

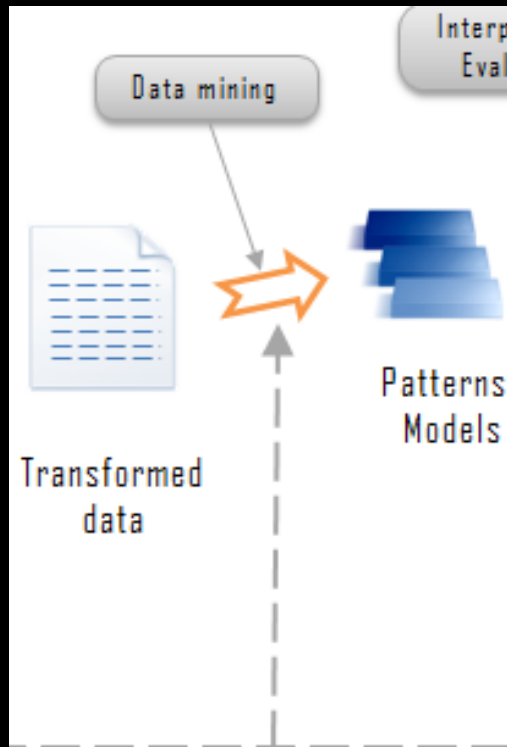
# MLP and CNN for dairy farming

Prof. Hélène Soyeurt



SensAIFood – May 2025 – Gembloux, Belgium

# WHAT IS THE OBJECTIVE ?



## Regression

- Quantitative y values
- Prediction

## Classification

- Quantitative y values
- Separation into several classes

## Clustering

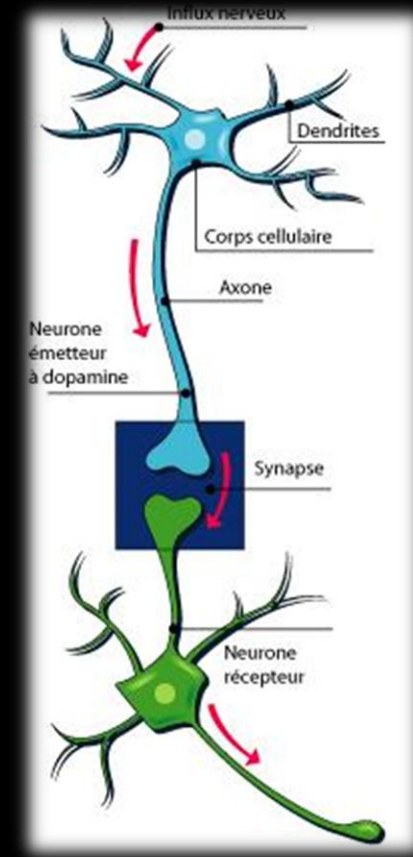
- Creation of sub-groups within the same dataset

Supervised

Not supervised

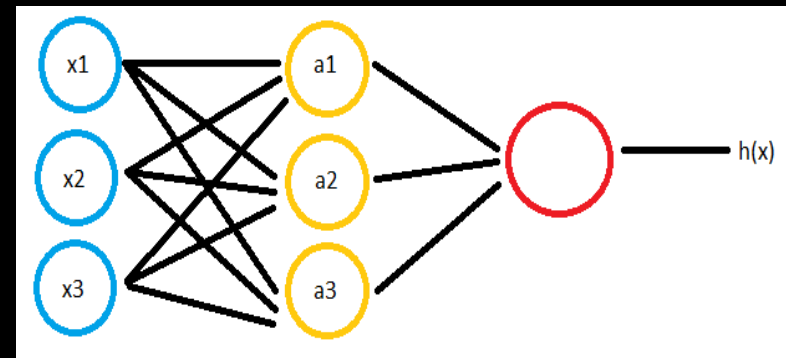
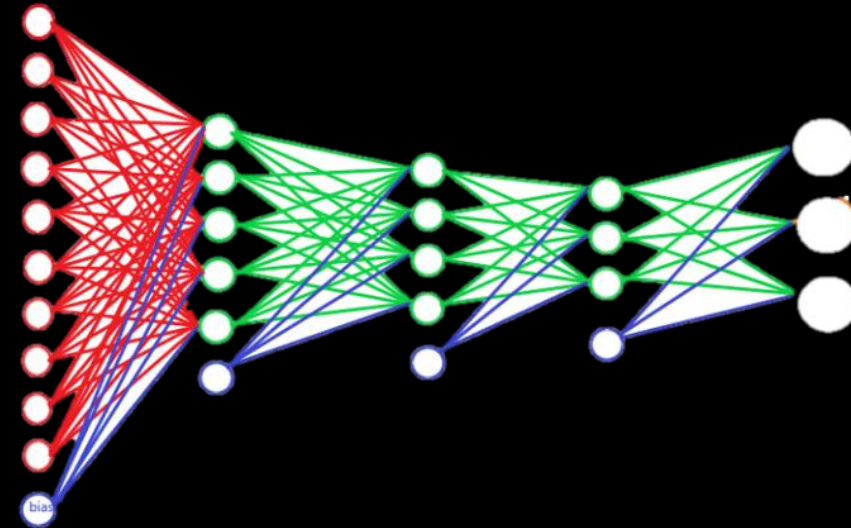
# INTRODUCTION

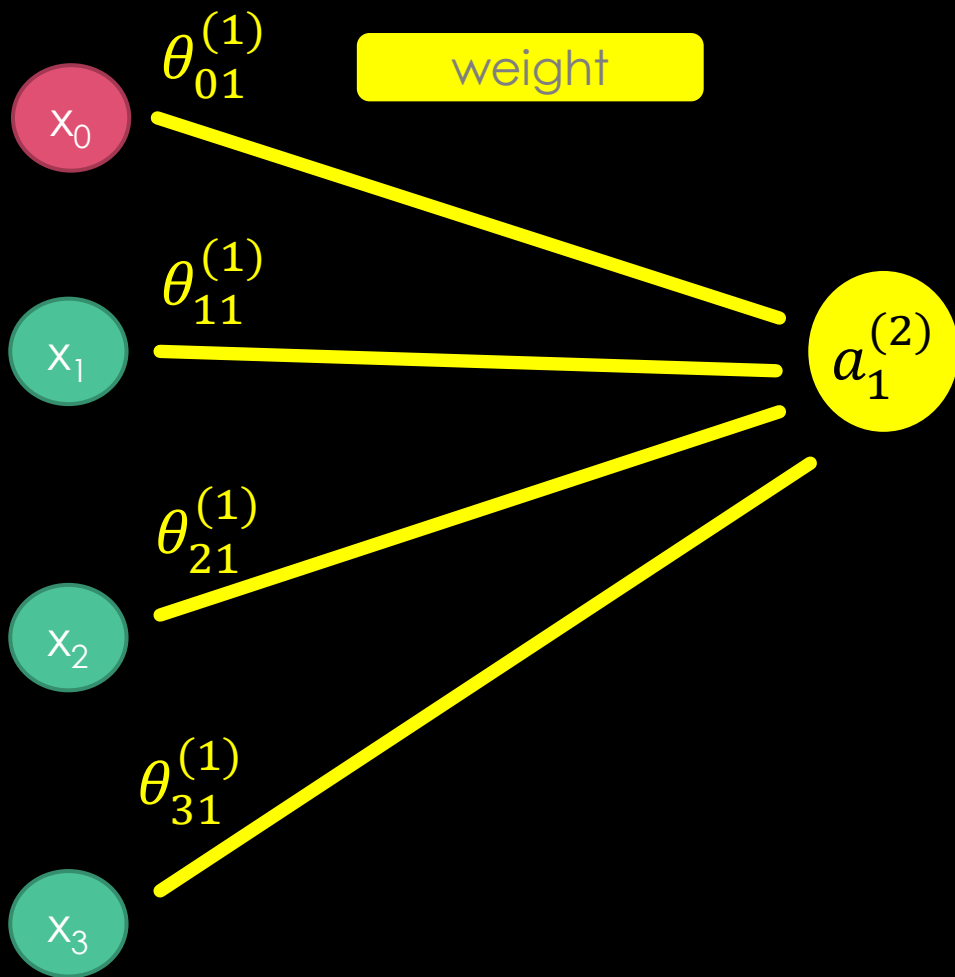
- ARTIFICIAL NEURAL NETWORK (ANN)
- BASIC OF DEEP LEARNING
- MCCULLOCH-PITTS (1943) REPRESENT THE NEURON AS A BINARY TOOL
- GREAT INTEREST IN THE 80'S AND 90'S
- MIMIC THE STRUCTURE OF BRAIN COMPOSED OF NEURONS AND SYNAPSES



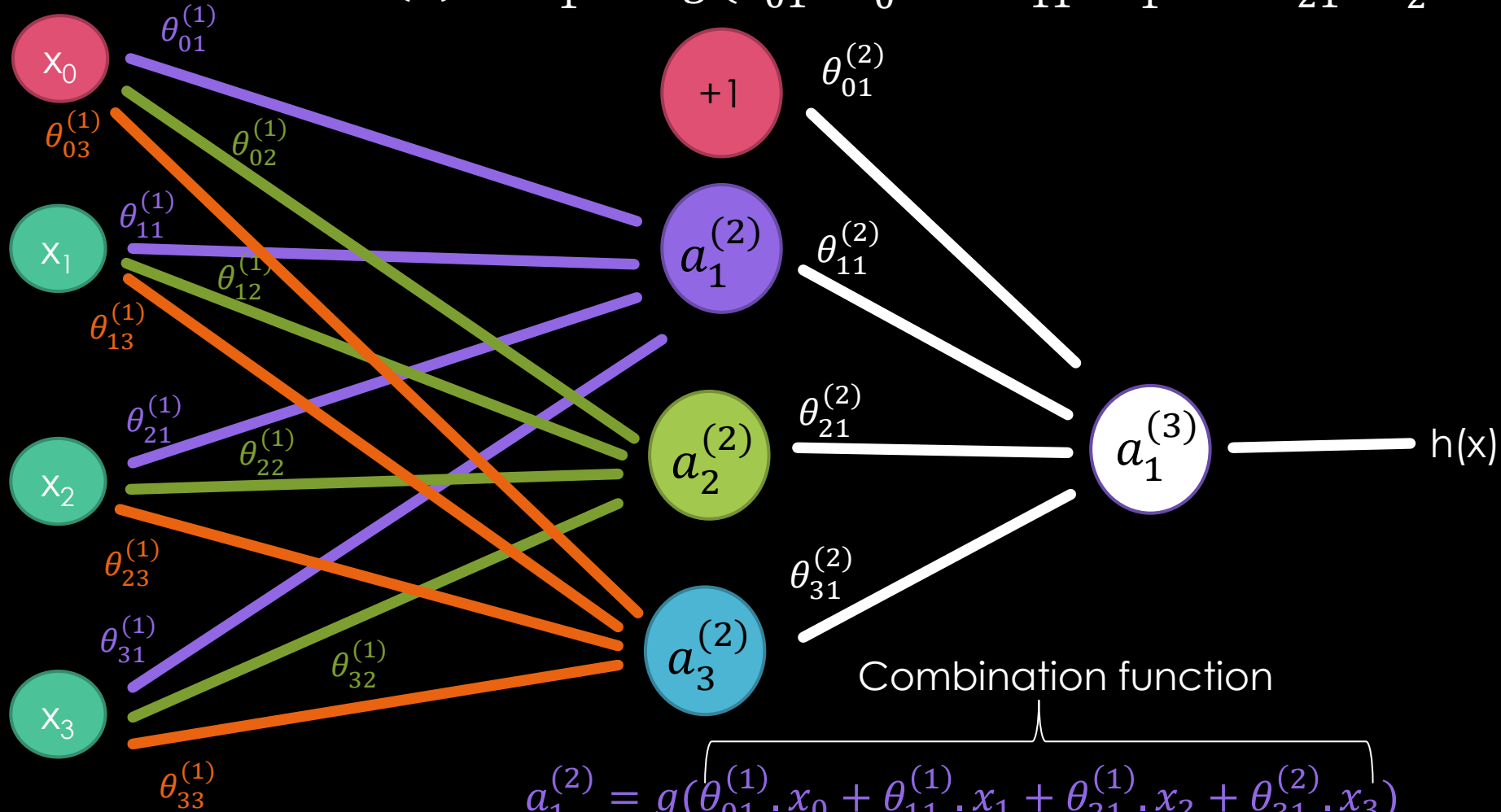
# MULTILAYER PERCEPTRON (MLP)

- UNITS/NODES/NEURONS
- FULLY CONNECTED OR NOT
- ONE OR MORE HIDDEN LAYERS





$$h(x) = a_1^{(3)} = g(\theta_{01}^{(2)} \cdot a_0^{(2)} + \theta_{11}^{(2)} \cdot a_1^{(2)} + \theta_{21}^{(2)} \cdot a_2^{(2)} + \theta_{31}^{(2)} \cdot a_3^{(2)})$$



Combination function

$$a_1^{(2)} = g(\theta_{01}^{(1)} \cdot x_0 + \theta_{11}^{(1)} \cdot x_1 + \theta_{21}^{(1)} \cdot x_2 + \theta_{31}^{(1)} \cdot x_3)$$


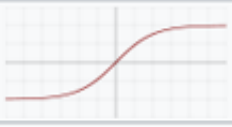

$$a_2^{(2)} = g(\theta_{02}^{(1)} \cdot x_0 + \theta_{12}^{(1)} \cdot x_1 + \theta_{22}^{(1)} \cdot x_2 + \theta_{32}^{(1)} \cdot x_3)$$

$$a_3^{(2)} = g(\theta_{03}^{(1)} \cdot x_0 + \theta_{13}^{(1)} \cdot x_1 + \theta_{23}^{(1)} \cdot x_2 + \theta_{33}^{(1)} \cdot x_3)$$

Activation/transfert function

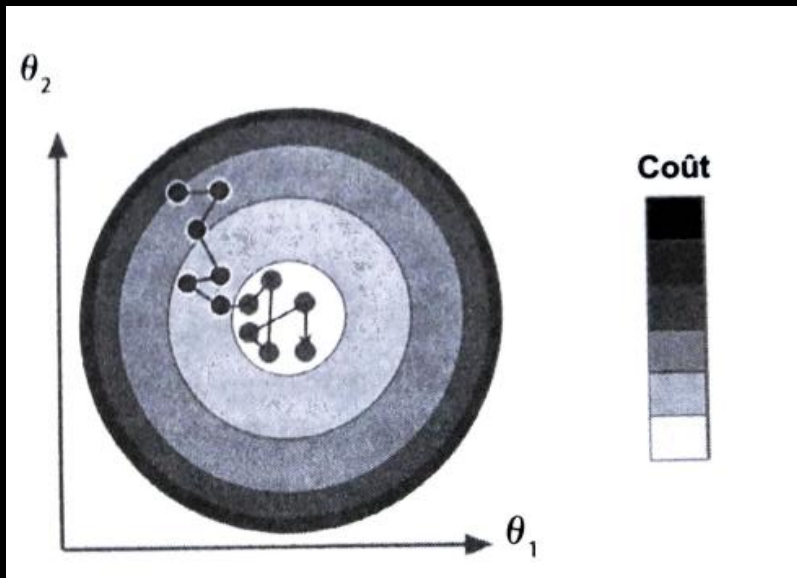
# ACTIVATION FUNCTIONS

## DIFFERENT FUNCTIONS

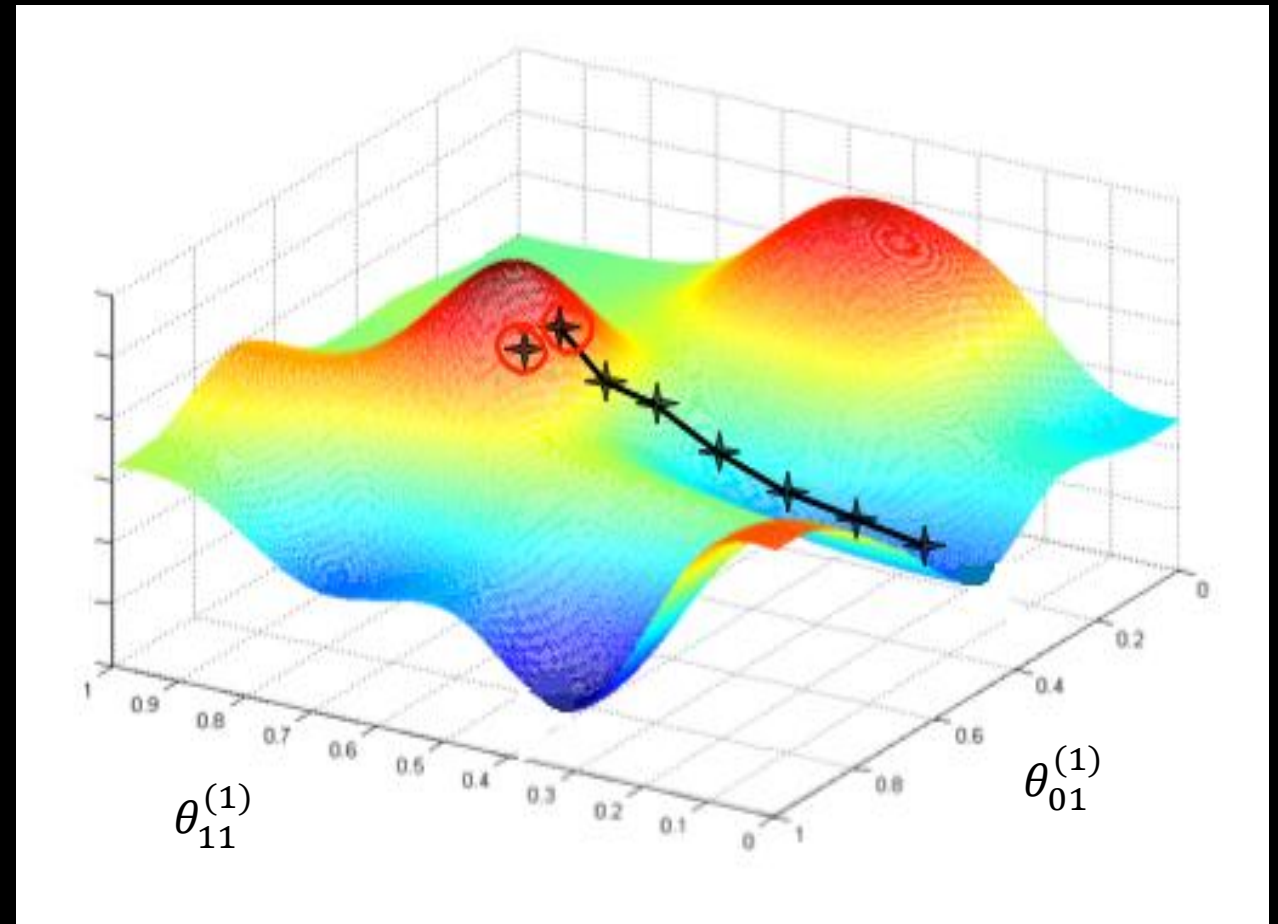
Logistic, sigmoid, or soft step		$\sigma(x) = \frac{1}{1 + e^{-x}}$ <sup>[1]</sup>	$f(x)(1 - f(x))$	$(0, 1)$
tanh		$\tanh(x) = \frac{e^x - e^{-x}}{e^x + e^{-x}}$	$1 - f(x)^2$	$(-1, 1)$
Rectified linear unit (ReLU) <sup>[7]</sup>		$\begin{cases} 0 & \text{if } x \leq 0 \\ x & \text{if } x > 0 \end{cases}$ $= \max\{0, x\} = x \mathbf{1}_{x>0}$	$\begin{cases} 0 & \text{if } x < 0 \\ 1 & \text{if } x > 0 \\ \text{undefined} & \text{if } x = 0 \end{cases}$	$[0, \infty)$

THE SOFTMAX FUNCTION IS RELATED TO THE SIGMOID FUNCTION. IF YOU HAVE 4 CLASSES, THE SOFTMAX FUNCTION WILL CALCULATE A PROBABILITY FOR EACH CLASS AND THE SUM OF THEM WILL BE EQUAL TO 1.

# GRADIENT DESCENT



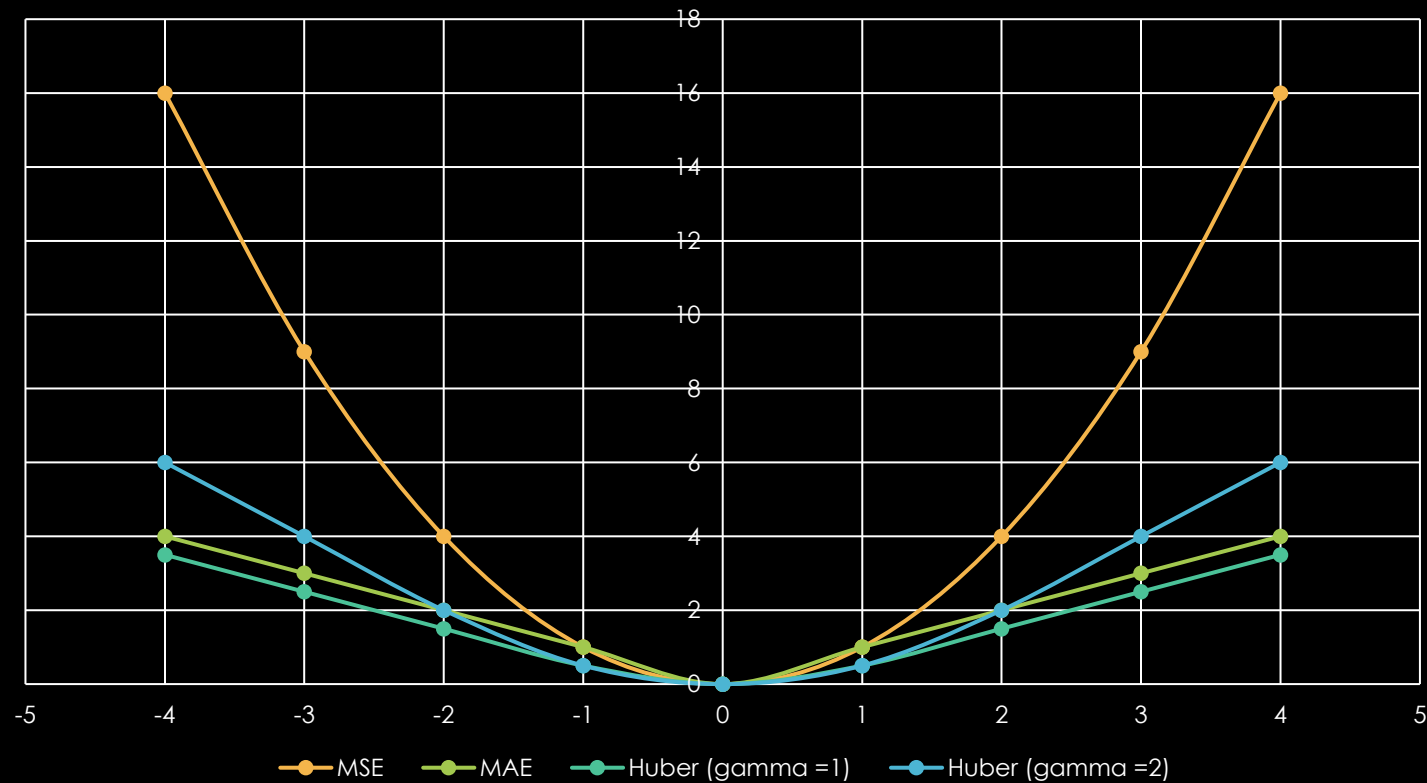
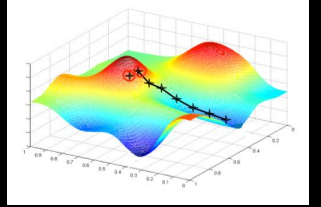
$J(\theta)$



Cost/loss function



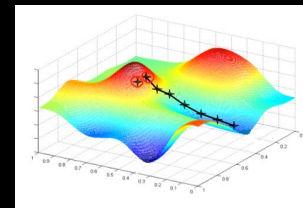
# LOSS/COST FUNCTION FOR REGRESSION



$$MAE = \frac{1}{n} \sum_{i=1}^{n_{sample}} |y_{obs} - y_{pred}|$$

$$MSE = \frac{1}{n} \sum_{i=1}^{n_{sample}} (y_{obs} - y_{pred})^2$$

$$Huber = \begin{cases} \frac{1}{2}(y_{obs} - y_{pred})^2 & \text{for } |y_{obs} - y_{pred}| \leq \delta \\ \delta|y_{obs} - y_{pred}| - \frac{1}{2}\delta^2 & \text{otherwise} \end{cases}$$



# LOSS/COST FUNCTION FOR CLASSIFICATION

Cross-entropy

$$H(p, q) = - \sum_x p(x) \ln(q(x))$$

p probability distribution

Dataset	
Animal	Human
0	1
0	1
1	0
1	0

q probability distribution

Predictions	
Animal	Human
0.2	0.8
0.1	0.9
0.6	0.4
0.9	0.1



Average cross-entropy = 0.236

Cross-entropy per sample

Sample

$$-(0 \cdot \ln(0.2) + 1 \cdot \ln(0.8)) = 0.223$$

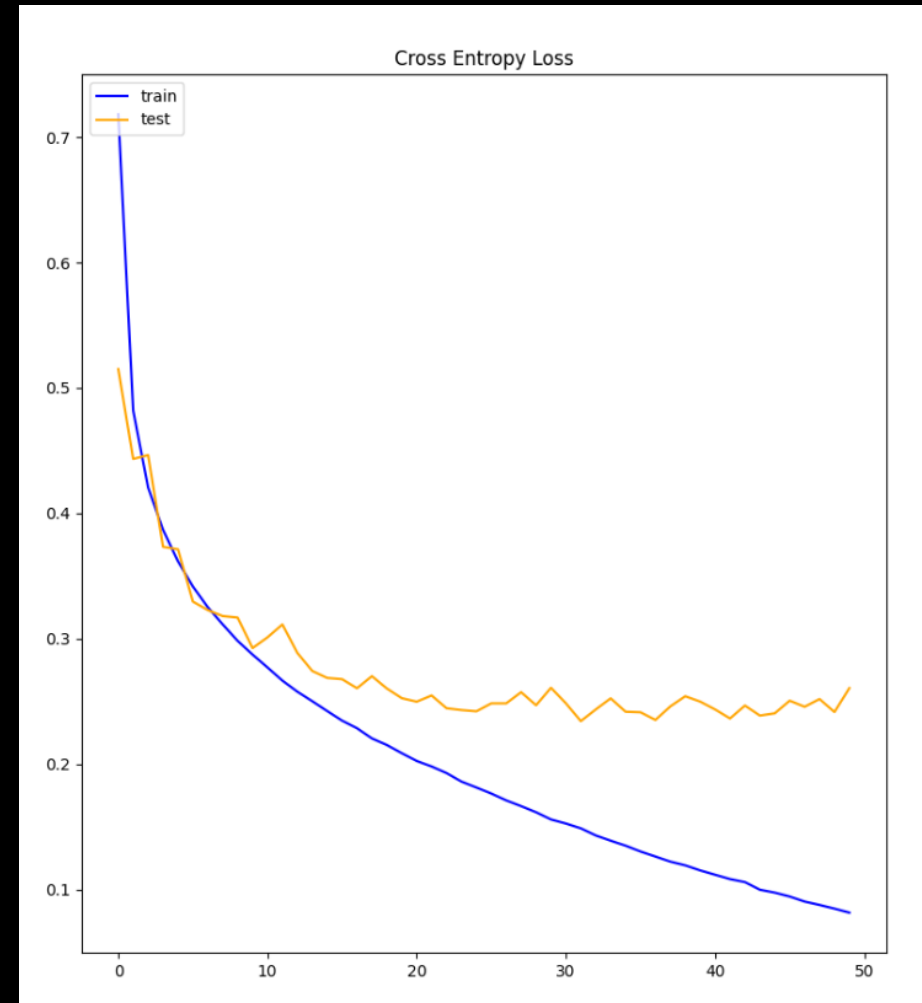
$$-(0 \cdot \ln(0.1) + 1 \cdot \ln(0.9)) = 0.105$$

$$-(1 \cdot \ln(0.6) + 0 \cdot \ln(0.4)) = 0.511$$

$$-(1 \cdot \ln(0.9) + 0 \cdot \ln(0.1)) = 0.105$$

# CROSS-ENTROPY

What is your conclusion ?

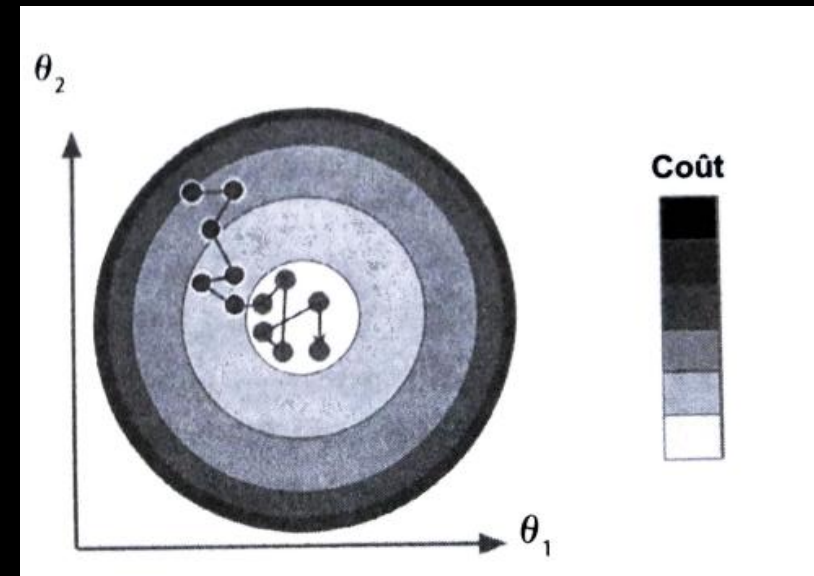
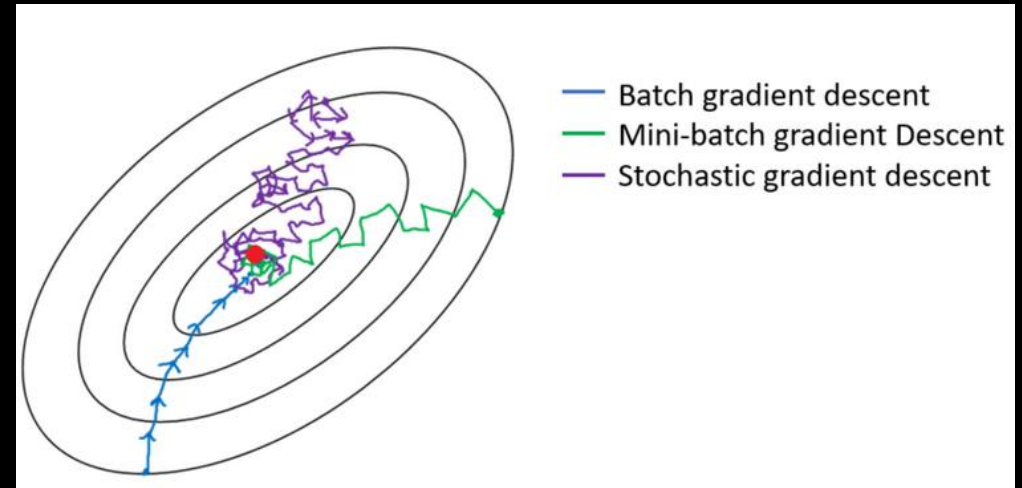


Epochs = iteration

# OPTIMIZER

- ORDINARY/BATCH GRADIENT DESCENT
  - SEEN IN THE LAST COURSE
  - ALL RECORDS
- STOCHASTIC GRADIENT DESCENT
  - RANDOM SELECTED RECORD
- MINI-BATCH GRADIENT DESCENT
  - BATCH OF USUALLY 32 RECORDS

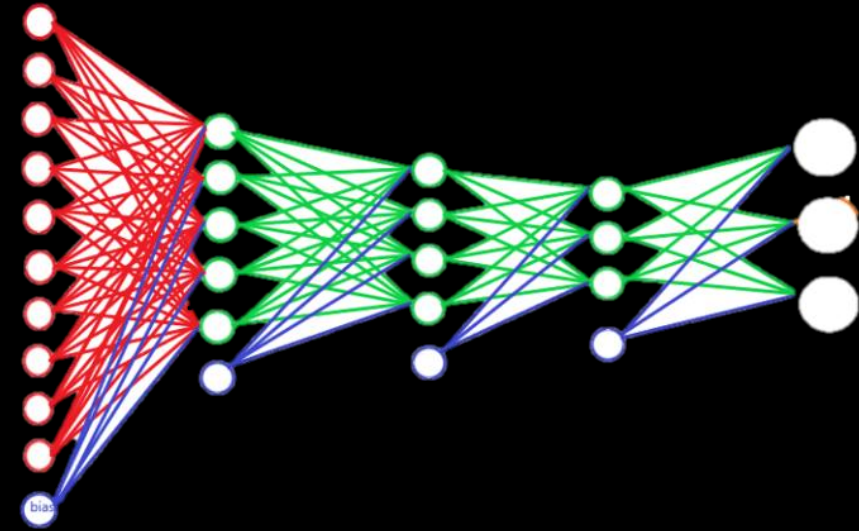
Others exist ... (adam...)



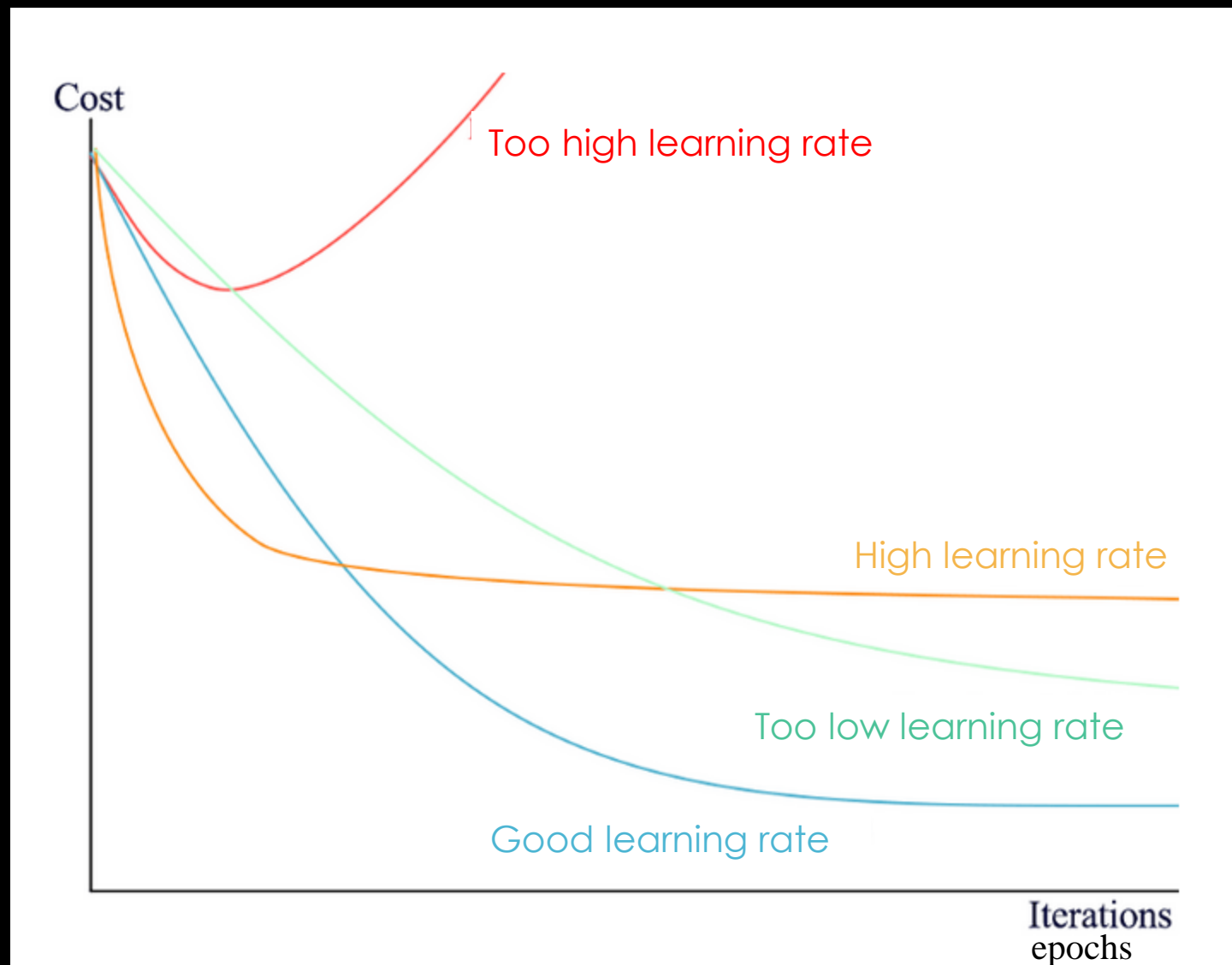
Do not reach the optimal value but are close

# MULTILAYER PERCEPTRON

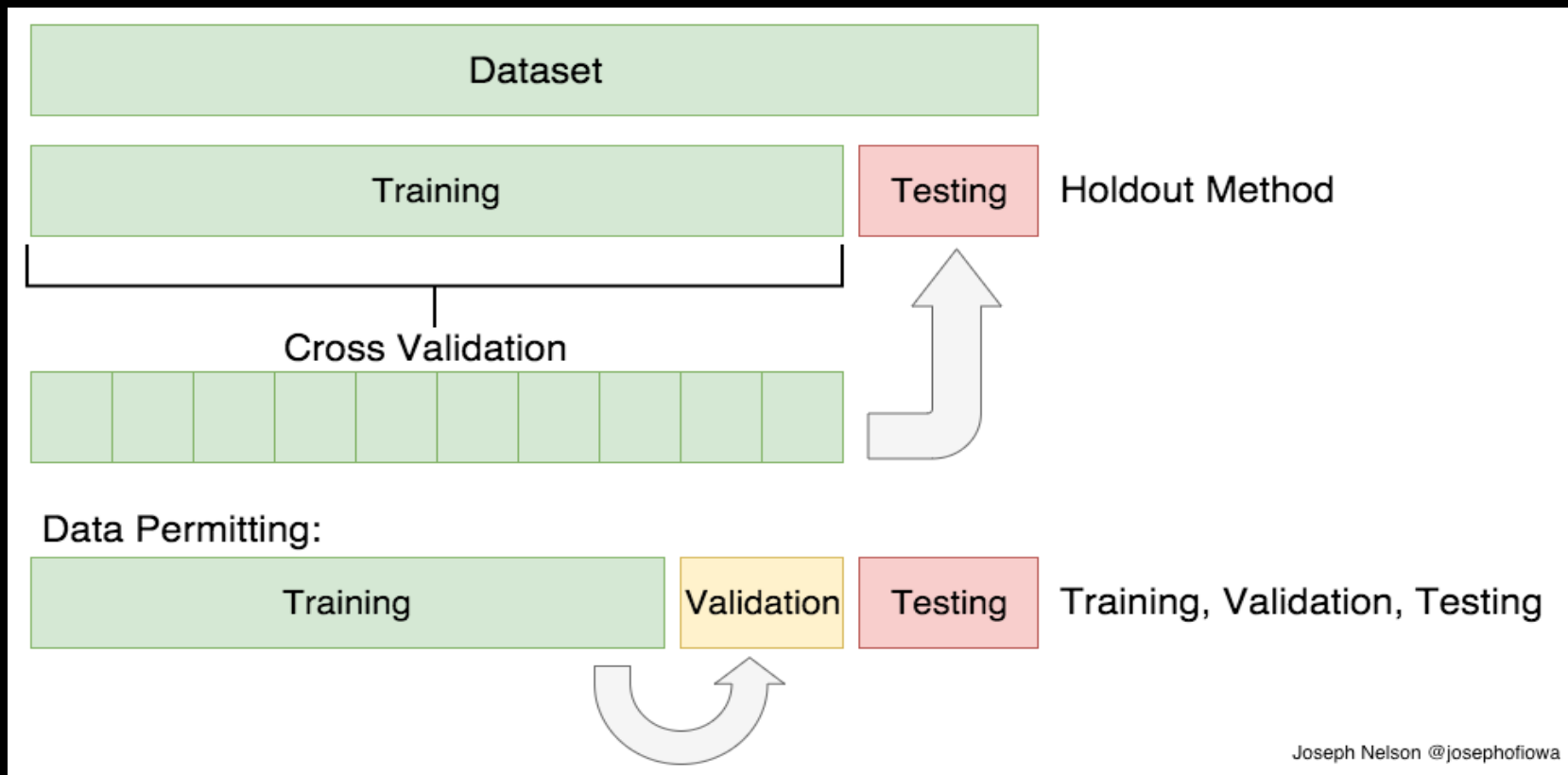
- HIGHER NEURONS  $\rightarrow$   $\uparrow$  COMPLEXITY
- HIGHER HIDDEN LAYERS  $\rightarrow$   $\uparrow$  COMPLEXITY
- $\neq$  RESULTS IF OTHER ACTIVATION FUNCTION
- $\neq$  RESULTS IF OTHER OPTIMIZER (MORE OR LESS IMPACTED BY LOCAL MINIMA)
- $\neq$  RESULTS IF OTHER LOSS FUNCTION
- $\neq$  RESULTS IF THE LEARNING RATE IS DIFFERENT
- YOU CAN ALSO ADD A REGULARISATION (L2 OR L1 NORM)



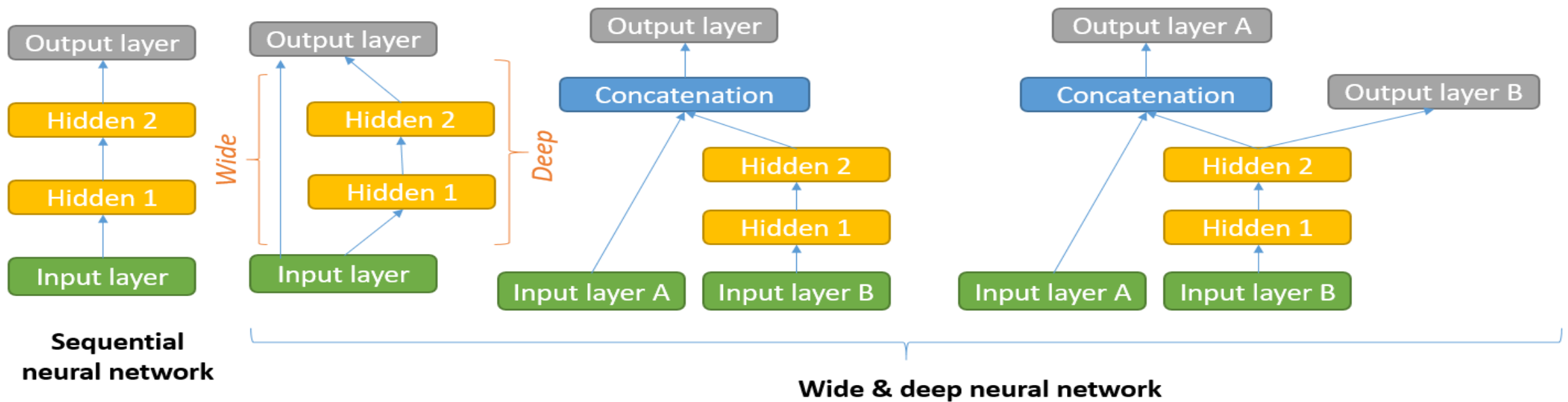
Take care to the overfitting



# HYPERPARAMETERS OPTIMIZATION



# WIDE & DEEP NEURAL NETWORK

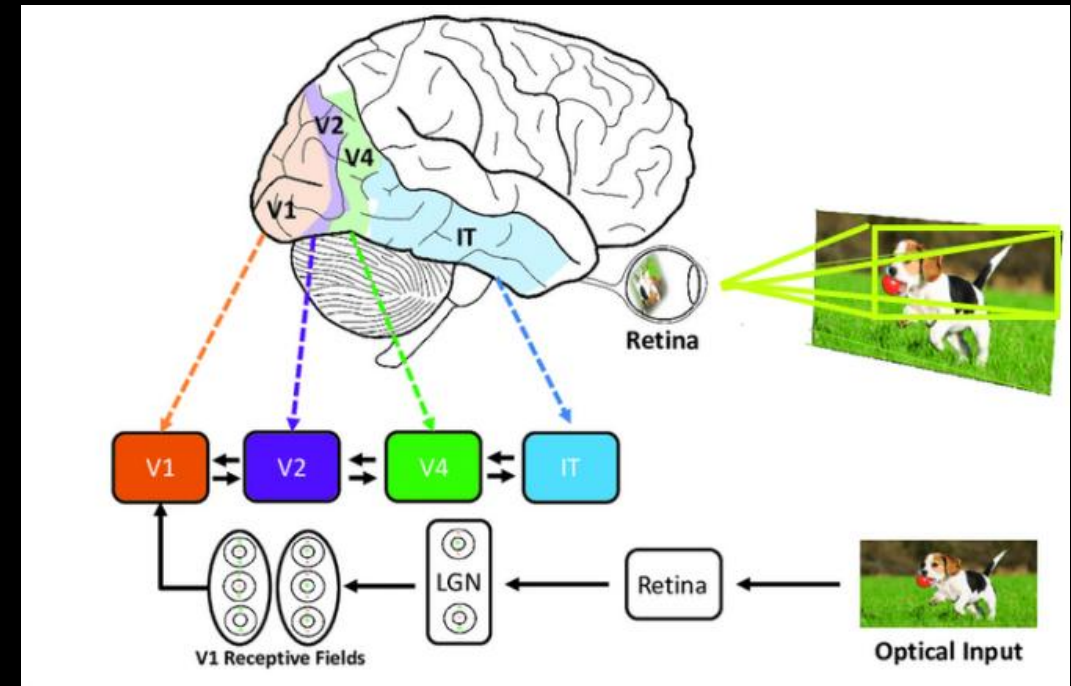




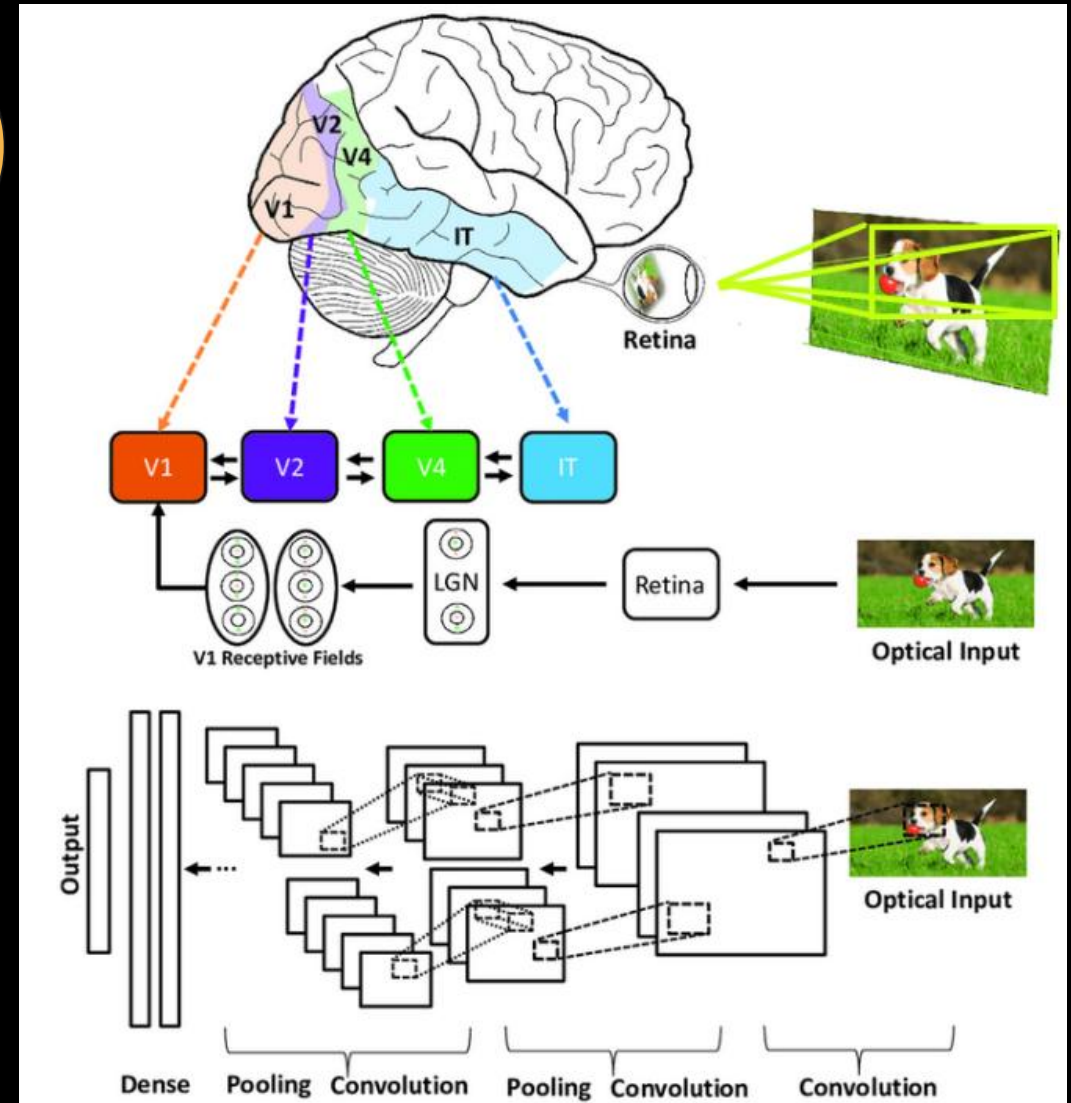
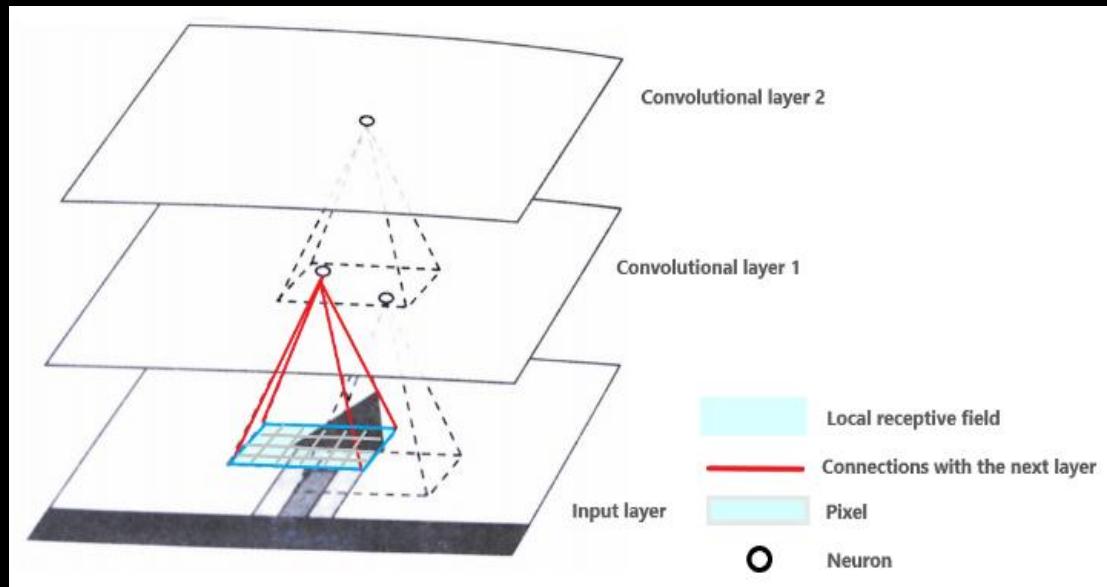
# CONVOLUTIONAL NEURAL NETWORK (CANN)

DAVID H. HUBEL AND TORSTEN WIESEL IN THE 50's OBSERVED THAT SEVERAL NEURONS IN THE VISUAL CORTEX OF CATS IN THE BRAIN FOCUS ON A RESTRICTED REGION OF THE VISUAL FIELD AND INTERACT ONLY IF A VISUAL STIMULI OCCURRED IN THIS REGION.

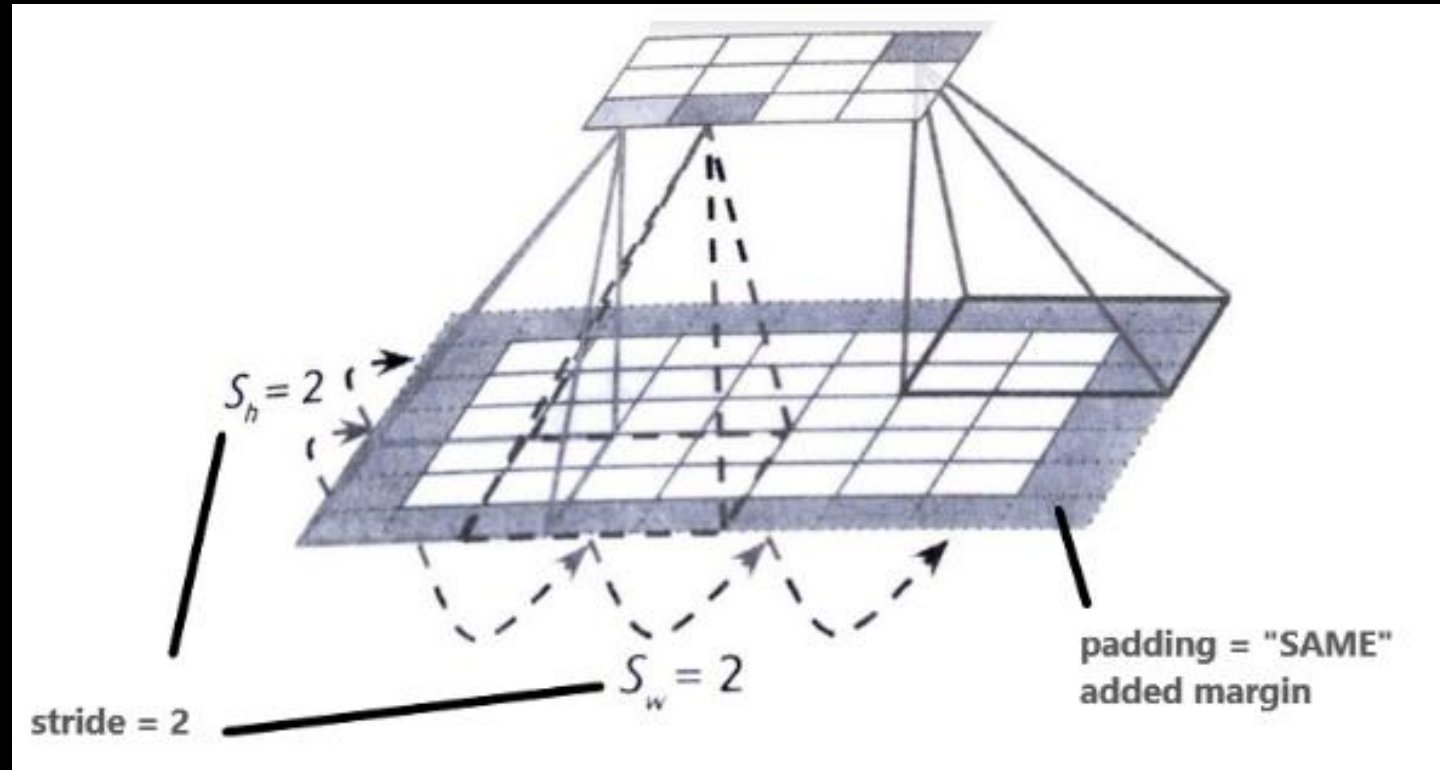
- No interest to link all neurons
- Local receptive field (LRF)



# LOCAL RECEPTIVE FIELD (LRF)



# PADDING AND STRIDE



# FILTER

2	0	1	1
0	1	0	0
0	0	1	0
0	3	0	0

Input image  
Padding = valid (no margin)  
Stride = 1

1	0	1
0	0	0
0	1	0

Filter (3x3)


Output image with  
stride = 1

**Multiplication**

$$\begin{aligned}
 & (2 \times 1) + (0 \times 0) + (1 \times 1) \\
 & + (0 \times 0) + (1 \times 0) + (0 \times 0) \\
 & + (0 \times 0) + (0 \times 1) + (1 \times 0) \\
 & = 3
 \end{aligned}$$

2	0	1	1
0	1	0	0
0	0	1	0
0	3	0	0

×

1	0	1
0	0	0
0	1	0

=

3	

2	0	1	1
0	1	0	0
0	0	1	0
0	3	0	0

×

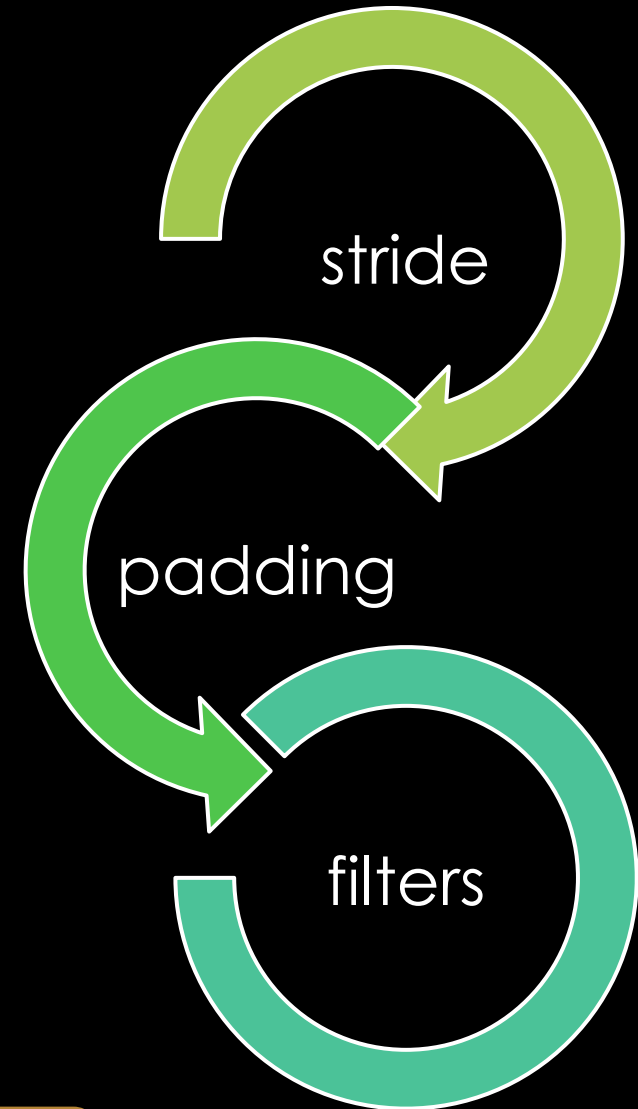
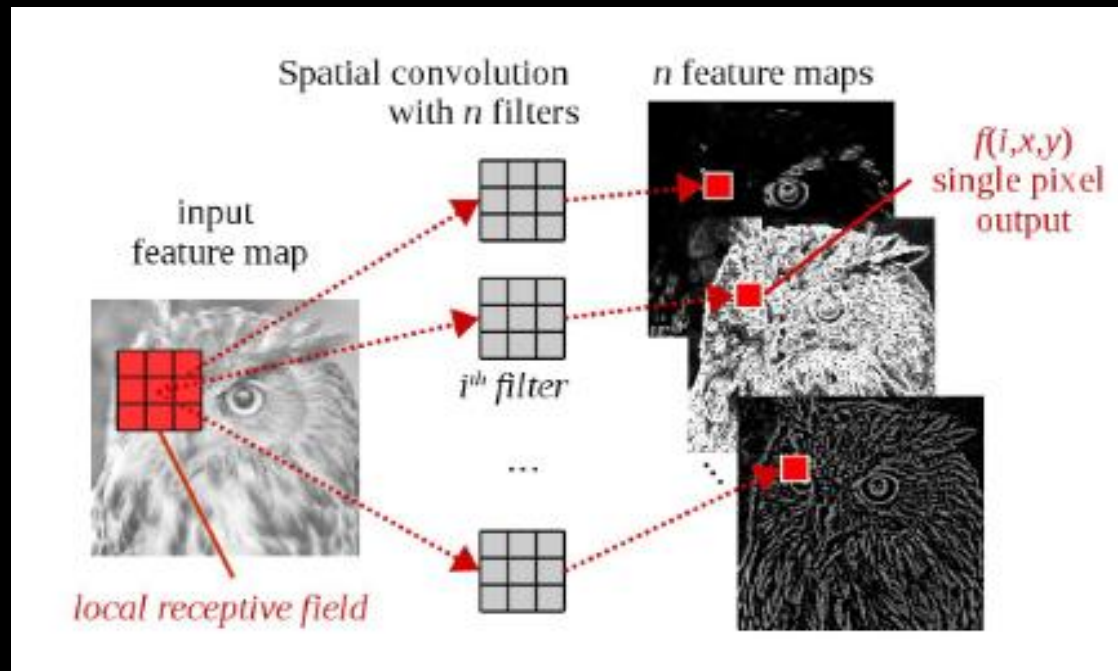
1	0	1
0	0	0
0	1	0

=

3	2
3	1

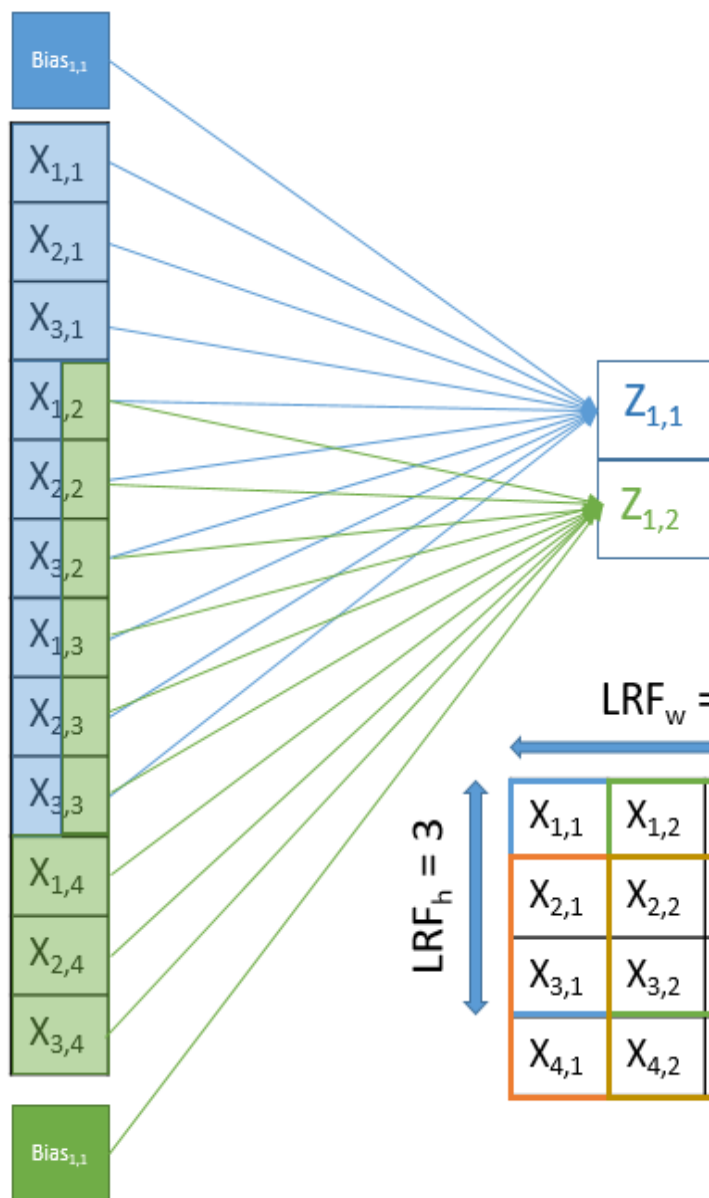
Final output image

# FEATURE MAPS



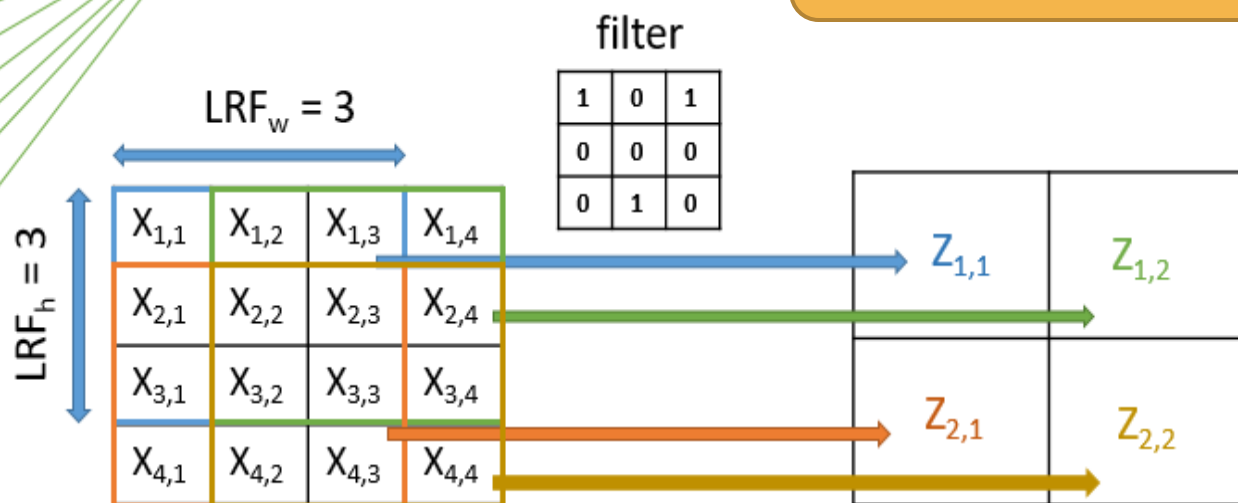
Where are the weights ?

# FEATURE MAP

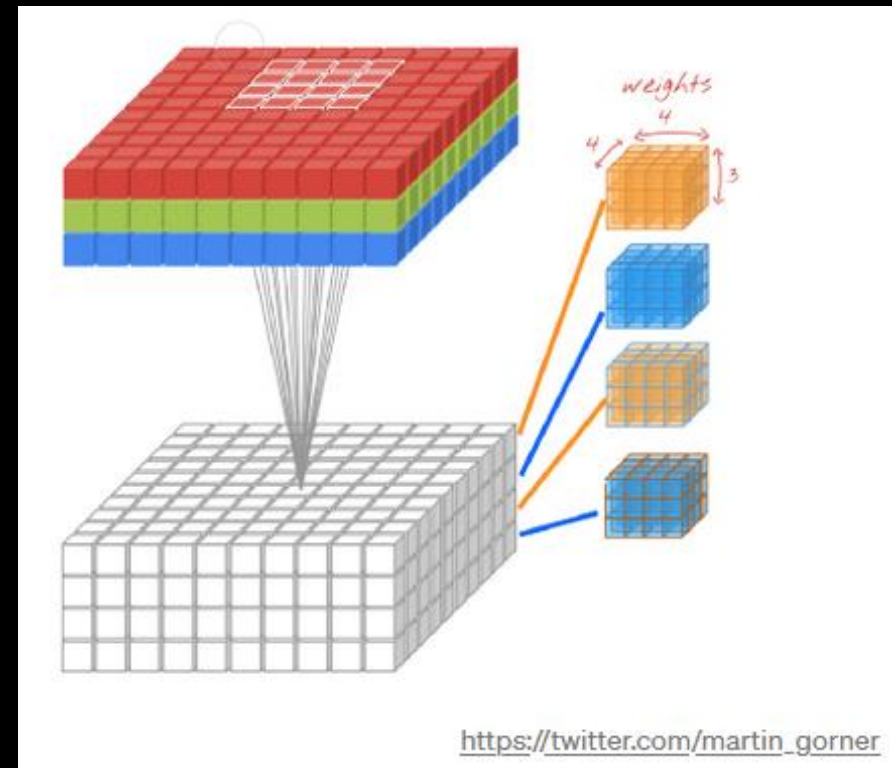
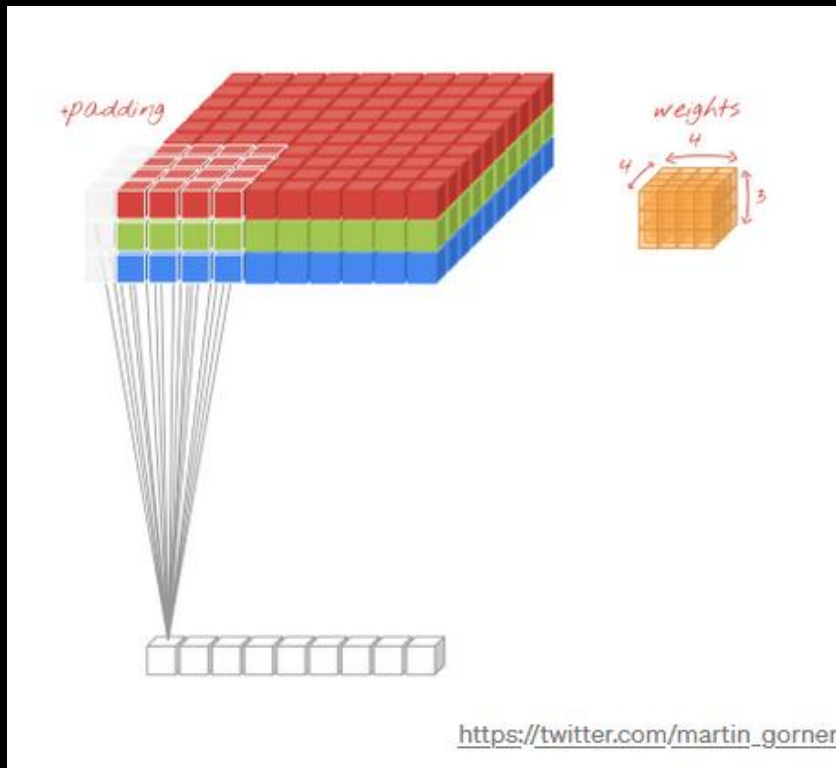


$$z_{1,1} = (f_{1,1} * w_{1,1} * x_{1,1}) + (f_{1,2} * w_{1,2} * x_{1,2}) + (f_{1,3} * w_{1,3} * x_{1,3}) \\ + (f_{2,1} * w_{2,1} * x_{2,1}) + (f_{2,2} * w_{2,2} * x_{2,2}) + (f_{2,3} * w_{2,3} * x_{2,3}) \\ + (f_{3,1} * w_{3,1} * x_{3,1}) + (f_{3,2} * w_{3,2} * x_{3,2}) + (f_{3,3} * w_{3,3} * x_{3,3}) \\ + bias_{1,1}$$

Not all neurons are connected



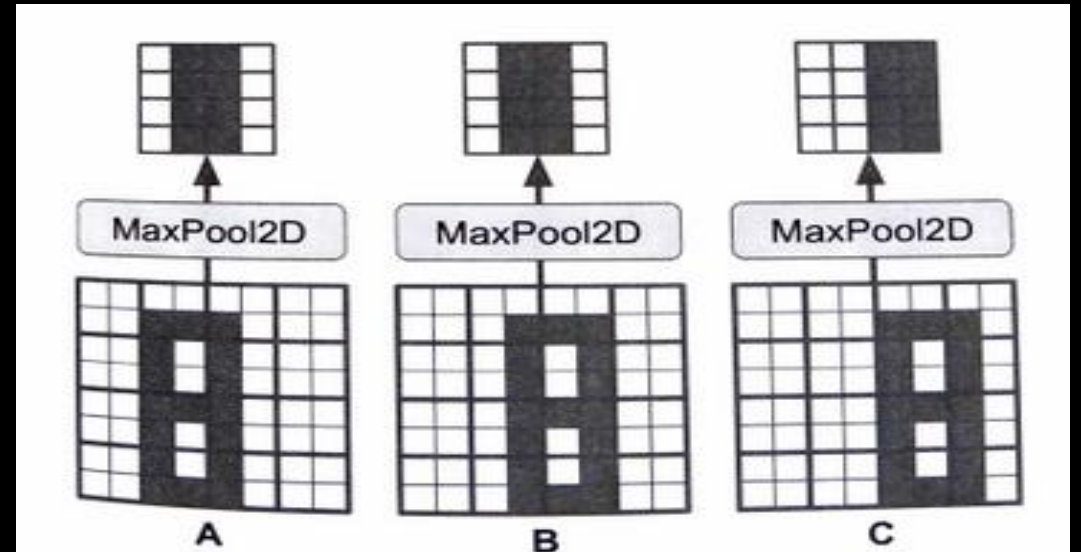
## 2D AND 3D CANN → FEATURE MAPS





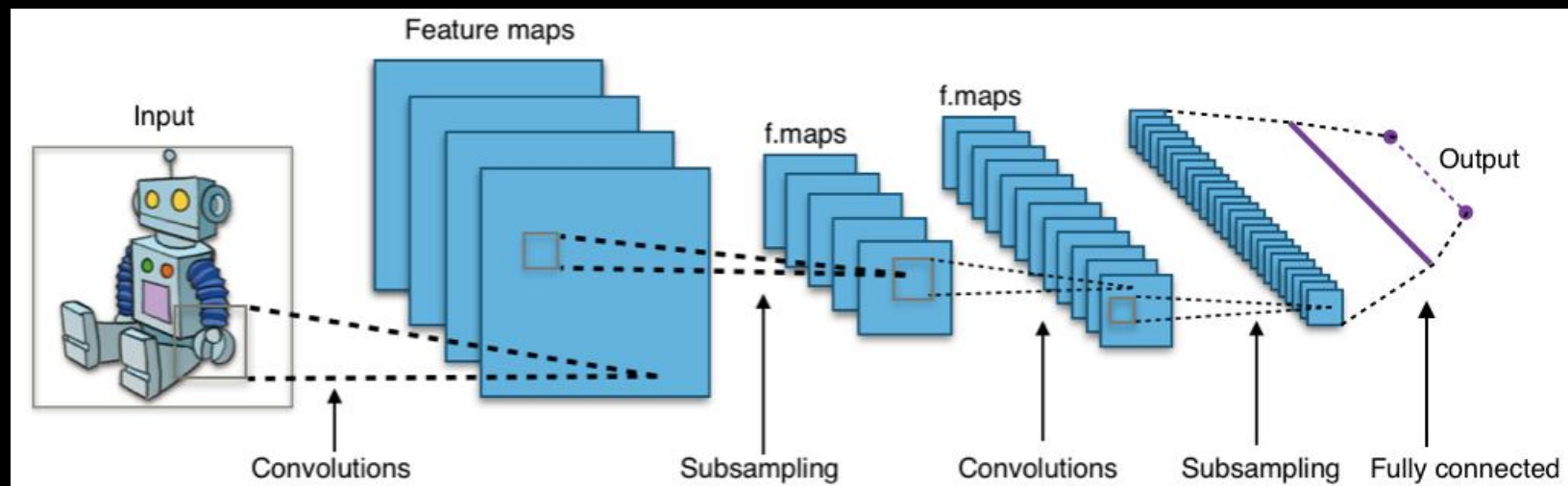
# POOLING LAYER

- DECREASE THE DIMENSIONALITY OF THE LAYER BY SUB-SETTING THE IMAGE
- $2 \times 2$  IS PREFERRED
- MAX OR MEAN IS USED





# CANN





# CURRENT CONCLUSIONS FOR DAIRY FARMING



# THE COMPLEXITY OF MODELS CHANGES

- Structure
- Number of parameters



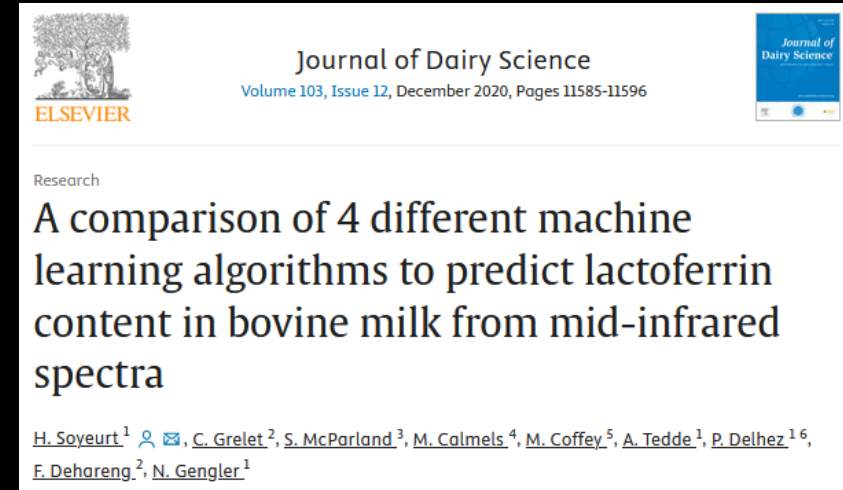


Table 1. The 10-fold cross-validation and external-validation performances for predicting lactoferrin content in milk using 4 different machine learning algorithms<sup>1</sup>

		PLSR	PLS + Linear SVR	PLS + Polynomial SVR	PLS + ANN
Selection function		oneSE	oneSE	best	best = oneSE <sup>2</sup>
Calibration (n = 5,541)	Parameters	nLV <sup>3</sup> = 23	C <sup>4</sup> = 5	degree = 3; scale = 0.01; C = 1	size = 4; decay = 0.5
	R <sup>2</sup> c	0.53	0.53	0.64	0.60
	RMSEc	140.94	144.32	125.89	130.59
Cross-validation	R <sup>2</sup> cv	0.51	0.53	0.56	0.55
	R <sup>2</sup> cv SD	0.03	0.03	0.03	0.03
	RMSEcv	144.31	144.60	138.40	139.01
	RMSEcv SD	5.77	5.61	8.08	5.05
	RPD	1.43	1.42	1.49	1.48
External validation (n = 836)	R <sup>2</sup> v	0.61	0.63	0.62	0.60
	RMSEv	163.76	174.92	166.75	162.17

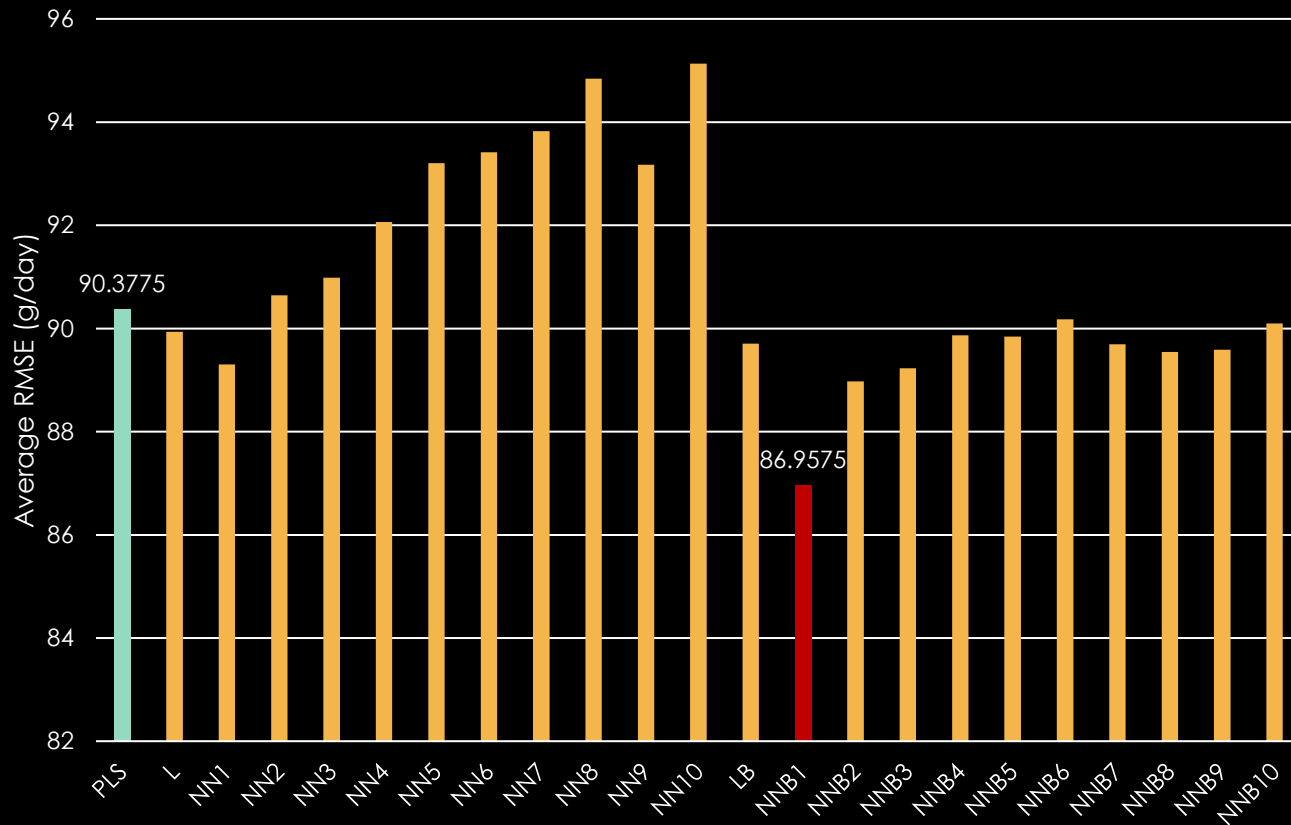
## Example : Lactoferrin

Mean : 260 mg/L

| Difference | PLS vs ANN : 1.59 mg/L → 0.61%  
 | Difference | PLS vs LSVM : 11.16 mg/L → 4.29%  
 | Difference | PLS vs PSVM : 2.99 mg/L → 1.15%  
 | Difference | ANN vs LSVM : 12.75 mg/L → 4.90%  
 | Difference | ANN vs PSVM : 4.58 mg/L → 1.76%  
 | Difference | PSVM vs LSVM : 8.17 mg/L → 3.14%

Small differences !





Research

## Predicting methane emission in Canadian Holstein dairy cattle using milk mid-infrared reflectance spectroscopy and other commonly available predictors via artificial neural networks

Saeed Shadpour<sup>1</sup>, Tatiane C.S. Chud<sup>1</sup>, Dagnachew Hailemariam<sup>2</sup>, Graham Plastow<sup>2</sup>, Hinayah R. Oliveira<sup>1</sup>, Paul Stothard<sup>2</sup>, Jan Lassen<sup>3</sup>, Filippo Miglior<sup>1,4</sup>, Christine F. Baes<sup>1</sup>, Dan Tulpan<sup>1</sup>, Flavio S. Schenkel<sup>1</sup>

Example : **Methane**


Mean : +/- 400g/jour

| Difference |<sub>PLS vs NN</sub> : 3g/day → 0.75%

Method	Predictor	Statistic					Method		Bias	RMSE	r	b	RPIQ
		Bias	RMSE	r	b	RPIQ							
Partial least squares	Previous						Neural networks	Previous					
	a.m.	-0.23 (8.97)	43.04 (2.36)	0.64 (0.03)	0.94 (0.11)	1.76 (0.16)		a.m.	0.12 (8.66)	42.50 (1.64)	0.64 (0.02)	0.90 (0.11)	1.76 (0.16)
	p.m.	0.03 (9.49)	43.73 (2.48)	0.63 (0.03)	0.93 (0.11)	1.74 (0.16)		p.m.	-0.11 (10.00)	43.15 (2.04)	0.64 (0.03)	0.87 (0.10)	1.73 (0.18)
	a.m. and p.m.	-0.01 (8.89)	42.15 (1.52)	0.65 (0.03)	0.94 (0.10)	1.77 (0.17)		a.m. and p.m.	-0.42 (8.26)	41.14 (1.86)	0.67 (0.02)	0.90 (0.09)	1.81 (0.16)
	Average							Average					
	a.m. and p.m.	-0.09 (8.81)	41.76 (1.98)	0.66 (0.03)	0.95 (0.11)	1.79 (0.18)		a.m. and p.m.	-0.42 (7.96)	41.45 (1.45)	0.66 (0.03)	0.91 (0.12)	1.80 (0.15)
	Following							Following					
	a.m.	-0.49 (9.34)	44.62 (1.46)	0.61 (0.02)	0.95 (0.09)	1.71 (0.11)		a.m.	-0.73 (8.93)	43.59 (1.42)	0.64 (0.02)	0.92 (0.11)	1.75 (0.11)
	p.m.	-0.06 (9.93)	45.23 (0.90)	0.60 (0.04)	0.93 (0.12)	1.69 (0.12)		p.m.	-0.36 (9.08)	44.36 (0.37)	0.62 (0.03)	0.89 (0.07)	1.72 (0.11)
	a.m. and p.m.	-0.20 (9.14)	44.14 (0.76)	0.62 (0.03)	0.94 (0.10)	1.73 (0.11)		a.m. and p.m.	-0.96 (8.84)	42.48 (1.02)	0.66 (0.03)	0.89 (0.08)	1.80 (0.13)
	Average							Average					
	a.m. and p.m.	-0.56 (9.43)	44.00 (1.40)	0.63 (0.03)	0.95 (0.10)	1.74 (0.11)		a.m. and p.m.	-0.46 (8.23)	42.33 (1.38)	0.66 (0.03)	0.92 (0.09)	1.81 (0.13)
	Flanking							Flanking					
	a.m.	-0.42 (9.08)	38.09 (1.87)	0.68 (0.03)	0.95 (0.11)	1.87 (0.17)		a.m.	0.16 (8.97)	38.23 (1.50)	0.68 (0.03)	0.92 (0.12)	1.86 (0.17)
	p.m.	-0.11 (9.71)	39.25 (1.57)	0.66 (0.03)	0.93 (0.12)	1.81 (0.15)		p.m.	-0.46 (9.27)	39.16 (0.94)	0.66 (0.02)	0.87 (0.07)	1.82 (0.14)
	a.m. and p.m.	-0.10 (8.83)	37.98 (1.36)	0.68 (0.03)	0.95 (0.11)	1.87 (0.16)		a.m. and p.m.	-0.77 (8.43)	37.17 (1.53)	0.70 (0.02)	0.91 (0.09)	1.91 (0.15)
	Average							Average					
	a.m. and p.m.	-0.25 (8.92)	37.56 (2.00)	0.69 (0.03)	0.96 (0.11)	1.90 (0.16)		a.m. and p.m.	-0.33 (7.95)	37.46 (4.01)	0.71 (0.03)	0.92 (0.11)	1.95 (0.16)


Example : **Methane**

Also low differences



Journal of Dairy Science

Volume 107, Issue 2, February 2024, Pages 978-991



Research

## Predicting methane emissions of individual grazing dairy cows from spectral analyses of their milk samples

S. McParland, M. Frizzarin, B. Lahart, M. Kennedy, L. Shalloo, M. Egan, K. Starsmore, D.P. Berry

# Identifying Health Status in Grazing Dairy Cows from Milk Mid-Infrared Spectroscopy by Using Machine Learning Methods

by Brenda Contla Hernández <sup>1</sup> , Nicolas Lopez-Villalobos <sup>2</sup>  and Matthieu Vignes <sup>1,\*</sup> 

<sup>1</sup> School of Fundamental Sciences, Massey University, Palmerston North 4442, New Zealand

<sup>2</sup> School of Agriculture and Environment, Massey University, Palmerston North 4442, New Zealand

\* Author to whom correspondence should be addressed.

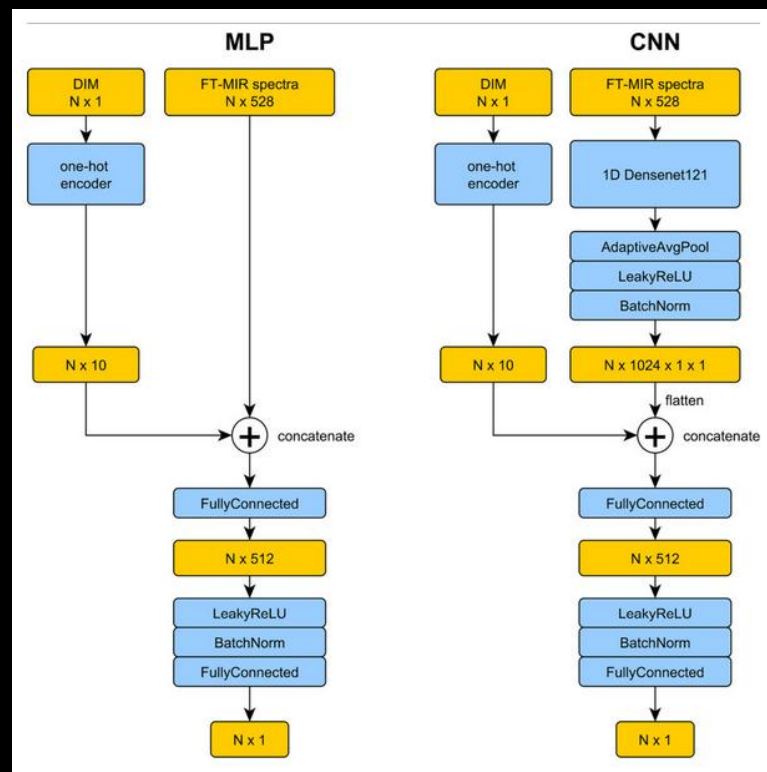
*Animals* **2021**, *11*(8), 2154; <https://doi.org/10.3390/ani11082154>

More nuanced in classification

**Table 2.** Performance of classification models obtained in 10 Monte Carlo cross-validation for classifying any health problem and healthy cows during lactation (early, mid and lactation) at two dairy farms during the 2016 production season <sup>1</sup>.

Models <sup>2</sup>	Sensitivity	Specificity	Accuracy	PPV	NPV	AUC	MCC
PLS-DA	65.60 ± 5.97	79.59 ± 2.36	78.85 ± 2.23	15.25 ± 3.07	97.66 ± 0.5	72.59 ± 3.27	0.24 ± 0.04
RF	46.22 ± 8.62	79.26 ± 2.15	77.51 ± 1.75	10.94 ± 1.88	96.38 ± 0.73	62.74 ± 3.78	0.14 ± 0.04
SVM	66.39 ± 6.80	76.39 ± 2.92	75.84 ± 2.42	13.48 ± 1.62	97.61 ± 0.61	71.39 ± 2.37	0.22 ± 0.02
NN	61.74 ± 15.99	97.00 ± 2.85	95.16 ± 3.26	59.99 ± 26.20	97.87 ± 0.87	79.37 ± 9.16	0.58 ± 0.22
CNN	57.02 ± 12.70	92.5 ± 5.27	90.63 ± 4.98	33.82 ± 13.41	97.5 ± 0.75	74.76 ± 6.88	0.39 ± 0.13
ESA	57.15 ± 12.38	87.61 ± 6.19	86.02 ± 6.21	24.06 ± 13.07	97.36 ± 0.77	72.38 ± 8.48	0.31 ± 0.16
ESMJ	60.75 ± 5.98	83.57 ± 2.56	82.36 ± 2.27	17.18 ± 3.21	97.46 ± 0.55	72.16 ± 2.9	0.25 ± 0.04
ESWA	56.43 ± 14.56	85.13 ± 7.41	83.61 ± 7.36	21.33 ± 14.18	97.22 ± 0.97	70.78 ± 9.71	0.27 ± 0.17

<sup>1</sup> These values correspond to the mean ± SD obtained by 10-fold Monte Carlo cross-validation for classifying any health problem (lameness, mastitis, reproductive disorder, etc.). From the cows' records, the positive cases were cows that had any illness (lameness, mastitis, reproductive disorder, etc.) and negative cases were cows who were healthy (no diagnosed disease); SD = Standard deviation; PPV = positive predicted value; NPV = negative predicted value; AUC = area under the receiver operating characteristic curve; MCC = Matthews correlation coefficient. <sup>2</sup> Models used to perform the classification: PLS-DA = partial least squares discriminant analysis, RF = random forest, SVM = support vector machine, NN = neural network, CNN = convolutional neural network, ESA = ensemble stacking average, ESMJ = ensemble stacking major voting and ESWA = ensemble stacking weighted average.



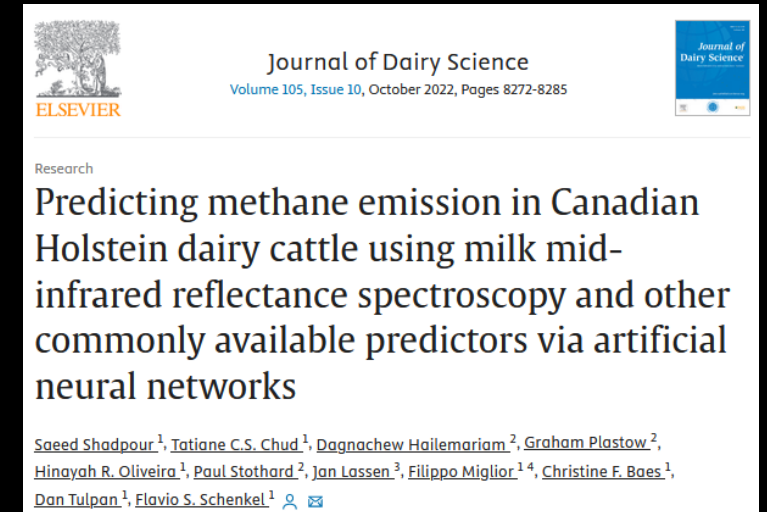
More nuanced in classification

Table 3. Model performance for multilayer perceptron (MLP) and convolutional neural network (CNN) approaches based on strategy 3 data<sup>1</sup>: accuracy (Acc), sensitivity (Sens), specificity (Spec), and area under the receiver operating characteristic curve (AUC) values within the training, herd-independent validation (VAL-Test) and pregnancy-associated glycoproteins validation (VAL-PAG) data sets

Deep learning approach <sup>2</sup> and model <sup>3</sup>	Training				Test validation (VAL-Test)				Glycoprotein-based validation (VAL-PAG)			
	Acc	Sens	Spec	AUC	Acc	Sens	Spec	AUC	Acc	Sens	Spec	AUC
MLP approach												
FT-MIR spectra	0.592	0.574	0.611	0.628	0.586	0.580	0.607	0.632	0.664	0.672	0.569	0.669
FT-MIR spectra + DIM	0.594	0.621	0.566	0.631	0.614	0.629	0.564	0.635	0.692	0.709	0.499	0.647
FT-MIR spectra (pre-adjusted for DIM)	0.559	0.554	0.564	0.583	0.562	0.567	0.547	0.581	0.554	0.547	0.636	0.636
CNN approach												
FT-MIR spectra	0.625	0.625	0.625	0.675	0.611	0.620	0.582	0.641	0.684	0.696	0.554	0.676
FT-MIR spectra + DIM	0.645	0.670	0.620	0.700	0.636	0.659	0.563	0.654	0.723	0.741	0.519	0.685
FT-MIR spectra (pre-adjusted for DIM)	0.982	0.975	0.988	0.998	0.668	0.790	0.273	0.551	0.759	0.805	0.266	0.564



The variability is bigger within the same model using different sets of features



Features	PLS	L	NN1	NN2	NN3	NN4	NN5	NN6	NN7	NN8	NN9	NN10	LB	NNB1	NNB2	NNB3	NNB4	NNB5	NNB6	NNB7	NNB8	NNB9	NNB10	SD
1	96.47	96.55	93.08	93.16	93.33	94.74	93.98	94.36	97.65	97.65	96.54	99.39	96.26	91.67	93.01	92.59	92.07	92.41	92.2	92.2	92.22	92.08	92.45	2.26
2	95.68	95.29	91.4	93.47	94.64	94.05	95.54	96.11	95.44	98.04	97.66	96.16	95.61	91.93	92.55	93.26	92.89	93.12	93.07	93.06	92.97	92.92	93.1	1.78
3	96.32	96.36	93.58	94.44	93.96	94.55	94.71	97.27	97.69	97.39	95.15	98.47	95.99	92.49	93.19	91.46	91.91	92.35	92.45	92.3	92.31	92.46	92.59	2.13
4	95.84	95.73	93.4	94.18	96.15	93.58	96.82	94.14	96.05	97.44	96.65	96.57	95.59	91.8	93.41	93.14	93.35	92.84	92.66	92.81	93.17	93.22	92.79	1.67
5	96.5	96.72	95.23	94.89	93.41	97.01	98.35	98.94	97.31	99.91	100.94	100.9	96.11	92.6	93.87	93.05	92.82	93.08	92.66	92.98	92.83	92.8	92.83	2.84
6	96.75	96.07	94.41	95.86	96.8	97.44	97.89	101.16	100.12	100.65	98.93	101.93	95.66	92.55	93.46	93.87	93.33	93.07	93.4	93.45	93.34	93.35	93.62	2.94
7	73.7	72.44	77.56	81.19	81.51	85.42	84.69	82.72	84.49	82.27	80.79	84.57	71.61	71.75	76.2	78.96	79.48	81.59	82.58	78.65	80.36	79.33	81.66	4.15
8	71.76	70.31	75.75	77.95	78.08	79.72	83.64	82.61	81.85	85.36	78.72	83.08	70.83	70.87	76.12	77.5	83.09	80.28	82.42	82.1	79.16	80.56	81.71	4.44
SD	10.91	11.48	7.90	6.94	7.08	6.20	5.78	7.02	6.76	6.96	8.48	7.26	11.42	9.67	7.92	6.83	5.41	5.52	4.75	5.84	6.06	5.98	5.20	

➔ Not only focus on the model type

# CONCLUSIONS

- **A lot of hyperparameters** to tune
- Infinite possibility of cANN and MLP structure
- Take care to the reproducibility of the model
- Limited interest for regression but high interest for classification and **image processing**
- However, **transfer learning** can solve problems of data sharing or allows to deal with large database



PYTHON SCRIPT EXAMPLE





```
#####
# MLP
#####

# MLP
ANN1 = tf.keras.Sequential([
    tf.keras.Input(shape=(100, 100, 3)),
    tf.keras.layers.Flatten(),
    tf.keras.layers.Dense(700, activation="relu"),
    tf.keras.layers.Dense(300, activation="relu"),
    tf.keras.layers.Dense(100, activation="relu"),
    tf.keras.layers.Dense(31, activation="softmax")
])

print("Perceptron with one hidden layer", ANN1.summary())
tf.keras.utils.plot_model(ANN1)

# network compilation
ANN1.compile(
    optimizer=tf.keras.optimizers.SGD(learning_rate=0.01, momentum=0.9),
    loss='categorical_crossentropy',
    metrics=['accuracy']
)

# Fit the network
history = ANN1.fit( x = xtrain,
                    y = ytrain,
                    epochs = 50, # 50 epochs
                    batch_size = 64, # mini-batch gradient descent with 32 samples
                    validation_split = 0.2,
                    callbacks = [tf.keras.callbacks.EarlyStopping(patience=3)]) # patience option is interesting to be sure that the minimum is not a local minimum.
```

```

# model creation : 64 filters - 256 neurons in the last hidden layer
cANN = tf.keras.Sequential([
    tf.keras.Input(shape=(100, 100, 3)), # declare input shape separately
    tf.keras.layers.Conv2D(10, 7, activation="relu", padding="same"),
    tf.keras.layers.MaxPooling2D(2),
    tf.keras.layers.Flatten(),
    tf.keras.layers.Dense(64, activation="relu"),
    tf.keras.layers.Dropout(0.5),
    tf.keras.layers.Dense(64, activation="relu"),
    tf.keras.layers.Dense(31, activation="softmax") # adjust output units to your number of classes
])

print("Convolutional neural network", cANN.summary())
tf.keras.utils.plot_model(cANN)

# network compilation
cANN.compile(tf.keras.optimizers.SGD(learning_rate=0.01, momentum=0.9), # optimizer "adam" "sgd"
             loss='categorical_crossentropy', #tf.keras.losses.SparseCategoricalCrossentropy(from_logits=True),
             metrics=['accuracy'])

# Fit the network
history2 = cANN.fit(x=xtrain,
                   y=ytrain,
                   batch_size=64,
                   epochs=50,
                   verbose=1,
                   validation_data=(xtest, ytest),
                   callbacks = [tf.keras.callbacks.EarlyStopping(patience=3)]) # patience option is interesting to be sure that the minimum is not a local minimum.

```