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Fissurina_nigrolabiata_DNA2852_Philippines_mtSSU-LSU
Fissurina_aggregatula_DNA3240_Peru_mtSSU-LSU
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Gyalideopsis_aff-rubescens_EL892c_Guadeloupe_mtSSU-LSU
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Gyalidea_aff-lecideopsis-var-eucarpa_Spribille39048_Alaska_RPB1
Gyalidea_fritzei_EB89_Sweden_mtSSU-LSU
Gyalidea_spec-nova-1_EL2299_Guadeloupe_mtSSU-LSU-RPB1
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Gyalideopsis_pallida_LOT00-37048B_Colombia_mtSSU-RPB1
Gyalidea_concava_Aptroot40947_Brazil_mtSSU-LSU-RPB1
Gyalidea_concava_Aptroot18285_Brazil_LSU-RPB1
Gyalidea_aff-mayaguezensis_EL1601_Guadeloupe_mtSSU
Gyalidea_aff-mayaguezensis_EL1564_Guadeloupe_mtSSU
Gyalidea_aff-mayaguezensis_EL1515_Guadeloupe_mtSSU
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Gyalidea_aff-mayaguezensis_EL1519_Guadeloupe_mtSSU-LSU-RPB1
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Taitaia_aurea_AF260_Kenya_LSU
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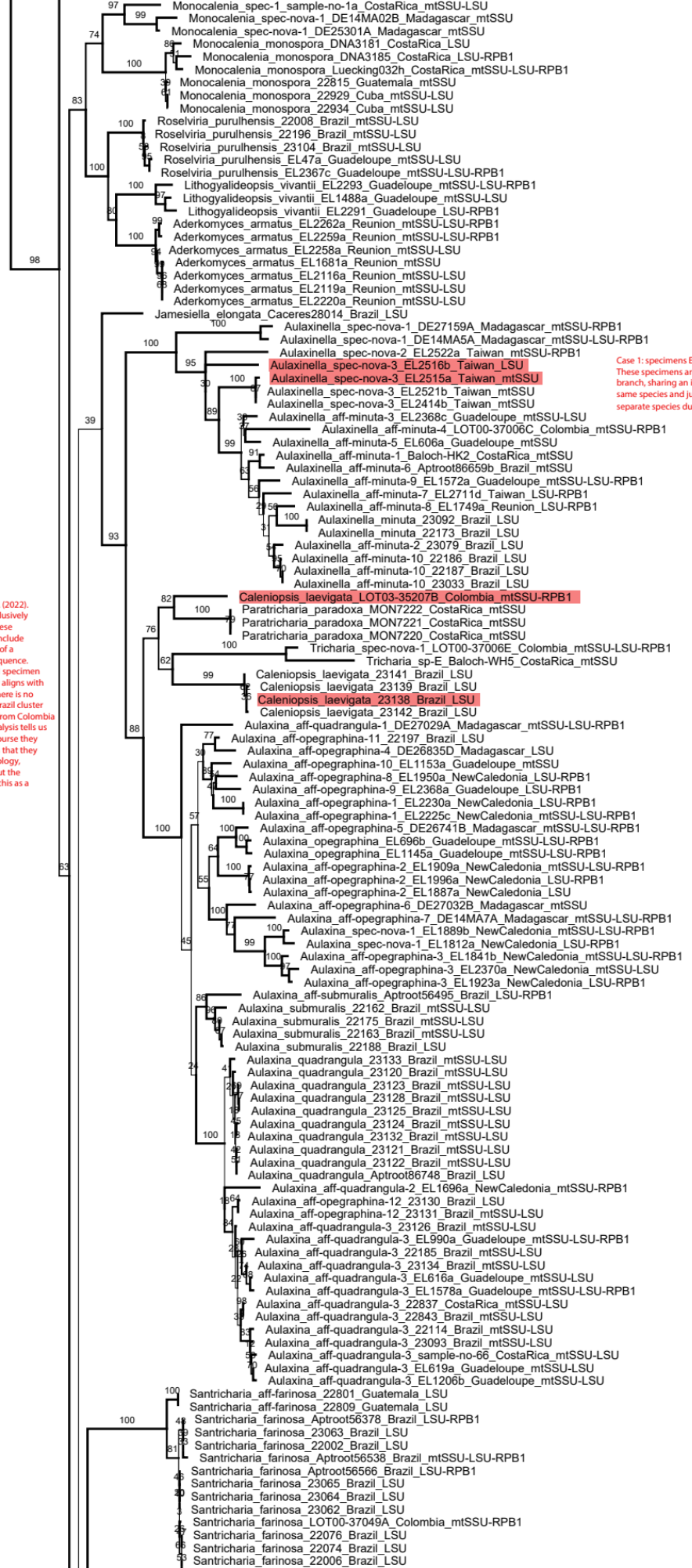
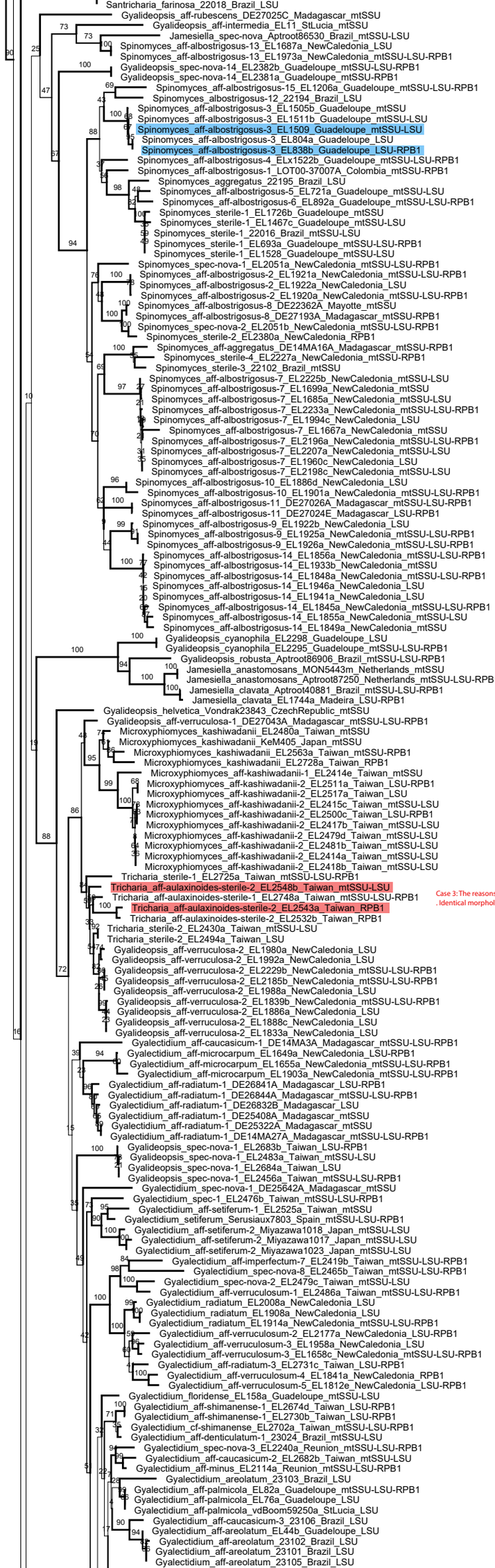
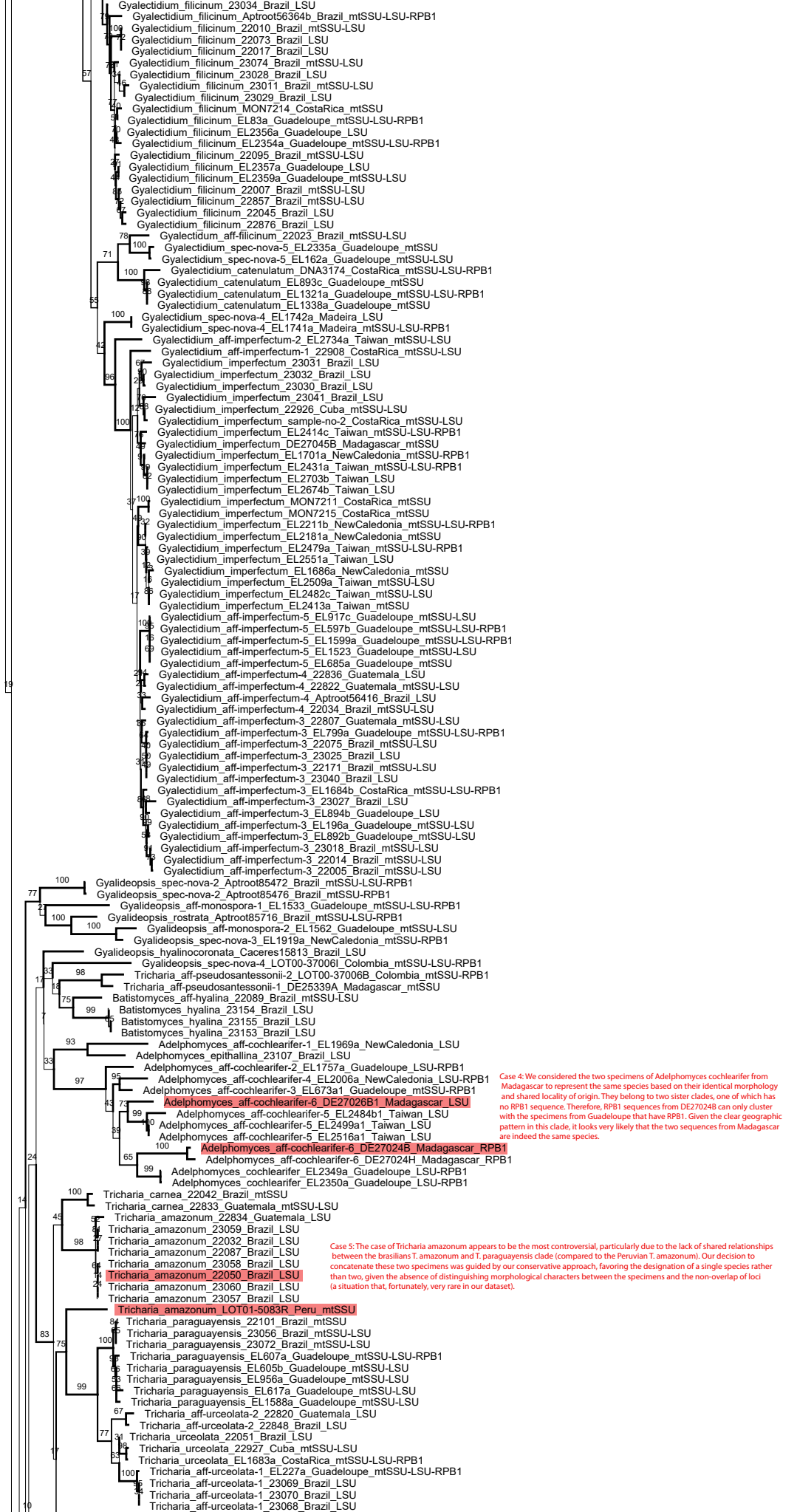


Case 1: specimens EL2516b and EL2515a are placed very close phylogenetically. These specimens are morphologically identical and were collected from the same tree branch, sharing an identical micro-locality. Its strongly suggest that they belong to the same species and justify concatenating their sequences rather than treating them as separate species due to non-overlapping loci.

Case 2: Caleniopsis genus is currently supported only by the LSU locus, as published in Xavier-Leite et al. (2022). Relationships between genera remain poorly resolved, as its sister genus, Paratricharia, is supported exclusively by the mtSSU locus, published in Lücking et al. (2025). Consequently, the phylogenetic positioning of these specimens within sister clades provides limited insight. Previously published sequences of C. laevigata include only the LSU, which presents challenges because this locus is difficult to amplify (necessitating the help of a direct PCR kit). Due to the small size of the specimen we collected, we were unable to obtain the LSU sequence. Collecting additional material for sequencing was not an option, as we had only collected a single, small specimen to preserve its morphological reference. Despite this limitation, the morphology of our specimen clearly aligns with C. laevigata s.lat., leaving little doubt about its identification. As can be seen on the phylogenetic tree, there is no specimen of Caleniopsis laevigata with overlapping loci (mtssu + RPB1 vs LSU). The 4 Caleniopsis from Brazil cluster in a clade with Tricharia spec nov 1, based on overlapping LSU sequence, and the Caleniopsis laevigata from Colombia cluster with Tricharia spec nov 1 based on overlapping mtSSU and RPB1 sequence. The phylogenetic analysis tells us that both Caleniopsis laevigata from Brazil and from Colombia are close to Tricharia spec nov 1, but of course they cannot group together because they have no sequence in common. Therefore, it looks convincing to us that they are the same thing. For these reasons, we attempted to concatenate sequences to improve the tree topology, particularly the relationships between sister genera. Furthermore, our conservative approach throughout the paper for species delimitation led us to treat the specimen as part of C. laevigata rather than proposing this as a cryptic species, pending further sequencing efforts.



Case 3:The reasons for concatenating these two specimens are similar to case 1 (see above) . Identical morphology, same microlocality and close in the phylogenetic tree.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Gyalectidium_filicinum_23011_Brazil_mtSSU-LSU
Gyalectidium_filicinum_23029_Brazil_LSU
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Gyalectidium_filicinum_EL2354a_Guadeloupe_mtSSU-LSU-RPB1
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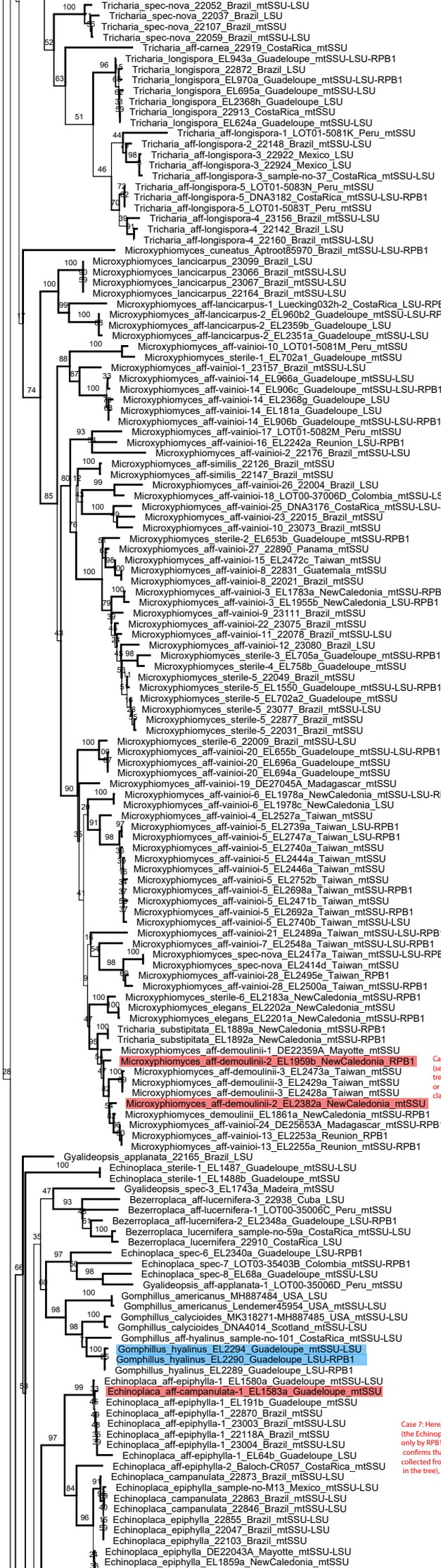
Case 4: We considered the two specimens of Adelphomyces cochlearifer from Madagascar to represent the same species based on their identical morphology and shared locality of origin. They belong to two sister clades, one of which has no RPB1 sequence. Therefore, RPB1 sequences from DE27024B can only cluster with the specimens from Guadeloupe that have RPB1. Given the clear geographic pattern in this clade, it looks very likely that the two sequences from Madagascar are indeed the same species.
Case 5: The case of Tricharia amazonum appears to be the most controversial, particularly due to the lack of shared relationships between the brasilians T. amazonum and T. paraguayensis clade (compared to the Peruvian T. amazonum). Our decision to concatenate these two specimens was guided by our conservative approach, favoring the designation of a single species rather than two, given the absence of distinguishing morphological characters between the specimens and the non-overlap of loci (a situation that, fortunately, very rare in our dataset).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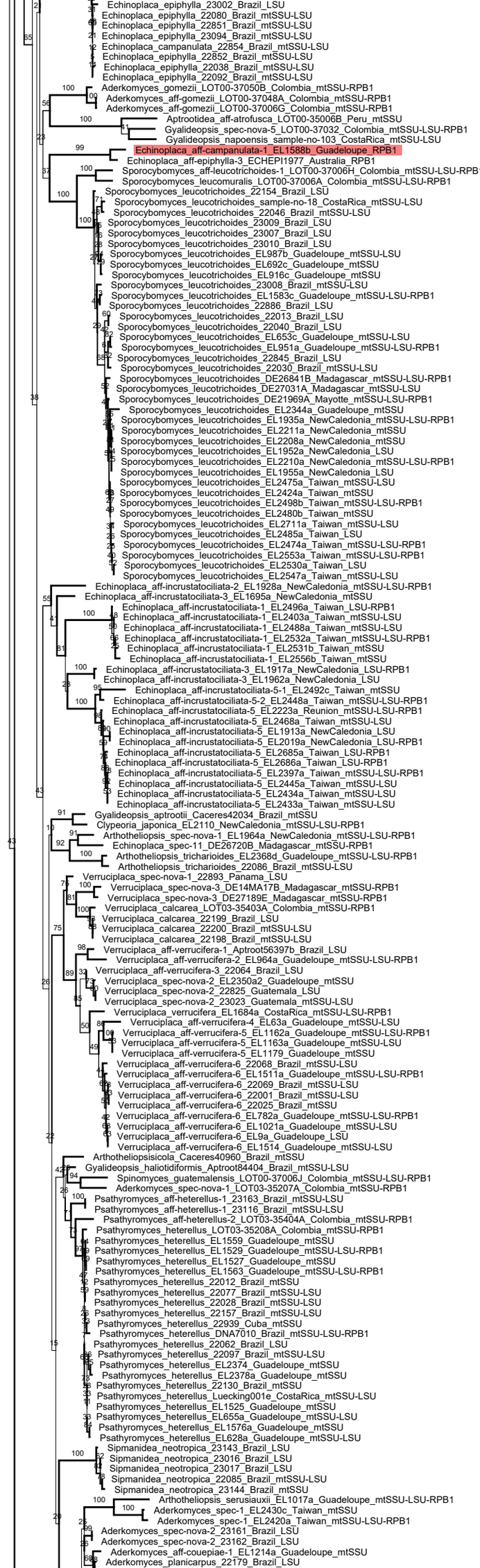
Echinoplaca_epiphylla_DE22043A_Mayotte_mtSSU-LSU
Echinoplaca_epiphylla_EL1859a_NewCaledonia_mtSSU
Case 6: The reasons for concatenating these two specimens are similar to case 1 (see above). Identical morphology, same microlocality and close in the phylogenetic tree. In this clade, all aff. demoulinii have non-overlapping sequences (either RPB1 or mtssu) so this pattern of them hanging into a polytomy between the two clades that have mtSSU + RPB1 suggests they are probably the same thing indeed.
Case 7: Here, the absence of overlap with the locus RPB1 for the other clades (the Echinoplaca epiphylla clades) results in the formation of a single branch supported only by RPB1. The GenBank sequence labeled "Echinoplaca epiphylla" from Australia confirms that EL1588b belongs to the E. epiphylla clade. Additionally, EL1583a was collected from the same tree branch as EL1588b (and EL1580a, specimen just above in the tree), sharing the same exact micro locality and morphology.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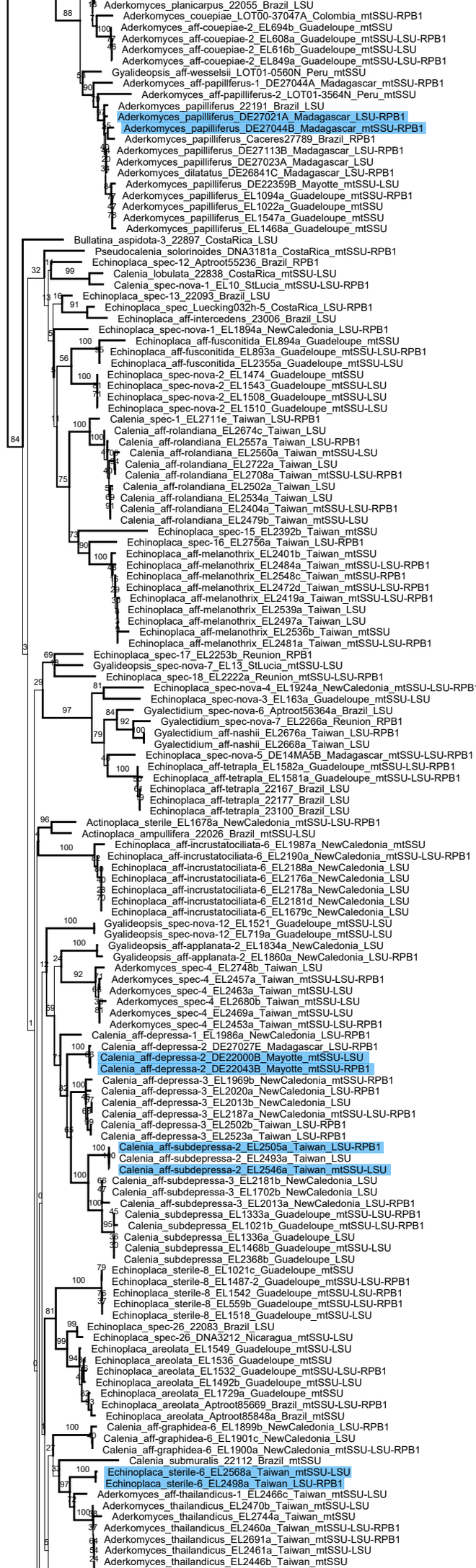
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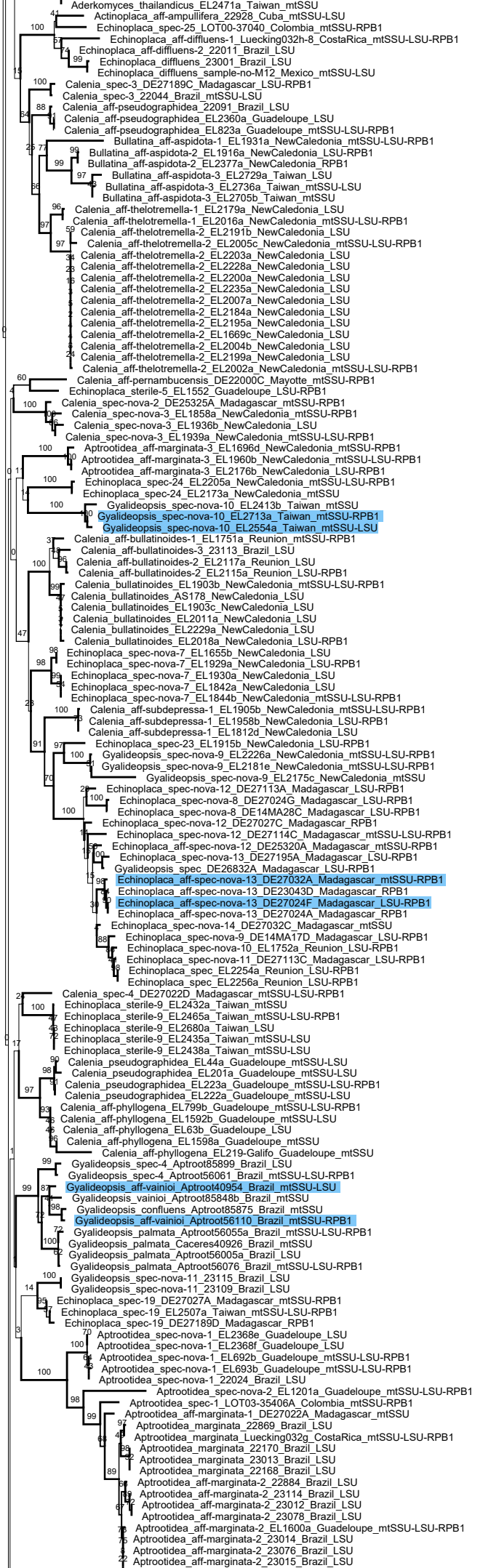
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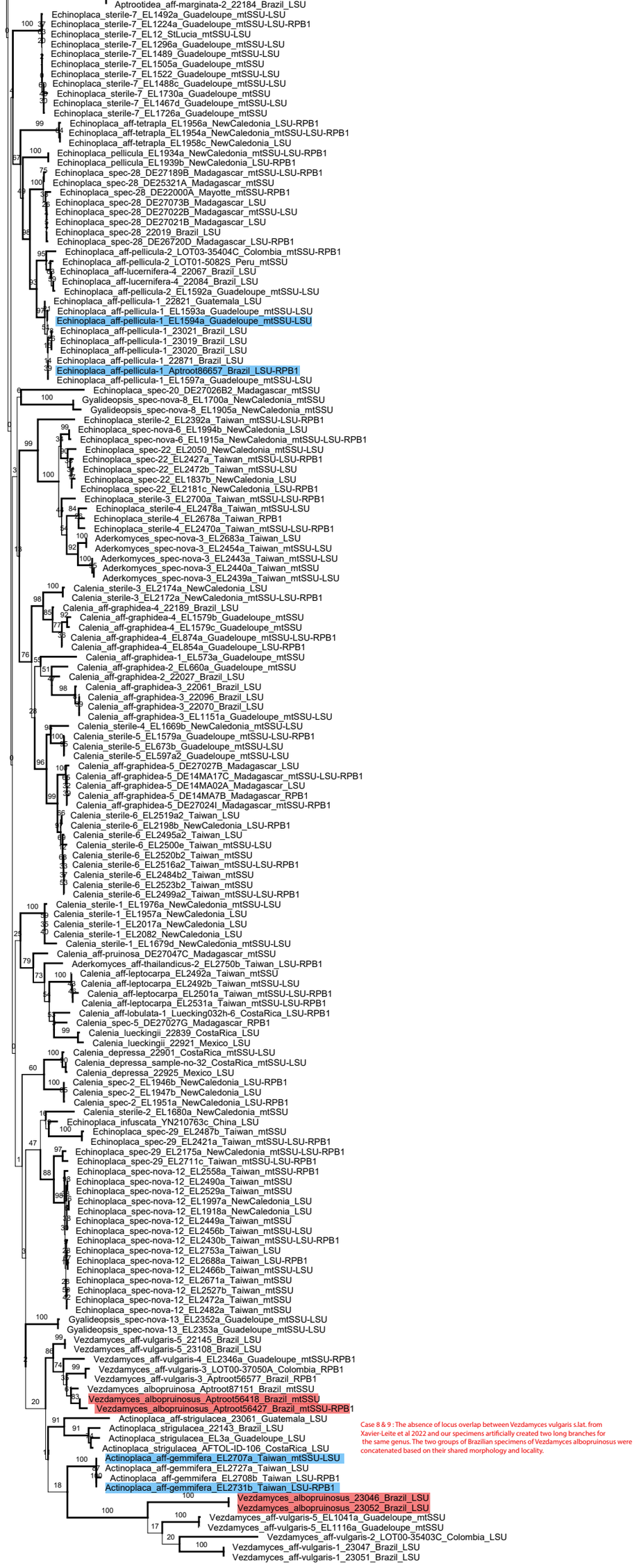

Fig. S1A. Phylogenetic tree on the complete dataset inferred by maximum likelihood analyses of the mtSSU, nuLSU and RPB1 region.
Specimens highlighted in red indicate the placement of the 18 specimens prior to concatenation into 9 specimens for species delimitation. Specimens marked in blue represent those concatenated after species delimitation, forming subset 1 used for BioGeoBEARS analyses.

Fissurina_astroisidiata_DNA1998_Mexico_mtSSU-LSU
Fissurina_insidiosa_AFTOL-ID-1662_USA_mtSSU-LSU
Fissurina_nigrolabiata_DNA2852_Philippines_mtSSU-LSU
Fissurina_aggregatula_DNA3240_Peru_mtSSU-LSU
Fissurina_spec_AFTOL-ID-2101_Gabon_mtSSU-RPB1
Gyalideopsis_aff-rubescens_EL892c_Guadeloupe_mtSSU-LSU
Gyalideopsis_aff-rubescens_EL1583b_Guadeloupe_mtSSU-LSU
Psorotheciopsis_cf-premneella_22933_Cuba_LSU
Gyalidea_aff-lecideopsis-var-eucarpa_Spribille39048_Alaska_RPB1
Gyalidea_fritzei_EB89_Sweden_mtSSU-LSU
Gyalidea_spec-nova-1_EL2299_Guadeloupe_mtSSU-LSU-RPB1
Gyalidea_hyalinescens_AFTOL-ID-332_CostaRica_mtSSU-LSU
Gyalideopsis_pallida_LOT00-37048B_Colombia_mtSSU-RPB1
Gyalidea_concava_Aptroot40947_Brazil_mtSSU-LSU-RPB1
Gyalidea_concava_Aptroot18285_Brazil_LSU-RPB1
Gyalidea_aff-mayaguezensis_EL1601_Guadeloupe_mtSSU
Gyalidea_aff-mayaguezensis_EL1564_Guadeloupe_mtSSU
Gyalidea_aff-mayaguezensis_EL1515_Guadeloupe_mtSSU
Gyalidea_aff-mayaguezensis_EL1519_Guadeloupe_mtSSU-LSU-RPB1
Gyalidea_aff-mayaguezensis_EL1512_Guadeloupe_mtSSU-LSU
Taitaia_aurea_EA-5103_Kenya_mtSSU-LSU
Taitaia_aurea_TU56327-TU56326_Kenya_mtSSU-LSU
Taitaia_aurea_TU56327_Kenya_LSU
Taitaia_aurea_AF260_Kenya_LSU
Corticifraga_peltigerae_RP282_India_mtSSU-LSU
Corticifraga_peltigerae_GM-2015-05-02-1_Luxembourg_LSU
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Caleniella_aff-triseptata-2_Aptroot86642_Brazil_mtSSU-LSU-RPB1
Caleniella_triseptata_LOT00-37006F_Colombia_mtSSU-LSU-RPB1
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Caleniella_triseptata_EL615a_Guadeloupe_mtSSU-LSU
Caleniella_triseptata_ELx1531a_Guadeloupe_mtSSU-LSU-RPB1
Caleniella_triseptata_ELx1532a_Guadeloupe_mtSSU-LSU-RPB1
Caleniella_triseptata_23136_Brazil_LSU
Caleniella_triseptata_23135_Brazil_LSU
Caleniella_triseptata_23137_Brazil_LSU
Caleniella_triseptata_23146_Brazil_LSU
Caleniella_triseptata_23148_Brazil_LSU
Corticifraga_ramalinae_Pinault-isoT_France_mtSSU
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Linhartia_aff-patellarioides-4_EL2410a_Taiwan_mtSSU-LSU-RPB1
Linhartia_aff-patellarioides-2_EL2660a_Taiwan_LSU
Linhartia_aff-patellarioides-2_EL2566a_Taiwan_LSU-RPB1
Linhartia_aff-patellarioides-1_VdBoom59341a_StLucia_LSU
Linhartia_aff-patellarioides-1_vdBoom59340b_StLucia_mtSSU-LSU-RPB1
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Asterothyrium_leucophthalmum_23081_Brazil_LSU
Asterothyrium_aff-leucophthalmum-2_22936_Cuba_LSU
Asterothyrium_monosporum_vdBoom59340a_StLucia_LSU-RPB1
Asterothyrium_tetrasporum_EL893d_Guadeloupe_LSU-RPB1
Asterothyrium_longisporum_sample-no-4_CostaRica_LSU
Asterothyrium_rotuliforme_22161_Brazil_LSU
Asterothyrium_rotuliforme_22814_Guatemala_LSU
Asterothyrium_rotuliforme_Galifo183_Guadeloupe_LSU
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Asterothyrium_rotuliforme_EL2378b_NewCaledonia_LSU
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Asterothyrium_aff-microsporum-4_EL2735a_Taiwan_mtSSU-LSU-RPB1
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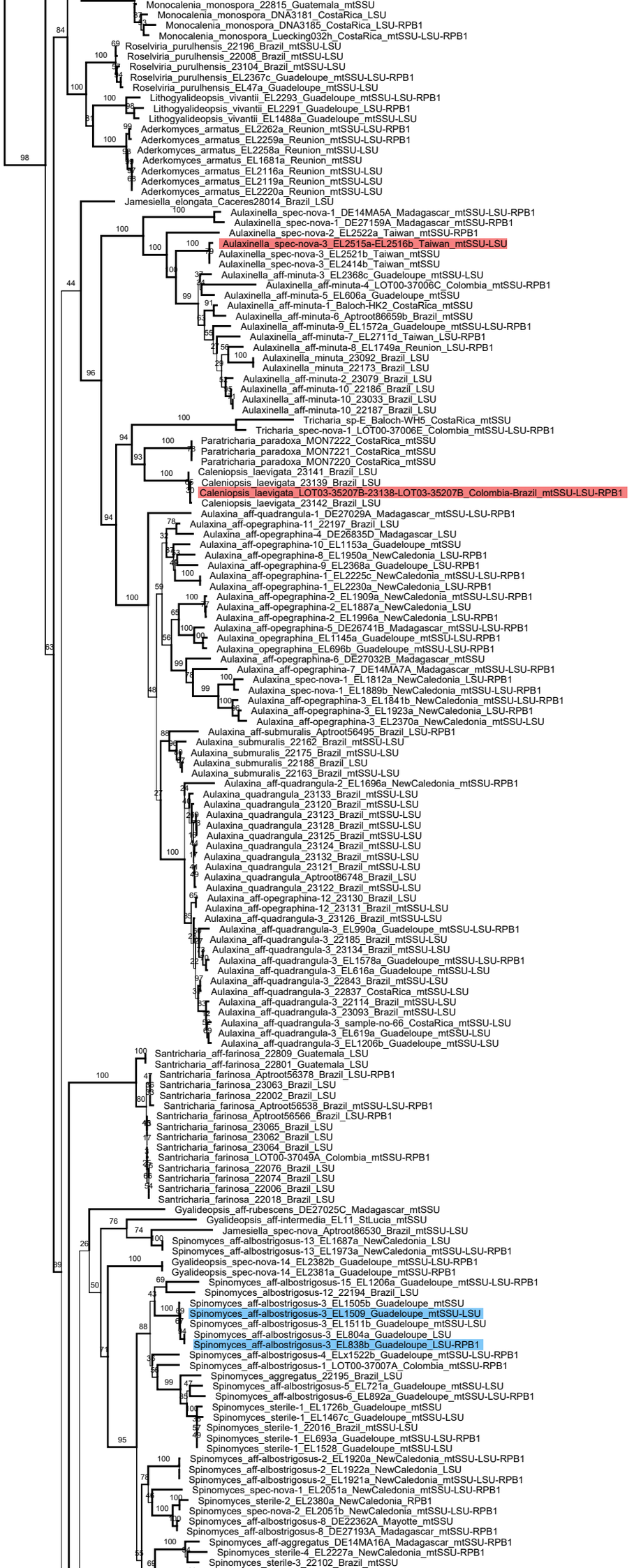
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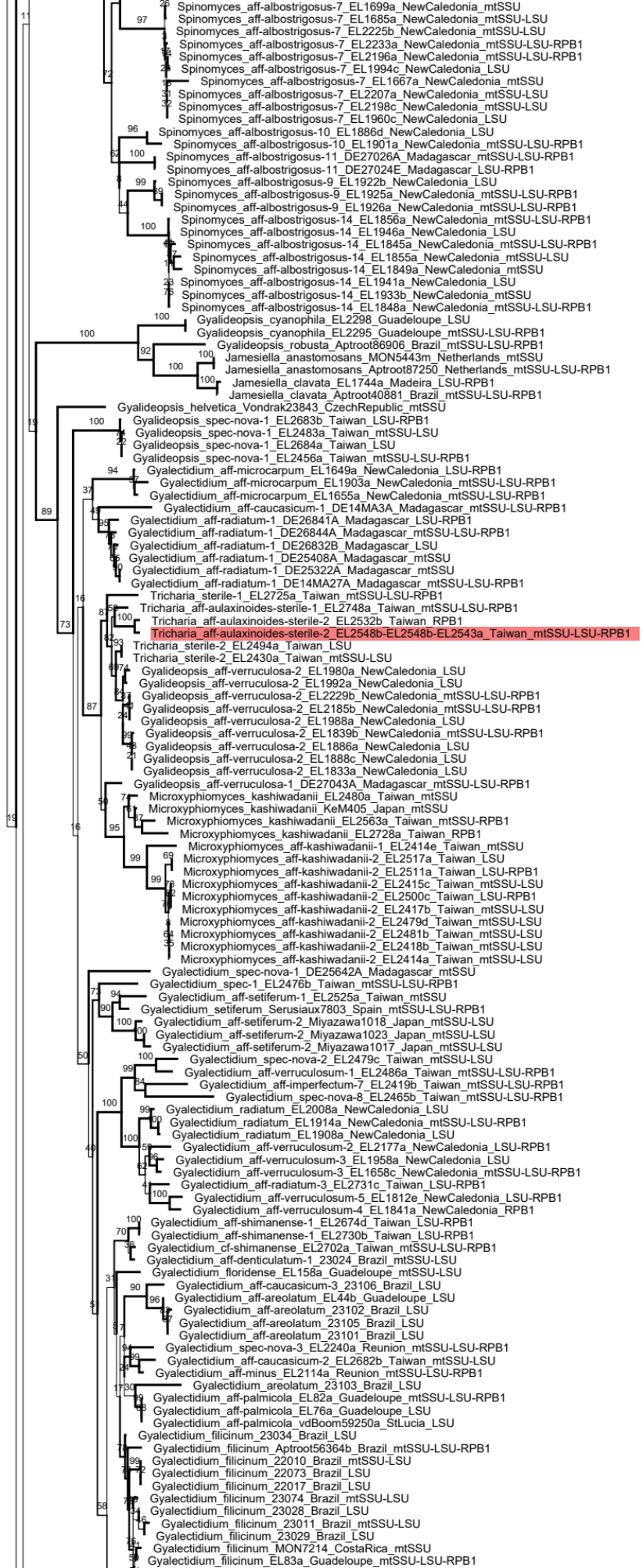
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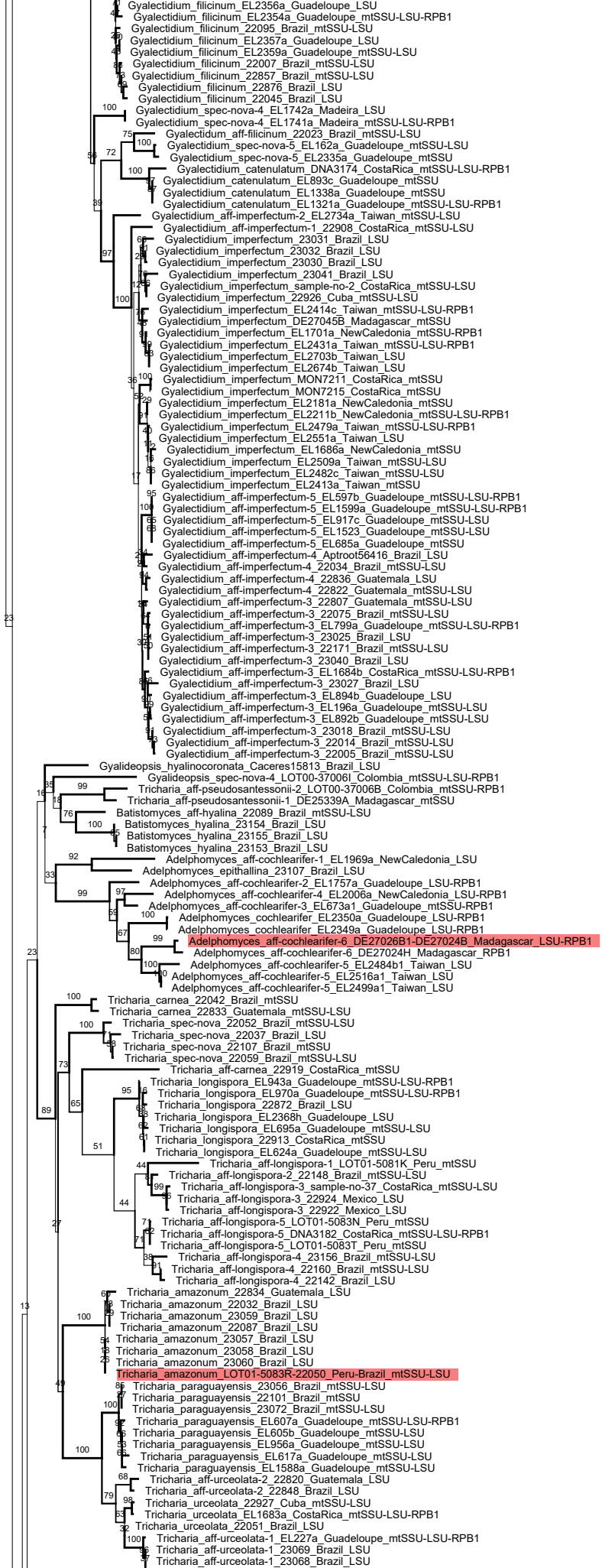
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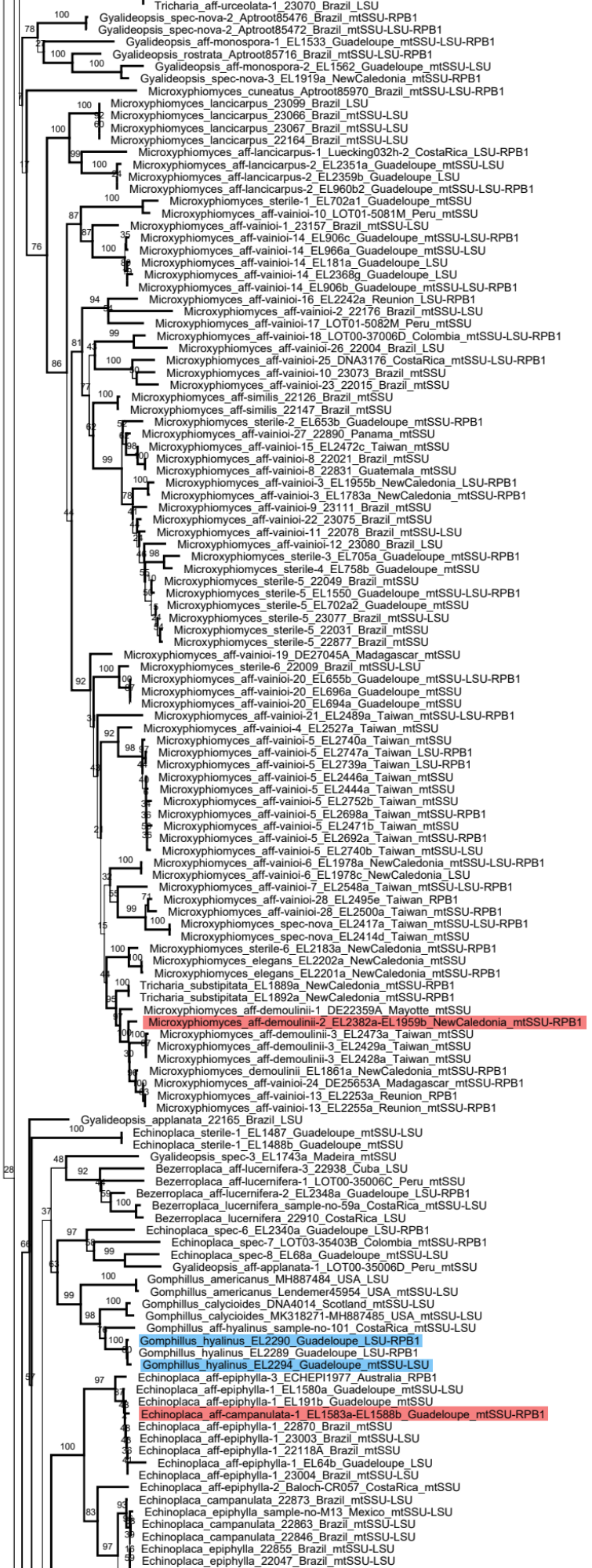
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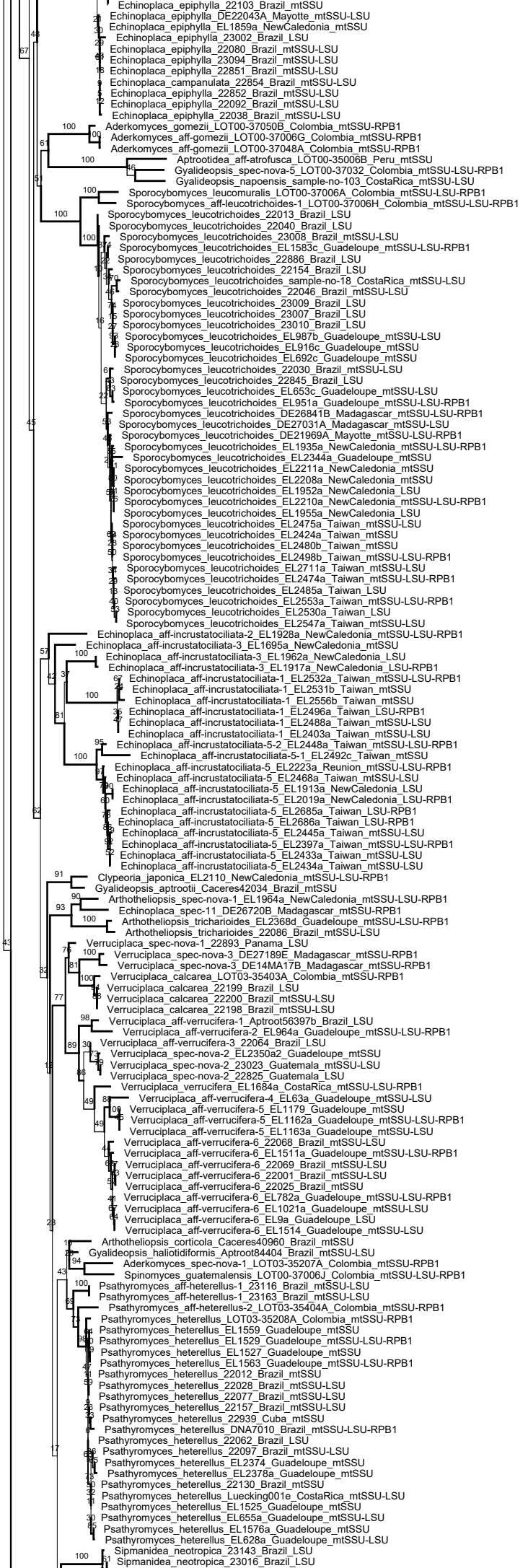
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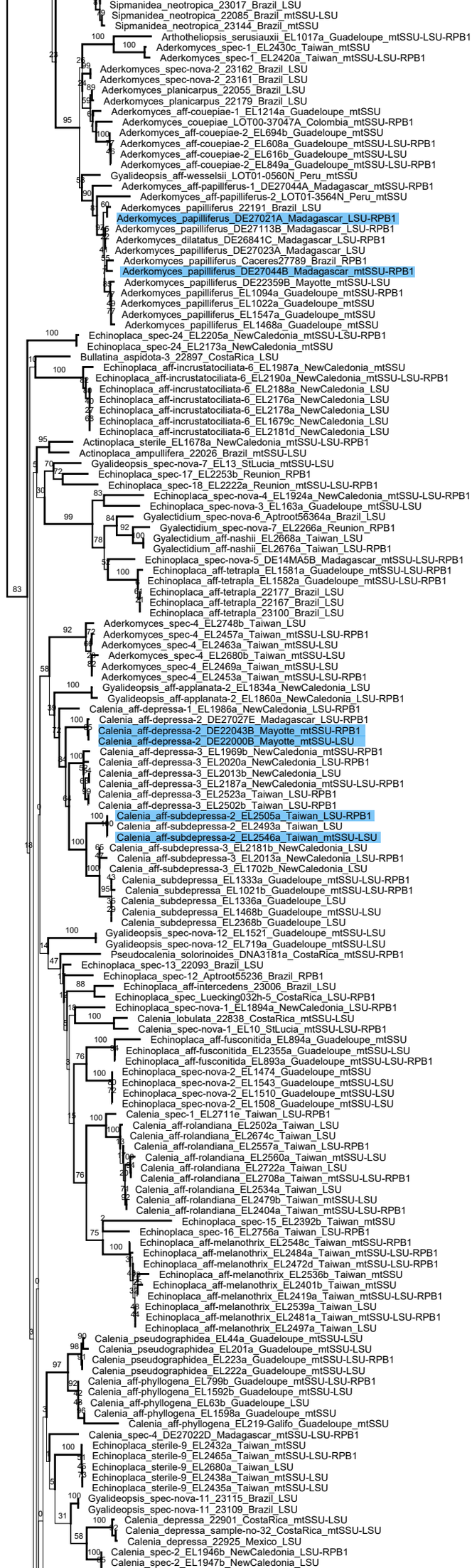
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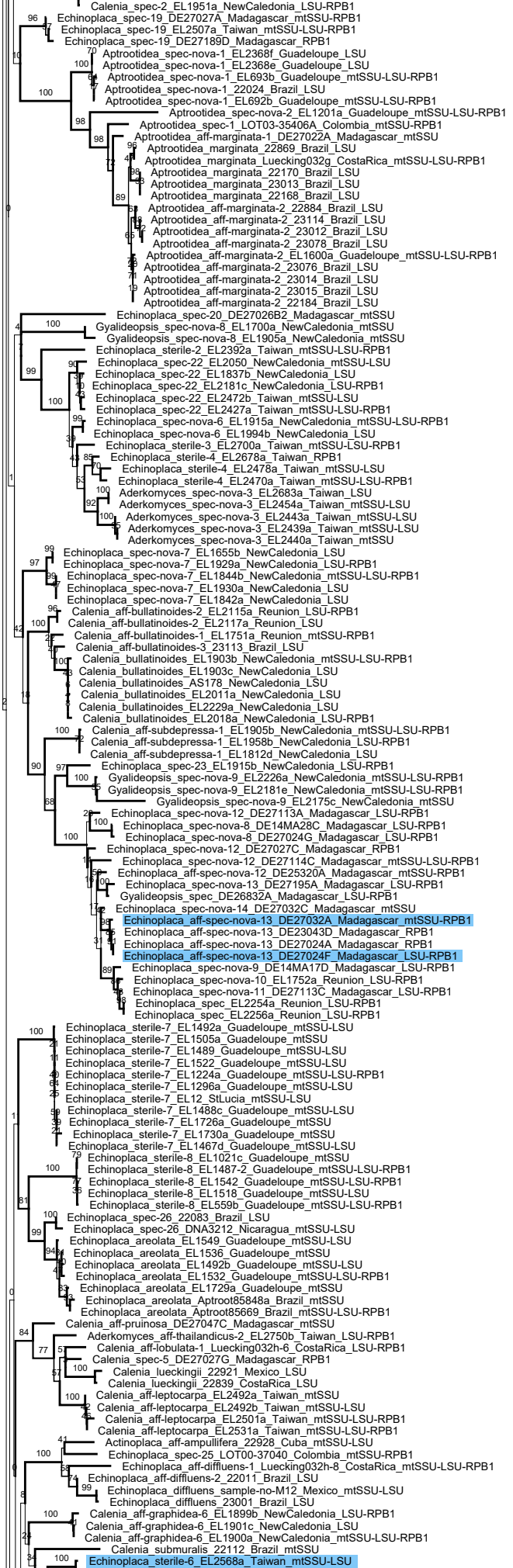
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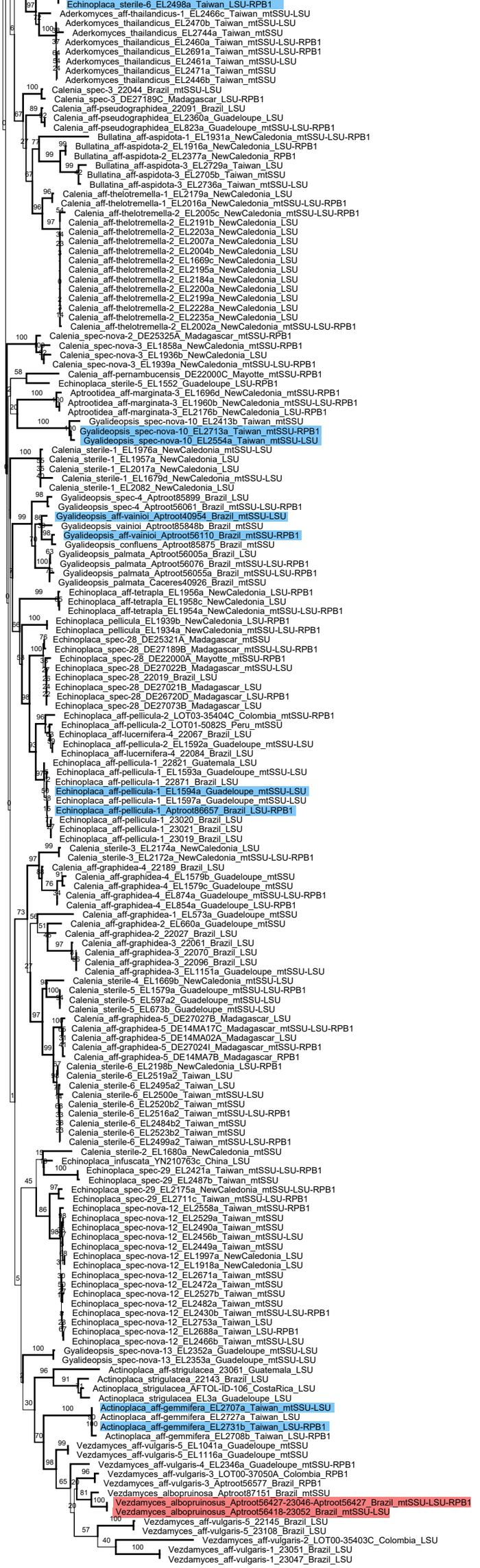

Fig. S1A. Phylogenetic tree on the complete dataset inferred by maximum likelihood analyses of the mtSSU, nuLSU and RPB1 region.
Specimens highlighted in red indicate the placement of **the 9 concatened specimens** for species delimitation. Specimens marked in blue represent those concatenated after species delimitation, forming subset 1 used for BioGeoBEARS analyses.