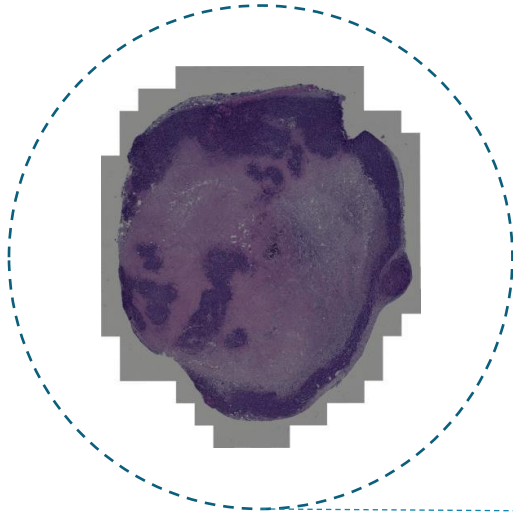


Linking spatial to functional proteomics on heterogeneous samples: The “pixel-by-pixel” shotgun proteomic approach

Matthieu HODEIGE, Christopher KUNE, Maximilien FLERON, Dominique BAIWIR, Nor Eddine SOUNNI, Gauthier EPPE,
Laurent GATTO, Gabriel MAZZUCHELLI

Belgian Proteomics Association Conference, Ghent, 05/12/24



Context and Biological Challenges

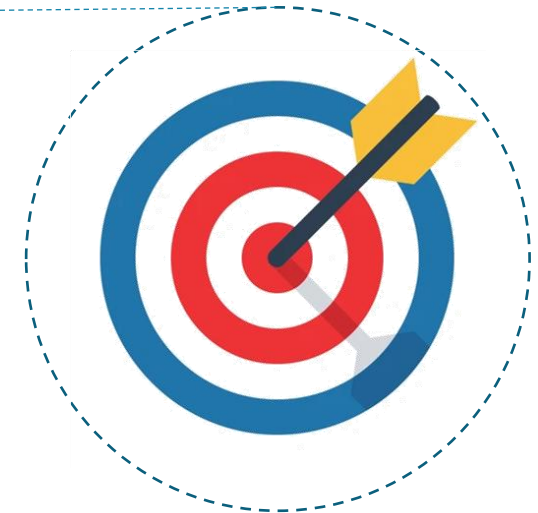
Tissue = complex functional assembly of cells and structures

- Cellular/phenotypic composition
- Impact of the microenvironment
- Complexity and interpretation of molecular signatures

Strategic Approach

Spatial and Functional Proteomics

- Mapping of cellular subpopulations
- Understanding biological mechanisms (physiological and pathological)

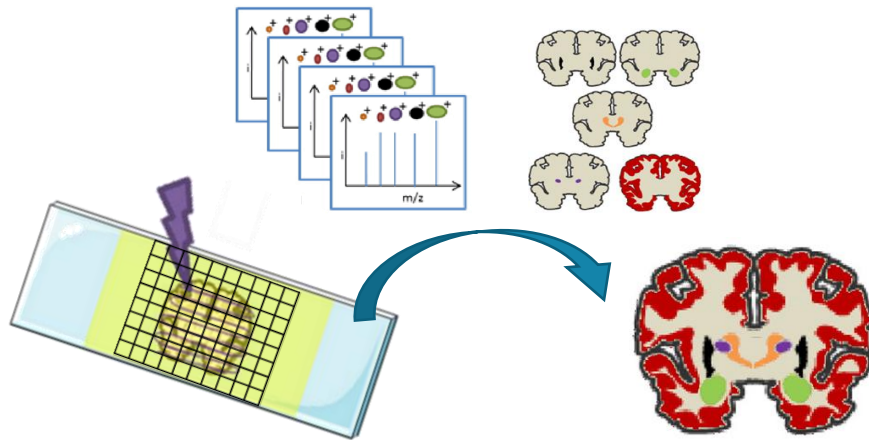


Spatial and functional proteomics

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Spatial proteomics

Mass Spectrometry Imaging (MSI)

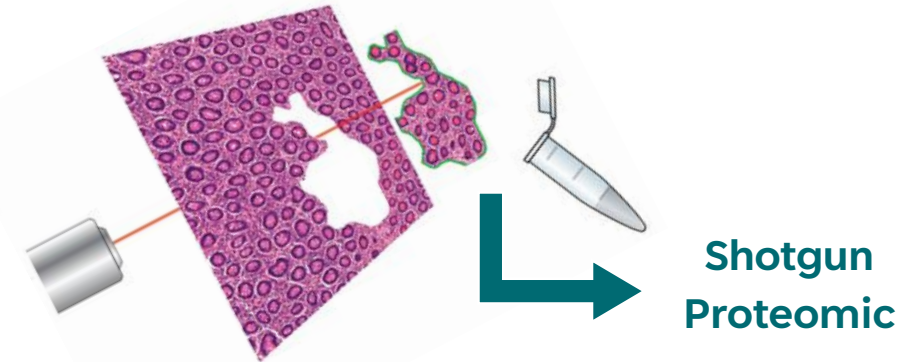


- ✓ High spatial resolution
- ❑ Low functional information
- Spatial molecular localization

Functional proteomics

Laser capture Microdissection (LCM) / LC-MS

Region
Of
Interest

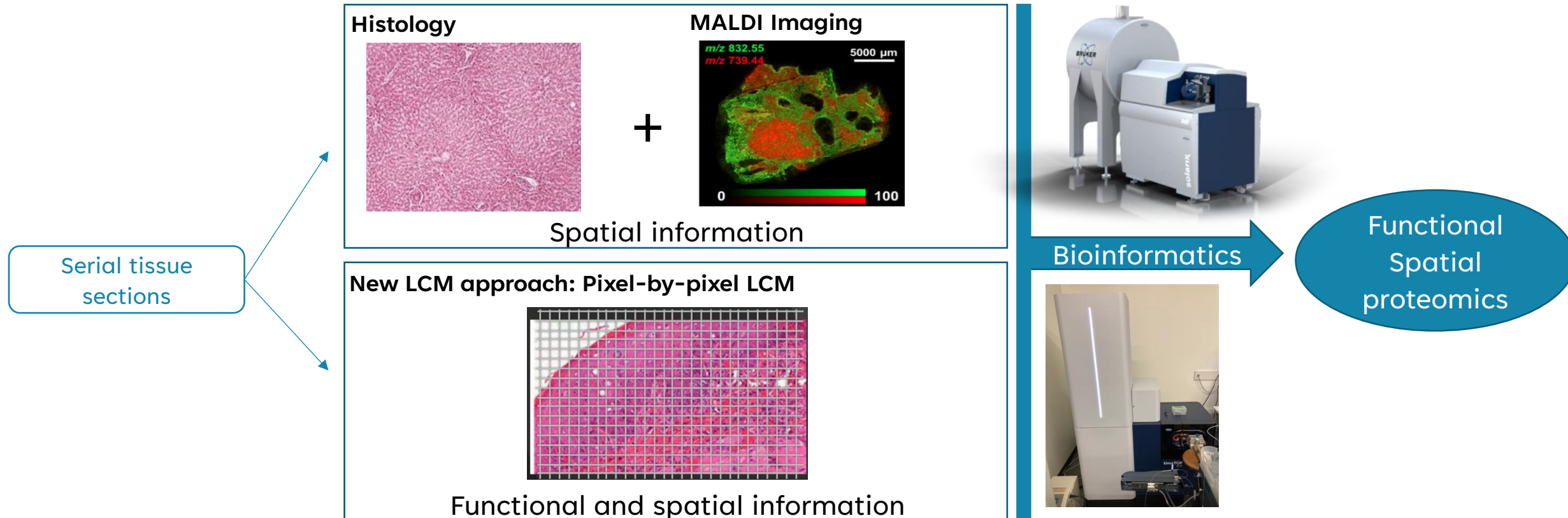


- ❑ Low spatial resolution
- ✓ High functional information
- Proteins function in region of interest

Strategy

CBIO

Develop spatial and functional proteomic method combining MALDI-MSI and LCM-LC-MS



Pixel-by-pixel LCM concept: unsupervised LCM proteomics at high spatial resolution

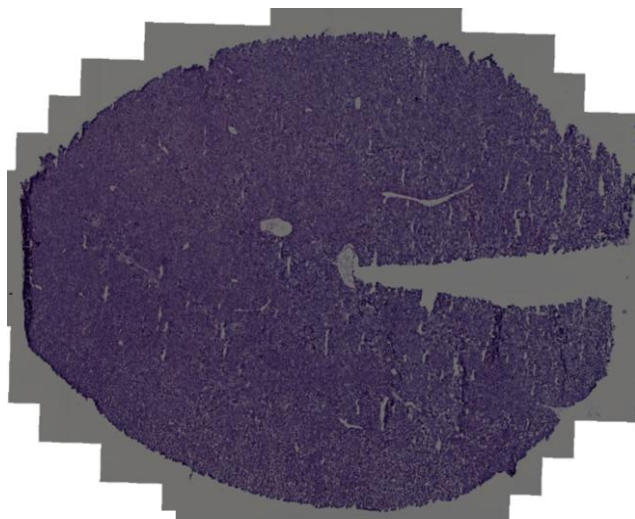
- Require method development and optimisation

Biological models

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Homogenous tissue

Mouse liver

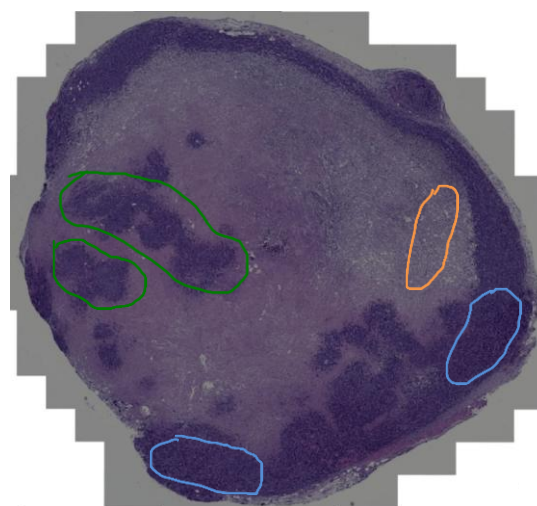


Methods optimisation

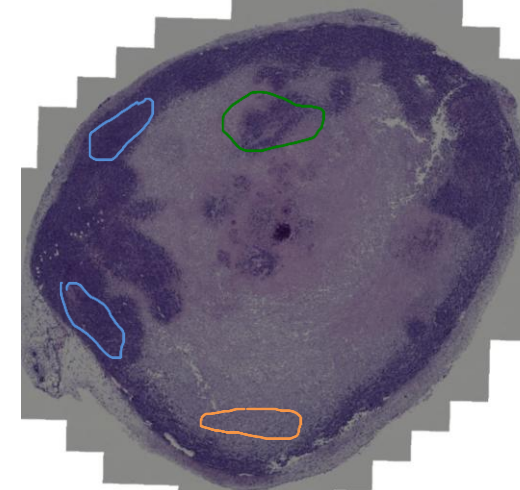
Heterogenous tissue

Triple negative breast cancer (TNBC)

(Patient Derived Xenograft (PDX) from human cancer in a mouse)



Non-treated



Anti-cancer drug treated

■ Peripheral region ■ Hypoxic region ■ Inflammatory region

Methods evaluation

- Heterogeneity study
- Region differential study
- Non-treated vs treated comparison

Serial sections strategy

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5-6 μm

5-6 μm

10-12 μm

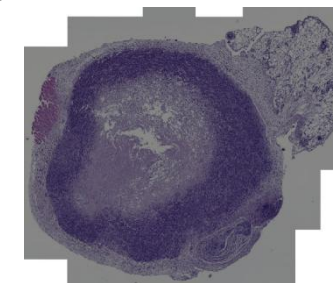
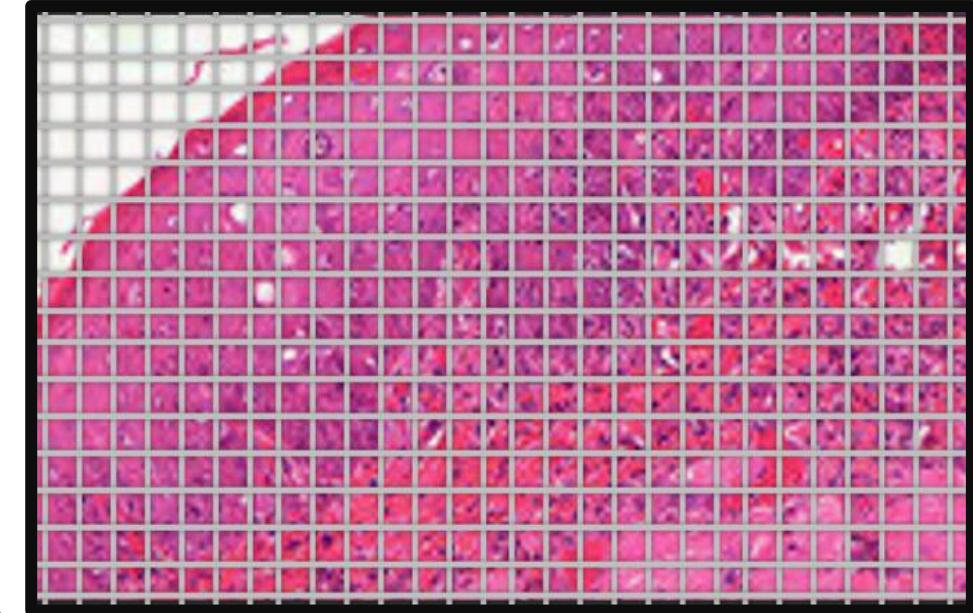
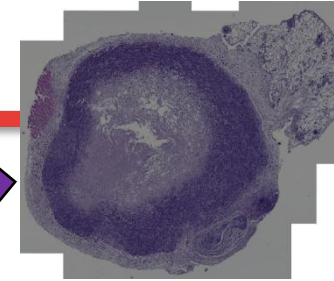
H&E

Pixel by pixel
proteomics

H&E

Molecular MALDI imaging

- A section for each experiment
- H&E sections, to visualise tissue structure variation

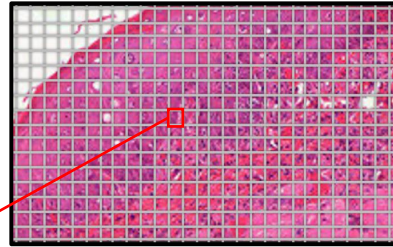


Analytical and bioinformatics workflow

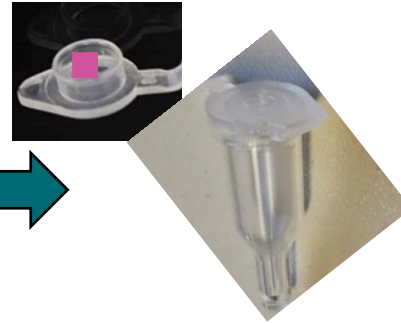
CBIO

MASS
LABORATORY
SPECTROMETRY

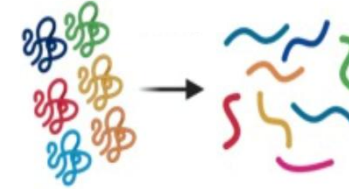
Systematical Laser Microdissection



FFPE and FF (Fresh Frozen) Tissue Sections



Reduction, alkylation, and trypsin digestion

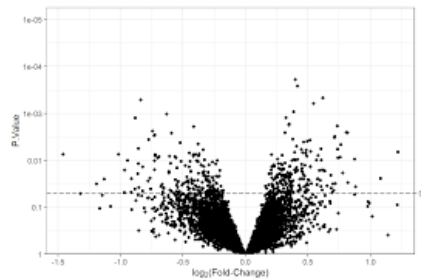
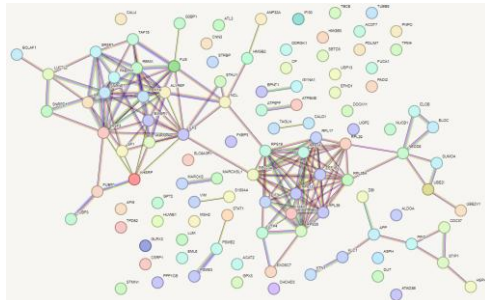


LC-MS/MS

60 minutes, 15cm Aurora column, DIA run

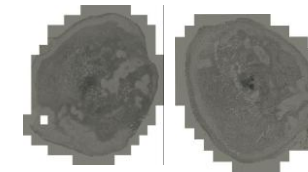
STRING

Biological interpretation



DaSER

In-house spectral library



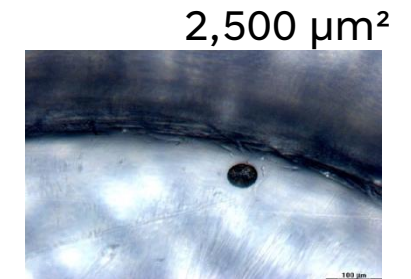
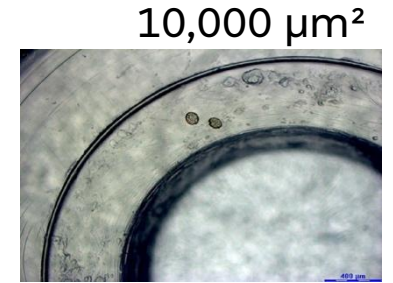
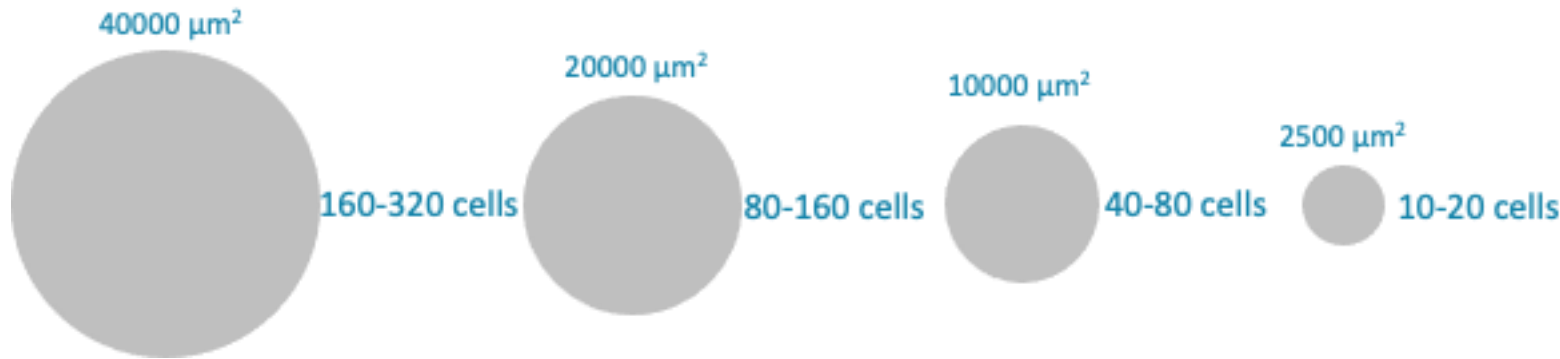
- DDA analysis of whole sections in triplicate
- 25 cm Aurora column and 100 min run
- New spectral library with 12000 proteins



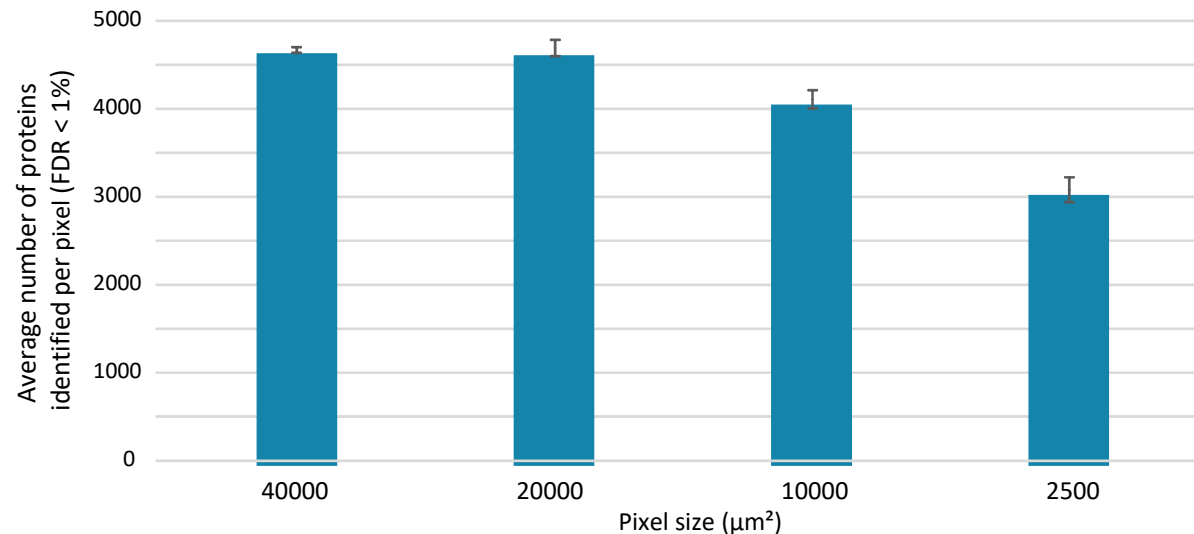
Pixel-by-pixel LCM: LCM optimisations

CBIO

Best spatial resolution (smallest pixel size) maintaining high proteomics performance



Proteomic performance vs Pixel size

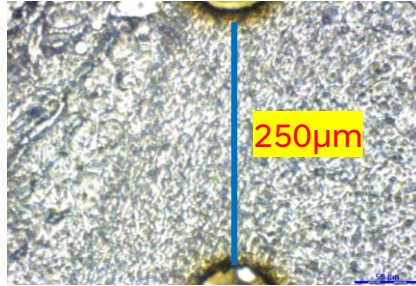
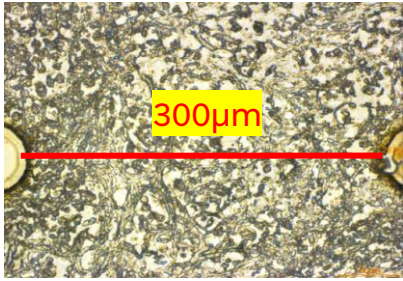


- Pixel surface 16 times smaller.
- Decrease in terms of identified proteins does not even correspond to a factor of 2.
- Biological material sufficient to achieve a fairly deep proteome.

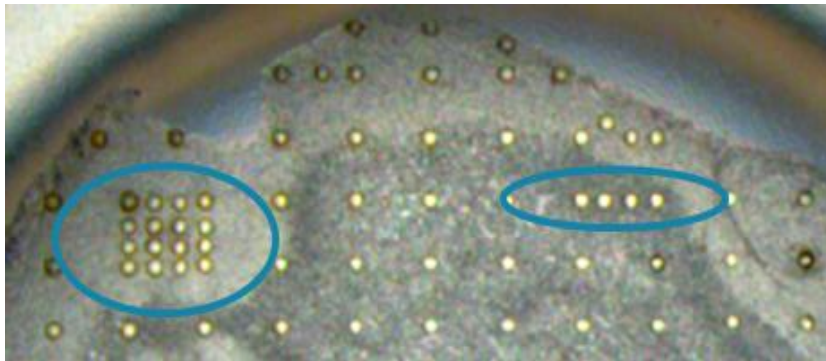
Pixel-by-pixel LCM: Two approaches of pixel sampling

CBIO

1) Systematic - grid

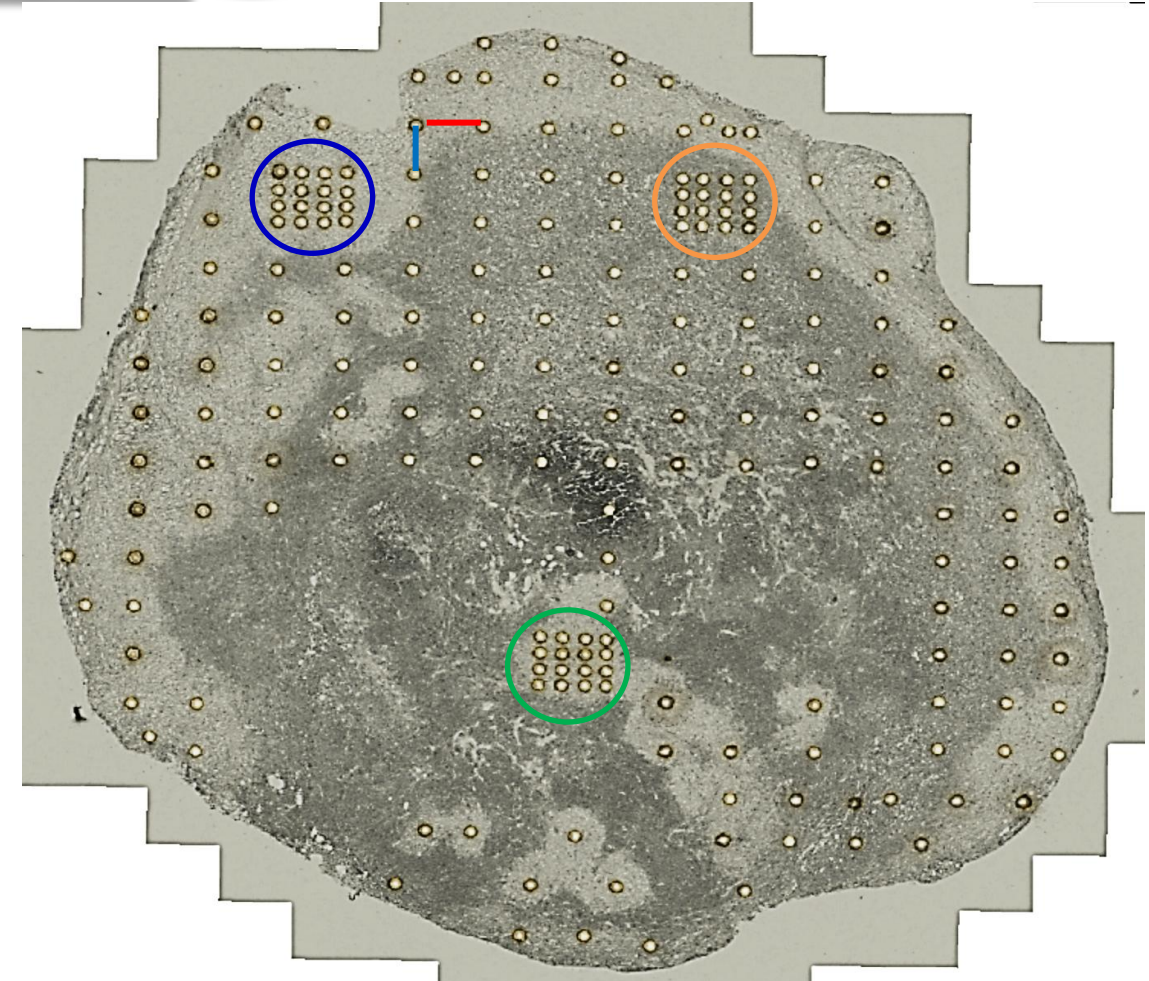


2) Areas of interest



- Pixels density higher in areas of histological interest
distance = 25 microns

Best approach (best resolution), but challenging without automation

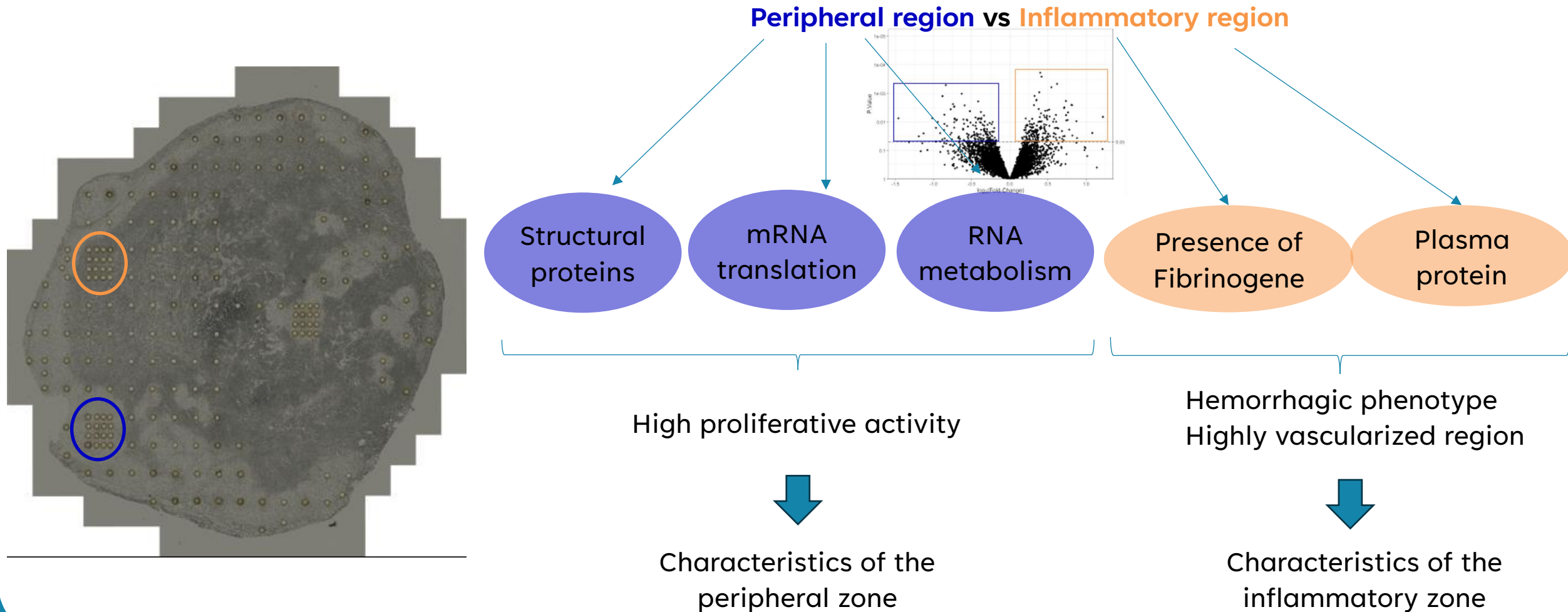


- Number of proteins ranges between 800 and 3000 per pixel
(2500µm² - 10 to 20 cells)
- Supervised analysis between areas of biological interest

Pixel-by-pixel LCM: Differential analysis on regions

CBIO

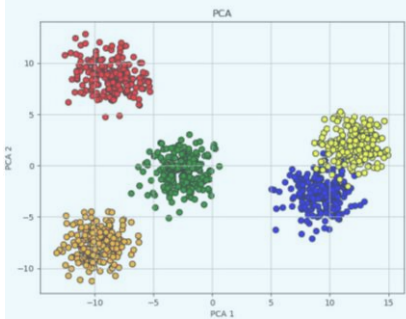
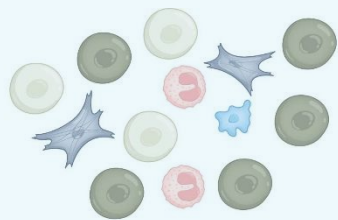
Differential protein abundances have been detected in the specific regions of biological interest :



Perspectives: towards single-cell spatial resolution

CBIO

SCP



Phénotype cellulaire

A

B

C

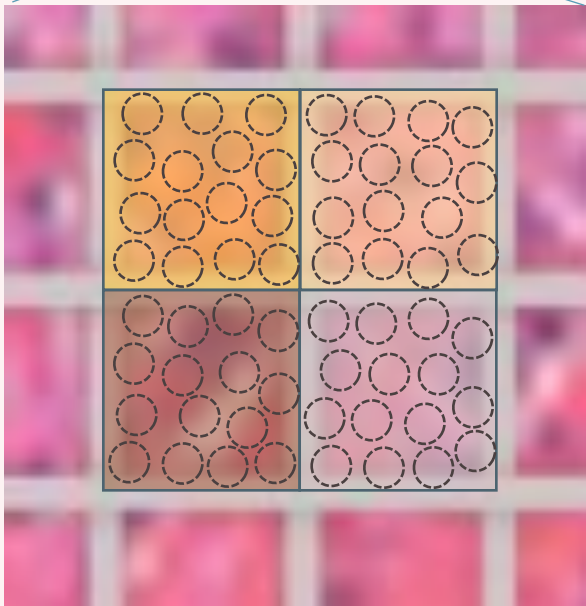
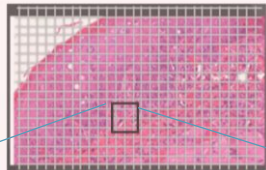
D

E



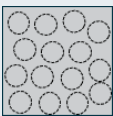
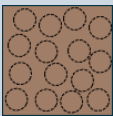
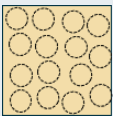
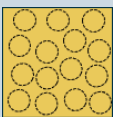
Deconvolution

P2P



Pixel

Cellular phenotypes



Cellular Spatio-functional



Take-home messages

- Development of a spatial method giving functional information with a systematic LCM sampling grid.
 - Pixel size = 10-20 cells = from 800 to 3000 identified proteins
 - 120 pixels /tissue section, 2 tissue sections analyzed in the current proof of concept study
 - A novel approach to reach single-cell spatial resolution
- Ongoing work: Bioinformatic pipeline development to combine single-cell experiment and the pixel-by-pixel approach



The End

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Thank you for your attention!

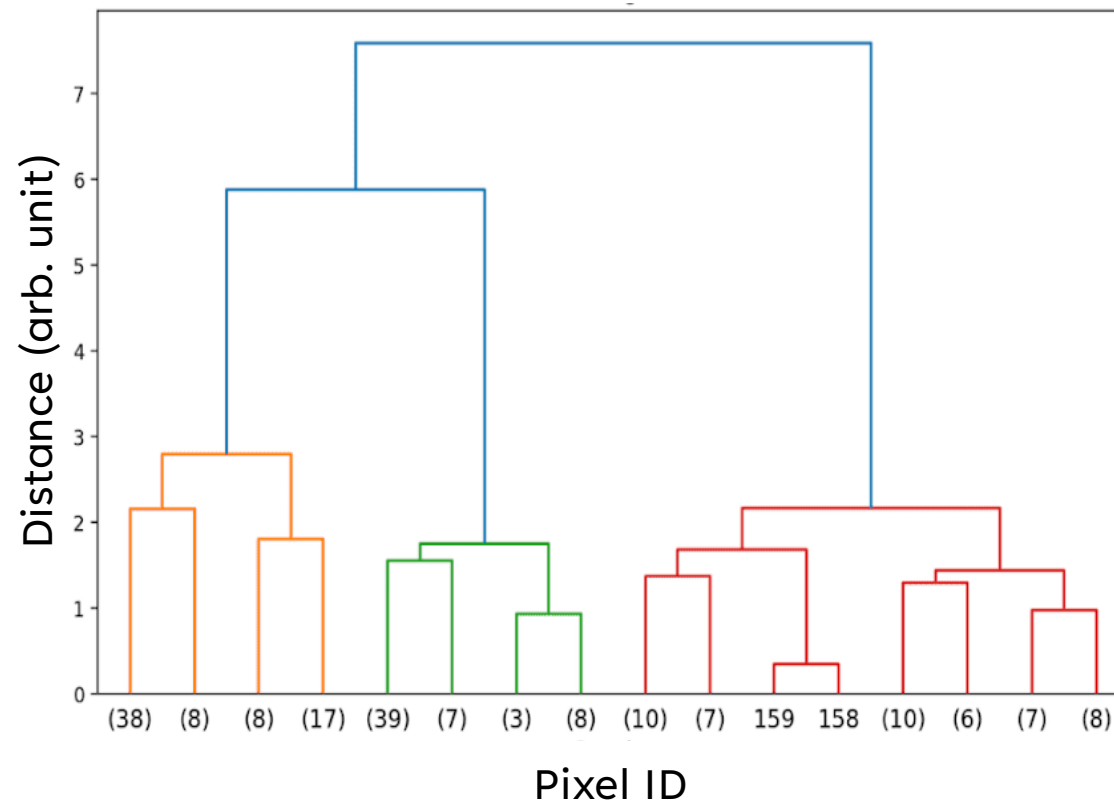
Do you have any questions?



BACK-UP

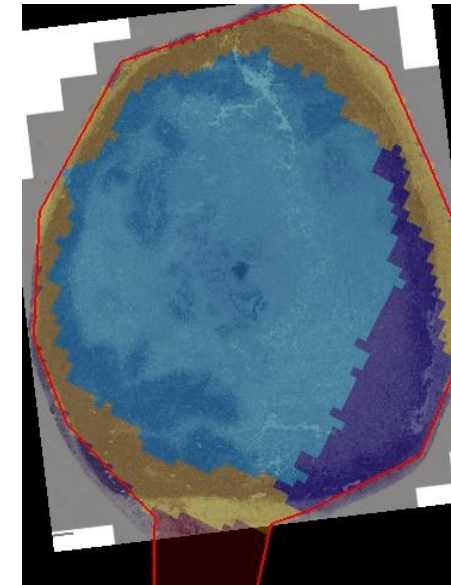
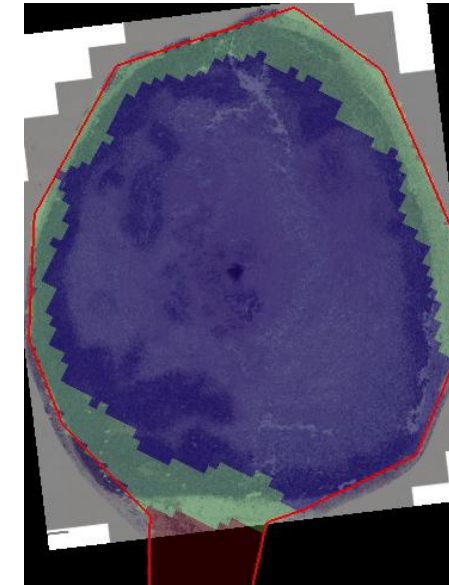
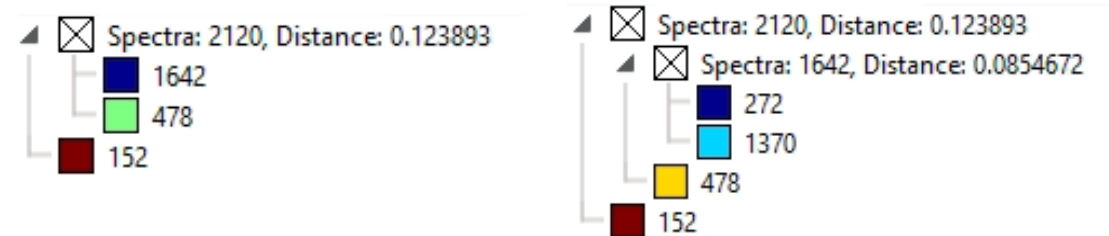
MALDI MSI Results: Hierarchical clustering

Hierarchical clustering concept :



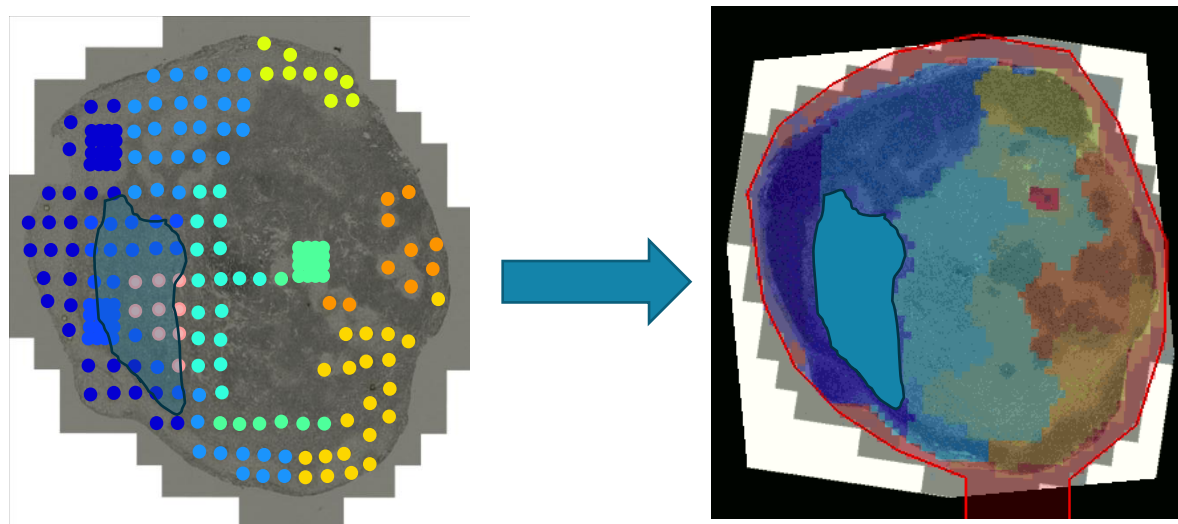
Pixel are clustered based on the difference on their MS spectrum

Pixel are colored based on their group



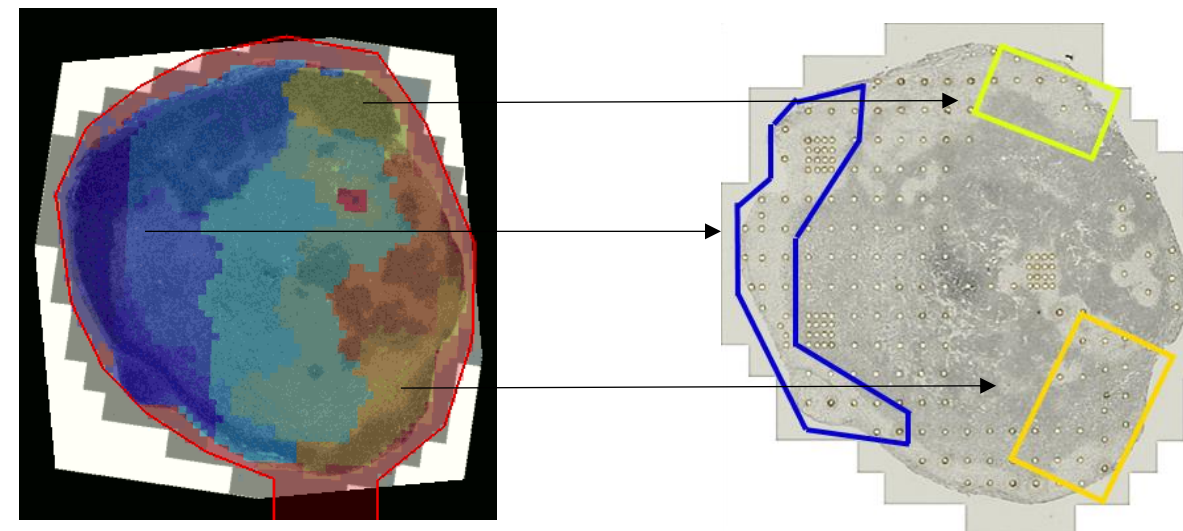
Perspectives

Unsupervised LCM pixel-by-pixel proteomics



- Obtaining zones by clustering LCM data with functional interpretation
- Compare with MALDI MSI molecular clustering

Supervised LCM pixel clustering

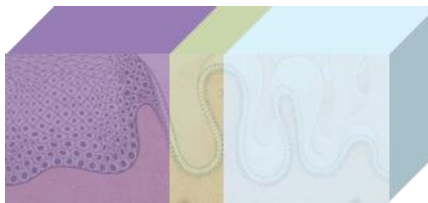
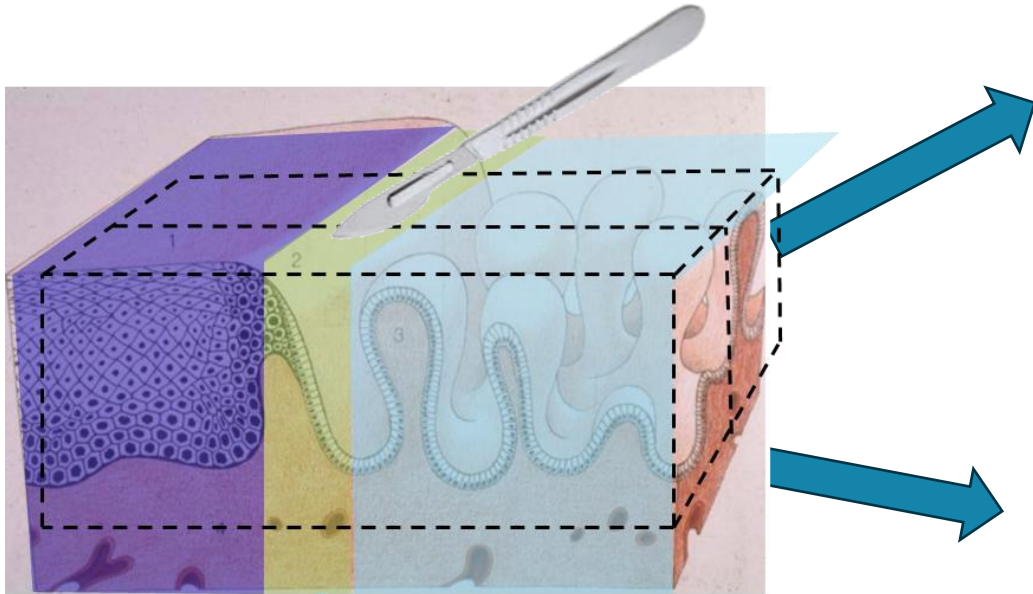


- Clustering pixels based on the zones sharing similar molecular MALDI profiles
- Supervised differential study based on MALDI imaging
- Allow futur revisiting of LCM-data

Objectif et approche

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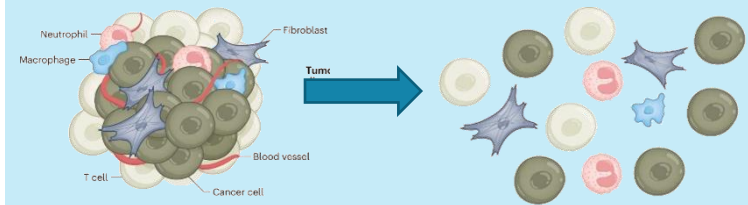
Méthode d'analyse de
protéomique spatiale
systématique à échelle
cellulaire



Approche fonctionnelle



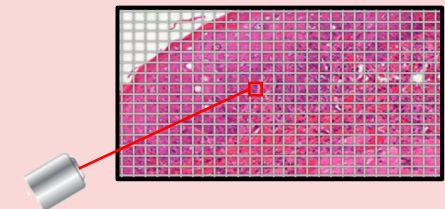
Single-cell Protéomique (SCP)



Approche spatiale

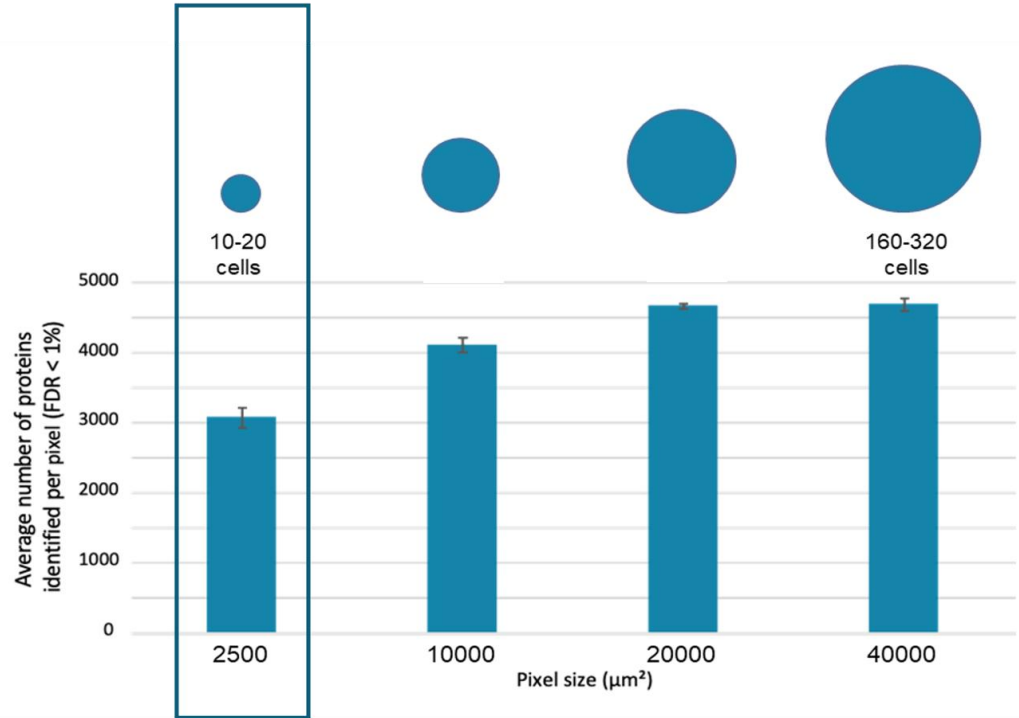
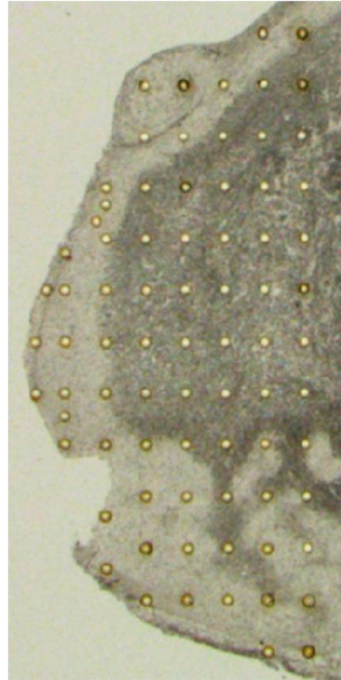


Pixel par pixel Protéomique (P2P)



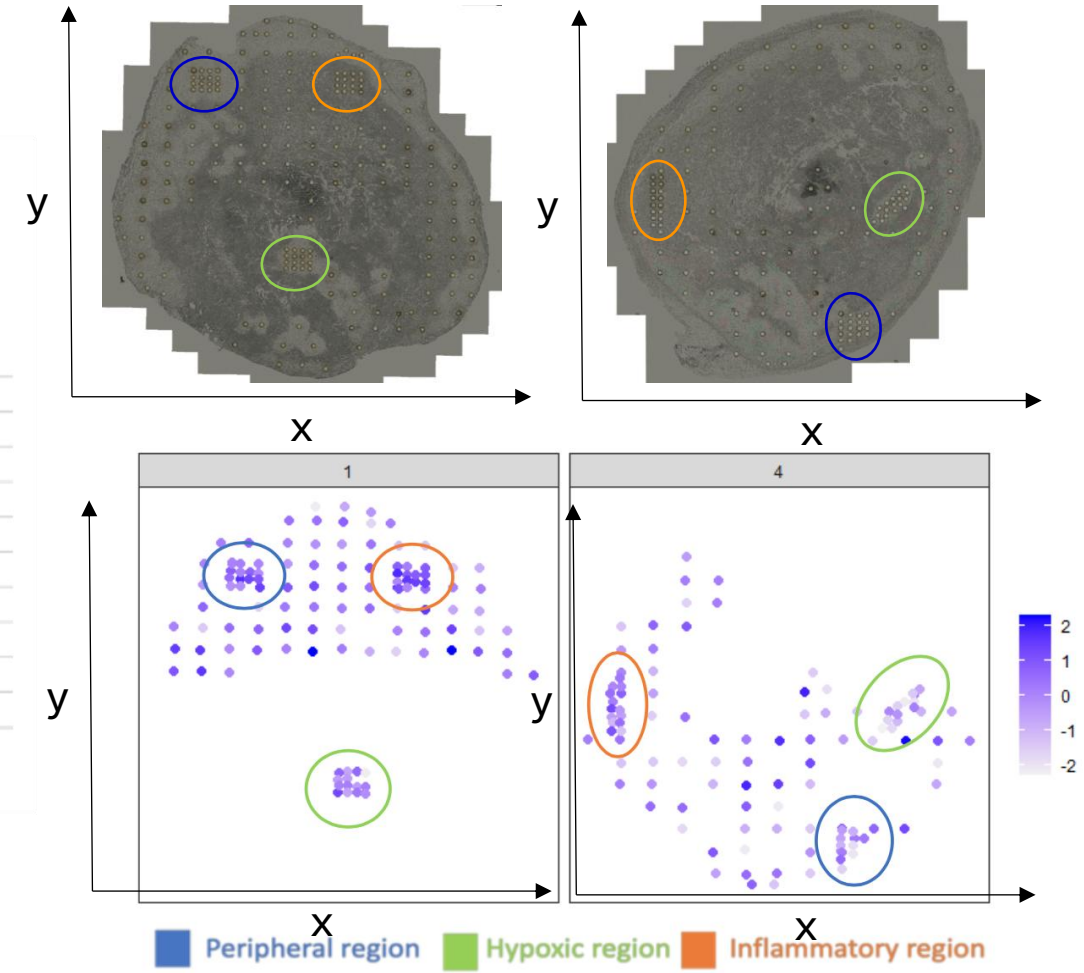
Résultats préliminaires

CBIO



✓ Microdissection systématique

↗ Densité pixels



✓ Protéomique spatiale (P²P)