Clustering individuals using INMTD: a novel versatile multi-view embedding framework integrating omics and imaging data

Zuqi Li^{1,2*}, Sam F. L. Windels³, Noël Malod-Dognin³, Seth M. Weinberg^{4,5}, Mary L. Marazita^{4,5}, Susan Walsh⁶, Mark D. Shriver⁷, David W. Fardo⁸, Peter Claes^{1,2,9,10}, Nataša Pržulj^{3,11,12}, Kristel Van Steen^{1,13}

 ¹ Department of Human Genetics, KU Leuven, Leuven, Belgium
 ² Medical Imaging Research Center, UZ Leuven, Leuven, Belgium
 ³ Barcelona Supercomputing Center, Barcelona, Spain
 ⁴ Center for Craniofacial and Dental Genetics, Department of Oral and Craniofacial Sciences, University of Pittsburgh, Pittsburgh, USA
 ⁵ Department of Human Genetics, University of Pittsburgh, Pittsburgh, USA
 ⁶ Department of Biology, Indiana University Purdue University Indianapolis, Indianapolis, USA
 ⁷ Department of Anthropology, Pennsylvania State University, University Park, USA
 ⁸ Sanders-Brown Center on Aging, University of Kentucky, Lexington, USA
 ⁹ Department of Electrical Engineering, ESAT/PSI, KU Leuven, Leuven, Belgium
 ¹⁰ Murdoch Children's Research Institute, Melbourne, Australia
 ¹¹ Department of Computer Science, University College London, London, UK
 ¹² Catalan Institution for Research and Advanced Studies (ICREA), Barcelona, Spain
 ¹³ GIGA-R Medical Genomics, University of Liège, Liège, Belgium

Abstract

Motivation: Combining omics and images, can lead to a more comprehensive clustering of individuals than classic single-view approaches. Among the various approaches for multi-view clustering, nonnegative matrix tri-factorization (NMTF) and nonnegative Tucker decomposition (NTD) are advantageous in learning low-rank embeddings with promising interpretability. Besides, there is a need to handle unwanted drivers of clusterings (i.e. confounders).

Results: In this work, we introduce a novel multi-view clustering method based on NMTF and NTD, named INMTD, that integrates omics and 3D imaging data to derive unconfounded subgroups of individuals. In the application to real-life facial-genomic data, INMTD generated biologically relevant embeddings for individuals, genetics and facial morphology. By removing confounded embedding vectors, we derived an unconfounded clustering with better internal and external quality; the genetic and facial annotations of each derived subgroup highlighted distinctive characteristics. In conclusion, INMTD can effectively integrate omics data and 3D images for unconfounded clustering with biologically meaningful interpretation.

Availability and implementation: https://github.com/ZuqiLi/INMTD

1 Introduction

Clustering is a crucial technique in data analysis, enabling the identification of intrinsic structures within complex datasets by grouping similar data points. In the field of medicine, clustering has been widely used for uncovering disease subtypes, tailoring personalized treatments, and improving early diagnosis (Ghosal et al. 2020). As data complexity grows, clustering methods based on a single view or single data source are often insufficient, necessitating the development of more sophisticated approaches. Multi-view clustering has emerged as a powerful solution, leveraging multiple data perspectives to enhance clustering quality and reveal richer patterns than single-view methods (Rappoport and Shamir 2018; Chauvel et al. 2020). As the data views commonly used in biomedical science to describe an individual, omics and imaging data have shown essential advantages in understanding various biological phenomena (Antonelli et al. 2019). For instance, Chen et al. obtained better prediction for subtypes of lung adenocarcinoma by integrating extracted features from histopathological images and omics data than using a single view (Chen et al. 2021). However, few people have worked on clustering individuals based on omics and imaging data. Moreover, processing images normally requires tensor methods due to their 3D format (Hériché, Alexander and Ellenberg 2019), e.g. a color image consists of pixels represented by height, width and color channel, and a 3D mesh consists of X, Y, Z coordinates for height, weight and depth.

Various multi-view clustering methods have been developed, which can be generally classified into three categories, based on the relationship between data integration and clustering (Rappoport and Shamir 2018): 1) early integration combines all datasets into a single one before building the model for clustering, 2) intermediate integration clusters a joint embedding learnt from all views, and 3) late integration computes a clustering from each dataset and then merges all clusterings together. Out of the three, intermediate integration approaches have shown superior performance in many applications possibly because they require a model specifically designed for multi-view clustering tasks (Khan and Maji 2019; Wang et al. 2020; Yun et al. 2021). By clustering subjects with multi-view data from a joint embedding, integrative nonnegative matrix factorization (intNMF) proposed by Chalise and Fridley has found similar cancer subtypes identified by previous studies (Chalise and Fridley 2017). This embedding represents the subjects with patterns that are naturally additive and hence easily interpretable (Lee and Seung 1999). A well-established extension of NMF model for better interpretation and clustering is nonnegative matrix tri-factorization (NMTF) that decomposes the input dataset into three smaller matrices (Ding et al. 2006). However, NMTF models only work with 2D matrices and cannot deal with data views of higher dimensions, e.g. a 3D tensor, which is the common data format for imaging or spatial data. A generalization of NMTF to tensors is the nonnegative Tucker decomposition (NTD) (Kim and Choi 2007). It decomposes the original input tensor into a core tensor with the same number of dimensions and one embedding matrix corresponding to each of its dimensions. There have been attempts to integrate multiple cross-linked 2D matrices into a tensor, which, however, does not work on originally 3D data (Luo et al. 2022). Another work by Broadbent et al. combined a tensor with a similarity matrix, while they treated the matrix as a graph regularization to the Tucker decomposition instead of a separate data view (Broadbent, Song and Kuang 2024).

Clustering real-world data is often complicated by the presence of confounders—factors that influence the observed data in an unwanted way (Liu *et al.* 2015). Confounders can obscure the true clustering structure, leading to spurious results (Schwarz *et al.* 2024). Addressing confounders is essential for accurate clustering, as their effects can mask the genuine patterns within the data. Effective clustering methods must account for these confounding variables to uncover the true underlying structure. A widely adopted approach is to regress out the confounding effects from every feature during pre-processing, but it comes with the potential loss of useful signals prior to the modelling (Pourhoseingholi, Baghestani and Vahedi 2012). Some other strategies include kernel conditional clustering which computes the final clustering conditioned on confounders (He *et al.* 2020). However, the conditional clustering is computationally expensive and cannot efficiently work on high-dimensional data.

Here in this paper, we propose a novel multi-view clustering method, integrative non-negative matrix and tensor decomposition (INMTD), which obtains unconfounded clustering jointly from 2D and 3D datasets. It learns an embedding matrix for each data dimension and subgroups the individuals from their embedding after removing vectors in the embedding space that are linked with confounders. Because the true cluster structure of real-life patient dataset is often unknown, we evaluated INMTD on a US cohort from healthy individuals (White *et al.* 2021), whose heterogeneity mainly comes from the population structure with confounders including age, sex, etc. Combining 2D genotypes and 3D facial morphology, our model computed biologically meaningful embeddings and connected well the facial and genetic embeddings. Furthermore, INMTD derived an unconfounded clustering of individuals with better intrinsic quality and clearer association with population structure than the original clustering. We also characterized each population subgroup with their enriched genetic pathways and highlighted facial areas.

2 Methods

2.1 INMTD: integrative non-negative matrix and tensor decomposition with correction for confounders

INMTD unifies NMTF and NTD to cluster subjects with multi-view data of 2D and 3D structure. We assume p_1 subjects described by two data views, a 2D matrix $X_{12} \in \mathbb{R}^{p_1 \times p_2}_+$ of p_2 features and a 3D tensor $\mathcal{X}_{134} \in \mathbb{R}^{p_1 \times p_3 \times p_4}_+$ of p_3 features in the 2nd dimension and p_4 features in the 3rd dimension, both nonnegative. The aim of our method is to jointly compute the embedding matrices for each dimension and cluster the p_1 subjects based on its own embedding (Fig. 1).



Fig. 1: Overview of INMTD model for integrating 2D and 3D data. The 2D matrix X_{12} is decomposed into two embedding matrices G_1 and G_2 and a core matrix S_{12} . The 3D tensor \mathcal{X}_{134} is decomposed into three embedding matrices G_1 , G_3 and G_4 and a core tensor S_{134} . INMTD integrates X_{12} and \mathcal{X}_{134} by jointly optimizing G_1 , which is shared by the two views. The orthogonality constraint on G_1 ensures disentanglement of embedding vectors.

For 2D matrix X_{12} , NMTF factorizes it into three nonnegative submatrices $G_1 \in \mathbb{R}^{p_1 \times r_1}_+$, $G_2 \in \mathbb{R}^{p_2 \times r_2}_+$ and $S_{12} \in \mathbb{R}^{r_1 \times r_2}_+$, so that $X_{12} \approx G_1 S_{12} G_2^T$. The objective of NMTF is to find the optimal G_1 , G_2 and S_{12} that minimize the reconstruction error:

$$\min_{G_1 \ge 0, G_2 \ge 0, S_{12} \ge 0} J = \|X_{12} - G_1 S_{12} G_2^T\|_F^2,$$
(1)

where $\|\cdot\|_F^2$ indicates the Frobenius norm and ≥ 0 for a matrix means all values in that matrix should be nonnegative. The multiplicative update rules to solve this objective function have been proposed by Ding et al. (Ding *et al.* 2006) G_1 and G_2 are low-rank embeddings for the p_1 subjects and p_2 features, respectively, where r_1 and r_2 are their ranks and normally $r_1 \ll p_1$ and $r_2 \ll p_2$. S_{12} is the core matrix that links G_1 with G_2 and can be considered as the compressed representation of X_{12} .

Similar to NMTF, the NTD model decomposes the 3D tensor \mathcal{X}_{134} into 3 embeddings $G_i \in \mathbb{R}^{p_i \times r_i}$ with $i \in \{1,3,4\}$, named the mode matrices, and a core tensor $S_{134} \in \mathbb{R}^{r_1 \times r_3 \times r_4}$ using the mode product of tensor:

$$\mathcal{X}_{134} \approx S_{134} \times_1 G_1 \times_2 G_3 \times_3 G_4, \tag{2}$$

where $S_{134} \times_n G_i$ is the mode-*n* product between tensor S_{134} and matrix G_i , resulting in a new tensor with its *n*-th dimension changed. The objective of NTD is to minimize the reconstruction error:

$$\min_{G_i \ge 0, S_{134} \ge 0} J = \|\mathcal{X}_{134} - S_{134} \times_1 G_1 \times_2 G_3 \times_3 G_4\|_F^2,$$
(3)

To jointly decompose X_{12} and X_{134} , we derive an integrative objective from the two views via NMTF and NTD:

$$\min_{G_i \ge 0, S_{12} \ge 0, S_{134} \ge 0} J = \|X_{12} - G_1 S_{12} G_2^T\|_F^2 + \|X_{134} - S_{134} \times_1 G_1 \times_2 G_3 \times_3 G_4\|_F^2,$$
(4)

where $G_i \in \mathbb{R}^{p_i \times r_i}$ are the low-rank embeddings corresponding to p_1 subjects, p_2 features of view 1, p_3 features of view 2 and p_4 channels of view 2. The rank parameters r_i are determined via the rule of thumb: $r_i = \sqrt{p_i/2}$ (Kodinariya and Makwana 2013). G_1 is shared by both terms in formula (4) and jointly learnt from both views. We further adopt an orthogonality constraint on G_1 for more rigorous clustering interpretation (Ding *et al.* 2006):

$$\min_{\substack{G_i \ge 0, S_{12} \ge 0, S_{134} \ge 0}} J = \|X_{12} - G_1 S_{12} G_2^T\|_F^2 + \|\mathcal{X}_{134} - S_{134} \times_1 G_1 \times_2 G_3 \times_3 G_4\|_F^2,$$
s.t. $G_1^T G_1 = I$
(5)

Because formula (5) has no analytic solution for G_i , S_{12} and S_{134} , we iteratively compute their values via the multiplicative update rules (Dissez *et al.* 2019) (see section 2.2). Furthermore, in each iteration, we normalize every column of G_1 after updating to further guarantee unit vectors and eliminate the scale indeterminacy as suggested by Li et al. (Bo Li, Guoxu Zhou and Cichocki 2015)

We apply k-means clustering (with 10 random initializations) on the joint embedding, G_1 , and select the best number of clusters based on Silhouette score. Silhouette score is a classic internal metric that measures how well a dataset is clustered. A higher value (close to 1) suggests a more valid clustering.

To assess how much G_1 is confounded by a set of known confounders, C, we conduct a linear model F-test between every confounder and every column of G_1 . The unconfounded clustering is then recomputed from the columns of G_1 that have no significant association with any confounders. This is done also by k-means and Silhouette score for the optimal number of clusters. The removal of confounded embedding components is based on the additive nature of NMF-based methods that the data is represented by the sum of all its embedding aspects (Lee and Seung 1999). Deconfounding at the embedding level is computationally less expensive and can better preserve meaningful information than the widely used approach, which is to regress out confounders from each feature at the input level.

2.2 Training procedure of INMTD

To solve INMTD, we use a fixed-point method that, starting from an initial solution, iteratively uses multiplicative update rules to converge towards a locally optimal solution. During the optimization process, all the embedding matrices and core matrix/tensor of INMTD are iteratively updated to minimize the objective function (formula (5)). Following the derivation procedure used to derive multiplicative update rules for orthogonal NMTF and NTD, we derive the update rules for INMTD:

$$G_{1(ij)} \leftarrow G_{1(ij)} \sqrt{\frac{\left(X_{12}G_2S_{12}^T + X_{134}^{(1)} \left[S \times_2 G_3 \times_3 G_4\right]^{(1)^T}\right)_{ij}}{\left(G_1G_1^T X_{12}G_2S_{12}^T + G_1G_1^T X_{134}^{(1)} \left[S \times_2 G_3 \times_3 G_4\right]^{(1)^T}\right)_{ij}}}$$
(6)

$$G_{2(ij)} \leftarrow G_{2(ij)} \sqrt{\frac{(X_{12}^T G_1 S_{12})_{ij}}{(G_2 S_{12}^T G_1^T G_1 S_{12})_{ij}}}$$
(7)

$$G_{3(ij)} \leftarrow G_{3(ij)} \sqrt{\frac{\left(X_{134}^{(2)} \left[\mathcal{S} \times_{1} G_{1} \times_{3} G_{4}\right]^{(2)^{T}}\right)_{ij}}{\left(G_{3} \left[\mathcal{S} \times_{1} G_{1} \times_{3} G_{4}\right]^{(2)} \left[\mathcal{S} \times_{1} G_{1} \times_{3} G_{4}\right]^{(2)^{T}}\right)_{ij}}}$$
(8)

$$G_{4(ij)} \leftarrow G_{4(ij)} \sqrt{\frac{\left(X_{134}^{(3)} \left[\mathcal{S} \times_{1} G_{1} \times_{2} G_{3}\right]^{(3)}^{T}\right)_{ij}}{\left(G_{4} \left[\mathcal{S} \times_{1} G_{1} \times_{2} G_{3}\right]^{(3)} \left[\mathcal{S} \times_{1} G_{1} \times_{2} G_{3}\right]^{(3)}^{T}\right)_{ij}}}$$
(9)

$$S_{12(ij)} \leftarrow S_{12(ij)} \sqrt{\frac{(G_1^T X_{12} G_2)_{ij}}{(G_1^T G_1 S_{12} G_2^T G_2)_{ij}}}$$
(10)

$$S_{134(ijk)} \leftarrow S_{134(ijk)} \sqrt{\frac{[\mathcal{X}_{134} \times_1 G_1^T \times_2 G_3^T \times_3 G_4^T]_{ijk}}{[\mathcal{S}_{134} \times_1 G_1^T G_1 \times_2 G_3^T G_3 \times_3 G_4^T G_4]_{ijk}}}$$
(11)

 $X_{134}^{(n)}$ denotes the mode-n matricization of \mathcal{X}_{134} , which reshapes \mathcal{X}_{134} to a 2D matrix along its n-th dimension.

We initialize G_i via singular value decomposition (SVD), which has shown better results than random initialization (Malod-Dognin *et al.* 2019). For X_{12} , the original matrix is decomposed by SVD and G_1 and G_2 are derived from the left and right matrices of SVD, respectively. Because \mathcal{X}_{134} is 3D, we run SVD for every slice along the p_4 channels and average all the right matrices to compute the initial G_3 and similarly run SVD for every slice along the p_3 features to initialize G_4 . As the update rules are multiplicative, to avoid entries in G_i to remain 0, we add an infinitesimal number (1e-5) so that these entries can be updated.

 S_{12} and S_{134} can also be initialized by SVD through the eigen values if the original data frames are symmetric. But we don't assume their symmetry in our framework, therefore, we apply the following rules:

$$S_{12} = G_1^T X_{12} G_2 \tag{12}$$

$$\mathcal{S}_{134} = \mathcal{X}_{134} \times_1 G_1^T \times_2 G_3^T \times_3 G_4^T \tag{13}$$

The initialized S_{12} and S_{134} are automatically nonnegative because all the multipliers are nonnegative.

To assess the goodness and convergence of INMTD, we track a metric along the optimization, which is the total relative error:

$$\text{Fotal Relative Error} = \frac{\|X_{12} - G_1 S_{12} G_2^T\|_F^2 + \|\mathcal{X}_{134} - S_{134} \times_1 G_1 \times_2 G_3 \times_3 G_4\|_F^2}{\|X_{12}\|_F^2 + \|\mathcal{X}_{134}\|_F^2}$$
(14)

The total relative error computes the fraction of the reconstruction errors of the two datasets X_{12} and X_{134} in their L2 norms. It is a nonnegative value and a lower total relative error indicates better reconstruction from the decomposed elements.

2.3 Association between embeddings

Linking two embeddings from different views can be achieved by mapping them to the same space of G_1 because G_1 is the embedding shared by both data types. More specifically, we project G_2 to the space of G_1 via the core matrix S_{12} , so that $\hat{G}_2 = G_2 S_{12}^T$. The new matrix \hat{G}_2 now has the same embedding size as G_1 and is in the same embedding space as G_1 . Similarly, G_3 is also projected to the space of G_1 via the core tensor S_{134} , resulting in $\hat{G}_3 = S_{134} \times_3 G_3$. In the special case when $r_4 = 1$ and hence S_{134} is of shape $r_1 \times r_3 \times 1$, this is equivalent to $\hat{G}_3 = G_3 S_{13}^T$, where S_{13} reshapes S_{134} to $r_1 \times r_3$. Subsequently, the relationship between feature (row) i in G_2 and feature (row) j in G_3 can be assessed by cosine similarity:

Cosine Similarity =
$$\frac{\hat{G}_{2(i)} \cdot \hat{G}_{3(j)}}{\|\hat{G}_{2(i)}\|_{F}^{2} \|\hat{G}_{3(j)}\|_{F}^{2}},$$
(15)

where $\hat{G}_{2(i)}$ indicates the *i*-th row of \hat{G}_2 and $\hat{G}_{3(j)}$ the *j*-th row of \hat{G}_3 . Cosine similarity measures the dot product between two vectors regardless of their magnitudes, providing good normalization when comparing between \hat{G}_2 and \hat{G}_3 . It ranges from -1 to 1 and the higher the more similar between the two vectors.

3 Experiments

3.1 Evaluation dataset

We apply INMTD to a multi-view dataset of 4,680 normal people with European ancestry, characterized by a 2D matrix and a 3D tensor (White et al. 2021). These people were recruited from three independent studies in the US, 3D Facial Norms cohort (PITT), Pennsylvania State University (PSU) and Indiana University-Purdue University Indianapolis (IUPUI). Every individual was described by 7,141,882 SNPs and a 3D mesh image which contains the X, Y, Z coordinates of 7,160 landmarks, namely $X_{12} \in \mathbb{R}^{4,680 \times 7,141,882}$ and $\mathcal{X}_{134} \in \mathbb{R}^{4,680 \times 7,160 \times 3}$, respectively. Due to the enormous number of SNPs the SVD initialization on X_{12} had to adopt randomized SVD for feasibility. The initialization on χ_{134} still used full SVD. To standardize genomic and facial data, we subtracted the mean from each view and divided every entry by the maximum of each view. We then took the absolute values to ensure nonnegativity. The rank r_1 of embedding G_1 was determined via the rule of thumb: $\sqrt{p_1/2} \approx 48$ where p_1 is the number of individuals, thus $G_1 \in \mathbb{R}^{4,680 \times 48}_+$. r_2 was chosen in a similar way but based on the number of protein-coding genes, namely 19,430, instead of SNPs to reduce the computational burden, and thus $G_2 \in \mathbb{R}^{7,141,882 \times 99}_+$. We had $G_3 \in \mathbb{R}^{7,160 \times 60}_+$ and $G_4 \in \mathbb{R}^{3 \times 1}_+$ because $r_3 = r_3 = r_3$ $\sqrt{p_3/2} \approx 60$ and $r_4 = \sqrt{p_4/2} \approx 1$ where $p_3 = 7,160$ and $p_4 = 3$. For every individual, we also collected a few covariates, including age, sex, height, weight, face size and camera system. BMI was derived via: BMI = weight(kg)/height(m)² as an additional covariate. Here, we

consider these covariates as confounders to population structure because they might hinder us in finding the population subgroups based on genetic heterogeneity. To measure the population structure of the cohort, White et al. have computed four ancestry axes by projecting the genomic data onto the principal component (PC) space of the SNPs from the 1000G Project (White *et al.* 2021).

3.2 Evaluation on the Embeddings

INMTD learns an embedding for SNPs, faces, and individuals. Here, we outline how we assess the biological validity for those embeddings separately and jointly.

To biologically validate our individual embedding, we assess if G_1 captures any heterogeneity of the cohort, including both population structure (ancestry axes) and confounding effects. Due to the orthogonality constraint, embedding vectors of G_1 can be considered as independent and characterize different aspects of the individuals. We, therefore, test the statistical association between every embedding vector and every ancestry axis or confounder. In particular, Kruskal-Wallis ANOVA is used for ancestry axes and continuous confounders (age, height, weight, BMI and face size) and chi-squared test is used for categorical confounders (sex and camera system). We apply Benjamini-Hochberg (BH) correction for the multiple testing.

To assess our SNP embedding, we cluster SNPs based on their embedding and use enrichment analysis to see if their space is functionally organised. As G_2 is not orthogonal, we use k-means (with 10 random initializations) on G_2 to subgroup SNPs into $r_2 = 99$ clusters. We first check how well those SNP clusters coincide with the 19,430 protein-coding genes given the fact that gene is a natural summary of SNPs and biological processes are usually interpreted on a gene level. A gene is defined to be enriched in a SNP cluster if SNPs located 2K base pairs around this gene are present in this cluster significantly more than in the background. Hypergeometric test is applied for the enrichment analysis and the BH procedure is used for multiple testing correction. To further check if these clusters have biological process. We annotate every SNP by GO terms if it is mapped to a gene that is annotated by a GO term and then test the overrepresentation of GO terms in every SNP cluster. Because GO terms are gene annotations, we annotate SNPs with GO terms that annotate their mapped genes. Only GO terms under biological process category are used and the BH procedure is applied for multiple testing correction.

To assess our facial embedding, facial landmarks are subgrouped by Ward's hierarchical clustering on G_3 into $r_3 = 60$ clusters, which segments the shape of face. The Ward's method has been shown outperforming other common linkage methods (Ferreira and Hitchcock 2009; Vijaya, Sharma and Batra 2019). We first compute the Ward distance between every pair of landmarks in G_3 , based on which we then construct a hierarchical tree and cut it at a height with 60 clusters. We adopt hierarchical clustering in order to compare with the hierarchical segmentation done on the same dataset by White et al. They segmented the facial shape from global to local into five levels with 63 segments. To compare the ability of the hierarchical tree of G_3 and the hierarchical segmentation by White et al. to faithfully capture the pairwise

dendrogrammatic distances between landmarks, we compute their cophenetic correlation coefficient:

Cophenetic Correlation =
$$\frac{\sum_{i < j} (D_{ij} - \overline{D}) (Z_{ij} - \overline{Z})}{\sqrt{\sum_{i < j} (D_{ij} - \overline{D})^2 \sum_{i < j} (Z_{ij} - \overline{Z})^2}},$$
(16)

where *i* and *j* are facial landmarks, *D* is the Euclidean distance matrix between landmarks, and *Z* is the cophenetic distance matrix between landmarks, denoting the heights at which two points are first merged in the dendrogram. \overline{D} and \overline{Z} are the mean of *D* and *Z*, respectively. A cophenetic correlation close to 1 indicates a high-quality hierarchical clustering.

To assess the association between the SNP and facial embeddings, we map them to the space of G_1 and compute cosine similarity between each SNP and each facial landmark (see section 2.3). For the SNPs closest to facial landmarks in the joint space, we apply the GREAT analysis (v.4.0.4), which finds biological meaning of the set of SNPs via the annotations of nearby genes. Technically speaking, GREAT analysis performs a binomial test for a set of SNPs to check whether the overlap between their associated genes and genes with a certain annotation is greater than random chance. To associate SNPs with genes, we apply the default and recommended settings (McLean *et al.* 2010), namely the 'basal plus extension' rule with 5kb upstream and 1kb downstream plus 1000kb extension. Note that one SNP can be associated with multiple genes and SNPs not associated with any genes are not included in this analysis.

3.3 Characterization of unconfounded population subgroups

The unconfounded population subgroups are characterized based on the projection of genetic and facial embeddings to the space of the sample embedding, enabling computing the similarity between population subgroup centroids and SNPs and facial landmarks. For each subgroup, we first select the top 0.1% SNPs with highest cosine similarity to its centroid in the joint space, to which the GREAT analysis is applied to reveal the most relevant phenotypes (HPO). The threshold of 0.1% is determined by balancing the number of genes selected per subgroup and the genomic coverage of genes selected for all subgroups (Supp. Fig. 1 and 2). We then visualize the cosine similarities of all facial landmarks to a subgroup centroid on the averaged face, in order to demonstrate how different areas of the face are associated with the corresponding subgroup.

4 Results

4.1 INMTD generates biologically meaningful embeddings from a real-life multiview dataset

We applied INTMD to a real-world facial-genomic cohort collected from the US for unconfounded population subgrouping (White *et al.* 2021). This dataset consists of two data types, $X_{12} \in \mathbb{R}^{4,680 \times 7,141,882}_{+}$ for 7,141,882 SNPs in 4,680 people and $\mathcal{X}_{134} \in \mathbb{R}^{4,680 \times 7,160 \times 3}_{+}$ for the 3D coordinates of 7,160 facial landmarks in the same 4,680 individuals (White *et al.* 2021). A few confounders were collected as well, which are age, sex, height, weight, camera system and face size. We also derived BMI (body mass index) from height and weight as a potential confounder. To assess the population structure of this cohort, White et al. computed

four ancestry axes by projecting the genotypes to a principal component space built from the 1000 Genomes Project data, in the manner of EIGENSTRAT (Price *et al.* 2006). We ran our INMTD model on this multi-view dataset for 1,000 iterations with SVD initialization and it converged in terms of the total relative error (Supp. Fig. 3). Due to the heuristic solver for formula (5), G_1 is only approximately orthonormal. Therefore, we further checked the independencies between the embedding vectors of G_1 based on its covariance matrix, namely $G_1^T G_1$ (Supp. Fig. 4).

Because the heterogeneity of a population can be largely described by population structure, age, sex, etc., we validated the information captured by G_1 based on the statistical association between every column vector of G_1 and every ancestry axis or confounder (Fig. 2). The results indicate that most vectors captured the information of population structure while being confounded. This is expected as, for instance, height has been reported to highly relate with different European ancestries (Cavelaars *et al.* 2000). Furthermore, the 48 embedding vectors have different association patterns with the ancestry axes and confounders. For instance, the 1st vector is significantly associated with most ancestry axes as well as confounders, while no association with the 6th vector is observed.



Fig. 2: Heatmap of P-values for the linear F test between every G_1 embedding vector and every confounder and ancestry axis. Non-significant P-values (larger than 0.05 after BH correction) were removed from the plot.

To assess the biological relevance of the SNP embedding from G_2 , we applied k-means on G_2 to derive 99 clusters of SNPs. We first mapped every SNP to genes if it falls within or 2k base pairs around a gene and ignored SNPs that do not map to any genes. 97 out of 99 SNP clusters have at least one overrepresented gene (P-value < 0.05 in a hypergeometric test with BH correction for multiple testing) with respect to the background of all the SNPs that can map to a gene. 98.6% of genes have been enriched in at least one SNP cluster while most genes were enriched in only a few clusters (Fig. 3). We then applied GO (gene ontology) enrichment analysis for each cluster after assigning GO annotations of a gene to all its belonging SNPs. 96.0% of GO terms have been enriched in at least one Cluster (Fig. 4). This result validated that the SNP clusters obtained from G_2 have both genomic specificity and biological relevance.



Fig. 3: Histogram showing the number of genes (y-axis) that are enriched in a given number of G_2 clusters (x-axis). An enrichment analysis is done between every gene and every G_2 (SNP) cluster. We then compute how many genes (y-axis) are found to be enriched in different No. of clusters of SNPs (x-axis).



Fig. 4: Histogram showing the number of GO terms (y-axis) that are enriched in a given number of G_2 clusters. An enrichment analysis is done between every GO term and every G_2 cluster. We then compute how many GO terms (y-axis) are found to be enriched in different No. of clusters of SNPs (x-axis).

To assess the quality of the facial embedding from G_3 , we aimed to obtain a hierarchical segmentation on G_3 and compare it with the work of White et al. (White *et al.* 2021) The chosen Ward's hierarchical clustering yielded 60 facial segments (Supp. Fig. 5), with a cophenetic correlation coefficient of 0.638, which is higher than that of the facial segmentation by White et al. on the same images (0.414). It suggested that the hierarchical segmentation from embedding G_3 better groups together facial landmarks that are close in 3D space than the one by White et al., defining more spatially coherent 'patches'. The visualization of these 60 clusters on an averaged face also showed that the landmarks within each cluster are spatially close to each other and the clustering is morphologically meaningful (Fig. 5).



Fig. 5: The 60 clusters derived from G_3 , illustrated on the mean face shape of all individuals. Not every cluster has a unique color due to the large number of clusters, but neighboring clusters are distinguished by different colors. Every face shape is symmetric along X-axis (left/right), so is the facial segmentation.

To further investigate the relationships between genetics and facial morphology, we mapped both G_2 and G_3 to the space of G_1 (Supp. Fig. 6). For each facial landmark in the joint space, we selected the closest SNP in terms of cosine similarity, resulting in 905 unique SNPs in total. To assess what biological traits are associated with the chosen SNPs, we conducted a GREAT analysis on their neighbouring genes (McLean *et al.* 2010), which found 17 significantly enriched human phenotype ontology (HPO) terms (Gargano *et al.* 2024) based on the adjusted binomial P-values (Fig. 6). Most of the enriched terms are highly linked to facial morphology (especially eyes), limb or spine morphology and embryonic development, depicting close biological relationship between the embeddings for SNPs and facial landmarks, namely G_2 and G_3 . Other enriched terms suggested high relatedness between facial morphology and myotonia. The results indicate that INMTD allows for uncovering biologically relevant associations between SNPs and facial landmarks.



Fig. 6: Dotplot of 17 significantly enriched HPO terms from 905 SNPs that are closest to facial landmarks in the joint space. GREAT analysis first mapped the 905 SNPs to 1,156 genes based on the default and recommended settings, and then ran a binomial test for the enrichment analysis with BH correction. X-axis shows the ratio between the number of observed genes and the number of genes annotated for each HPO term. The dot color indicates the significance of the adjusted binomial P-value in the form of -log10.

4.2 INMTD finds unconfounded population subgroups characterized by their genetic and facial information

To derive the optimal population subgroups, we applied k-means on G_1 with different number of clusters and found that the clustering with 48 clusters achieved the highest Silhouette score (0.169) (Supp. Fig. 7). As mentioned before, many vectors of G_1 are significantly confounded, potentially disturbing correct interpretation and characterization of derived subgroups. In order to deal with the confounding effect, we removed the 28 columns in G_1 that are significantly associated with any confounders, and clustered individuals based on the remaining 20 embedding vectors (Supp. Fig. 8). We then applied k-means on the unconfounded G_1 with different number of clusters and the clustering with 20 clusters achieved the highest Silhouette score (0.329) (Supp. Fig. 9). This score is statistically higher (empirical P-value = 0.02 from 1000 repetitions, Supp. Fig. 10) than clusterings with the same number of clusters from 20 randomly sampled G_1 vectors, indicating better intrinsic validity of the unconfounded clustering.

To validate our reduced space is unconfounded and leads to a better capturing of the population structure, we assessed the statistical association between the derived clustering and the ancestry axes and confounders. The Kruskal-Wallis test (nonparametric ANOVA) showed both the original and the unconfounded clusterings are significantly associated with all ancestry axes (P-value < 0.05), suggesting their relationships with population structure (Table 1). Yet the original clustering also has significant associations with most confounders, especially age and camera system, while the unconfounded clustering has no significant associations with any confounders, validating our unconfounding strategy. The effective reduction of the influence by camera system, which resembles the batch effect, also indicates the strength of the confounder correction.

Variable	Statistical Test	Adj. P-value (original)	Adj. P-value (unconfounded)
Ancestry axis 1	Kruskal-Wallis	7.24e-40	2.99e-9
Ancestry axis 2	Kruskal-Wallis	1.04e-11	2.29e-7
Ancestry axis 3	Kruskal-Wallis	6.07e-11	2.69e-3
Ancestry axis 4	Kruskal-Wallis	8.11e-22	8.16e-11
Age	Kruskal-Wallis	9.36e-8	0.888
Sex	Chi-squared	0.118	0.899
Height	Kruskal-Wallis	8.38e-2	0.888
Weight	Kruskal-Wallis	5.51e-2	0.951
BMI	Kruskal-Wallis	2.19e-3	0.899
Camera system	Chi-squared	3.01e-78	0.899
Face size	Kruskal-Wallis	3.72e-2	0.888

Table 1: Adjusted P-values of the statistical tests between the clustering of G_1 and every ancestry axis and confounder. Kruskal-Wallis test was used for continuous variables and Chisquared test for categorical variables. All P-values have been corrected for multiple testing via the Benjamini-Hochberg (BH) procedure. Column 3 are adjusted P-values from the original clustering based on all vectors in G_1 while column 4 from the unconfounded clustering. Adjusted P-values lower than 0.05 (threshold for significance) are in bold.

To investigate if the derived subgroups from the unconfounded G_1 clustering capture well the population structure, we adopted 3,519 European ancestry informative markers (EuroAIMs) found by Tian et al. (Tian *et al.* 2009), which are SNPs capable of distinguishing European subpopulations. We first mapped the SNP embedding G_2 to the space of G_1 and then, in the joint space, selected 3,519 SNPs with highest cosine similarities to the centroids of derived subgroups, which drive the clustering of individuals. We selected the same number of SNPs as the EuroAIMs for better comparison. Because only 13 EuroAIMs were originally included in our dataset, we looked at the gene level and found that 856 out of the 3,519 selected SNPs are located in the same genes as the EuroAIMs. A hypergeometric test showed that this fraction is significantly (P-value = 2.15e-63) higher than a random selection from all SNPs in our dataset, indicating that the genetic basis of the derived subgrouping is statistically significantly associated with the European population structure. We also checked for the subgroups derived from the original G_1 clustering and found only 40 out of 3,519 selected SNPs that are located in the EuroAIM genes. The P-value of 1 from the hypergeometric test also implied this fraction is not statistically higher than a random selection. This result further proved that our deconfounding strategy has successfully led to population subgroups that could better highlight the population structure.

After showing the validity of our unconfounded population subgrouping, we focused on the characterization of two subgroups that are most associated with the four ancestry axes (Supp. Table 1), namely subgroup 2 and 7. The top 0.1% SNPs selected for population subgroup 2 were significantly enriched in over 500 HPO terms, with the top 20 terms clearly related to skeletal morphology or bone formation (Fig. 7A). It is in line with the highlighted areas on the face (Fig. 7B), e.g., the nasal bone, the zygomatic bone, maxilla, and most part of the mandible. This result characterized population subgroup 2 with facial skeleton. Note that the frontal bone does not show up as much as the facial bones, implying the different morphology of cranial and facial skeleton (Anderson et al. 2024). Some other enriched HPO terms involve anemia, telangiectasia and neutrophil, which are related to the blood. Whereas the genetic representation of population subgroup 7 found 72 significantly associated HPO terms in total, and many of the top 20 terms are strongly involved in kidney function (Fig. 7C). Meanwhile, the eye area was remarkably underlined on the mean face (Fig. 7D), which supports the common embryogenic stage of eyes and kidneys and the reported relationship between eye and kidney diseases (Bodaghi, Massamba and Izzedine 2014). Therefore, population subgroup 7 is likely characterized by eye morphology related to kidney function. Other enriched terms indicate the involvement of kidney function in diabetes, hair development, tooth development, etc.



Fig. 7: Genetic and facial representation for population subgroup 2 (top, A and B) and 7 (bottom, C and D). The genetic representation was obtained via GREAT analysis (left) for the top 20 HPO terms enriched in the top 0.1% SNPs highlighted for population subgroup 2 (A) and 7 (C). We used the default and recommended settings of GREAT analysis with binomial test and BH correction. For the facial representation, the cosine similarity between each facial landmark and the centroid of subgroup 2 (B) and 7 (D) separately was plotted on the mean face. Red color indicates higher value while blue color indicates lower value.

5 Discussion and conclusion

In this study, we proposed INMTD, a framework that integrates both 2D matrices and 3D tensors for unconfounded clustering, and applied it to a real-life facial-genomic dataset to evaluate its performance and find an unconfounded subgrouping for European population structure. We derived 20 unconfounded population subgroups with their representative genetic and facial characteristics, providing the potential for more precise healthcare towards each subpopulation.

This work applies INMTD to facial and genomic data, which reflect a large fraction of the population structure. A comprehensive subgrouping of population structure has the potential to facilitate precision medicine where individuals in each population subgroup may receive tailored medical decisions based on their intrinsic characteristics (Saria and Goldenberg 2015; Loh, Cao and Zhou 2019). For instance, special medical treatment may be given to individuals in subgroup 2 for their different skeleton development and individuals in subgroup 7 for their distinct kidney function.

While classic matrix or tensor decomposition models only focus on a single dataset and current integrative matrix factorization models cannot deal with higher-dimensional data structures, INMTD is able to jointly decompose both matrix and tensor data. Another key feature of INMTD is its orthogonality constraint on G_1 for better clustering and interpretation. An orthogonal matrix has all its vectors independent to each other, and therefore, every vector can be investigated specifically for its characteristics. We further normalized the vectors of G_1 to make them orthonormal, resembling naturally a cluster indicator matrix. Optionally, we can impose orthogonality on any embedding if needed.

As an extension of joint NMF model, INMTD has the potential to predict facial images from genotypes or vice versa of new samples, in a similar fashion as Akata et al. (Akata, Thurau and Bauckhage 2011) In the former case, given the genotypes x_{12} of an individual that has not been seen by the model, we first solve $\min_{g_1 \ge 0} J = ||x_{12} - g_1 S_{12} G_2^T||_F^2$ for g_1 and then predict its facial image $x_{134} = S_{134} \times_1 g_1 \times_2 G_3 \times_3 G_4$. Alternatively, we could find the closest image from the cohort to x_{134} according to cosine similarity or the image whose corresponding embedding vector in G_1 space is closest to g_1 . In addition, a new sample can be classified into one of the derived subgroups by assigning g_1 to its closest cluster centroid.

Even though INMTD was illustrated in a facial-genomic dataset for population subgrouping, it is not restricted to facial images and genomic data. INMTD can be applied to any 2D and 3D datasets for joint clustering, as long as they are non-negative or can be converted to non-negative values, e.g. transcriptomics or epigenomics as 2D matrices and CT scans or time series data as 3D tensors. It is also possible to further extend INMTD to deal with more than two data types or data of higher dimensions, e.g. a moving 3D image (4D), by adding extra embeddings to the model and objective function.

There are two main limitations of INMTD for future improvements. The first one is that it determines the ranks of each embedding based on a rule of thumb because of the extremely large scale of genomic dataset. Nevertheless, in most cases the data dimensionality would be feasible for INMTD model on a modern computer or computing server and the user could choose the optimal ranks via cross-validation or other non-heuristic methods. The other potential deficiency comes from the post-hoc confounder correction, which removes confounded vectors but in a fairly strict manner with the risk of 'over-correcting'. A more compromising strategy could be adding a regularization term in the objective function to iteratively minimize the confounding effect in G_1 , which indicates our future work.

In conclusion, with the surge in biological data in diverse formats and the growing demand for personalized medicine with Big Data, INMTD is envisaged to become an essential tool for

integrating multi-view datasets of varying dimensions, enabling meaningful and unconfounded clustering. We are confident that INMTD has the potential of widespread adoption in the future due to its exceptional performance and ease-of-interpretation.

Data availability

For the 3D Facial Norms dataset, genotypic markers are available to the research community through the dbGaP controlled-access repository (http:// www.ncbi.nlm.nih.gov/gap) at accession #phs000929.v1.p1. The raw source data for the phenotypes - the 3D facial surface models in .obj format - are available through the FaceBase Consortium (https://www.facebase.org) at accession #FB00000491.01. Access to these 3D facial surface models requires proper institutional ethics approval and approval from the FaceBase data access committee. The PSU and IUPUI datasets were not collected with broad data sharing consent. Code for INMTD is publicly available on GitHub: https://github.com/ZuqiLi/INMTD.

Acknowledgements

This project has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Sklodowska-Curie grant agreements No. 813533 (MLFPM) and No 860895 (TranSYS). We also want to acknowledge the European Research Council (ERC) Consolidator Grant No. 770827, the Spanish State Research Agency and the Ministry of Science and Innovation MCIN grant PID2022-141920NB-I00 / AEI /10.13039/501100011033/ FEDER, UE, and the Department of Research and Universities of the Generalitat de Catalunya code 2021 SGR 01536.

Conflict of interest

None declared.

References

- Akata Z, Thurau C, Bauckhage C. Non-negative Matrix Factorization in Multimodality Data for Segmentation and Label Prediction. 2011.
- Anderson BW, Kortz MW, Black AC *et al.* Anatomy, Head and Neck, Skull. *StatPearls*. Treasure Island (FL): StatPearls Publishing, 2024.
- Antonelli L, Guarracino MR, Maddalena L *et al.* Integrating imaging and omics data: A review. *Biomedical Signal Processing and Control* 2019;**52**:264–80.
- Bo Li, Guoxu Zhou, Cichocki A. Two Efficient Algorithms for Approximately Orthogonal Nonnegative Matrix Factorization. *IEEE Signal Process Lett* 2015;**22**:843–6.
- Bodaghi B, Massamba N, Izzedine H. The eye: a window on kidney diseases. *Clin Kidney J* 2014;**7**:337–8.
- Broadbent C, Song T, Kuang R. Deciphering high-order structures in spatial transcriptomes with graph-guided Tucker decomposition. *Bioinformatics* 2024;**40**:i529–38.
- Cavelaars AE, Kunst AE, Geurts JJ *et al.* Persistent variations in average height between countries and between socio-economic groups: an overview of 10 European countries. *Ann Hum Biol* 2000;**27**:407–21.

- Chalise P, Fridley BL. Integrative clustering of multi-level 'omic data based on non-negative matrix factorization algorithm. Peddada SD (ed.). *PLoS ONE* 2017;**12**:e0176278.
- Chauvel C, Novoloaca A, Veyre P *et al.* Evaluation of integrative clustering methods for the analysis of multi-omics data. *Briefings in Bioinformatics* 2020;**21**:541–52.
- Chen L, Zeng H, Xiang Y *et al.* Histopathological Images and Multi-Omics Integration Predict Molecular Characteristics and Survival in Lung Adenocarcinoma. *Front Cell Dev Biol* 2021;**9**:720110.
- Ding C, Li T, Peng W *et al.* Orthogonal nonnegative matrix t-factorizations for clustering. *Proceedings of the 12th ACM SIGKDD International Conference on Knowledge Discovery and Data Mining.* Philadelphia PA USA: ACM, 2006, 126–35.
- Dissez G, Ceddia G, Pinoli P *et al.* Drug Repositioning Predictions by Non-Negative Matrix Tri-Factorization of Integrated Association Data. *Proceedings of the 10th ACM International Conference on Bioinformatics, Computational Biology and Health Informatics.* Niagara Falls NY USA: ACM, 2019, 25–33.
- Ferreira L, Hitchcock DB. A Comparison of Hierarchical Methods for Clustering Functional Data. *Communications in Statistics Simulation and Computation* 2009;**38**:1925–49.
- Gargano MA, Matentzoglu N, Coleman B *et al.* The Human Phenotype Ontology in 2024: phenotypes around the world. *Nucleic Acids Res* 2024;**52**:D1333–46.
- Ghosal A, Nandy A, Das AK *et al.* A Short Review on Different Clustering Techniques and Their Applications. In: Mandal JK, Bhattacharya D (eds.). *Emerging Technology in Modelling and Graphics*. Singapore: Springer Singapore, 2020, 69–83.
- He X, Gumbsch T, Roqueiro D *et al.* Kernel conditional clustering and kernel conditional semisupervised learning. *Knowl Inf Syst* 2020;**62**:899–925.
- Hériché J-K, Alexander S, Ellenberg J. Integrating Imaging and Omics: Computational Methods and Challenges. *Annu Rev Biomed Data Sci* 2019;**2**:175–97.
- Khan A, Maji P. Low-Rank Joint Subspace Construction for Cancer Subtype Discovery. *IEEE/ACM Trans Comput Biol and Bioinf* 2019:1–1.
- Kim Y-D, Choi S. Nonnegative Tucker Decomposition. 2007 IEEE Conference on Computer Vision and Pattern Recognition. Minneapolis, MN, USA: IEEE, 2007, 1–8.
- Kodinariya TM, Makwana PR. Review on determining number of Cluster in K-Means Clustering. 2013.
- Lee DD, Seung HS. Learning the parts of objects by non-negative matrix factorization. *Nature* 1999;**401**:788–91.
- Liu J, Brodley CE, Healy BC *et al.* Removing confounding factors via constraint-based clustering: An application to finding homogeneous groups of multiple sclerosis patients. *Artif Intell Med* 2015;**65**:79–88.

- Loh W-Y, Cao L, Zhou P. Subgroup identification for precision medicine: A comparative review of 13 methods. *WIREs Data Mining and Knowledge Discovery* 2019;**9**:e1326.
- Luo J, Liu Y, Liu P *et al.* Data Integration Using Tensor Decomposition for the Prediction of miRNA-Disease Associations. *IEEE J Biomed Health Inform* 2022;**26**:2370–8.
- Malod-Dognin N, Petschnigg J, Windels SFL *et al.* Towards a data-integrated cell. *Nat Commun* 2019;**10**:805.
- McLean CY, Bristor D, Hiller M *et al.* GREAT improves functional interpretation of cisregulatory regions. *Nat Biotechnol* 2010;**28**:495–501.
- Pourhoseingholi MA, Baghestani AR, Vahedi M. How to control confounding effects by statistical analysis. *Gastroenterol Hepatol Bed Bench* 2012;**5**:79–83.
- Price AL, Patterson NJ, Plenge RM *et al.* Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet* 2006;**38**:904–9.
- Rappoport N, Shamir R. Multi-omic and multi-view clustering algorithms: review and cancer benchmark. *Nucleic Acids Research* 2018;**46**:10546–62.
- Saria S, Goldenberg A. Subtyping: What It is and Its Role in Precision Medicine. *IEEE Intell* Syst 2015;**30**:70–5.
- Schwarz M, Geryk J, Havlovicová M *et al.* Body mass index is an overlooked confounding factor in existing clustering studies of 3D facial scans of children with autism spectrum disorder. *Sci Rep* 2024;**14**:9873.
- Tian C, Kosoy R, Nassir R *et al.* European population genetic substructure: further definition of ancestry informative markers for distinguishing among diverse European ethnic groups. *Mol Med* 2009;**15**:371–83.
- Vijaya, Sharma S, Batra N. Comparative Study of Single Linkage, Complete Linkage, and Ward Method of Agglomerative Clustering. 2019 International Conference on Machine Learning, Big Data, Cloud and Parallel Computing (COMITCon). Faridabad, India: IEEE, 2019, 568–73.
- Wang X, Sun Z, Zhang Y *et al.* BREM-SC: a bayesian random effects mixture model for joint clustering single cell multi-omics data. *Nucleic Acids Research* 2020;**48**:5814–24.
- White JD, Indencleef K, Naqvi S *et al.* Insights into the genetic architecture of the human face. *Nat Genet* 2021;**53**:45–53.
- Yun Y, Xia W, Zhang Y *et al.* Self-representation and Class-Specificity Distribution Based Multi-View Clustering. *Neurocomputing* 2021;**437**:9–20.