

Breast cancer progression: insights into multifaceted matrix metalloproteinases

Vincent Chabottaux & Agnès Noel

Laboratory of Tumor and Developmental Biology, Center for Experimental Cancer Research (CRCE), Groupe Interdisciplinaire de Génomprotéomique Appliquée (GIGA-Research), University of Liege, Tour de Pathologie (B23), Sart-Tilman, Liège 4000, Belgium

Abstract: The restricted view of matrix metalloproteinases (MMPs) as simple destroyers of extracellular matrix components has largely ignored their substantial contribution in many aspects of cancer development and metastatic dissemination. Over the last few years, the relevance of MMPs in the processing of a large array of extracellular and cell surface-associated proteins has grown considerably. Our knowledge about the complex functions of MMPs and how their contribution may differ throughout cancer progression is rapidly expanding. These new findings provide several explanations for the lack of success of MMP inhibition in clinical trials. A complete understanding of MMP biology is needed before considering them, their substrates or their products as therapeutic targets. In this review, we explore the different faces of MMP implication in breast cancer progression by considering both clinical and fundamental aspects.

Keywords: Angiogenesis, Breast cancer, Cancer invasion, Degradome, Matrix metalloproteinases, Metastases; Stromal proteases

Introduction

Tumorigenesis and cancer progression rely on the acquisition by tumor cells of novel capacities which are shared by most if not all cancer types. According to Hanahan and Weinberg, six essential alterations in cellular physiology dictate malignant growth: (1) production of autocrine growth signals; (2) insensitivity to growth-inhibitory signals; (3) escape from apoptosis; (4) limitless replicative potential; (5) sustained angiogenesis and (6) tissue invasion and metastatic dissemination [1]. Initially, Matrix Metalloproteinases (MMPs) were claimed to be important in late stages of tumor progression by controlling tumor cell migration, invasion and metastasis through ECM degradation. However, due to the rapid development of innovative biochemical techniques [2-4] and the expanding use of transgenic and knockout mice [5, 6], it became obvious that the action of MMPs is not restricted to the massive destruction of physiological matrix barriers [7]. MMPs are now viewed as key regulators of the multiple cellular functions which dictate malignant growth. Although some MMPs are produced by tumor cells (e.g. MMP-7), most MMPs are rather produced by stromal cells and therefore might be considered as molecular determinants of the "seed and soil" concept proposed by Paget in 1889 [8]. Breast carcinomas are often characterized by a stromal reaction that consists of modifications in the composition of both cellular elements (infiltration of fibroblastic cells, endothelial cells and inflammatory cells) and the extracellular matrix (ECM) [9, 10]. An expansion of the tumor stroma and an increased deposition of ECM known as desmoplasia is often associated to invasive breast carcinomas [11]. Fibroblasts within the tumour stroma have acquired a modified phenotype similar to that of fibroblasts observed in wound healing [12]. Such "activated" fibroblasts named peritumoral fibroblasts, reactive stromal fibroblasts, carcinoma-associated fibroblasts (CAF) or tumor-associated fibroblasts [10, 13] actively control the malignant progression of breast cancers, at least through their capability to secrete MMPs. The present review aims at describing the emerging functions of MMPs which appear more and more as multifunctional enzymes tightly controlling proteolysis both at the cell surface and in the pericellular environment. Using examples of studies performed in animal models of breast cancers, we explore the mechanisms of MMP action with a special emphasis on the contribution of stromal MMPs. Although of great importance, the contribution of MMP in cancer-associated inflammation will not be addressed in this review and reader is referred to previous reviews [14-17].

The MMP family

MMPs are a family of 24 human zinc-binding endopeptidases that can degrade virtually all ECM components, release and activate/inactivate a growing number of modulators of cell functions [6, 7, 15, 16]. MMPs are multidomain proteins characterised by at least three conserved regions: (1) a zinc binding motif (HEXXHXXGXXH) required for proteolytic activity, (2) a propeptide cysteine site (PRCGXPD) whose cysteine residue interacts with the zinc ion in the zymogen form and (3) a "methionine turn" (XXMXP) which likely maintains the zinc-binding site integrity [15]. The activation of these proteases secreted as zymogens requires an amino-terminal cleavage of the pro-domain in the *trans* golgi network by furin-like convertases or extracellularly after their secretion (Fig. 1). The MMP production is precisely regulated at transcriptional and translational

levels [18, 19]. Once switched on, MMP proteolytic activity is under the control of various physiological inhibitors such as tissue inhibitors of metalloproteinases (TIMPs), the plasma inhibitor α 2-macroglobulin and the reversion-inducing cysteine-rich protein with Kazal motifs (RECK) [20-22]. Most of the MMPs are secreted as soluble enzyme but six of them are membrane-type MMPs (MT-MMPs) which are associated with the cell membrane by either a COOH-terminal transmembrane domain (MT1-, MT2-, MT3-, MT5-MMPs) or a glycosylphosphatidyl-inositol (GPI) anchor (MT4-, MT6-MMPs) [23] (Fig. 1). MT1-MMP (MMP-14), one of the most studied MMPs displays pleiotropic functions during both physiological and pathological processes. Although most MMP-knockout mice generated up to now do not present any obvious phenotype without challenging, MT1-MMP-deficiency is associated with growth delay and leads to a lethal phenotype after birth [5, 24, 25]. MT1-MMP activates pro-MMP-2 [26] and pro-MMP-13 [27] and has a very wide range of matrix substrates [6, 23, 28]. Activation of pro-MMP-2 by MT1-MMP requires the tissue inhibitor of metalloproteinases-2 (TIMP-2) which acts as an adaptor molecule mediating pro-MMP-2 binding to MT1-MMP [29, 30].

An increasing number of in vitro studies, mouse models and human clinical studies demonstrate the implication of MMPs in all steps of cancer progression including tumor growth, angiogenesis and metastasis [7, 8, 18]. The increasing diversity in both substrates and functions of MMPs makes them central regulators in different steps of cancer progression and invasion. Now, some MMPs such as MMP-3, -8, -9, -11, -12, -19 and -26 are expected to have dual functions in tumor progression and even in some cases anti-tumor properties [17, 31]. Some of the known substrates of MMPs include ECM components, growth factors, chemokines, cytokines, cell surface proteins and adhesion molecules [6, 7, 17]. Thanks to the development of novel powerful proteomic techniques, a dedicated effort is currently underway to identify the key in vivo substrates of individual MMPs [2, 4, 32] (Fig. 2).

The multiple functions of MMPs in cancer

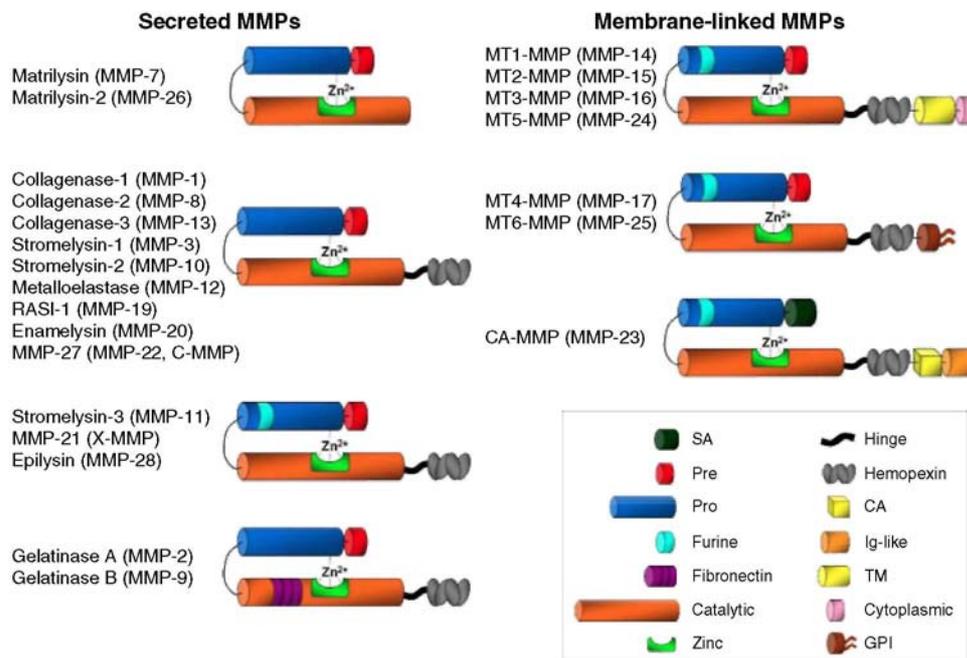
The recent identification of a large panel of matrix and non matrix substrates of MMPs revealed that aside their initial roles as ECM modulators, these proteases can regulate cellular physiology through several mechanisms. In early stages of cancer, the proteolytic processing of bioactive molecules contributes to the elaboration of a permissive microenvironment that promotes malignant transformation and tumor growth. MMP-3 can induce the expression of an alternative spliced form of Rac1 which causes an increase in cellular reactive oxygen species and genomic instability [33]. When bound, growth factors such as Transforming Growth Factor- β (TGF β), insulin-like growth factor (IGF), Fibroblast Growth Factor (FGF) and Heparin Binding Epidermal Growth Factor-like growth factor (HB-EGF) are unable to interact with their receptor and to transduce a signal. Several MMPs control tumor cell proliferation by releasing growth factor bound to specific binding proteins or to matrix components. For instances, bioactive IGF is generated by the action of MMP-3 [34] or MMP-7 [35]. In addition, MMP-7 activates HB-EGF by cleaving its precursors anchored at the cell surface [36]. MT1-MMP confers a proliferative advantage to tumor cells when they are embedded in a 3D collagen-matrix [37]. Opposite effect on cell proliferation can be achieved by the shedding of growth factor receptors such as FGF receptor-1 (FGF-R1) [6, 38]. The cleavage of membrane bound Fas Ligand (mFasL) to soluble FasL (sFasL) by MMP-7 increases apoptosis in normal surrounding cells [39]. However, it permits tumor cells to escape from apoptosis [40, 41] since most cancer cells are relatively resistant to Fas-mediated apoptosis due to abnormalities in the signal transduction cascade [42]. Similarly, MMP-11 inhibits cancer cell death [43].

Loss of E-cadherin-mediated cell-cell adhesion is a prerequisite for tumor cell invasion and metastasis. Proteolytic degradation of E-cadherin by MMP-3 or MMP-7 is one of the mechanisms through which epithelial cell invasion is promoted by disrupting cell aggregation [44]. Proteolysis of E-cadherin and the release of free β catenin play a crucial role in epithelial to mesenchymal transition (EMT), a conversion of epithelial cells to an altered cellular phenotype which is associated with the acquisition of mesenchymal features and aggressive malignant behaviour [45, 46].

MMP-mediated degradation of ECM facilitates angiogenesis, tumor invasion and metastasis [7, 47]. Carcinoma cells were anticipated to produce by themselves proteolytic enzymes in order to degrade basement membrane for invading surrounding tissue. However, it is remarkable that individual tumor cells can cross ECM barriers through non proteolytic processes by exerting physical and mechanical forces that are capable of distorting matrix architecture [48]. Among several MMPs tested, only membrane-associated MMPs (MT1-MMP, MT2-MMP and MT3-MMP) can serve as direct-acting proteases that are able of dissolving BM during cell migration [49]. MT1-MMP is a key enzyme in fibrillar collagen processing and its deletion in mice leads to severe connective tissue defect [25, 50]. Of interest is the recent finding that collective migration of human breast cancer cells and multicellular strand formation is controlled by MT1-MMP through ECM remodelling [51]. Importantly membrane-associated MT-MMPs focus proteolytic activity on specific sites on the cell surface that

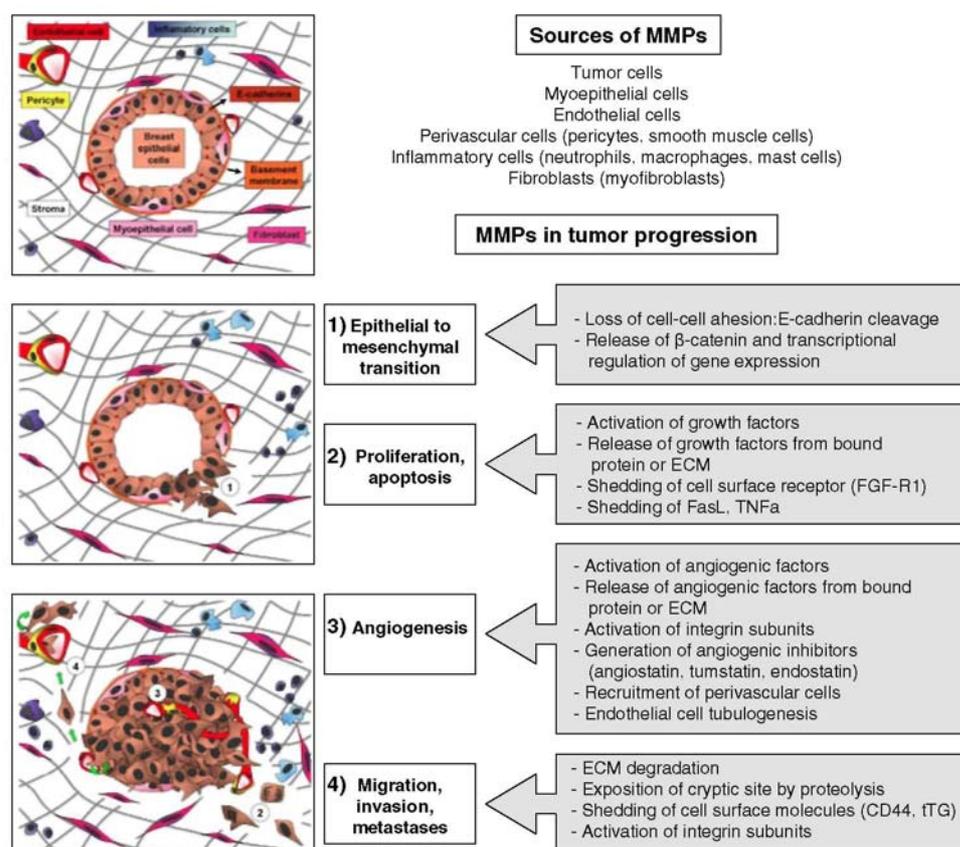
are involved in cell migration [23, 28]. In addition to its fibrinolytic and collagenolytic activities, MT1-MMP stimulates cell motility through the processing of cell adhesion molecules CD44 [52, 53], integrin subunits (pro α -integrin, β 3 subunit) [54, 55] and tissue transglutaminase (tTG) [56]. It is also worth noting that MMP cleavage of ECM components such as laminin 5 or type IV collagen can expose cryptic sites that promote cell migration [57-59].

Fig. 1: Structure of MMPs. *Matrilysins are the minimal-domain MMPs. They contain a signal peptide (Pre) for secretion and a propeptide (Pro) that maintains the enzyme in an inactive form by interacting with the Zinc binding site (Zinc) of the catalytic domain. Collagenases, stromelysins, metalloelastase, Enamelysin and MMP-27 are composed of these minimal domains and a hemopexin-like domain (hemopexin) connected to the catalytic domain with a hinge. The hemopexin domain allows the interaction with substrates and inhibitors. In addition of these domains, gelatinases display fibronectin type II modules (Fibronectin) improving collagen/gelatin degradation, and stromelysin-3, MMP-21, epilysin have a furin-like cleavage site allowing their intracellular activation. Membrane-Type MMPs (MT-MMPs) are linked to the cell membrane with either a transmembrane (TM) domain followed by a short cytoplasmic tail (Cytoplasmic) (MT1-, MT2-, MT3-, MT5-MMPs) or with a glyco-sylphosphatidyl-inositol (GPI) anchor (MT4- and MT6-MMPs). CA-MMP is a type II transmembrane MMP which is characterized by a N-terminal signal anchor (SA) targeting it to the membrane, a unique cysteine array (CA) and immunoglobulin-like (Ig-like) domains in C-terminal*



Several MMPs contribute to angiogenesis through different mechanisms [8, 28, 47, 60, 61]. They include at least the fibrinolytic activity [62], the collagenolytic activity [37], the morphogenesis of endothelial cell (tube formation or tubulogenesis) [63-65], the activation of α v β 3 integrin [66], the transcriptional regulation of Vascular Endothelial Growth Factor (VEGF) [67-69], the release of VEGF sequestered in the ECM [70] or bound to connective tissue growth factor (CTGF) [71], the post-translational processing of VEGF [72], the mural cell investment through a control of PDGF receptor function and the recruitment of perivascular cells contributing to vessel stabilization [8, 73, 74]. The role of MMPs in angiogenesis is dual and complex, some MMPs acting as positive regulators (MMP-1, MMP-2, MMP-9, MT-MMPs) [8, 47, 70, 75, 76] and other as negative regulators (MMP-19) [77] sometimes involved in vessel regression (MMP-10) [61]. Abrogation of angiogenesis can rely on the production of protein fragments endowed with anti-angiogenic activities. For instances, degradation of ECM components (collagen types IV, XVIII) or plasminogen can generate angiogenic inhibitors (tumstatin, endostatin, angiostatin) [78, 79].

Fig. 2: Implication of MMPs in cancer progression. MMPs are implicated in all steps of cancer progression including tumor growth, angiogenesis and metastases via the degradation of extracellular matrix (ECM) components, the release and/or activation of growth factors sequestered in the matrix or complexed to associated proteins, the cleavage of cell surface receptors and the shedding of adhesion molecules. Although not indicated in this schematic representation, MMPs are also key regulators of the inflammatory reaction associated to cancer progression



MMPs in human breast cancers

With the aim of finding new powerful and earlier breast cancer prognostic bio-markers and new targets for cancer treatment, MMPs and MMP inhibitors (MMPIs), respectively, have been extensively investigated in human breast cancer clinical studies [80-82]. MMPs and TIMPs, are frequently overexpressed in human cancer tissues [15]. At least, MMP-1, -2, -9, -11, MT1-MMP, TIMP-1 and -2 levels have been largely investigated in breast cancer tissues by RT-PCR, immunohistochemistry, ELISA, in situ hybridization or zymography analyses (for a review see [80, 81, 83-88]). Despite some conflicting results regarding MMP-9 [89], in most of these studies, the tissue levels of MMPs and TIMPs have been correlated with poor outcome of breast cancer patients [81, 89, 90]. Additionally to their individual level of expression and activity, the ratio of MMP-2/TIMP-2 or MMP-9/TIMP-1, expected to reflect the proteolytic potential, has already been suggested as an early indicator of lymph-node metastases and prognosis [91, 92]. Regarding disease-free survival (DFS) and overall survival (OS) of breast cancer patients, MT1-MMP (MMP-14) mRNA [86, 93] but not protein levels [94, 95], stromal MMP-9 but not tumoral protein expression [95], MMP-2 protein [96] and MMP-7 mRNA expression [97] seem to have an unfavourable prognostic significance. In sharp contrast, MMP-26 has been proposed as a favorable prognostic factor [98]. MMP and TIMP levels in body fluids such as blood and urine of breast cancer patients have been extensively assessed in many pathological processes [99, 100] including breast cancer [80]. Up to now, MMP-2, -7, -9, TIMP-1, -2 concentrations and MMP-2, -9 activities have been analyzed by ELISA and gelatin zymography or immuno-capture assay, respectively, in blood and urine of breast cancers patients [80]. Despite some divergent data, many of these studies have linked circulating MMPs or TIMPs with breast cancer presence, disease status, lymph-node metastasis or other clinicopathological parameters of patients suggesting their potential use in breast cancer screening, follow-up and risk of metastasis establishment. MMP-2 and MMP-9 appear to have clinical value as diagnostic factors for breast cancer or predictive factors of metastases. In addition, proportions between the different forms or between MMPs and their tissue inhibitors (TIMPs), in term of concentration or activity could provide useful clinical information on breast cancer disease and classification [88, 101-104].

Recently, the emphasis has been to reveal the gene expression signatures of primary tumors, which have been associated with their metastatic potential [105-107]. MMP-1 and MMP-9 are involved in the 70 genes identifying the "gene-expression signature" able to predict distant metastasis in lymph-node negative breast cancer patients [106]. Moreover, MMP-1 and MMP-2 have been described as genes that selectively mediate lung metastasis in a mouse model of breast cancer [107] and as members of a lung metastasis gene signature for human breast cancers [108]. Accordingly, MMP-1 has been identified as a useful marker to predict breast cancer development from ductal hyperplasia tissues by global gene expression analysis [109]. These data suggest that, in addition to their prognostic values, MMPs could be used as diagnostic factors to early predict breast lesions that may develop into cancer [80]. Interestingly, these global gene analyses have pinpointed the importance of stroma-derived genes [105, 108] and it is worth noting that peritumoral fibroblasts and inflammatory cells are mainly responsible for the production of tumor-associated MMPs, rather than tumor cells themselves. Peritumoral fibroblasts are the main producers of MMP-1 (interstitial collagenase), MMP-2, MMP-3 (stromelysin-1), MMP-11 (stromelysin-3), MMP-13, MMP-14 in breast cancers [83, 86, 87, 110-112]. The expression of MMP-13 has been co-localized with that of MT1-MMP and MMP-2 suggesting their contribution in a proteolytic cascade [87], [113]. MMP-2 produced by fibroblasts can bind the cell surface of tumor cells through interaction with for instance MT1-MMP and integrin $\alpha_v\beta_3$ [87, 114, 115]. In this context, it is worth noting that in patients with invasive breast carcinomas, mRNA [93, 116-118] and membranous—but not cytoplasmic—protein expression levels of MT1-MMP [95, 119] have been correlated with lymph-node metastasis.

MMPs in experimental models of breast cancer

Several genetically engineered mouse models have been developed to mimic tumor initiation and progression processes of different types of cancer. These models allow a better understanding of cellular and molecular mechanisms underlying cancer progression and can provide useful information for anti-cancer drug development [120]. In breast cancer, these models consist in targeting the expression of oncogenes such as ErbB-2, Ras, Wnt1 or the polyomavirus middle T antigen (PymT) in the mammary epithelium under the control of specific promoters including the mouse mammary tumor virus long terminal repeat (MMTV) and the whey acid protein (WAP) promoters [121]. The availability of these transgenic mice, together with others that are deficient for a specific MMP or that are overexpressing a MMP has been useful in attributing specific functions to individual MMPs in different steps of cancer progression [5, 6, 18]. Breast carcinogenesis can be achieved by crossing the transgenic mice lacking or over-expressing an MMP with mice expressing an oncogene in mammary glands, or by inducing mammary tumors chemically through the oral administration of 7,12-dim-ethylbenzanthracene (DMBA) [122].

Expression of MMP-3 and MT1-MMP in the mammary gland is sufficient to stimulate the development of invasive tumors [123, 124]. MMTV-MMP-3 and WAP-MMP-3 expressing mice display altered spontaneous or DMBA-induced tumor initiation [123, 125, 126]. Moreover, MMTV-MMP-7 expressing mice develop pre-malignant nodules and increased oncogene-induced (MMTV-ErbB-2) mammary tumors. In contrast, mice lacking MMP-7 expression with a mutated *Apc* allele show a transient reduction of mammary tumors [127, 128]. The transgenic deficiency in MMP-2 or MMP-9 expression, the transgenic expression of TIMP-1 or -2 (MMTV-TIMP) and the treatment with a MMPI are also reported to affect mammary tumorigenesis and lung metastases induced in MMTV-PymT or MMTV-Wnt1 models [122]. Altogether, these data implicate MMPs and their inhibitors in mammary tumor development. However, the situation is rendered even more complex by the fact that some MMPs appear to function as dual modulators of tumor progression. Indeed, MMP-11-deficient mice show a decreased DMBA-induced mammary carcinogenesis [129] and a decrease of tumor incidence/tumor growth [130]. However, MMP-11^{-/-}/MMTV-ras mice develop more lung metastases than their wild type counterpart [130]. Therefore, MMP-11 function differs throughout cancer progression, it is an enhancer for primary tumor development, but a repressor for metastatic dissemination.

Xenografts of human cancer cells transfected with one or other MMP cDNA is extensively used to investigate the behaviour of human cells in an in vivo environment. Indeed, different studies in which immunodeficient mice are injected with breast cancer cells over/down-expressing MMPs or TIMPs, demonstrate their implication in breast cancer progression and especially in development of metastases. Although most MMPs including for instance MMP-2 [131], MMP-11 [132, 133], MMP-3 [123], MT1-MMP [67, 134] and MT4-MMP [135] are generally positive regulators of cancer progression (tumor promoters), some of them such as MMP-8 [136] negatively regulate metastasis in breast cancer models. Similarly, as mentioned above, MMP-11 represses metastatic dissemination, while it enhances primary tumor development [130]. These opposite tumor/metastasis-promoting effects of different MMPs or of the same MMP at different stages of cancer progression is one of the explanations why clinical trials of broad spectrum MMP inhibitors have failed, underlining the importance to

develop more specific inhibitors of MMPs.

Since fibroblasts constitute the majority of stromal cells within a breast carcinoma and since they are a primary source of MMP, a co-implantation tumor xenograft model has been used to investigate the interplay between fibroblasts and breast carcinoma cells [10, 137]. The tumor promoting effect of fibroblasts in xenografts is blocked by TIMP2 or synthetic MMP inhibitor [137, 138]. Interestingly, MMP-11-null fibroblasts [129] or MT1-MMP-null fibroblasts [139] do not support *in vivo* growth of tumor cells whereas corresponding wild-type fibroblasts enhance tumor development. MMP-11 is a stromal factor which promotes the primary implantation of cancer cells in an aberrant environment [110]. In MMP-11-deficient mice, the number of apoptotic cancer cells is increased in primary tumors, indicating that host MMP-11 helps tumor cells in escaping apoptosis [43].

Cancer cells can stimulate fibroblasts to synthesize MMPs in a paracrine manner through the secretion of interleukins, interferons, growth factors and Extracellular Matrix Metalloproteinase Inducer (EMMPRN) [19, 140, 141]. The pathologic consequence of elevated EMMPRN is supported by the accelerated growth and increased invasiveness exhibited by breast cancer cells overexpressing EMMPRN [23, 142]. Interestingly, carcinoma-associated fibroblasts (CAF)s extracted from human breast carcinomas are better promoters of human breast adenocarcinoma cell growth in xenograft than normal primary fibroblasts derived from the same patient [143]. It is worth noting that the upregulation of MMPs is one of the physiological changes that occur when fibroblasts undergo senescence. This likely promotes the generation of a pro-oncogenic microenvironment that contributes to the increased incidence of cancers observed with age [144, 145]. Accordingly, fibroblasts that have been forced into senescence by DNA damage increased the growth of cancer cells in a MMP-dependent manner [145]. The tumor microenvironment can be a potent carcinogen, not only by facilitating cancer progression, but also by stimulating tumor formation. A stromal enzyme such as MMP-3 can cause sustained EMT and malignant transformation in cultured cells and genomically unstable mammary carcinomas in transgenic mice [33, 123].

Conclusions

Based on the fact that MMPs were initially viewed as invasion-associated proteases, preclinical studies of MMP inhibition were performed in different mouse cancer models. The success of these studies led to the rapid development of synthetic MMP inhibitors (MMPIs) and their assessment in clinical trials. However, the results obtained in phase III trials were disappointing with many adverse side effects [15, 18, 140, 146, 147]. The failure of MMP inhibition in cancer therapy is now better understood [146, 148]. One explanation, among others, is that clinical trials have been performed in advanced stages of cancers whereas MMPs are more expected to play crucial role in early steps of cancer progression. In addition, broad spectrum MMPI block the activity of all metalloproteases (including ADAMs and ADAMTS) and it is now well known that different MMPs can have opposite effects or different effects at different stage of cancer progression. Therefore, some MMPs are viewed as "drug targets", while others are considered as "anti-targets" for cancer therapy [17]. The initial concept of MMPs as simple modulators of ECM remodeling has been replaced by the consideration of MMPs as multifaceted proteases able to tightly control the bioavailability and activity of a large panel of proteins. In addition, it is possible that substrates and products of MMPs could be preferred as targets for treating cancer rather than MMPs themselves. However, such strategies depend on better knowledge on how individual MMPs are contributing to tumor growth and metastatic dissemination. In this context, the complementarity between human clinical studies and mouse models is of great importance.

Acknowledgements:

This work was supported by grants from the European Union Framework Programme 6 projects, the Fonds National de la Recherche Scientifique (F.N.R.S., Belgium), the Fédération contre le Cancer, the D.G.T.R.E. («Région Wallonne», Belgium) and the Interuniversity Attraction Poles Programme—Belgian Science Policy (Belgium).

References

1. Hanahan D, Weinberg RA (2000) The hallmarks of cancer. *Cell* 100:57-70
2. Lopez-Otin C, Overall CM (2002) Protease degradomics: a new challenge for proteomics. *Nat Rev Mol Cell Biol* 3:509-519

3. Overall CM, Tam EM, Kappelhoff R et al (2004) Protease degradomics: mass spectrometry discovery of protease substrates and the CLIP-CHIP, a dedicated DNA microarray of all human proteases and inhibitors. *Biol Chem* 385:493-504
4. Greenlee KJ, Corry DB, Engler DA et al (2006) Proteomic identification of in vivo substrates for matrix metalloproteinases 2 and 9 reveals a mechanism for resolution of inflammation. *J Immunol* 177:7312-7321
5. Page-McCaw A, Ewald AT, Werb Z (2007) Matrix metalloproteinases and the regulation of tissue remodelling. *Nat Rev Mol Cell Biol* 8:221-233
6. Cauwe B, Van den Steen PE, Opdenakker G (2007) The biochemical, biological, and pathological kaleidoscope of cell surface substrates processed by matrix metalloproteinases. *Crit Rev Biochem Mol Biol* 42:113-185
7. Overall CM, Dean RA (2006) Degradomics: systems biology of the protease web. Pleiotropic roles of MMPs in cancer. *Cancer Metastasis Rev* 25:69-75
8. Noel A, Jost M, Maquoi E (2007) Matrix metalloproteinases at cancer tumor-host interface. *Semin Cell Dev Biol*. doi:10.1016/j.semcdb.2007.05.011
9. Noel A, Foidart JM (1998) The role of extracellular matrix and fibroblasts in breast carcinoma growth in vivo. *J Mammary Gland Biol Neoplasia* 3:215-225
10. Kalluri R, Zeisberg M (2006) Fibroblasts in cancer. *Nat Rev Cancer* 6:392-401
11. Shekhar MPV, Pauley R, Heppner G (2003) Host microenvironment in breast cancer development—extracellular matrix-stromal cell contribution to neoplastic phenotype of epithelial cells in the breast. *Breast Cancer Res* 5:130-135
12. Dvorak HF (1986) Tumors: wounds that do not heal. Similarities between tumor stroma generation and wound healing. *N Engl J Med* 315:1650-1659
13. Mueller MM, Fusenig NE (2004) Friends or foes—bipolar effects of the tumour stroma in cancer. *Nat Rev Cancer* 4:839-849
14. Balkwill F, Coussens LM (2004) Cancer: an inflammatory link. *Nature* 431:405-406
15. Egeblad M, Werb Z (2002) New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2:161-174
16. Greenlee KJ, Werb Z, Kheradmand F (2007) Matrix metalloproteinases in lung: multiple, multifarious, and multifaceted. *Physiol Rev* 87:69-98
17. Overall CM, Kleinfeld O (2006) Tumour microenvironment— opinion: validating matrix metalloproteinases as drug targets and anti-targets for cancer therapy. *Nat Rev Cancer* 6:227-239
18. Folgueras AR, Pendas AM, Sanchez LM, Lopez-Otin C (2004) Matrix metalloproteinases in cancer: from new functions to improved inhibition strategies. *Int J Dev Biol* 48:411-424
19. Sternlicht MD, Werb Z (2001) How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol* 17:463-516
20. Oh J, Takahashi R, Kondo S et al (2001) The membrane-anchored MMP inhibitor RECK is a key regulator of extracellular matrix integrity and angiogenesis. *Cell* 107:789-800
21. Baker AH, Edwards DR, Murphy G (2002) Metalloproteinase inhibitors: biological actions and therapeutic opportunities. *J Cell Sci* 115:3719-3727
22. Visse R, Nagase H (2003) Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. *Circ Res* 92:827-839
23. Zucker S, Pei D, Cao J, Lopez-Otin C (2003) Membrane type-matrix metalloproteinases (MT-MMP). *Curr Top Dev Biol* 54:1-74
24. Zhou Z, Apte SS, Soininen R et al (2000) Impaired endochondral ossification and angiogenesis in mice deficient in membrane-type matrix metalloproteinase I. *Proc Natl Acad Sci USA* 97:4052-4057
25. Holmbeck K, Bianco P, Caterina J et al (1999) MT1-MMP-deficient mice develop dwarfism, osteopenia, arthritis, and connective tissue disease due to inadequate collagen turnover. *Cell* 99:81-92
26. Sato H, Takino T, Okada Y et al (1994) A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature* 370:61-65
27. Knauper V, Will H, Lopez-Otin C et al (1996) Cellular mechanisms for human procollagenase-3 (MMP-13) activation. Evidence that MT1-MMP (MMP-14) and gelatinase a (MMP-2) are able to generate active enzyme. *J Biol Chem* 271:17124-17131

28. Sounni NE, Noel A (2005) Membrane type-matrix metalloproteinases and tumor progression. *Biochimie* 87:329-342
29. Sounni NE, Janssen M, Foidart JM, Noel A (2003) Membrane type-1 matrix metalloproteinase and TIMP-2 in tumor angiogenesis. *Matrix Biol* 22:55-61
30. Seiki M, Koshikawa N, Yana I (2003) Role of pericellular proteolysis by membrane-type 1 matrix metalloproteinase in cancer invasion and angiogenesis. *Cancer Metastasis Rev* 22:129-143
31. Lopez-Otin C, Matrisian LM (2007) Emerging roles of proteases in tumour suppression. *Nat Rev Cancer* 7:800-808
32. Overall CM, McQuibban GA, Clark-Lewis I (2002) Discovery of chemokine substrates for matrix metalloproteinases by exo-site scanning: a new tool for degradomics. *Biol Chem* 383:1059-1066
33. Radisky DC, Levy DD, Littlepage LE et al (2005) Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability. *Nature* 436:123-127
34. Manes S, Mira E, Barbacid MD et al (1997) Identification of insulin-like growth factor-binding protein-1 as a potential physiological substrate for human stromelysin-3. *J Biol Chem* 272:25706-25712
35. Miyamoto S, Nakamura M, Yano K et al (2007) Matrix metalloproteinase-7 triggers the matricrine action of insulin-like growth factor-II via proteinase activity on insulin-like growth factor binding protein 2 in the extracellular matrix. *Cancer Sci* 98:685-691
36. Yu Q, Stamenkovic I (1999) Localization of matrix metalloproteinase 9 to the cell surface provides a mechanism for CD44-mediated tumor invasion. *Genes Dev* 13:35-48
37. Hotary KB, Allen ED, Brooks PC, Datta NS, Long MW, Weiss SJ (2003) Membrane type I matrix metalloproteinase usurps tumor growth control imposed by the three-dimensional extracellular matrix. *Cell* 114:33-45
38. Powers CJ, McLeskey SW, Wellstein A (2000) Fibroblast growth factors, their receptors and signaling. *Endocr Relat Cancer* 7:165-197
39. Powell WC, Fingleton B, Wilson CL, Boothby M, Matrisian LM (1999) The metalloproteinase matrilysin proteolytically generates active soluble Fas ligand and potentiates epithelial cell apoptosis. *Curr Biol* 9:1441-1447
40. Mitsiades N, Yu WH, Poulaki V, Tsokos M, Stamenkovic I (2001) Matrix metalloproteinase-7-mediated cleavage of Fas ligand protects tumor cells from chemotherapeutic drug cytotoxicity. *Cancer Res* 61:577-581
41. Ii M, Yamamoto H, Adachi Y, Maruyama Y, Shinomura Y (2006) Role of matrix metalloproteinase-7 (matrilysin) in human cancer invasion, apoptosis, growth, and angiogenesis. *Exp Biol Med* 231:20-27
42. O'Connell J, Bennett MW, O'Sullivan GC, Collins JK, Shana-han F (1999) The Fas counterattack: cancer as a site of immune privilege. *Immunol Today* 20:46-52
43. Boulay A, Masson R, Chenard MP et al (2001) High cancer cell death in syngeneic tumors developed in host mice deficient for the stromelysin-3 matrix metalloproteinase. *Cancer Res* 61:2189-2193
44. Noe V, Fingleton B, Jacobs K et al (2001) Release of an invasion promoter E-cadherin fragment by matrilysin and stromelysin-1. *J Cell Sci* 114:111-118
45. Gilles C, Newgreen DF, Sato H, Thompson EW (2004) Matrix metalloproteinases and epithelial-to-mesenchymal transition: implications for carcinoma metastasis. In: Savagner P (ed) *Rise and fall of epithelial phenotype*. Kluwer Academic/Plenum Publishers, pp 233-251
46. Christofori G (2007) Cancer—Division of labour. *Nature* 446:735-736
47. van Hinsbergh VW, Engelse MA, Quax PH (2006) Pericellular proteases in angiogenesis and vasculogenesis. *Arterioscler Thromb Vasc Biol* 26:716-728
48. Wolf K, Muller R, Borgmann S, Brocker EB, Friedl P (2003) Amoeboid shape change and contact guidance: T-lymphocyte crawling through fibrillar collagen is independent of matrix remodeling by MMPs and other proteases. *Blood* 102:3262-3269
49. Hotary K, Li XY, Allen E, Stevens SL, Weiss SJ (2006) A cancer cell metalloprotease triad regulates the basement membrane transmigration program. *Genes Dev* 20:2673-2686
50. Holmbeck K, Bianco P, Yamada S, Birkedal-Hansen H (2004) MT1-MMP: a tethered collagenase. *J Cell Physiol* 200:11-19
51. Wolf C, Wu YI, Liu Y et al (2007) Multi-step pericellular proteolysis controls the transition from individual to collective cancer cell invasion. *Nature Cell Biol* 9:893-904
52. Kajita M, Itoh Y, Chiba T et al (2001) Membrane-type 1 matrix metalloproteinase cleaves CD44 and promotes cell migration. *J Cell*

Biol 153:893-904

53. Suenaga N, Mori H, Itoh Y, Seiki M (2005) CD44 binding through the hemopexin-like domain is critical for its shedding by membrane-type 1 matrix metalloproteinase. *Oncogene* 24:859-868
54. Deryugina EI, Bourdon MA, Jungwirth K, Smith JW, Strongin AY (2000) Functional activation of integrin alpha V beta 3 in tumor cells expressing membrane-type 1 matrix metalloproteinase. *Int J Cancer* 86:15-23
55. Deryugina EI, Ratnikov BI, Postnova TI, Rozanov DV, Strongin AY (2002) Processing of integrin alpha(v) subunit by membrane type 1 matrix metalloproteinase stimulates migration of breast carcinoma cells on vitronectin and enhances tyrosine phosphorylation of focal adhesion kinase. *J Biol Chem* 277:9749-9756
56. Belkin AM, Akimov SS, Zaritskaya LS, Ratnikov BI, Deryugina EI, Strongin AY (2001) Matrix-dependent proteolysis of surface transglutaminase by membrane-type metalloproteinase regulates cancer cell adhesion and locomotion. *J Biol Chem* 276:18415-18422
57. Gilles C, Polette M, Coraux C et al (2001) Contribution of MT1-MMP and of human laminin-5 gamma2 chain degradation to mammary epithelial cell migration. *J Cell Sci* 114:2967-2976
58. Giannelli G, Falk-Marzillier J, Schiraldi O, Stetler-Stevenson WG, Quaranta V (1997) Induction of cell migration by matrix metalloprotease-2 cleavage of laminin-5. *Science* 277:225-228
59. Xu J, Rodriguez D, Petitclerc E et al (2001) Proteolytic exposure of a cryptic site within collagen type IV is required for angiogenesis and tumor growth in vivo. *J Cell Biol* 154:1069-1079
60. Handsley MM, Edwards DR (2005) Metalloproteinases and their inhibitors in tumor angiogenesis. *Int J Cancer* 115:849-860
61. Davis GE, Saunders WB (2006) Molecular balance of capillary tube formation versus regression in wound repair: role of matrix metalloproteinases and their inhibitors. *J Invest Dermatol* 11:44-56
62. Hotary KB, Yana I, Sabeh F et al (2002) Matrix metalloproteinases (MMPs) regulate fibrin-invasive activity via MT1-MMP-dependent and -independent processes. *J Exp Med* 195:295-308
63. Hotary K, Allen E, Punturieri A, Yana I, Weiss SJ (2000) Regulation of cell invasion and morphogenesis in a three-dimensional type I collagen matrix by membrane-type matrix metalloproteinases 1, 2, and 3. *J Cell Biol* 149:1309-1323
64. Lafleur MA, Handsley MM, Knauper V, Murphy G, Edwards DR (2002) Endothelial tubulogenesis within fibrin gels specifically requires the activity of membrane-type-matrix metalloproteinases (MT-MMPs). *J Cell Sci* 115:3427-3438
65. Plaisier M, Kapiteijn K, Koolwijk P et al (2004) Involvement of membrane-type matrix metalloproteinases (MT-MMPs) in capillary tube formation by human endometrial microvascular endothelial cells: role of MT3-MMP. *J Clin Endocrinol Metab* 89:5828-5836
66. Deryugina EI, Ratnikov B, Monosov E et al (2001) MT1-MMP initiates activation of pro-MMP-2 and integrin alphavbeta3 promotes maturation of MMP-2 in breast carcinoma cells. *Exp Cell Res* 263:209-223
67. Sounni NE, Devy L, Hajitou A et al (2002) MT1-MMP expression promotes tumor growth and angiogenesis through an up-regulation of vascular endothelial growth factor expression. *FASEB J* 16:555-564
68. Deryugina EI, Soroceanu L, Strongin AY (2002) Up-regulation of vascular endothelial growth factor by membrane-type 1 matrix metalloproteinase stimulates human glioma xenograft growth and angiogenesis. *Cancer Res* 62:580-588
69. Noel A, Maillard C, Rocks N et al (2004) Membrane associated proteases and their inhibitors in tumour angiogenesis. *J Clin Pathol* 57:577-584
70. Bergers G, Brekken R, McMahon G et al (2000) Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol* 2:737-744
71. Hashimoto G, Inoki I, Fujii Y, Aoki T, Ikeda E, Okada Y (2002) Matrix metalloproteinases cleave connective tissue growth factor and reactivate angiogenic activity of vascular endothelial growth factor 165. *J Biol Chem* 277:36288-36295
72. Lee S, Jilani SM, Nikolova GV, Carpizo D, Iruela-Arispe ML (2005) Processing of VEGF-A by matrix metalloproteinases regulates bioavailability and vascular patterning in tumors. *J Cell Biol* 169:681-691
73. Chantraine CF, Shimada H, Jodele S et al (2004) Stromal matrix metalloproteinase-9 regulates the vascular architecture in neuroblastoma by promoting pericyte recruitment. *Cancer Res* 64:1675-1686
74. Jodele S, Chantraine CF, Blavier L et al (2005) The contribution of bone marrow-derived cells to the tumor vasculature in neuroblastoma is matrix metalloproteinase-9 dependent. *Cancer Res* 65:3200-3208
75. Itoh T, Tanioka M, Yoshida H, Yoshioka T, Nishimoto H, Ito-hara S (1998) Reduced angiogenesis and tumor progression in gelatinase

A-deficient mice. *Cancer Res* 58:1048-1051

76. Masson V, de la Ballina LR, Munaut C et al (2005) Contribution of host MMP-2 and MMP-9 to promote tumor vascularization and invasion of malignant keratinocytes. *FASEB J* 18:234-236

77. Jost M, Folgueras AR, Frerart F et al (2006) Earlier onset of tumoral angiogenesis in matrix metalloproteinase-19-deficient mice. *Cancer Res* 66:5234-5241

78. Nyberg P, Xie L, Kalluri R (2005) Endogenous inhibitors of angiogenesis. *Cancer Res* 65:3967-3979

79. Hamano Y, Kalluri R (2005) Tumstatin, the NCI domain of alpha 3 chain of type IV collagen, is an endogenous inhibitor of pathological angiogenesis and suppresses tumor growth. *Bio-chem Biophys Res Commun* 333:292-298

80. Chabottaux V, Noel A (2007) Matrix metalloproteinases to predict breast cancer metastases. *Clin Lab Int* 31:8-10. <http://www.clin-online.com>

81. Duffy MJ, Maguire TM, Hill A, McDermott E, O'Higgins N (2000) Metalloproteinases: role in breast carcinogenesis, invasion and metastasis. *Breast Cancer Res* 2:252-257

82. Martin M, Matrisian L (2004) Matrix metalloproteinases as prognostic factors for cancer. *Clin Lab Int* 28:16-18. <http://www.clin-online.com>

83. Basset P, Bellocq JP, Wolf C et al (1990) A novel metalloproteinase gene specifically expressed in stromal cells of breast carcinomas. *Nature* 348:699-704

84. Polette M, Gilbert N, Stas I et al (1994) Gelatinase A expression and localization in human breast cancers. An in situ hybridization study and immunohistochemical detection using confocal microscopy. *Virchows Arch* 424:641-645

85. Remacle A, McCarthy K, Noel A et al (2000) High levels of TIMP-2 correlate with adverse prognosis in breast cancer. *Int J Cancer* 89:118-121

86. Tetu B, Brisson J, Wang CS et al (2006) The influence of MMP-14, TIMP-2 and MMP-2 expression on breast cancer prognosis. *Breast Cancer Res* 8:R28

87. Bisson C, Blacher S, Polette M et al (2003) Restricted expression of membrane type 1-matrix metalloproteinase by myofibroblasts adjacent to human breast cancer cells. *Int J Cancer* 105:7-13

88. Jinga D, Stefanescu M, Blidaru A, Condrea I, Pistol G, Matache C (2004) Serum levels of matrix metalloproteinases MMP-2 and MMP-9 and their tissue natural inhibitors in breast tumors. *Roum Arch Microbiol Immunol* 63:141-158

89. Turpeenniemi-Hujanen T (2005) Gelatinases (MMP-2 and -9) and their natural inhibitors as prognostic indicators in solid cancers. *Biochimie* 87:287-297

90. Matrisian LM (1990) Metalloproteinases and their inhibitors in matrix remodeling. *Trends Genet* 6:121-125

91. Onisto M, Riccio MP, Scannapieco P et al (1995) Gelatinase A/ TIMP-2 imbalance in lymph-node-positive breast carcinomas, as measured by RT-PCR. *Int J Cancer* 63:621-626

92. Jinga DC, Blidaru A, Condrea I et al (2006) MMP-9 and MMP-2 gelatinases and TIMP-1 and TIMP-2 inhibitors in breast cancer: correlations with prognostic factors. *J Cell Mol Med* 10:499-510

93. Jiang WG, Davies G, Martin TA et al (2006) Expression of membrane type-1 matrix metalloproteinase, MT1-MMP in human breast cancer and its impact on invasiveness of breast cancer cells. *Int J Mol Med* 17:583-590

94. Ishigaki S, Toi M, Ueno T et al (1999) Significance of membrane type 1 matrix metalloproteinase expression in breast cancer. *Jpn J Cancer Res* 90:516-522

95. Mylona E, Nomikos A, Magkou C et al (2007) The clinico-pathological and prognostic significance of membrane type 1 matrix metalloproteinase (MT1-MMP) and MMP-9 according to their localization in invasive breast carcinoma. *Histopathology* 50:338-347

96. Talvensaari-Mattila A, Paakko P, Turpeenniemi-Hujanen T (2003) Matrix metalloproteinase-2 (MMP-2) is associated with survival in breast carcinoma. *Br J Cancer* 89:1270-1275

97. Jiang WG, Davies G, Martin TA et al (2005) Targeting matrix-lysin and its impact on tumor growth in vivo: the potential implications in breast cancer therapy. *Clin Cancer Res* 11:6012-6019

98. Savinov AY, Remacle AG, Golubkov VS et al (2006) Matrix metalloproteinase 26 proteolysis of the NH2-terminal domain of the estrogen receptor beta correlates with the survival of breast cancer patients. *Cancer Res* 66:2716-2724

99. Zucker S, Hymowitz M, Conner C et al (1999) Measurement of matrix metalloproteinases and tissue inhibitors of metalloproteinases in blood and tissues. Clinical and experimental applications. *Ann N Y Acad Sci* 878:212-227
100. Zucker S, Doshi K, Cao J (2004) Measurement of matrix metalloproteinases (MMPs) and tissue inhibitors of metalloproteinases (TIMP) in blood and urine: potential clinical applications. *Adv Clin Chem* 38:37-85
101. Giannelli G, Erriquez R, Fransvea E et al (2004) Proteolytic imbalance is reversed after therapeutic surgery in breast cancer patients. *Int J Cancer* 109:782-785
102. Kuvaja P, Talvensaari-Mattila A, Paakko P, Turpeenniemi-Hujanen T (2006) Low serum level of pro-matrix metalloproteinase 2 correlates with aggressive behavior in breast carcinoma. *Hum Pathol* 37:1316-1323
103. Somiari SB, Somiari RI, Heckman CM et al (2006) Circulating MMP2 and MMP9 in breast cancer—potential role in classification of patients into low risk, high risk, benign disease and breast cancer categories. *Int J Cancer* 119:1403-1411
104. Somiari SB, Shriver CD, Heckman C et al (2006) Plasma concentration and activity of matrix metalloproteinase 2 and 9 in patients with breast disease, breast cancer and at risk of developing breast cancer. *Cancer Lett* 233:98-107
105. Ramaswamy S, Ross KN, Lander ES, Golub TR (2003) A molecular signature of metastasis in primary solid tumors. *Nat Genet* 33:49-54
106. van't Veer LJ, Dai HY, van de Vijver MJ et al (2002) Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 415:530-536
107. Minn AJ, Gupta GP, Siegel PM et al (2005) Genes that mediate breast cancer metastasis to lung. *Nature* 436:518-524
108. Gupta GP, Nguyen DX, Chiang AC et al (2007) Mediators of vascular remodelling co-opted for sequential steps in lung metastasis. *Nature* 446:765-770
109. Poola I, DeWitty RL, Marshalleck JJ, Bhatnagar R, Abraham J, Leffall LD (2005) Identification of MMP-1 as a putative breast cancer predictive marker by global gene expression analysis. *Nat Med* 11:481-483
110. Rio MC (2005) From a unique cell to metastasis is a long way to go: clues to stromelysin-3 participation. *Biochimie* 87:299-306
111. Okada A, Bellocq JP, Rouyer N et al (1995) Membrane-type matrix metalloproteinase (MT-MMP) gene is expressed in stromal cells of human colon, breast, and head and neck carcinomas. *Proc Natl Acad Sci USA* 92:2730-2734
112. Polette M, Nawrocki B, Gilles C et al (1996) MT-MMP expression and localisation in human lung and breast cancers. *Virchows Arch* 428:29-35
113. Ala-Aho R, Kahari VM (2005) Collagenases in cancer. *Biochimie* 87:273-286
114. Polette M, Birembaut P (1998) Membrane-type metalloproteinases in tumor invasion. *Int J Biochem Cell Biol* 30:1195-1202
115. Brooks PC, Stromblad S, Sanders LC et al (1996) Localization of matrix metalloproteinase MMP-2 to the surface of invasive cells by interaction with integrin alpha v beta 3. *Cell* 85:683-693
116. Ueno H, Nakamura H, Inoue M et al (1997) Expression and tissue localization of membrane-types 1, 2, and 3 matrix metalloproteinases in human invasive breast carcinomas. *Cancer Res* 57:2055-2060
117. Mimori K, Ueo H, Shirasaka C, Mori M (2001) Clinical significance of MT1-MMP mRNA expression in breast cancer. *Oncol Rep* 8:401-403
118. Yao GY, Yang MT, Rong TH, He P (2004) Significance of membrane type-1 matrix metalloproteinase expression in breast cancer. *Ai Zheng* 23:1482-1486
119. Jones JL, Glynn P, Walker RA (1999) Expression of MMP-2 and MMP-9, their inhibitors, and the activator MT1-MMP in primary breast carcinomas. *J Pathol* 189:161-168
120. Singh M, Johnson L (2006) Using genetically engineered mouse models of cancer to aid drug development: an industry perspective. *Clin Cancer Res* 12:5312-5328
121. Shen Q, Brown PH (2005) Transgenic mouse models for the prevention of breast cancer. *Mutat Res Fundam Mol Mech Mutagen* 576:93-110
122. Almholt K, Green KA, Juncker-Jensen A, Nielsen BS, Lund LR, Romer J (2007) Extracellular proteolysis in transgenic mouse models of breast cancer. *J Mammary Gland Biol Neoplasia* 12:83-97

123. Sternlicht MD, Lochter A, Sympon CJ et al (1999) The stromal proteinase MMP3/stromelysin-1 promotes mammary carcinogenesis. *Cell* 98:137-146
124. Ha HY, Moon HB, Nam MS et al (2001) Overexpression of membrane-type matrix metalloproteinase-1 gene induces mammary gland abnormalities and adenocarcinoma in transgenic mice. *Cancer Res* 61:984-990
125. Witty JP, Lempka T, Coffey RJ, Matrisian LM (1995) Decreased tumor-formation in 7,12-dimethylbenzanthracene-treated stromelysin-1 transgenic mice is associated with alterations in mammary epithelial-cell apoptosis. *Cancer Res* 55: 1401-1406
126. Witty EP, Wright JH, Matrisian LM (1995) Matrix metallo-proteinases are expressed during ductal and alveolar mammary morphogenesis, and misregulation of stromelysin-1 in transgenic mice induces unscheduled alveolar development. *Mol Biol Cell* 6:1287-1303
127. Rudolph-Owen LA, Chan R, Muller WJ, Matrisian LM (1998) The matrix metalloproteinase matrilysin influences early-stage mammary tumorigenesis. *Cancer Res* 58:5500-5506
128. Hulboy DL, Gautam S, Fingleton B, Matrisian LM (2004) The influence of matrix metalloproteinase-7 on early mammary tumorigenesis in the multiple intestinal neoplasia mouse. *Oncol Rep* 12:13-17
129. Masson R, Lefebvre O, Noel A et al (1998) In vivo evidence that the stromelysin-3 metalloproteinase contributes in a paracrine manner to epithelial cell malignancy. *J Cell Biol* 140:1535-1541
130. Andarawewa KL, Boulay A, Masson W et al (2003) Dual stromelysin-3 function during natural mouse mammary tumor virus-ras tumor progression. *Cancer Res* 63:5844-5849
131. Tester AM, Waltham M, Oh SJ et al (2004) Pro-matrix metalloproteinase-2 transfection increases orthotopic primary growth and experimental metastasis of MDA-MB-231 human breast cancer cells in nude mice. *Cancer Res* 64:652-658
132. Noel AC, Lefebvre O, Maquoi E et al (1996) Stromelysin-3 expression promotes tumor take in nude mice. *J Clin Invest* 97: 1924-1930
133. Noel A, Boulay A, Kebers F et al (2000) Demonstration in vivo that stromelysin-3 functions through its proteolytic activity. *Oncogene* 19:1605-1612
134. Sounni NE, Roghi C, Chabottaux V et al (2004) Up-regulation of vascular endothelial growth factor-A by active membrane-type 1 matrix metalloproteinase through activation of Src-tyrosine kinases. *J Biol Chem* 279:13564-13574
135. Chabottaux V, Sounni NE, Pennington CJ et al (2006) Membrane-type 4 matrix metalloproteinase promotes breast cancer growth and metastases. *Cancer Res* 66:5165-5172
136. Montel V, Kleeman J, Agarwal D, Spinella D, Kawai K, Tarin D (2004) Altered metastatic behavior of human breast cancer cells after experimental manipulation of matrix metalloproteinase 8 gene expression. *Cancer Res* 64:1687-1694
137. Noel A, Hajitou A, L'Hoir C et al (1998) Inhibition of stromal matrix metalloproteases: Effects on breast-tumor promotion by fibroblasts. *Int J Cancer* 76:267-273
138. Maquoi E, Sounni NE, Devy L et al (2004) Anti-invasive, antitumoral, and antiangiogenic efficacy of a pyrimidine-2,4,6-trione derivative, an orally active and selective matrix metallo-proteinases inhibitor. *Clin Cancer Res* 10:4038-4047
139. Zhang WY, Matrisian LM, Holmbeck K, Vick CC, Rosenthal EL (2006) Fibroblast-derived MT1-MMP promotes tumor progression in vitro and in vivo. *Bmc Cancer* 6:52
140. Overall CM, Lopez-Otin C (2002) Strategies for MMP inhibition in cancer: innovations for the post-trial era. *Nat Rev Cancer* 2:657-672
141. Zigrino P, Loffek S, Mauch C (2005) Tumor-stroma interactions: their role in the control of tumor cell invasion. *Biochimie* 87:321-328
142. Foda HD, Zucker S (2001) Matrix metalloproteinases in cancer invasion, metastasis and angiogenesis. *Drug Discov Today* 6:478-482
143. Orimo A, Gupta PB, Sgroi DC et al (2005) Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. *Cell* 121:335-348
144. Bavik C, Coleman I, Dean JP, Knudsen B, Plymate S, Nelson PS (2006) The gene expression program of prostate fibroblast senescence modulates neoplastic epithelial cell proliferation through paracrine mechanisms. *Cancer Res* 66:794—802
145. Liu D, Hornsby PJ (2007) Senescent human fibroblasts increase the early growth of xenograft tumors via matrix metalloproteinase secretion. *Cancer Res* 67:3117-3126
146. Coussens LM, Fingleton B, Matrisian LM (2002) Matrix metalloproteinase inhibitors and cancer: trials and tribulations. *Science* 295:2387-2392

147. Zucker S, Cao J, Chen WT (2000) Critical appraisal of the use of matrix metalloproteinase inhibitors in cancer treatment. *Oncogene* 19:6642-6650
148. Overall CM, Kleinfeld O (2006) Towards third generation matrix metalloproteinase inhibitors for cancer therapy. *Brit J Cancer* 94:941-946