# ORIGINAL

# RESEARCH Storage stability of traditional Tunisian butter oil produced from spontaneous fermentation of cow's milk

# BALI OLFA, MOHAMED ALI AYADI\* and HAMADI ATTIA

Département de biologie, Unité Analyses Alimentaires, Ecole Nationale d'Ingénieurs de Sfax, Route de Soukra, 3038 Sfax, Tunisia

Physicochemical characteristics of traditional Tunisian butter (TTB) and thermal stability of the butter oil were studied. Thermal stability of samples was estimated by using the accelerated shelf life testing method. Effect of heating on some quality characteristics of traditional Tunisian butter oil (TTBO) was investigated and compared at different temperatures (60 $^{\circ}$ C, 100 $^{\circ}$ C and 130 $^{\circ}$ C). Induction period of sample heated at  $60^{\circ}$ C was important compared to that at  $100^{\circ}$ C and  $130^{\circ}$ C. This result may indicate the sensitivity of TTBO to elevated temperature. Absorption at 232 and 270 nm, acidity and PV increased rapidly after reaching the oxidation induction time. The temperature had a significant effect on the formation of oxidation products in traditional butter oil (TTBO). Fatty acid composition of TTBO was also changed after heating. This may explain the observed modification on some quality characteristics of TTBO such as viscosity, hardness, cohesiveness and colour. As a consequence, heat treatment produced alterations in the oxidative status of the butter oil that could affect the shelf life.

Keywords Physicochemical composition, Traditional Tunisian butter, Butter oil, Thermal stability.

#### INTRODUCTION

Lactic acid fermentation, as a means of food preservation, is probably one of the oldest biotechnological processes rooted in the cultural history of mankind (Tamang et al. 2005). Traditional fermentation of milk remains one of the most practical and widely applied empirical methods for preserving and often enhancing the organoleptic and nutritional quality of milk. A variety of fermented milk products are prepared in Tunisia, mostly by rural women using their traditional knowledge of fermentation for the bio-preservation of milk for storage and future consumption. Two products result from the spontaneous fermentation of milk in Tunisia: the first is rich in proteins and locally called 'Leben'. The second is rich in fat and is locally called 'traditional butter' or 'Zebda beldi'. Traditional Tunisian Butter (TTB) is widely produced and consumed, and is still a popular Tunisian product. Its characteristic aroma plays an important role in its popularity. According to De Noni and Resmini (2005), the term 'traditional' may be used, together with the name 'butter', provided the cream is obtained directly from milk by either centrifugation or natural creaming. Churning is the most important step for butter-making, during which the oil-in water emulsion is broken, leading to the formation of water-in oil emulsion (Rousseau 2000). Butter oil is obtained from traditional butter after the separation of the fat from the milk serum by melting or heating. A major portion of butter oil is utilised for culinary cooking and for the frying of different foods. Deterioration (lipolysis and oxidation) of milk fat, due to several factors, causes flavour impairment, lowers nutritional quality, and creates serious problems for storage stability (Özkanli and Kaya 2005). Oxidation has a significant financial impact due to the development of positive flavours as well as rancid flavours that reduces the sensory characteristics, deteriorates essential fatty acids and heat-sensitive vitamins, and forms oxidised products which may be hazardous to health. Several studies have been carried out to determine the effects of cooking (e.g. method, temperature, time and end-point temperature) on lipid oxidation (Varela 1998; Romero et al. 2000; Özkanli and Kaya 2005; Ramírez and Cava 2005), High cooking temperatures, above 100°C, have been reported to reduce the development of lipid oxidation (Spanier et al. 1998). This effect has been attributed to the antioxidant action of Maillard reaction products (MRPs) that is induced at elevated cooking temperatures (Bailey 1998). Additionally, the frying process causes modifications in the colour of food due to the formation of low molecular weight chromophores closely related to MRPs (Hofmann 1998). The onset of rancidity in butter may be due to the oxidation of unsaturated glycerides leading to the development of peroxides and/or due to hydrolysis of glycerides resulting in increased levels of free

\*Author for correspondence. E-mail: ayadimedali@ yahoo.fr

© 2009 Society of Dairy Technology

International Journal of  $\overline{{\it D} a iry~T}echnology$ 

fatty acids (Amr 1991; Joshi and Thakar 1994). It has been reported that, both storage time and type of treatment have highly significant effects on the peroxide value and free fatty acid content of anhydrous butter oil (Amr 1991).

The aim of this paper was to determine the physicochemical properties of TTB and to evaluate heat treatment effects on oxidation indicators to determine thresholds for each heat treatment.

## MATER IALS AND METHODS

#### Raw butter production

Fresh raw cows' milk (Holstein breed) used in this study was obtained from a local farm in the Southern of Tunisia. Cows' milk was collected, kept at  $\approx$ 4°C and transported to our laboratory within 24 h.

Raw milk was left at room temperature in a jar  $(25 \pm 2^{\circ}\text{C})$  until coagulation occurred  $(\sim)18$  hours). Microorganisms growing in the raw milk during fermentation steps depend on the environmental conditions (result not published). On gelation, the product is called 'rayeb'. By churning the 'rayeb' is separated into an aqueous fraction (protein, lactose, mineral…) called 'Leben' and raw butter called 'Zebda beldi'. Traditionally, churning, takes place in a goat skin bag called 'Checoua'. The churning is achieved by hanging the Checoua filled with 'rayeb' and vigorously shaking it back and forth until fluidisation of the contents and coalescence of the fat globules. The end of churning is discerned by the sound of the butter lumps when shaking. Warm water  $(40^{\circ} \text{ C},$  $10\%$  v/v) was added at the beginning of the churning period to enhance the coalescing of the fat globules and, thus, to increase the yield of butter (zebda beldi).

For this work triplicate jars were used to perform fermentation steps (about 15 L of milk in each jar) and a goat skin bag (traditional checoua) was used to perform churning process. Duplicate analyses were performed on each replicate.

#### Raw butter analysis

Total nitrogen (TN) was achieved using the Kjeldahl method (AFNOR 1993) with a Büchi 325 apparatus (Büchi, Flawil, Switzerland). Dry matter, ash, lactose and fat contents were determined according to standard methods (AFNOR 1993). NaCl (chloride content) was measured using the International Dairy Federation method (IDF 1969). Titrable acidity was determined by titration of sample with  $N/9$  sodium hydroxide to pink endpoint using phenolphthalein as indicator and pH was determined using a pH metre (744 pH Meter Metrohm) (AFNOR 1993).

Traditional Tunisian butter (TTB) samples were placed in glass jars open to the atmosphere in temperature controlled ovens at  $60^{\circ}$ C,  $100^{\circ}$ C and 130°C (Memmert-GmbH+Co.KG, Germany) to perform stability tests. Instantaneous separation occurs (density phenomena) in heated TTB sample leading to two phases: an aqueous phase [containing protein, minerals, lactose…) and an oil phase (traditional Tunisian butter oil (TTBO)]. In addition, TTBO was thoroughly separated by centrifugation and filtration. Samples of TTBO was collected and used to perform heat stability. Peroxide value (PV), free fatty acid (FFA), coefficient extinctions, colour, viscosity, texture and fatty acid composition were measured at different periods of heating to evaluate the deterioration state of traditional Tunisian butter oil (TTBO).

AOCS (1997) official methods were used for the determination of the peroxide and acidity values (method number Cd 8-53 and Cd 3d-63, respectively).

 $\varepsilon_{232}$  and  $\varepsilon_{270}$  extinction coefficients were calculated from absorption at the corresponding waves length, with an UV spectrophotometre (UV mini 1240, UV-Vis SPECTROPHOTOMETER, Schimadzu, JAPAN), using a 1% solution of oil in cyclohexane and a path length of 1 cm.

The CieLab coordinates  $(L^*, a^*, b^*)$  were directly read with a spectrophotometre MS⁄Y-2500 (Hunterlab, In., Reston, VA, USA), calibrated with a white tile. Under the tristimulus colour coordinate system, the L\* value is a measure of lightness and varies from  $-100$  (black) to 100 (white), the a\* value varies from  $-100$  (green) to  $+100$  (red), and the b<sup>\*</sup> value varies from  $-100$  (blue) to  $+100$  (yellow).

Viscosity of the butter oil samples was followed at 37C with a Stress Tech Rheologica Rheometre (Rheologica Instruments AB, Lund, Sweden) conducted with a steel cone-plate (C40/4) under a constant shear rate of  $100/s$ .

Textural analyses were performed using uniaxial compression test (Texture analyser: LLOYD instruments, England). For each temperature ( $60^{\circ}$ C,  $100^{\circ}$ C and  $130^{\circ}$ C) three samples of TTBO were taken at different heating time and stored at 4C. All textural analyses were performed at solidified TTBO and at 4°C. A Textural Procedure Analysis was performed under the following conditions: cylindrical probe (diametre =  $1 \text{ cm}^2$ ), compression speed of 40 mm⁄min and a compression rate of 2 cm. textural test were performed at triplicate.

Fatty acid composition was performed using Gas Chromatography (GC). TTBO samples were warmed to  $37^{\circ}$ C immediately prior to analysis, then vortexed vigorously to achieve sample uniformity. Fatty acid methyl esters (FAMES) were prepared according to Maxwell and Warner (1983). Fifty microlitre of the butter oil were converted to methyl esters using  $2 \text{ mL}$  hexane and  $200 \mu\text{L}$  of 2 N KOH in methanol. Aqueous fraction were recuperated and mixed with 200 µL of sodium acetate solution. After centrifugation, organic fraction was washed with 0.5 mL of distilled water. Fatty acid methyl esters (FAMES) were presented in the organic phase for GC analyses.

Gas Chromatography analyses were performed on a Shimadzu, GC 17 A chromatograph, equipped with a flame hydrogen ionisation detector and a capillary column (FFAD, 50 m  $\times$  0.32 mm  $\times$ 0.5 µm, PERICHROM Sarl, France). The oven temperature was programmed as follow: the initial temperature (100 $^{\circ}$ C) was raised to 150 $^{\circ}$ C at a rate of 30C⁄min and held at this temperature for 5 min, then increasing at  $10^{\circ}$ C/min to 190 $^{\circ}$ C and held at this temperature for 14 min, and then increasing at  $5^{\circ}$ C/min to  $255^{\circ}$ C and held at this temperature for 10 min. The injector and detector temperatures were  $255^{\circ}$ C and  $270^{\circ}$ C, respectively. Nitrogen was the carrier gas. The identification of the peaks was achieved by retention times and by comparing them with authentic standards analysed under the same conditions. Peak areas of triplicate injections were measured with a HP computing integrator. Results were expressed as  $w/w$  (%) total fatty acid (Sağdiç et al. 2004).

#### Statistical analysis

Values of different tests were expressed as the mean  $\pm$  standard deviation (x  $\pm$  SD). SPSS packet program for Windows was used for the statistical analysis. Significant differences between mean  $(P < 0.05)$  were determined by using a one-way ANOVA (Duncan's test).

# RESULTS AND DISCUSSION

For the production of traditional butter, the physicochemical characteristics of fresh raw cow's milk samples obtained from local farm are presented in Table 1. The fresh raw milk sample gave suitable technological properties.



All values given are means of three determinations.

## Physical and chemical composition

Table 1 shows the physicochemical properties of the traditional Tunisian butter (TTB). The fat content of the TTB  $(65.70 \pm 2.16\%)$  was lower compared to the content reported with most common butters. This result could be explained by the low dry matter content in TTB  $(70.60 \pm 0.15\%)$ . In deed, the traditional churning process (with 'chekwa') doesn't allow a good separation performance. In addition, mean Chloride content was less than those reported by Hayaloğlu (1999) for other butters. Mean pH value (4.7) and titratable acidity value  $(22.5°D)$  were similar to those reported by Filkensen (1987) and Sağdiç et al. (2004) respectively.

#### Heat stability

TTBO oxidation was followed by testing a TTBO sample at regular intervals using PV,  $\varepsilon_{232}$  and  $\varepsilon_{270}$ extinction coefficients, acidity, viscosity, texture, colour values determinations and fatty acids composition.

Figure 1 shows the change in TTBO PV values at  $60^{\circ}$ C,  $100^{\circ}$ C and  $130^{\circ}$ C. Hydroperoxide is the primary product of lipid oxidation; therefore, the determination of the peroxide value can be used as an oxidative index for the early stage of lipid oxidation (Ramadan and Mörel 2004). The initial stage of slow oxidation (the induction period) can be measured as the time required to reach an end point of oxidation corresponding either to a detectable level of rancidity (a defined peroxide value) or to a sudden change in oxidation rate (Fearon et al. 1998). The peroxide values obtained for TTBO stored at 60°C (Figure 1a) proceeded at a lower rate initially. This period of time is called the induction period (IP) or induction time (IT) (Nissiotis and Tasioula-Margari 2002). After that, the oxidation rate increases considerably. Absence of induction time and an increase of the slope were observed during heating at  $100^{\circ}$ C and  $130^{\circ}$ C (Figure 1b). Temperature had a significant effect on the formation of oxidation products in TTBO, with higher PV levels obtained at higher temperatures (Figure 1b) and low PV levels obtained at lower temperature (Figure 1a). Indeed, the induction period is 14 days for TTBO stored at  $60^{\circ}$ C with the peroxide values reaching approximately 1.495 mequiv. $O_2$  *Kg fat. However, for higher* temperature (100 $^{\circ}$ C and 130 $^{\circ}$ C), absence of induction period was observed. The peroxide values of TTBO at  $100^{\circ}$ C and  $130^{\circ}$ C increased gradually during the whole storage period. After 8 hours  $(0.3 \text{ day})$  of heating at  $100^{\circ}$ C the measured value was  $4.45$  mequiv.  $O_2/Kg$  fat. In contrast, 4.95 mequiv. $O_2$ /kg fat was identified after 2 hours of heating at 130°C. Consequently, TTBO shows the highest stability at lower heating temperature. Ndjouenkeu and Ngassoum (2002) reported that



Figure 1 Peroxide value of TTBO during storage: (a) at  $60^{\circ}$ C; (b) at  $100^{\circ}$ C and  $130^{\circ}$ C.

PV values must normally increase during heating because of the oxidative activation. This result could be explained by the fact that reactions and mechanisms taking place in the degradation of TTBO was dependant on the heating treatment. Gertz et al. (2000) studied the oxidative stability of fats and oils heated at frying temperature  $(170^{\circ}C)$ and reported that for temperatures  $\leq 120^{\circ}$ C, main oxidation reactions are: either hydrolysis or oxidation induced by moisture and atmospheric oxygen. Whereas, for temperature above  $120^{\circ}$ C, the most obvious and important reactions consisted in polymerisation. Whereas, Özkanli and Kaya (2005) studied the stability of pasteurised and nonpasteurised butter oil, and reported that stability of pasteurised butter oil can be attributed to pasteurisation. Thus, through pasteurisation and heat treatment of milk, microorganisms and enzyme activities, which could initiate lipid oxidation, are eliminated. Whereas heat treatment was not applied to butter oil produced from nonpasteurised milk in the melting process. This result leads the authors to suppose that butter oil is more susceptible to enzyme oxidation.

It can be seen that the traditional method for determining PV serves as an indicator of butter oil quality does not distinguish between various unsaturated fatty acids that undergo oxidation; It also does not supply information about the secondary oxidation products formed by hydro peroxide decomposition. However, it can generally be stated that the PV is an indicator of the primary level of oxidation.

The formation of hydro peroxides is accompanied by the generation of conjugated compound, measured by absorption at a wavelength of 232- 234 nm (Guillén and Ruiz 2004). The hydro peroxide and the conjugated compound reflect the degree of primary products formation during lipid oxidation (Guillén and Ruiz 2004). Figure 2 illustrates evolution of absorption at 232 nm during the storage period at different temperatures:  $60^{\circ}$ C (Figure 2a),  $100^{\circ}$ C and  $130^{\circ}$ C (Figure 2b). Absortivity at 232 nm of TTBO at different storage temperatures evolves in the same way. The primary products of lipid oxidation of TTBO heated at  $130^{\circ}$ C shows a higher absorption at 232 nm compared to samples heated at  $60^{\circ}$ C and  $100^{\circ}$ C. This



Figure 2 Absorptivity at 232 nm of TTBO during storage: (a) at  $60^{\circ}$ C; (b) at  $100^{\circ}$ C and  $130^{\circ}$ C.



Figure 3 Absorptivity at 270 nm of TTBO during storage: (a) at  $60^{\circ}$ C, (b) at  $100^{\circ}$ C and  $130^{\circ}$ C.

result could explain the higher sensitivity of TTBO to temperature during storage, leading to a higher rate of the oxidation primary products content. The coincidence in the changes in absorption at 232 nm with hydro peroxide formation was observed. As can be seen, absorption at 232 nm increased immediately for samples heated at  $130^{\circ}$ C (Figure 2b). However, induction period was more important for samples stored at  $60^{\circ}$ C (Figure 2a) and  $100^{\circ}$ C (Figure 2b).

The primary products of oxidation are not stable and their degradation could lead to formation of oxidation secondary product with absorption occurring at 270 nm (Vieira and Regitano d'Arce 2001). These secondary products have a role in the break-up of the acyl group chains as was suggested by Guillén and Ruiz (2004). Figure 3 shows that specific extinction at 270 nm increased during heating storage at different temperature. It can be seen clearly that secondary compounds of oxidation were detected before reaching the oxidation induction time for the TTBO.

Figure 4 illustrate evolution of TTBO acidity vs storage time at  $60^{\circ}$ C,  $100^{\circ}$ C and  $130^{\circ}$ C. Acid value of TTBO remains relatively lower and constant (stable) during 14 days of heating at  $60^{\circ}$ C (Figure 4a). This means that the TTBO hydrolysis is not sufficient to compensate or to even increase the acid functions blocked by polymerisation (Ndjouenkeu and Ngassoum 2002). Acidity of samples heated at  $60^{\circ}$ C exhibit a low increasing. However during heating treatment at  $100^{\circ}$ C and 130°C, acidity of TTBO increases immediately. In fact, acidity reaches 2.8 after 30 days of heating at  $60^{\circ}$ C, 2.79 after only 2 days heating at 100 $^{\circ}$ C and 2.9 after 5 hours heating at  $130^{\circ}$ C. The acid values increased as the storage time and temperature increased. This could be attributed to the initiation of (Tunisian traditional butter oil) TTBO degradation with a high heating treatment. Indeed, butter oil undergoes lipolysis releasing free fatty acids and increasing acidity.

Table 2 presents free fatty acid profiles of TTBO at different time of heating at  $60^{\circ}$ C,  $100^{\circ}$ C and 130°C. Nontreated TTBO were characterised by the presence of four major fatty acids [palmitic  $(C_{16.0})$ , stearic  $(C_{18.0})$ , myristic  $(C_{14.0})$ , and oleic  $(C_{18:1})$  acids]. Palmitic acid was the major fatty



Figure 4 Acidity of TTBO during storage: at 60°C, (b) at 100°C and 130°C.

Table 2 Fatty acid composition (%) of traditional Tunisian butter oil produced from spontaneous fermentation of cows' milk and stored at different temperature (60 $\degree$ C, 100 $\degree$ C and 130 $\degree$ C)

		After	After 30	After 3 hours	After 720 hours	After 60 hours	After 6 hours
Fatty	TTBO before	360 hours	hours of heating	of heating	of heating	of heating	of heating
acid	heating	of heating at $60^{\circ}$ C	at $100^{\circ}C$	at $130^{\circ}$ C	at $60^{\circ}C$	at $100^{\circ}$ C	at $130^{\circ}C$
$C_{4:0}$	$2.01 \pm 0.20^a$	$1.98 \pm 0.20^{\circ}$	$2.30 \pm 0.13^a$	$2.11 \pm 0.15^a$	$1.98 \pm 0.13^{\text{a}}$	$2.02 \pm 0.21^{\text{a}}$	$2.31 \pm 0.18^a$
$C_{6:0}$	$1.20 \pm 0.11^a$	$1.22 \pm 0.12^a$	$1.41 \pm 0.17^b$	$1.36 \pm 0.14^b$	$1.21 \pm 0.11^a$	$1.27 \pm 0.12^a$	$1.46 \pm 0.15^b$
$C_{8:0}$	$2.80 \pm 0.22^{\text{a}}$	$2.93 \pm 0.21^a$	$3.26 \pm 0.14^a$	$3.30 \pm 0.19^a$	$2.90 \pm 0.12^a$	$3.03 \pm 0.23^{\text{a}}$	$3.41 \pm 0.11^a$
$C_{10:0}$	$0.31 \pm 0.13^{\text{a}}$	$0.31 \pm 0.24^{\text{a}}$	$0.33 \pm 0.22^{\text{a}}$	$0.34 \pm 0.16^a$	$0.28 \pm 0.14^a$	$0.32 \pm 0.22^{\text{a}}$	$0.34 \pm 0.21^a$
$C_{12:0}$	$3.30 \pm 0.23^{\text{a}}$	$3.53 \pm 0.21^{\circ}$	$3.79 \pm 0.22^{\rm a}$	$3.96 \pm 0.23^{\text{a}}$	$3.44 \pm 0.25^{\text{a}}$	$3.60 \pm 0.23^{\text{a}}$	$4.00 \pm 0.24^b$
$C_{14:0}$	$11.38 \pm 0.40^a$	$12.53 \pm 0.33^b$	$12.83 \pm 0.32^{b,c}$	$12.98 \pm 0.36^b$	$11.74 \pm 0.28^b$	$12.70 \pm 0.31^b$	$13.05 \pm 0.30^{\circ}$
$C_{14:1}$	$1.14 \pm 0.10^a$	$1.21 \pm 0.11^a$	$1.21 \pm 0.10^a$	$1.21 \pm 0.13^a$	$1.08 \pm 0.14^a$	$1.18 \pm 0.11^a$	$1.19 \pm 0.12^a$
$C_{16:0}$	$32.04 \pm 0.61^a$	$36.31 \pm 0.65^{\rm b}$	$34.37 \pm 0.60^b$	$34.7 \pm 0.61^{\rm b}$	$34.61 \pm 0.59^b$	$34.62 \pm 0.57^b$	$34.86 \pm 0.61^b$
$C_{16:1}$	$1.92 \pm 0.09^{\rm a}$	$2.15 \pm 0.10^a$	$2.08 \pm 0.12^a$	$2.00 \pm 0.11^a$	$2.28 \pm 0.14^a$	$2.10 \pm 0.10^a$	$2.01 \pm 0.11^a$
$C_{18:0}$	$18.80 \pm 0.54$ <sup>a</sup>	$9.06 \pm 0.56^b$	$9.92 \pm 0.49^b$	$9.31 \pm 0.43^b$	$9.82 \pm 0.40^b$	$9.86 \pm 0.39^b$	$9.35 \pm 0.51^{\rm b}$
$C_{18:1}$	$21.62 \pm 0.42^a$	$24.73 \pm 0.43^b$	$24.58 \pm 0.46^b$	$24.33 \pm 0.40^b$	$26.88 \pm 0.41^b$	$25.17 \pm 0.42^b$	$24.14 \pm 0.40^b$
$C_{18:2}$	$2.41 \pm 0.10^a$	$2.86 \pm 0.11^a$	$2.82 \pm 0.14^a$	$3.31 \pm 0.20^{\rm a}$	$2.51 \pm 0.11^a$	$3.06 \pm 0.14^a$	$2.76 \pm 0.10^8$
Total	98.93	98.82	98.90	98.92	98.73	98.93	98.88
Saturated (S)	71.84	67.87	68.21	68.06	65.98	67.42	68.78
Unsaturated $(U)$	27.09	30.95	30.69	30.85	32.75	31.51	30.10
U/S	0.38	0.46	0.45	0.45	0.49	0.47	0.44

Data are the mean of three measurements. Different letters in the same line indicate significant differences ( $P < 0.05$ ).

acid found (32.04%) in TTBO. These results are in agreement with previous studies on different butter oil produced from cow's milk. Indeed, Glew et al. (1999) reported that saturated fatty acid level in 'Fulani butter oil' made from cow's milk was 53.3% and the major fatty acid was palmitic acid (30.2%). Whereas, percentage of saturated fatty acid in traditional Turkish butter was 67.06% and palmitic acid was the major one  $(33.72%)$  (Sagdic et al. 2004). In addition, Fatouh et al. (2007) reported that saturated fatty acid rate in butter oil made from buffalo milk was 70.72% and palmitic acid was also the major fatty acid (31.89%).

Butter oil made from milk of other animal species was studies (Sağdiç et al. 2004; Özkanlı and Kaya 2005). Özkanli and Kaya (2005) found a lower saturated fatty acid level (59.13%) and the major fatty acid was oleic acid (31.08%) in butter produced from sheep's milk. Sağdiç et al. (2004) reported that percentage of saturated fatty acid was 73.88% and 69.10% in butter oil made from Goats' and Ewes' milk, respectively. These authors reported too that palmitic acid was the major one for the two butter oils.

Milk fatty acid composition depends on several factors as animal species and feed, climate and the environmental conditions. TTBO showed higher saturated fatty acid content (especially stearic acid) than butter oil produced from sheep's milk (Özkanlı and Kaya 2005).

Changes were observed in fatty acid composition of traditional Tunisian butter oil (TTBO) during heat treatment. Indeed, a decrease in the relative percentages (%) of stearic acid ( $\sim$ 50%) and an increase in the percentage of palmitic  $(C_{16:0})$ , myristic  $(C_{14:0})$  and oleic  $(C_{18:1})$  acids were detected. These changes in the fatty acid composition could be attributed to the degradation of fat matter under heating and oxidation. Özkanli and Kaya (2005) reported that fatty acid composition of oil butter made from sheeps' milk varies slightly under heating treatment. Ozkanli and Kaya (2005) attribute this stability to temperature effect which inhibits lipolitic activity.

CieLab coordinates (L\*, a\*, b\*) of the TTBO during oxidation were given in Figure 5. The initial colour  $(t = 0$  h) of TTBO was reported to be yellow, and this is due to its richness in yellow pigments (carotenoids). Figures 5a,b show that heating gives immediately a considerable increase in L<sup>\*</sup> parameter at a rate of  $\sim$  50% from the initial value. Furthermore, b\* (Figures 5c,d) values increase at the beginning of the heating at a rate of  $\sim 60\%$  from the initial rate, and then decrease during treatment. However, what was observed for a\* (Figures 5e,f) parameter was different: a decrease in values was observed at the beginning of heating, then values remained practically constant and finally increased considerably at the end of the treatment. This colour change (darkness and green-brown colour) was essentially marked by the high loss of yellow colour and then of yellow pigments, essentially  $\beta$ -carotenes, beyond the oxidation induction period. On the other hand, the colour formation in traditional Tunisian butter oil (TTBO), during heating processes, could be attributed to both nonenzymatic browning and phospholipids degradation during heating (Husain



Figure 5 CieLab coordinates (L\*, a\*, b\*) of TTBO during storage: (a) L\* at  $60^{\circ}C$ , (b) L\* at  $100^{\circ}C$  and  $130^{\circ}C$ , (c) b\* at  $60^{\circ}C$ , (d) b\* at  $100^{\circ}$ C and  $130^{\circ}$ C, (e) a\* at  $60^{\circ}$ C, (f) a\* at  $100^{\circ}$ C and  $130^{\circ}$ C.

et al. 1986). Nonenzymatic browning is favoured by heat treatment and includes a wide number of reactions such as Maillard reaction, caramilisation and chemical oxidation of phenols (Manzocco et al. 2001). TTBO could contain compound which would result in formation of Maillard reaction products (MRPs). In fact, TTBO could contain a trace of lactose and milk protein depends on the separation operation between aqueous and oil phases. Thus, colour formation during heat treatment is partly due to the formation of coloured MRPs. These MRPs correspond to the compound of low-molecular-weight and melanoidins with high-molecular-weight (Ames 1992).

At  $60^{\circ}$ C and  $100^{\circ}$ C, L<sup>\*</sup> and b<sup>\*</sup> parameters remain constant at the beginning of the experiment. This result, not being the case for a temperature of 130°C, could be explained by the effect of the oxidation reaction, induced essentially by atmospheric oxygen at lower heat temperature  $(60^{\circ}$ C and  $100^{\circ}$ C). Whereas for temperature above  $120^{\circ}$ C, oxidative colour change could be essentially attributed to the polymerisation reactions induced by high treatment temperature.

Table 3 shows viscosity value, cohesiveness, springiness and hardness of TTBO after different time of heating at  $60^{\circ}$ C,  $100^{\circ}$ C and  $130^{\circ}$ C. The low initial viscosity of TTBO (5.85 mPa.s) could

 14710307, 2010, 1, Downloaded from https://onlinelibrary.wiley.com/doi/10.1111/j.1471-0307.2009.00537.x by Université De Liège, Wiley Online Library on [16/12/2024]. See the Terms and Conditions (https://onlinelibrary.wiley.com/terms-and-conditions) on Wiley Online Library for rules of use; OA articles are governed by the applicable Creative Commons License1470300, 1, Downloaded from https://om/doi/0.1111/j.1471-0307200900337.x by Université De Lièray on [16122024]. See the Tems and Conditions (https://online Library on [16122024]. See the Tems and Conditions (https://online

mentation of cows' milk and stored at different temperature (60 $^{\circ}$ C, 100 $^{\circ}$ C and 130 $^{\circ}$ C)					Table 3 Viscosity, cohesiveness, springiness, adhesiveness and firmness coefficients of Tunisian traditional butter oil produced from spontaneous fer-		
Textural parameters	TTBO before heating	After 360 hours of heating at $60^{\circ}C$	After 30 hours of heating at $100^{\circ}C$	After 3 hours of heating at $130^{\circ}$ C	After 720 hours of heating at $60^{\circ}$ C	After 60 hours of heating at $100^{\circ}C$	After 6 hours of heating at $130^{\circ}C$
Viscosity (mPa/s) Cohesiveness Springiness (mm) Hardness (N)	$5.85 \pm 0.90^{\rm a}$ $0.13 \pm 0.10^a$ $4.99 \pm 0.22^{\text{a}}$ $101.87 \pm 5.12^{\text{a}}$	$2.07 \pm 0.60^{\rm b}$ $0.21 \pm 0.12^b$ $4.93 \pm 0.33^{\text{a}}$ $63.28 \pm 2.15^b$	$1.19 \pm 0.40^b$ $0.21 \pm 0.22^b$ $4.90 \pm 0.32^{\text{a}}$ $47.29 \pm 4.10^b$	$9.22 \pm 1.2^{\circ}$ $0.29 \pm 0.21^b$ $4.98 \pm 0.33^{\text{a}}$ $175.47 \pm 6.22^{\circ}$	$1.2 \pm 0.31^{\rm b}$ $0.25 \pm 0.24^b$ $4.96 \pm 0.31^{\text{a}}$ $56.39 \pm 4.11^b$	$1.12 \pm 0.10^b$ $0.28 \pm 0.13^b$ $4.88 \pm 0.32^{\rm a}$ $46.99 \pm 4.50^b$	$6.33 \pm 0.85^{a,c}$ $0.25 \pm 0.12^b$ $4.97 \pm 0.35^{\text{a}}$ $42.96 \pm 3.81^b$
					Data are the mean of three measurements. Different letters in the same line indicate significant differences ( $P < 0.05$ ).		
		be explained by the relatively level count of mono- unsaturated and polyunsaturated fatty acids as pre- viously shown (Table 2) and by the presence of high medium and short-chain fatty acids content. These results reinforce the finding of Geller and Goodrum (2000), concerning the existence of a strong relationship between fatty acid chain length and viscosity. During heat stability tests, viscosity			induction period was 14 days for samples stored $60^{\circ}$ C and absence of this induction period for sample. ples stored at 100°C and 130°C. Change in colo parameters observed depending on storage temperature Indeed, TTBO stored at 130°C exhibit a change colour parameters and in viscosity and hardne values.	and texture	characteristics we

Table 3 Viscosity, cohesiveness, springiness, adhesiveness and firmness coefficients of Tunisian traditional butter oil produced from spontaneous fermentation of cows

and viscosity. During heat stability tests, viscosity and hardness of TTBO decreases, then remains constant till the end of the oxidation process, except for the sample stored at  $130^{\circ}$ C for about 3 hours. In addition, during storage cohesiveness increases then remains constant till the end of oxidation process. Initially, butter seemed to be harder and then became softer after heat treatment at  $60^{\circ}$ C and 100°C. These results could be explained by the change in the fatty acid composition. Indeed, the decrease of saturated fatty rate was compensated by the increase of the unsaturated one during the heat stability experiments (Table 2).

An increase in viscosity and hardness value of traditional butter was observed after 3 hours heating at 130°C. This significant increase could be attributed to the polymerisation reaction and the formation of high-molecular-weight compounds including carbon–carbon bonds and carbon–oxygen–carbon bridges between fatty acids (Stevenson et al. 1984).

Spreadability of the butter could be evaluated by both cohesiveness and hardness values. However springiness is a characteristic for the materials' elasticity. Table 3 shows that springiness' value during heat stability test, remains constant for all storage temperatures. These results could be explained by the absence of milk proteins, which are responsible of the elasticity characteristic, in dairy products (Sandoval-Castilla et al. 2004).

# CONCLUSION

To conclude, the present study has allowed physico-chemical characterisation and heat stability evaluation of Tunisian traditional butter (TTB). Fatty acids compositions were also determined. Predominant fatty acids in Tunisian traditional butter oil (TTBO) are: myristic, palmitic and oleic acids. TTBO Heat stability tests show that

vas 14 days for samples stored at of this induction period for sam-<sup>o</sup>C and 130<sup>o</sup>C. Change in colour texture characteristics were ding on storage temperature.  $I$  at 130 $^{\circ}$ C exhibit a change in and in viscosity and hardness values.

 14710307, 2010, 1, Downloaded from https://onlinelibrary.wiley.com/doi/10.1111/j.1471-0307.2009.00537.x by Université De Liège, Wiley Online Library on [16/12/2024]. See the Terms and Conditions (https://onlinelibrary.wiley.com/terms-and-conditions) on Wiley Online Library for rules of use; OA articles are governed by the applicable Creative Commons License1470300, 1, Downloaded from https://wipy.com/doi/0.1111/j.1471-03072000037.x by Universite De Liberay willey Online Liberay on [16/122024]. See the Temas and Conditions (https://online Liberay on [16/122024]. See the Temas

# REFERENCES

- AFNOR. (1993) Contrôle de la qualité des produits alimentaires. In Lait et produits laitiers. Paris: Association Francaise de Normalisation.
- Ames J M (1992) The maillard reaction. In Biochemistry of Food Proteins, pp 99–153. Hudson B J F, ed. London: Elsevier Applied Science.
- Amr S A (1991) Effectiveness of synthetic and potential natural antioxidants in improving the stability of sheep's anhydrous butter fat during long-term storage. Journal of the Science of Food and Agriculture 55 75–85.
- AOCS. (1997) Official Methods and Recommended Practices of the American Oil Chemists' Society, 5th ed. Champaign, IL: AOCS Press.
- Bailey M E (1998) Inhibition of warmed-over flavour with emphasis on maillard reaction products. Food Technology 42 123–126.
- De Noni D I and Resmini P (2005) Identification of rennetwhey solids in 'traditional butter' by means of HPLC⁄ ESI-MS of non-glycosylated caseinomacropeptide A. Food Chemistry 93 65–72.
- Fatouh A E, Mahran G A, El Gandour M A and Singh R K (2007) Fractionation of buffalo butter oil by supercritical carbon dioxide. LWT – Food Science and Technology 40 1687–1693.
- Fearon A M, Mayne C S and Charlton C T (1998) Effect of naked oats in the cow's diet on the oxidative stability of the milk fat. Journal of the Science of Food and Agriculture 76 546–552.
- Filkensen W E (1987) Production proportions and product quality. Nordisk Mejeriindustri 14 414–416.
- Geller D P and Goodrum J W (2000) Rheology of vegetable oil analogs and triglycerides. Journal of the American Oil Chemists' Society 77 111–114.
- Gertz C, Klostermann S and Kochhar S P (2000) Testing and comparing oxidative stability of vegetable oils and fats at frying temperature. European Journal of Lipid Science and Technology 102 543–552.
- Glew R H, Okolo S N, Chuang Lu T Huang Y S and Vander-Jagt D J (1999) Fatty acid composition of Fulani butter oil made from cow's milk. Journal of Food composition and Analysis 12 235–240.
- Guillén M D and Ruiz A (2004) Formation of hydroperoxyand hydroxyalkenals during thermal oxidative degradation of sesame oil monitored by proton NMR. Journal of Lipid Science and Technology 106 680-687.
- Hayaloğlu A A (1999) A comparative study on physicochemical, microbiological and organoleptic qualities of butter produced from cream and yoghurt in Malatya region. PhD Thesis. Institute of Natural and Applied Sciences, Cukurova University, Adana, Turkey.
- Hofmann T (1998) Characterization of the chemical structure of novel coloured maillard reaction products from furan-2-carbox-aldehyde and amino acids. Journal of Agriculture and Food Chemistry 46 932–940.
- Husain S R, Terao J and Matsushita S (1986) Effect of browning reaction products of phospholipids on autoxidation of methyl linoleate. Journal of the American oil Chemists' Society 63 1457–1560.
- IDF. (1969) Standard 12A Determination of Salt (sodium Chloride) Content Butter. Brussels, Belgium: International Dairy Federation.
- Joshi N S and Thakar P N (1994) Methods to evaluate deterioration of milk fat – a critical appraisal. Journal of Food Science & Technology 31 181–196.
- Manzocco L, Calligaris S, Mastrocola D, Nicoli M C and Lerici C R (2001) Review of nonenzymatic browning and antioxidant capacity in processed foods. Trends in Food Science & Technology 11 340–346.
- Maxwell R J and Warner W N (1983) Fatty acid analysis: phospholipids-rich analysis. Lipids 18 453–459.
- Ndjouenkeu R and Ngassoum M (2002) Etude comparative de la valeur en friture de quelques huiles végétales (Comparative study of frying behaviour of some vegetable oils). Journal of Food Engineering 52 121–125.
- Nissiotis M and Tasioula-Margari M (2002) Changes in antioxidant concentration of virgin olive oil during thermal oxidation. Food Chemistry 77 371–376.
- Özkanli O and Kaya A (2005) Storage stability of butter oils produced from sheep's non-pasteurized and pasteurised milk. Food Chemistry 100 1026–1031.
- Ramadan M F and Mörel J-T (2004) Oxidative stability of black cumin (Nigella sativa L.), coriander (Coriandrum sativum L.) and niger (Guizotia abyssinica Cass.) crude seed oils upon stripping. European Journal of Lipid Science and Technology 106 35–43.
- Ramírez M R and Cava R (2005) Changes in colour, lipid oxidation and fatty acid composition of pork loin chops as affected by the type of culinary frying fat.  $LWT - Food$ science and Technology 38 726-734.
- Romero A, Sanchez-Muniz F J and Cuesta C (2000) Deep fat frying of frozen foods in sunflower oil. Fatty acid composition in fryer oil and frozed prefied potaoes. Journal of the Science of Food and Agriculture 80 2185–2192.
- Rousseau D (2000) Fat crystals and emulsion stability a review. Food Research International 33 3–14.
- Sağdiç O, Dönmez M and Demirci M (2004) Comparison of characteristics and fatty acid profiles of traditional Turkish yanik butters produced from goats', ewes' or cows' milk. Food Control 15 485–490.
- Sandoval-Castilla O, Lobato-Calleros C, Aguirre-Mandujano E and Vernon-Carter E J (2004) Microstructure and texture of yogurt as influenced by fat replacers. International Dairy Journal 14 151–159.
- Spanier A M, Edwards J and Dupuy H P (1998) The warmedover flavor process in deef: a study of meat proteins and peptides. Food Technology 46 110–118.
- Stevenson S G, Vaisey-Genser M and Eskin N A (1984) Quality control in the use of deep frying oils. Journal of the American Oil Chemists' Society 61 1102–1108.
- Tamang J P, Tamang B, Schillinger U, Franz C A P, Gores M and Holzapfel W H (2005) Identification of predominant lactic acid bacteria isolated from traditionally fermented vegetable products of the Eastern Himalayas. International Journal of Food Microbiology 105 347–356.
- Varela G (1998) Current facts about frying of food. In Frying of Food Principals Changes, New Approaches. Varela G, Bender A E and Morton I D, eds. Chichester, UK: Ellis Horwood.
- Vieira T M F S and Regitano d'Arce M A B (2001) Canola oil thermal oxidation during oven test and microwave heating. Lebensmittel-Wissenschaft und-Technologie 34 215–221.