

Impact of lipid-polymer type and content on Lipid Nanoparticles (LNPs) physico-chemical properties and protein corona formation

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Introduction

Since COVID-19 pandemic, Lipid Nanoparticles (LNPs) encapsulating mRNA have emerged as a new class of therapeutic agent as well as Onpatro[®], the first FDA-approved drug based on LNPs and siRNA for a hepatic disease [1]. Indeed, siRNA and mRNA need vectors to reach the target undamaged. This is the reason why LNPs have garnered attention in this context (Figure 1). To target beyond the liver, notably for cancer treatment, LNPs need to be protected from blood biomolecules because of the formation of a protein corona (Figure 2), affecting their efficacy. In this context, polyethylene glycol (PEG) is used to prevent this phenomenon, but its issues initiate the search for alternatives [2,3]. Recently, alternatives to PEG such as polysarcosine and Poly(N-methyl-N-vinylacetamide) (PNMVA) have been studied and seem to be promising [4,5].

Lipid Nanoparticle (LNP)

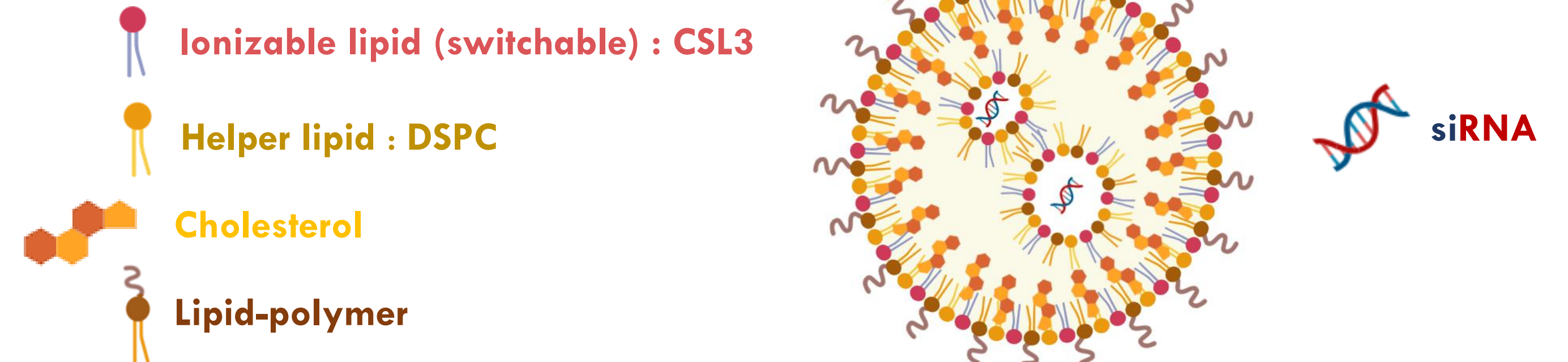


Figure 1 : Lipid Nanoparticle (LNP) structure.

Protein corona

Interactions between LNP and proteins from the blood stream
Adsorption of proteins on LNP surface

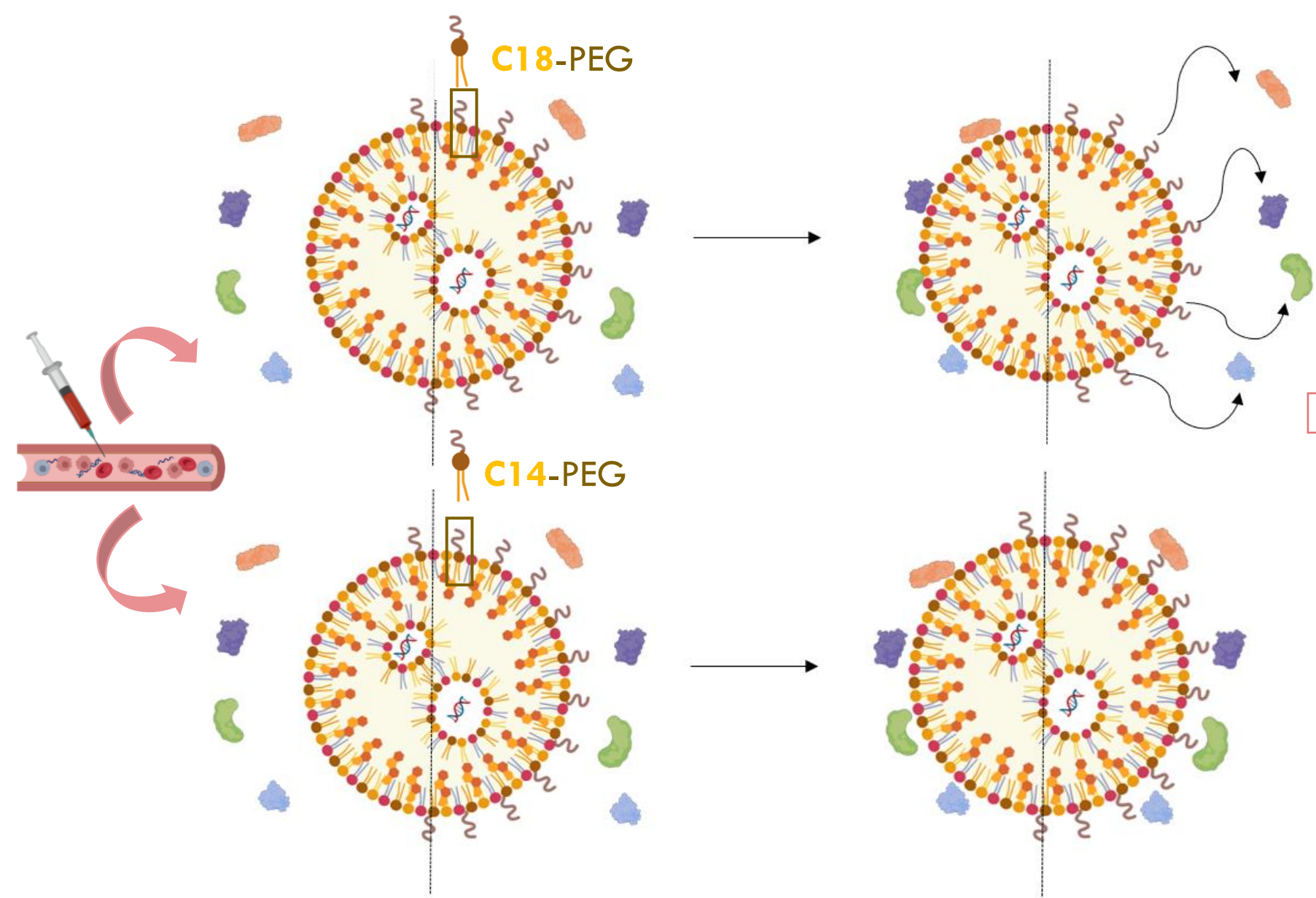


Figure 2 : Illustration of protein corona formation on LNP surface composed of long lipid-PEG (C18) and short lipid-PEG (C14).

Objectives

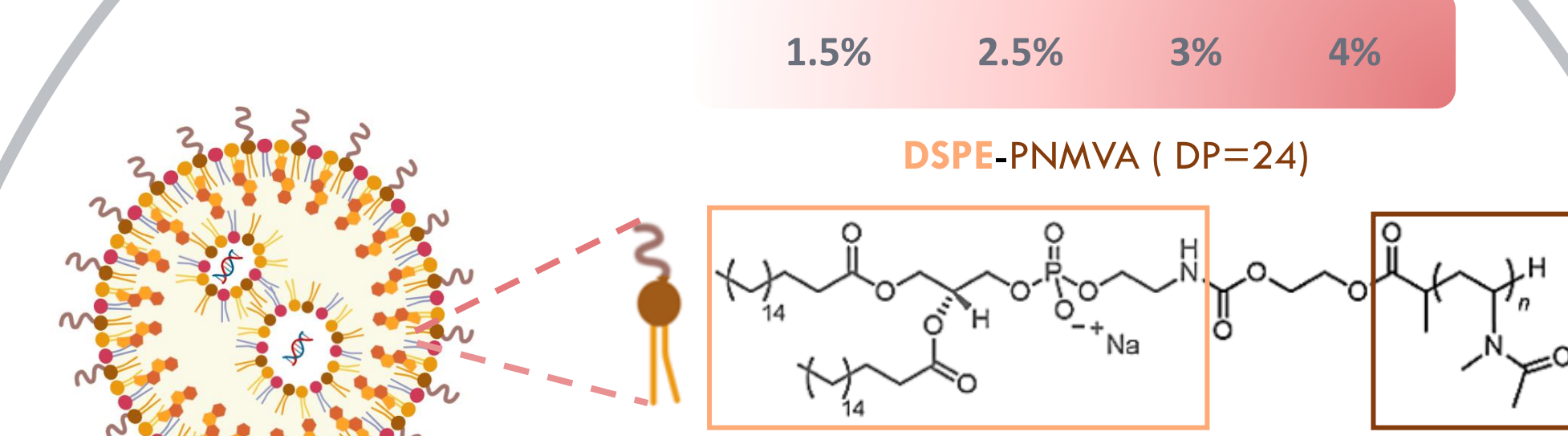


Figure 3 : Structure of DSPE-PNMVA₂₄ used at different contents for LNP formulation.

- To produce LNPs by the rapid-mixing method with different types of lipid-polymers and contents (Figure 3-4)
- To analyze the impact of these modifications on physico-chemical properties and protein corona formation (Figure 2-4)
- To compare LNPs with different contents of DSPE-PNMVA₂₄ (recently patented) and DSPE-PEG₂₀₀₀ to then compare it to another commercial polymer (polysarcosine)

Development of safer and efficient LNP formulations for an antitumoral application

Materials and methods

The LNPs, produced by rapid-mixing, are composed of CSL3, DSPC, cholesterol and different types of C18 lipid-polymers (DSPE-PEG₂₀₀₀, DOPE-PEG₂₀₀₀, DOPE-Psar₂₅, DSPE-PNMVA₂₄) and DMG-PEG₂₀₀₀ as C14 control at a molar ratio respectively of 50:10:37.5:2.5 (Figure 4). The lipid-polymer content was decreased until 1.5% for all the formulations and increased until 3 to 4% by varying the cholesterol content only for DSPE-PEG₂₀₀₀ and DSPE-PNMVA₂₄ formulations.

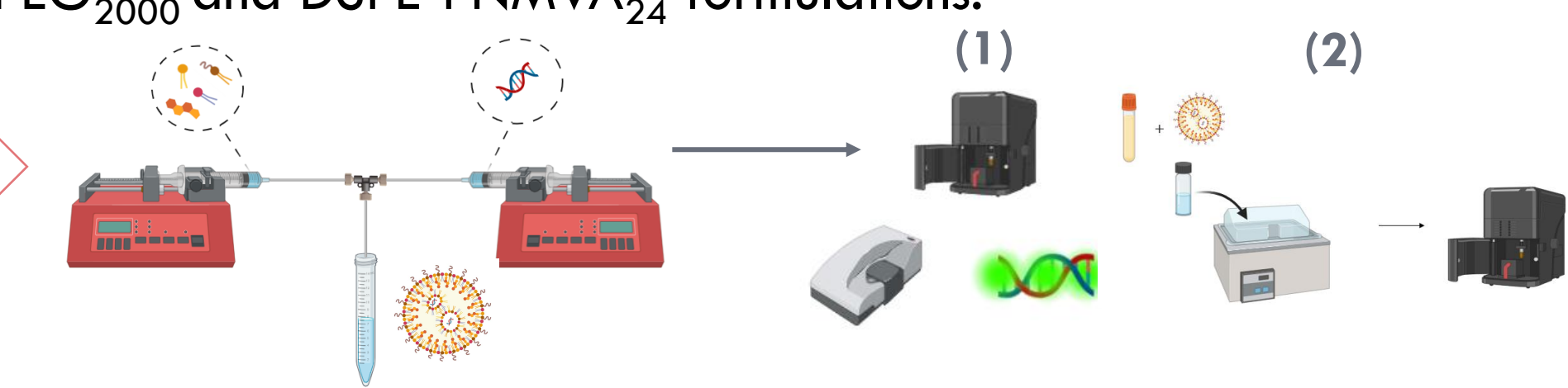


Figure 4 : Production of LNP by rapid-mixing followed by the techniques used for LNP analysis.

Key properties such as size, Pdl and surface charge were analyzed by DLS and NTA while siRNA encapsulation efficiency was evaluated by Ribogreen[®] assay (Figure 4 (1)). The goal was to meet intravenous administration standards: size < 150 nm, Pdl < 0.2 and maximum encapsulation efficiency. Protein corona formation was evaluated using NTA method after incubation in 33.33% of FBS at 37°C (Figure 4 (2)).

Results and discussion

I. Impact of DSPE-PNMVA₂₄ and DSPE-PEG₂₀₀₀ content on LNP properties

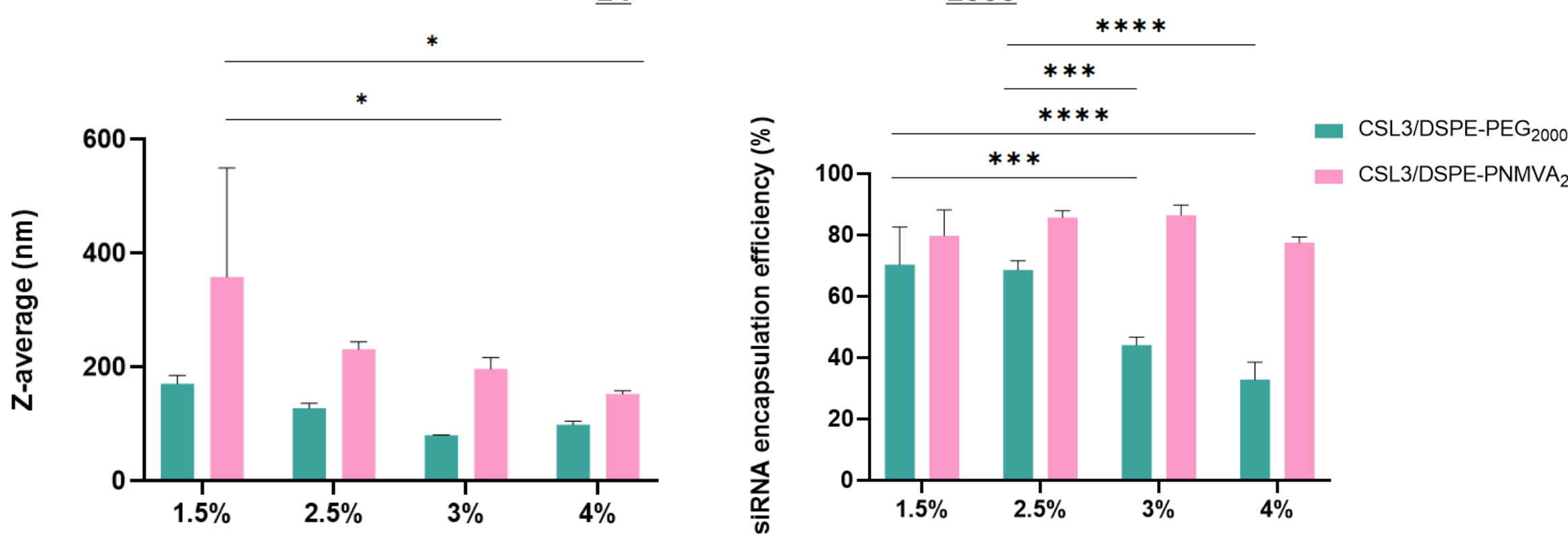


Figure 5 : Physico-chemical properties of different LNP formulations.

It has been observed that particle size and Pdl were significantly decreased while the encapsulation efficiency remained constant when increasing DSPE-PNMVA₂₄ content. By contrast, DSPE-PEG₂₀₀₀ didn't show a decreased Pdl and showed a significant decrease of siRNA encapsulation efficiency when increasing the content, demonstrating instability (Figure 5). Moreover, the NTA test confirmed that DSPE-PEG₂₀₀₀ at 2,5% hinders protein corona formation by contrast to DMG-PEG₂₀₀₀ at 2,5%, demonstrated by an increased particle concentration and constant size for DSPE-PEG₂₀₀₀ (due to the non-adsorption of proteins) and a decreased particle concentration and increased size for DMG-PEG₂₀₀₀ (due to protein adsorption because of the loss of the C14 anchor).

By increasing DSPE-PEG₂₀₀₀ content, profiles seemed to demonstrate less protection against proteins at the opposite of DSPE-PNMVA₂₄ at 4% that demonstrated a similar profile to the C18 control at 2,5%, with refined peaks (Figure 6).

II. Impact of DOPE-Psar₂₅ and DOPE-PEG₂₀₀₀ content on LNP properties

Compared to its lipid-PEG analogous, DOPE-Psar₂₅ followed the same tendencies for size and Pdl when increasing the content : a decrease of the size and an increase of the Pdl. In contrast, the encapsulation efficiency remained constant when increasing DOPE-Psar₂₅ content while DOPE-PEG₂₀₀₀ showed a decrease in encapsulation efficiency (not significant) as well as DSPE-PNMVA₂₄. DOPE-Psar₂₅ showed disturbed profiles at 1.5% and 2.5% (broad peaks) demonstrating the potential need of a higher content. In contrast, DOPE-PEG₂₀₀₀ already showed protein corona formation at 2,5% demonstrated by flattening of the curves (as a C14 lipid-PEG) starting to 3h of incubation (Figure 7).

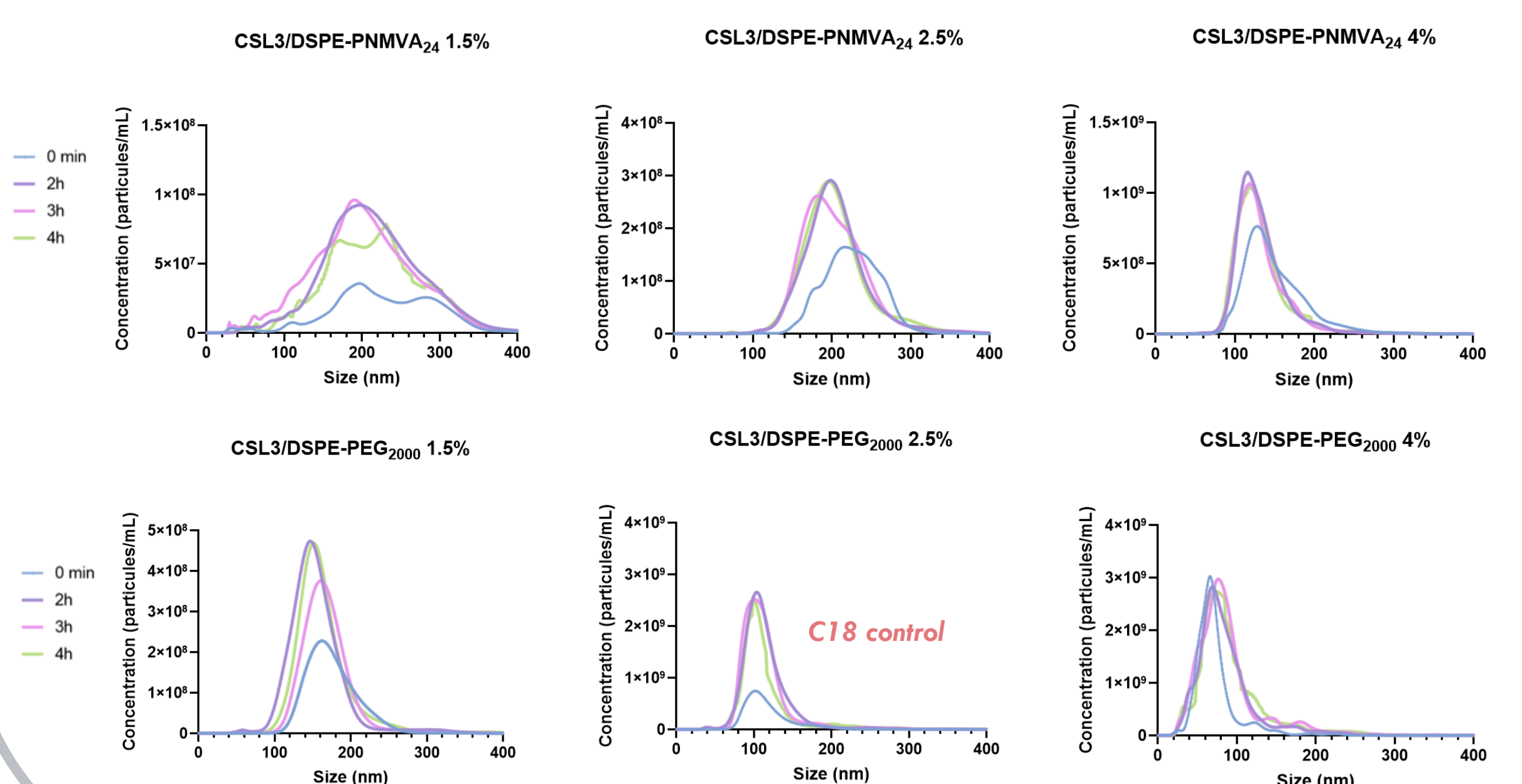


Figure 6 : NTA profiles (FBS 33.33%) of different LNP formulations.

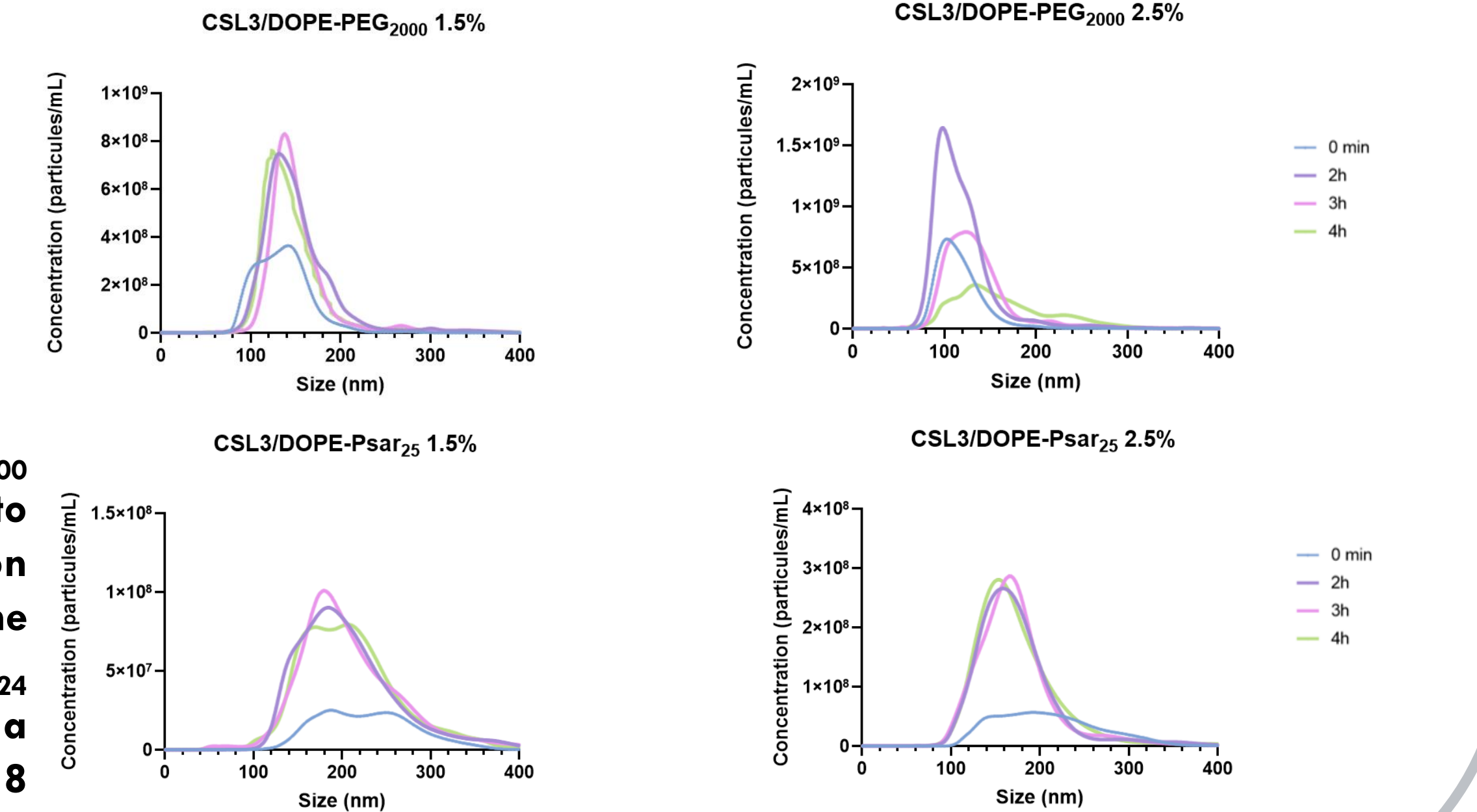


Figure 7 : NTA profiles (FBS 33.33%) of different LNP formulations.

Conclusion and perspectives

In conclusion, the promising potential of PNMVA as an alternative to PEG in LNPs has been confirmed, with findings indicating the need for a slightly higher concentration than previously used. The comprehensive set of tests clearly demonstrates its effectiveness. These conclusions have been transferred to another commercial polymer for further optimization. Moreover, *in vitro* tests will be realized to confirm that a higher content of PNMVA allows siRNA delivery and that it is not cytotoxic.

References

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