

CAR-T and nanoCAR-T for multiple myeloma: Does smaller mean smarter ?

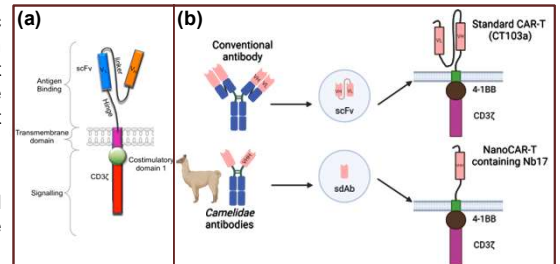
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INTRODUCTION & OBJECTIVE

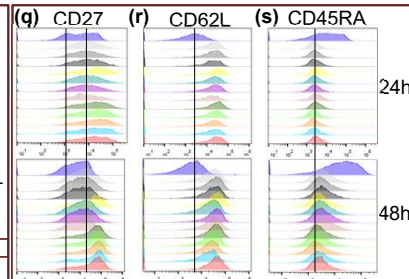
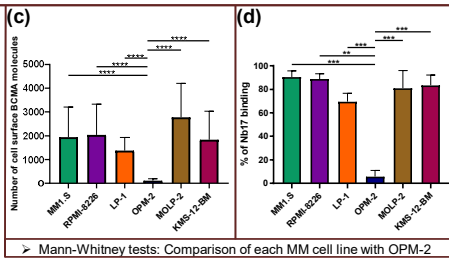
❖ **Multiple myeloma (MM)** is an incurable hematologic malignancy of plasma cells. **Chimeric antigen receptor T lymphocyte (CAR-T)** immunotherapy from second generation (a) has shown remarkable results in relapse patients. **Single-domain antibody (sdAb)** offers an excellent alternative to scFv due to the advantage of having a small and stable folding structure (b). Despite the development of numerous nanoCAR-Ts, there is limited data demonstrating the direct differences and their mechanisms of action.



❖ This project aims to characterize the activity of a **nanoCAR-T** (containing sdAb, called Nb17) and a **standard CAR-T** (called CT103a) to elucidate the underlying mechanisms driving these differences. These findings may reveal new potential targets for novel CAR-T design strategies.

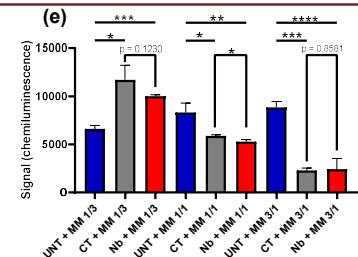
METHODS & RESULTS

1) Cell surface BCMA was **quantified** in six MM cell lines using the BD Quantibrite™Beads-PE kit, using OPM-2 as a negative control (c). Then, the **binding capacity** of the sdAb (Nb17) to BCMA cell surface was analyzed on each MM cell lines (d).

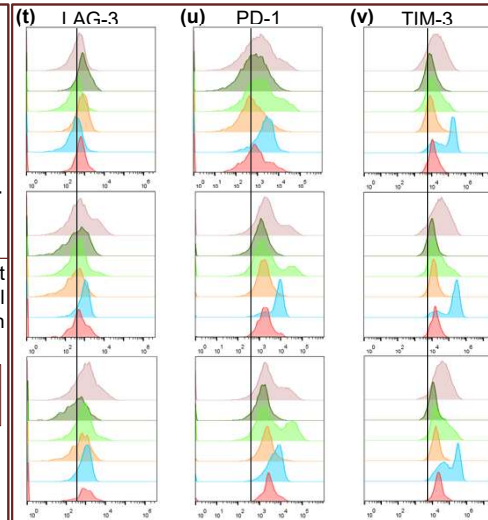
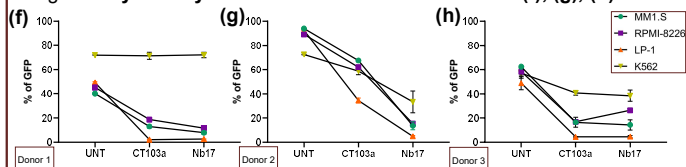


3) After 24h and 48h of co-culture with MM1.S using different ratios, **differentiation markers** were analyzed by flow cytometry. Compared to unactivated T cells from PBMCs, UNT and both CAR-Ts express CD27 (q) and CD62L (r) but do not express CD45RA (s). Thus, cells are differentiated into **central memory T cells**.

2) **Killing assays** were first performed by co-cultures of MM1.S with untransduced T cells (UNT) or CAR-Ts using different ratios (1/3, 1/1, 1/3) and analyses were done by **luciferase assay** (e).

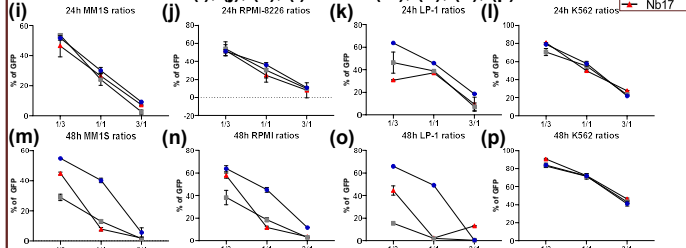


Secondly, using a ratio 1/1 UNT and CAR-Ts were co-cultured with different MM (MM1.S, RPMI-8226, LP-1) expressing BCMA or lymphoma (K562) cell lines (n=3 for each of the 3 donor). Co-cultured were performed for 48h using **flow cytometry** with GFP transduced tumoral cells (f), (g), (h).



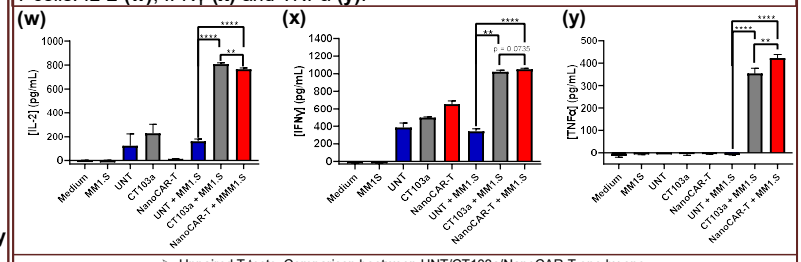
4) **Exhaustion markers** of UNT and both CAR-Ts were analyzed using flow cytometry after being co-cultured with MM1.S for 24h, 48h, or after rechallenge of 48h after 48h of current co-culture. Following the timepoints the expression of LAG-3, PD-1 and TIM-3 is increasing (t), (u), (v).

In a third time each cell lines was co-cultured with UNT, CT103a or NanoCAR-T for 24h (i), (j), (k), (l) or 48h (m), (n), (o), (p).



➤ These data show CAR-Ts are killing tumoral cells but more **specifically** BCMA cell surface positive cell lines.

5) **ELISAs** were performed for different donors after 24h of co-cultures with MM1.S cells. Both CT103a and NanoCAR-T produce cytokines in comparison to untransduced T cells: IL-2 (w), IFNγ (x) and TNFα (y).



CONCLUSION

The nanoCAR-T generated in our lab demonstrates excellent anti-cancer efficacy equivalent to the standard CAR-T CT103a. Further investigation of activation and degranulation markers and transcriptomic levels may reveal mechanistic differences and potential improvements to CAR-T therapy.