBV	2 (5.3)	0
HCV	2 (5.3)	0
Cholestatic disease	4 (10.5)	4 (11.1)
NASH	2 (5.3)	10 (27.8)
Cryptogenic	1 (2.6)	0
Implantation duration, median (IQR), min <sup>h</sup>	39.5 (35.0-48.0)	38.5 (34.5-45.0)
ICU LOS, median (IQR), d	3 (1.5-6.5)	4 (2.5-7.0)

Abbreviations: CDA, combined drug approach; CIT, cold ischemia time; CVA, cerebrovascular accident; DBD, donation after brain death; DCD, donation after circulatory death; HBV, hepatitis B virus; HCC, hepatocellular carcinoma; HCV, hepatitis C virus; HTK, histidine tryptophan ketoglutarate; ICU, intensive care unit; IGL, Institut Georges Lopez; LOS, length of stay; MELD, Model for End-stage Liver Disease; NASH, nonalcoholic steatohepatitis; UW, University of Wisconsin.

<sup>a</sup> Data are presented as number (percentage) of participants unless otherwise indicated.

<sup>b</sup> Time from withdrawal of life-sustaining therapy to start of cold aortic perfusion in the donor (applied only to DCD liver transplants).

<sup>c</sup> Time from withdrawal of life-sustaining therapy to circulatory arrest (mean arterial pressure <30 mm Hg).

<sup>d</sup> Time between the brain death diagnosis and the cold aortic perfusion.

<sup>e</sup> Time between the start of aortic cold perfusion and completion of the donor hepatectomy, when the liver was placed in ice water on the back table.

<sup>f</sup> Time between the start of cold aortic perfusion and removal of the liver from cold storage before implantation.

<sup>g</sup> Possible score range of 4 to 40, with higher scores indicating a higher risk of mortality at 3 months.

<sup>h</sup> Time required for graft reperfusion between the liver leaving the ice water and completion of portal vein anastomosis.

## Discussion

In this RCT, we did not find evidence that an add-on perioperative combination drug regimen delivered to the graft immediately before implantation and to the recipient attenuated the degree of IRI. In addition, no differences were found in postreperfusion syndrome, histological IRI score, liver function, and the frequencies of severe surgical complications, IC, AKI, and acute cellular rejection, and graft and patient survival were similar between groups.

These findings were in contrast with those in the proof-of-concept study,<sup>10</sup> in which a substantial improvement in clinical outcomes was observed, with attenuation of IRI and complete prevention of primary graft nonfunction in a stringent model of porcine DCD liver transplant. The use of pleiotropic treatment strategies is well described and validated in other clinical fields, such as in oncological treatments<sup>59</sup> or immunosuppressive regimens after transplant.<sup>60</sup> The outcome of this study was unexpected given the increasingly accepted view that simultaneous targeting of different

**Table 3. Primary and Secondary End Points**

End point	CDA group	Control group	Treatment effect (95% CI)	P value
<b>Primary end point<sup>a</sup></b>				
Peak AST level, geometric mean (95% CI), U/L				
PPS results (n = 72)				
Unadjusted	1262.9 (946.3-1685.4)	1451.2 (1087.4-1936.7)	0.87 (0.58-1.31)	.49
Adjusted	1261.0 (964.1-1649.5)	1453.3 (1111.0-1901.0)	0.87 (0.59-1.27)	.46
DBD subgroup				
Unadjusted	1295.6 (907.7-1849.5)	1754.4 (1219.7-2523.6)	0.74 (0.44-1.22)	.23
Adjusted	1327.6 (946.7-1861.8)	1710.3 (1210.7-2416.1)	0.77 (0.48-1.26)	.29
FAS results (n = 74)				
Unadjusted	1223.3 (922.6-1622.1)	1451.2 (1086.0-1939.2)	0.84 (0.56-1.26)	.40
Adjusted	1232.5 (950.5-1598.0)	1439.8 (1102.5-1880.2)	0.86 (0.59-1.24)	.41
<b>Secondary end points</b>				
Early allograft dysfunction, No./total No. (%) <sup>b</sup>	13/36 (36.1)	17/36 (47.2)	0.63 (0.24-1.62)	.34
Ischemic cholangiopathy at 1 y, % (95% CI) <sup>c,d</sup>	11 (3-24)	5 (1-16)	2.05 (0.40-10.45)	.38
Acute kidney injury score, No./total No. (%) <sup>b,e</sup>				
0	27/34 (79.4)	31/35 (88.5)		
1	4/34 (11.7)	1/35 (2.9)	0.49 (0.31-1.88)	.30
2	2/34 (5.8)	3/35 (8.5)	0.98 (0.12-7.80)	>.99
3	1/34 (2.9)	0	ND	.49
Clinically relevant graft rejection at 1 y, % (95% CI) <sup>c</sup>	13 (5-27)	5 (1-16)	2.5 (0.52-12.05)	.25
IRI score, median (IQR) <sup>f</sup>	2 (2-4)	2 (1-4)	0 (1-0)	.35
Postreperfusion syndrome, No./total No. (%) <sup>b,g</sup>	7/29 (24.1)	6/29 (20.7)	1.22 (0.35-4.20)	.75
Severe surgical complication, No./total No. (%) <sup>b</sup>	10/36 (27.8)	8/36 (22.2)	1.35 (0.46-3.93)	.79

Abbreviations: AST, aspartate aminotransferase; CDA, combined drug approach; DBD, donation after brain death; FAS, full analysis set; IRI, ischemia-reperfusion injury; ND, not determined because of an absence of events in 1 group; PPS, per protocol set analysis.

SI conversion factor: To convert AST to  $\mu\text{kat/L}$ , multiply by 0.0167.

<sup>a</sup> Treatment effect is expressed as the geometric mean ratio. P values are based on unadjusted and adjusted linear regression analysis on log-transformed peak AST values. For the adjusted analysis, the ratio of the geometric means was adjusted for cold ischemia time and Model for End-stage Liver Disease score.

<sup>b</sup> Treatment effect is expressed as the unadjusted odds ratio and 95% CI. In cases with less than 5 events in at least 1 group, the exact 95% CI and exact P value (Fisher test) are reported.

<sup>c</sup> Treatment effect is expressed as an unadjusted hazard ratio from a Fine-Gray model treating mortality as a competing risk. In each group, the percentage at 1 year derived from the cumulative incidence curve is given.

<sup>d</sup> Data were missing for 4 in the CDA group and 4 in the control group.

<sup>e</sup> An acute kidney injury score of 1 indicates risk; 2, injury; and 3, failure. The ordinal character of the score was kept (ie, the reported odds ratios and P values refer to the comparison of the probability of having a score of >0, >1, or 3, respectively). Data were missing for 2 in the CDA group and 1 in the control group.

<sup>f</sup> The treatment effect is the Hodges-Lehmann estimator and equals the median of all pairwise differences. P value from a Mann-Whitney U test. Data were missing for 11 in the CDA group and 13 in the control group.

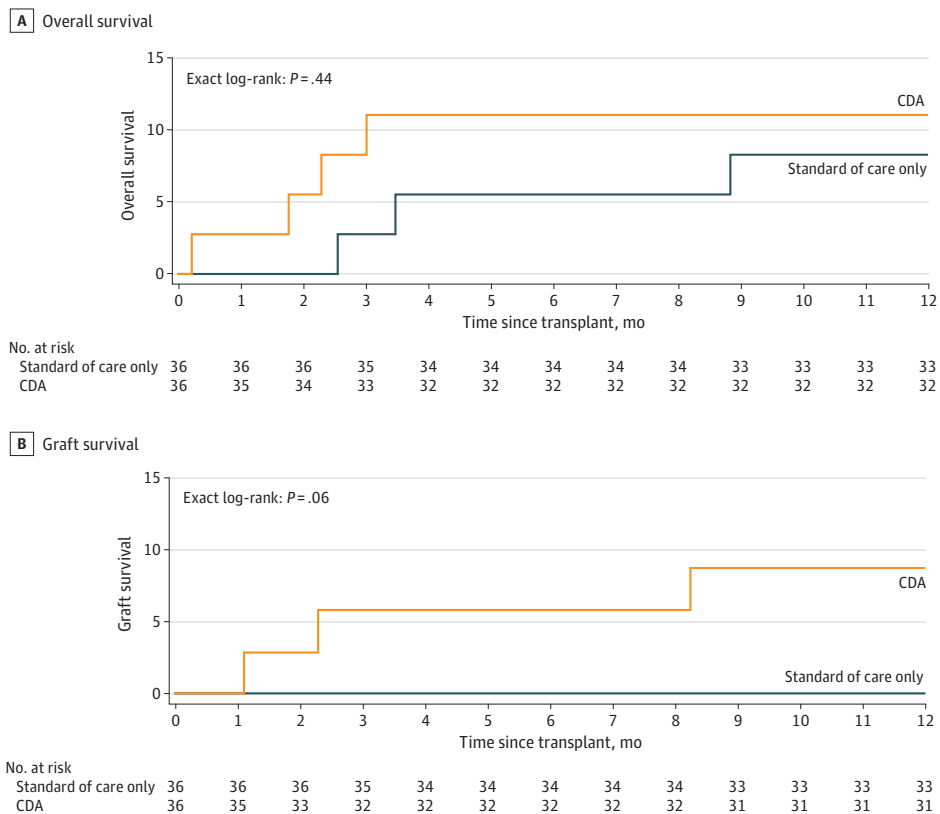
<sup>g</sup> Data were missing for 7 each in the CDA and control groups.

mechanistic pathways of IRI, through its cumulative and synergistic effects, is more efficient than a single compound approach.<sup>16,20,21,61-63</sup> Several variations, however, arose from the translation of the preclinical proof-of-concept model to the clinical setting and may explain the lack of effect observed in this RCT.

Physiological differences between pigs and humans and the highly controlled and reproducible conditions of experimental models as opposed to the variability in IRI severity in the clinical setting may explain why the protective effect observed in pigs was not reproduced in this study of humans. Another factor to explain the discrepancy between the porcine model and this RCT is that the porcine study<sup>10</sup> used a primary nonfunction model, which is the extreme clinical manifestation of severe IRI. Such extreme conditions were not encountered in this RCT. In addition, the RCT was powered on a surrogate end point (peak AST) rather than a hard clinical outcome, such as primary nonfunction.

Perhaps the design of the CDA, adapted to the clinical setting, may be another plausible explanation for the difference in results. In the proof-of-concept study,<sup>10</sup> the CDA was also administered in situ to the donor during the flush prior to static cold storage (streptokinase and epoprostenol) as an upstream strategy (and subsequently during the transplant in the recipient as a downstream strategy) to improve the graft quality during the procurement and preservation. This more upstream strategy could not be applied in this RCT since many donor livers were imported. Of note, at the time when the RCT was conducted, dynamic preservation was still in its infancy. Therefore, a downstream strategy limited to the post-static cold storage phase may not have been sufficient to halt IRI pathways that have already been triggered or the damage that was already established as a result of the ischemic insult during procurement and preservation.<sup>8,11</sup> Of note, the downstream strategy used in this RCT, which targeted the post-static cold storage graft and the graft recipient, aimed to scavenge ROS and mitigate the ensuing inflammatory cascade but was unable to achieve attenuation of IRI. The addition of an upstream strategy that prevents the formation of

Figure 2. One-year Cumulative Patient Mortality and Graft Loss in the Combined Drug Approach (CDA) and Control Groups



mitochondrial ROS by targeting the donor graft before and/or during static cold preservation may be more efficient.<sup>11</sup> Meanwhile, the importance of maintaining the integrity of mitochondrial function and replenishment of ATP during preservation has been demonstrated in settings where ex situ normothermic<sup>49,64</sup> and hypothermic oxygenated dynamic preservation<sup>65</sup> could attenuate liver IRI.

To translate the CDA regimen from the experimental to clinical setting, changes in drug composition were made, which may also have contributed to a variability in effect. The different agents, doses, and time regimens were judiciously chosen based on published evidence of their biological efficacy and pharmacokinetics (Supplement 1). In brief, instead of a streptokinase and epoprostenol flush of the donor pig liver before static cold storage, the human donor liver was perfused via the portal vein with epoprostenol after cold storage only with the goal to improve microcirculatory perfusion.<sup>30-33</sup> Since  $\alpha$ -tocopherol cannot be used clinically as an intravenous formulation, it was given to the recipient orally prior to transportation to the operative theater, and melatonin was added as a synergistic free-radical scavenger to increase the endogenous antioxidant capacity as it becomes overpowered by the burst of mitochondrial ROS after liver graft reperfusion.<sup>25-29</sup> The anti-inflammatory agents glycine and mitogen-activated protein kinase inhibitor FR167653 and the antioxidant plasma protein  $\alpha_1$ -acid glycoprotein used in the porcine model are not readily available for clinical use. For this study, they were therefore replaced with infliximab, which is used as part of an immunomodulatory regimen<sup>38</sup> in intestinal transplant to attenuate ROS production and leukocyte infiltration enhancement,<sup>39,40</sup> and with recombinant erythropoietin- $\beta$  to prevent apoptosis, inflammation, and oxidative stress in liver IRI.<sup>42,43</sup> Antithrombin III was added to reduce microcirculatory disorders and tissue injury,<sup>34-37</sup> and complement C1-inhibitor was added to reduce neutrophil infiltration and production of ROS.<sup>44-47</sup> Finally, as in the preclinical model, glutathione was included to improve the vascular antioxidant capacity and hepatocyte injury as previously described,<sup>66,67</sup> and apotransferrin was given to attenuate redox-active iron, which after liver graft reperfusion, is known to exceed the iron-binding capacity of transferrin and to contribute to liver graft failure.<sup>9,10,41</sup>

Recent developments in liver transplantation have included significant progress in the preservation of liver grafts by using dynamic preservation strategies. Preclinical and clinical studies have shown that compared with static cold storage, dynamic preservation allows for a reduction in IRI severity, frequencies of biliary complications and EAD, hospital stay, and graft and patient survival.<sup>49,68-73</sup> As a platform to deliver an upstream conditioning treatment strategy, dynamic preservation may counter or prevent ischemia and maintain mitochondria function<sup>11</sup> and probably represents a more promising strategy than multidrug strategies targeting post-cold storage grafts and recipients only.

### Limitations

This study has some limitations. First, it was a single-center RCT. Second, individual effects of each component of the CDA in decreasing IRI were not assessed separately. As the CDA specifically aims to achieve synergistic effects, the parallel testing of individual drugs was considered not useful and, in the clinical setting, extremely complex given the multitude of possible combinations. Third, despite the fact that CDA components were administered according to a specific timing in relation to their pharmacokinetic data with the aim to reach a peak concentration during reperfusion, blood levels of each component were not determined. Fourth, although the study was powered to detect differences in peak AST, the small sample size did not allow a claim on the absence of an effect.

### Conclusions

In this RCT, we did not find evidence that a combined drug approach targeting the post-cold storage graft and the recipient was sufficient to decrease IRI. Targeting the graft before and/or during preservation may also be necessary. The advent of dynamic preservation may bring the opportunity to combine a downstream CDA in the recipient with upstream conditioning of the donor liver.

## ARTICLE INFORMATION

**Accepted for Publication:** January 11, 2023.

**Published:** February 28, 2023. doi:10.1001/jamanetworkopen.2023.0819

**Open Access:** This is an open access article distributed under the terms of the [CC-BY License](#). © 2023 Meurisse N et al. *JAMA Network Open*.

**Corresponding Author:** Diethard Monbaliu, MD, PhD, Department of Abdominal Transplant Surgery and Transplant Coordination, University Hospitals Leuven, Herestraat 49, B-3000 Leuven, Belgium ([diethard.monbaliu@uzleuven.be](mailto:diethard.monbaliu@uzleuven.be)).

**Author Affiliations:** Laboratory of Abdominal Transplantation, Transplantation Research Group, Department of Microbiology, Immunology and Transplantation, KU Leuven, Leuven, Belgium (Meurisse, Mertens, Fieuws, Gilbo, Jochmans, Pirenne, Monbaliu); Department of Abdominal Transplant Surgery and Transplant Coordination, University Hospitals Leuven, Leuven, Belgium (Meurisse, Mertens, Fieuws, Gilbo, Jochmans, Pirenne, Monbaliu); Department of Abdominal Surgery and Transplantation, CHU de Liège, University of Liège, Liège, Belgium (Meurisse); Interuniversity Institute for Biostatistics and Statistical Bioinformatics, KU Leuven—University of Leuven, Leuven, Belgium (Fieuws).

**Author Contributions:** Dr Monbaliu had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

**Concept and design:** Meurisse, Jochmans, Pirenne, Monbaliu.

**Acquisition, analysis, or interpretation of data:** Meurisse, Mertens, Fieuws, Gilbo, Pirenne, Monbaliu.

**Drafting of the manuscript:** Meurisse, Mertens, Monbaliu.

**Critical revision of the manuscript for important intellectual content:** All authors.

**Statistical analysis:** Mertens, Fieuws.

**Obtained funding:** Monbaliu.

**Administrative, technical, or material support:** Meurisse, Gilbo.

**Supervision:** Jochmans, Pirenne, Monbaliu.

**Conflict of Interest Disclosures:** Dr Jochmans reported receiving speaker fees paid to the institution from XVIVO Perfusion outside the submitted work and having a patent pending on methods and applications of analyzing the perfusate of an ex situ perfused kidney. No other disclosures were reported.

**Funding/Support:** This research was funded by IWT grant 100790 from the Research Foundation Flanders (Dr Monbaliu), which allowed for payment of the research nurse and research costs.

**Role of the Funder/Sponsor:** The Research Foundation Flanders had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

**Data Sharing Statement:** See [Supplement 3](#).

**Additional Contributions:** Ilse Senesaël, Sarah Mertens, Donna Peerboom, Veerle Heedfeld, Tine Wylin, PhD, and Sofie Vets assisted with data collection and analysis; Anne Kaiser assisted with preparation of the manuscript; and An Billiau, MD, PhD, provided medical writing and editing support. We thank the members of the data safety monitoring board: Johan Fevery, PhD, Patrick Ferdinande, PhD, Peter Lauwers, PhD, and Kris Bogaerts, PhD, all from KU Leuven. None of these individuals received compensation.

## REFERENCES

1. Eurotransplant. Annual report 2019. June 11, 2020. Accessed November 9, 2021. <https://www.eurotransplant.org/annual-report/annual-report-2019>
2. Wall SP, Plunkett C, Caplan A. A potential solution to the shortage of solid organs for transplantation. *JAMA*. 2015;313(23):2321-2322. doi:10.1001/jama.2015.5328
3. Dar WA, Sullivan E, Bynon JS, Eltzschig H, Ju C. Ischaemia reperfusion injury in liver transplantation: cellular and molecular mechanisms. *Liver Int*. 2019;39(5):788-801. doi:10.1111/liv.14091
4. Briceño J, Marchal T, Padillo J, Solórzano G, Pera C. Influence of marginal donors on liver preservation injury. *Transplantation*. 2002;74(4):522-526. doi:10.1097/00007890-200208270-00015
5. Zhai Y, Petrowsky H, Hong JC, Busuttill RW, Kupiec-Weglinski JW. Ischaemia-reperfusion injury in liver transplantation—from bench to bedside. *Nat Rev Gastroenterol Hepatol*. 2013;10(2):79-89. doi:10.1038/nrgastro.2012.225
6. Jiménez-Castro MB, Cornide-Petronio ME, Gracia-Sancho J, Peralta C. Inflammasome-mediated inflammation in liver ischemia-reperfusion injury. *Cells*. 2019;8(10):1131-1157. doi:10.3390/cells8101131

7. Eltzhischig HK, Eckle T. Ischemia and reperfusion—from mechanism to translation. *Nat Med*. 2011;17(11):1391-1401. doi:10.1038/nm.2507
8. Chouchani ET, Pell VR, James AM, et al. A unifying mechanism for mitochondrial superoxide production during ischemia-reperfusion injury. *Cell Metab*. 2016;23(2):254-263. doi:10.1016/j.cmet.2015.12.009
9. Monbaliu D, van Pelt J, De Vos R, et al. Primary graft nonfunction and Kupffer cell activation after liver transplantation from non-heart-beating donors in pigs. *Liver Transpl*. 2007;13(2):239-247. doi:10.1002/lt.21046
10. Monbaliu D, Vekemans K, Hoekstra H, et al. Multifactorial biological modulation of warm ischemia reperfusion injury in liver transplantation from non-heart-beating donors eliminates primary nonfunction and reduces bile salt toxicity. *Ann Surg*. 2009;250(5):808-817. doi:10.1097/SLA.0b013e3181bdd787
11. Pell VR, Chouchani ET, Murphy MP, Brookes PS, Krieg T. Moving forwards by blocking back-flow: the yin and yang of MI therapy. *Circ Res*. 2016;118(5):898-906. doi:10.1161/CIRCRESAHA.115.306569
12. de Vries Y, von Meijnenfeldt FA, Porte RJ. Post-transplant cholangiopathy: classification, pathogenesis, and preventive strategies. *Biochim Biophys Acta Mol Basis Dis*. 2018;1864(4 pt B):1507-1515. doi:10.1016/j.bbadis.2017.06.013
13. Nastos C, Kalimeris K, Papoutsidakis N, et al. Global consequences of liver ischemia/reperfusion injury. *Oxid Med Cell Longev*. 2014;2014:906965. doi:10.1155/2014/906965
14. Jochmans I, Meurisse N, Neyrinck A, Verhaegen M, Monbaliu D, Pirenne J. Hepatic ischemia/reperfusion injury associates with acute kidney injury in liver transplantation: prospective cohort study. *Liver Transpl*. 2017;23(5):634-644. doi:10.1002/lt.24728
15. Zhai Y, Busuttill RW, Kupiec-Weglinski JW. Liver ischemia and reperfusion injury: new insights into mechanisms of innate-adaptive immune-mediated tissue inflammation. *Am J Transplant*. 2011;11(8):1563-1569. doi:10.1111/j.1600-6143.2011.03579.x
16. Soares ROS, Losada DM, Jordani MC, Évora P, Castro-E-Silva O. Ischemia/reperfusion injury revisited: an overview of the latest pharmacological strategies. *Int J Mol Sci*. 2019;20(20):5034. doi:10.3390/ijms20205034
17. Lang JD Jr, Smith AB, Brandon A, et al. A randomized clinical trial testing the anti-inflammatory effects of preemptive inhaled nitric oxide in human liver transplantation. *PLoS One*. 2014;9(2):e86053. doi:10.1371/journal.pone.0086053
18. D'Amico F, Vitale A, Piovan D, et al. Use of N-acetylcysteine during liver procurement: a prospective randomized controlled study. *Liver Transpl*. 2013;19(2):135-144. doi:10.1002/lt.23527
19. Busuttill RW, Lipshutz GS, Kupiec-Weglinski JW, et al. rPSGL-Ig for improvement of early liver allograft function: a double-blind, placebo-controlled, single-center phase II study. *Am J Transplant*. 2011;11(4):786-797. doi:10.1111/j.1600-6143.2011.03441.x
20. de Rougemont O, Dutkowski P, Clavien PA. Biological modulation of liver ischemia-reperfusion injury. *Curr Opin Organ Transplant*. 2010;15(2):183-189. doi:10.1097/MOT.0b013e3283373ced
21. Rossello X, Yellon DM. The RISK pathway and beyond. *Basic Res Cardiol*. 2017;113(1):2. doi:10.1007/s00395-017-0662-x
22. Vekemans K, Monbaliu D, Balligand E, et al. Improving the function of liver grafts exposed to warm ischemia by the Leuven drug protocol: exploring the molecular basis by microarray. *Liver Transpl*. 2012;18(2):206-218. doi:10.1002/lt.22446
23. World Medical Association. World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. *JAMA*. 2013;310(20):2191-2194. doi:10.1001/jama.2013.281053
24. Evrard P; Belgian Working Group on DCD National Protocol. Belgian modified classification of Maastricht for donors after circulatory death. *Transplant Proc*. 2014;46(9):3138-3142. doi:10.1016/j.transproceed.2014.09.169
25. Giakoustidis D, Papageorgiou G, Iliadis S, et al. Intramuscular administration of very high dose of alpha-tocopherol protects liver from severe ischemia/reperfusion injury. *World J Surg*. 2002;26(7):872-877. doi:10.1007/s00268-002-6271-2
26. Bartels M, Biesalski HKHK, Engelhart K, Sendlhofer G, Rehak P, Nagel E. Pilot study on the effect of parenteral vitamin E on ischemia and reperfusion induced liver injury: a double blind, randomized, placebo-controlled trial. *Clin Nutr*. 2004;23(6):1360-1370. doi:10.1016/j.clnu.2004.05.003
27. Moussavian MR, Scheuer C, Schmidt M, et al. Multidrug donor preconditioning prevents cold liver preservation and reperfusion injury. *Langenbecks Arch Surg*. 2011;396(2):231-241. doi:10.1007/s00423-010-0668-4
28. Zaouali MA, Reiter RJ, Padriisa-Altés S, et al. Melatonin protects steatotic and nonsteatotic liver grafts against cold ischemia and reperfusion injury. *J Pineal Res*. 2011;50(2):213-221.

29. Vairetti M, Ferrigno A, Bertone R, et al. Exogenous melatonin enhances bile flow and ATP levels after cold storage and reperfusion in rat liver: implications for liver transplantation. *J Pineal Res*. 2005;38(4):223-230. doi:10.1111/j.1600-079X.2004.00193.x
30. Kim YI, Kai T, Kitano S, et al. Hepatoprotection by a PGI2 analogue in complete warm ischemia of the pig liver: prostanoic release from the reperfused liver. *Transplantation*. 1994;58(8):875-879. doi:10.1097/00007890-199410270-00002
31. Kishida A, Kurumi Y, Kodama M. Efficacy of prostaglandin I2 analog on liver grafts subjected to 30 minutes of warm ischemia. *Surg Today*. 1997;27(11):1056-1060. doi:10.1007/BF02385788
32. Klein M, Geoghegan J, Wangemann R, Böckler D, Schmidt K, Scheele J. Preconditioning of donor livers with prostaglandin I2 before retrieval decreases hepatocellular ischemia-reperfusion injury. *Transplantation*. 1999;67(8):1128-1132. doi:10.1097/00007890-199904270-00007
33. Pirenne J, Monbaliu D, Aerts R, et al. Biliary strictures after liver transplantation: risk factors and prevention by donor treatment with epoprostenol. *Transplant Proc*. 2009;41(8):3399-3402. doi:10.1016/j.transproceed.2009.09.026
34. Ozden A, Sarioglu A, Demirkan NC, Bilgihan A, Düzcan E. Antithrombin III reduces renal ischemia-reperfusion injury in rats. *Res Exp Med (Berl)*. 2001;200(3):195-203.
35. Ostrovsky L, Woodman RC, Payne D, Teoh D, Kubes P. Antithrombin III prevents and rapidly reverses leukocyte recruitment in ischemia/reperfusion. *Circulation*. 1997;96(7):2302-2310. doi:10.1161/01.CIR.96.7.2302
36. Harada N, Okajima K, Kushimoto S, Isobe H, Tanaka K. Antithrombin reduces ischemia/reperfusion injury of rat liver by increasing the hepatic level of prostacyclin. *Blood*. 1999;93(1):157-164. doi:10.1182/blood.V93.1.157
37. Fertmann JM, Wimmer CD, Arbogast HP, et al. Single-shot antithrombin in human pancreas-kidney transplantation: reduction of reperfusion pancreatitis and prevention of graft thrombosis. *Transpl Int*. 2006;19(6):458-465. doi:10.1111/j.1432-2277.2006.00325.x
38. Ceulemans LJ, Braza F, Monbaliu D, et al. The Leuven immunomodulatory protocol promotes T-regulatory cells and substantially prolongs survival after first intestinal transplantation. *Am J Transplant*. 2016;16(10):2973-2985. doi:10.1111/ajt.13815
39. Colletti LM, Remick DG, Burtch GD, Kunkel SL, Strieter RM, Campbell DA Jr. Role of tumor necrosis factor- $\alpha$  in the pathophysiologic alterations after hepatic ischemia/reperfusion injury in the rat. *J Clin Invest*. 1990;85(6):1936-1943. doi:10.1172/JCI114656
40. Rüdiger HA, Clavien PA. Tumor necrosis factor  $\alpha$ , but not Fas, mediates hepatocellular apoptosis in the murine ischemic liver. *Gastroenterology*. 2002;122(1):202-210. doi:10.1053/gast.2002.30304
41. de Vries B, Walter SJ, von Bonsdorff L, et al. Reduction of circulating redox-active iron by apotransferrin protects against renal ischemia-reperfusion injury. *Transplantation*. 2004;77(5):669-675. doi:10.1097/01.TP.0000115002.28575.E7
42. Shimoda M, Sawada T, Iwasaki Y, et al. Erythropoietin strongly protects the liver from ischemia-reperfusion injury in a pig model. *Hepatology*. 2009;56(90):470-475.
43. Kato M, Sawada T, Kita J, Shimoda M, Kubota K. Erythropoietin ameliorates early ischemia-reperfusion injury following the Pringle maneuver. *World J Gastroenterol*. 2010;16(38):4838-4845. doi:10.3748/wjg.v16.i38.4838
44. Heijnen BHM, Straatsburg IH, Padilla ND, Van Mierlo GJ, Hack CE, Van Gulik TM. Inhibition of classical complement activation attenuates liver ischaemia and reperfusion injury in a rat model. *Clin Exp Immunol*. 2006;143(1):15-23. doi:10.1111/j.1365-2249.2005.02958.x
45. Bergamaschini L, Gobbo G, Gatti S, et al. Endothelial targeting with C1-inhibitor reduces complement activation in vitro and during ex vivo reperfusion of pig liver. *Clin Exp Immunol*. 2001;126(3):412-420. doi:10.1046/j.1365-2249.2001.01695.x
46. Saidi RF, Rajeshkumar B, Sharifabrizi A, Dresser K, Walter O. Human C1 inhibitor attenuates liver ischemia-reperfusion injury and promotes liver regeneration. *J Surg Res*. 2014;187(2):660-666. doi:10.1016/j.jss.2013.09.009
47. Fattouch K, Bianco G, Speziale G, et al. Beneficial effects of C1 esterase inhibitor in ST-elevation myocardial infarction in patients who underwent surgical reperfusion: a randomised double-blind study. *Eur J Cardiothorac Surg*. 2007;32(2):326-332. doi:10.1016/j.ejcts.2007.04.038
48. Kamath PS, Kim WR; Advanced Liver Disease Study Group. The model for end-stage liver disease (MELD). *Hepatology*. 2007;45(3):797-805. doi:10.1002/hep.21563
49. Nasralla D, Coussios CC, Mergental H, et al; Consortium for Organ Preservation in Europe. A randomized trial of normothermic preservation in liver transplantation. *Nature*. 2018;557(7703):50-56. doi:10.1038/s41586-018-0047-9

50. Jochmans I, Monbaliu D, Pirenne J. The beginning of an end point: peak AST in liver transplantation. *J Hepatol*. 2014;61(5):1186-1187. doi:10.1016/j.jhep.2014.07.021
51. Hilmi I, Horton CN, Planinsic RM, et al. The impact of postreperfusion syndrome on short-term patient and liver allograft outcome in patients undergoing orthotopic liver transplantation. *Liver Transpl*. 2008;14(4):504-508. doi:10.1002/lt.21381
52. Suzuki S, Toledo-Pereyra LH, Rodriguez FJ, Cejalvo D. Neutrophil infiltration as an important factor in liver ischemia and reperfusion injury: modulating effects of FK506 and cyclosporine. *Transplantation*. 1993;55(6):1265-1272. doi:10.1097/00007890-199306000-00011
53. Olthoff KM, Kulik L, Samstein B, et al. Validation of a current definition of early allograft dysfunction in liver transplant recipients and analysis of risk factors. *Liver Transpl*. 2010;16(8):943-949. doi:10.1002/lt.22091
54. Clavien PA, Barkun J, de Oliveira ML, et al. The Clavien-Dindo classification of surgical complications: five-year experience. *Ann Surg*. 2009;250(2):187-196. doi:10.1097/SLA.0b013e3181b13ca2
55. Meurisse N, Vanden Bussche S, Jochmans I, et al. Outcomes of liver transplantations using donations after circulatory death: a single-center experience. *Transplant Proc*. 2012;44(9):2868-2873. doi:10.1016/j.transproceed.2012.09.077
56. Bellomo R, Ronco C, Kellum JA, Mehta RL, Palevsky P; Acute Dialysis Quality Initiative workgroup. Acute renal failure—definition, outcome measures, animal models, fluid therapy and information technology needs: the Second International Consensus Conference of the Acute Dialysis Quality Initiative (ADQI) Group. *Crit Care*. 2004;8(4):R204-R212. doi:10.1186/cc2872
57. Demetris AJ, Bellamy C, Hübscher SG, et al. 2016 Comprehensive update of the Banff working group on liver allograft pathology: introduction of antibody-mediated rejection. *Am J Transplant*. 2016;16(10):2816-2835. doi:10.1111/ajt.13909
58. Ormonde DG, de Boer WB, Kierath A, et al. Banff schema for grading liver allograft rejection: utility in clinical practice. *Liver Transpl Surg*. 1999;5(4):261-268. doi:10.1002/lt.500050418
59. Wu C. Systemic therapy for colon cancer. *Surg Oncol Clin N Am*. 2018;27(2):235-242. doi:10.1016/j.soc.2017.11.001
60. Di Maira T, Little EC, Berenguer M. Immunosuppression in liver transplant. *Best Pract Res Clin Gastroenterol*. 2020;46-47:101681. doi:10.1016/j.bpg.2020.101681
61. Selzner N, Rudiger H, Graf R, Clavien PA. Protective strategies against ischemic injury of the liver. *Gastroenterology*. 2003;125(3):917-936. doi:10.1016/S0016-5085(03)01048-5
62. Menger MD, Vollmar B. Pathomechanisms of ischemia-reperfusion injury as the basis for novel preventive strategies: is it time for the introduction of pleiotropic compounds? *Transplant Proc*. 2007;39(2):485-488. doi:10.1016/j.transproceed.2007.01.022
63. Yamanaka K, Houben P, Bruns H, Schultze D, Hatano E, Schemmer P. A systematic review of pharmacological treatment options used to reduce ischemia reperfusion injury in rat liver transplantation. *PLoS One*. 2015;10(4):e0122214. doi:10.1371/journal.pone.0122214
64. Xu H, Berendsen T, Kim K, et al. Excorporeal normothermic machine perfusion resuscitates pig DCD livers with extended warm ischemia. *J Surg Res*. 2012;173(2):e83-e88. doi:10.1016/j.jss.2011.09.057
65. Schlegel A, Muller X, Dutkowski P. Hypothermic liver perfusion. *Curr Opin Organ Transplant*. 2017;22(6):563-570. doi:10.1097/MOT.0000000000000472
66. Schauer RJ, Gerbes AL, Vonier D, et al. Glutathione protects the rat liver against reperfusion injury after prolonged warm ischemia. *Ann Surg*. 2004;239(2):220-231. doi:10.1097/01.sla.0000110321.64275.95
67. Schauer RJ, Kalmuk S, Gerbes AL, et al. Intravenous administration of glutathione protects parenchymal and non-parenchymal liver cells against reperfusion injury following rat liver transplantation. *World J Gastroenterol*. 2004;10(6):864-870. doi:10.3748/wjg.v10.i6.864
68. Monbaliu D, Brassil J. Machine perfusion of the liver: past, present and future. *Curr Opin Organ Transplant*. 2010;15(2):160-166. doi:10.1097/MOT.0b013e328337342b
69. Jia JJ, Li JH, Yu H, et al. Machine perfusion for liver transplantation: a concise review of clinical trials. *Hepatobiliary Pancreat Dis Int*. 2018;17(5):387-391. doi:10.1016/j.hbpd.2018.06.003
70. van Leeuwen OB, de Vries Y, Fujiyoshi M, et al. Transplantation of high-risk donor livers after ex situ resuscitation and assessment using combined hypo- and normothermic machine perfusion: a prospective clinical trial. *Ann Surg*. 2019;270(5):906-914. doi:10.1097/SLA.0000000000003540

71. Nickkholgh A, Nikdad M, Shafie S, et al. Ex situ liver machine perfusion as an emerging graft protective strategy in clinical liver transplantation: the dawn of a new era. *Transplantation*. 2019;103(10):2003-2011. doi:10.1097/TP.0000000000002772

72. Dutkowsky P, Polak WG, Muiesan P, et al. First comparison of hypothermic oxygenated perfusion versus static cold storage of human donation after cardiac death liver transplants: an international-matched case analysis. *Ann Surg*. 2015;262(5):764-770. doi:10.1097/SLA.0000000000001473

73. van Rijn R, Schurink IJ, de Vries Y, et al; DHOPE-DCD Trial Investigators. Hypothermic machine perfusion in liver transplantation—a randomized trial. *N Engl J Med*. 2021;384(15):1391-1401. doi:10.1056/NEJMoa2031532

#### SUPPLEMENT 1.

##### Trial Protocol

#### SUPPLEMENT 2.

eTable 1. Surgical Complications

eTable 2. Adverse Events and Serious Adverse Events

#### SUPPLEMENT 3.

##### Data Sharing Statement