Combined *in situ* niche analysis and pulse-chase labelling experiments unravel energy acquisition strategies in cold-water corals

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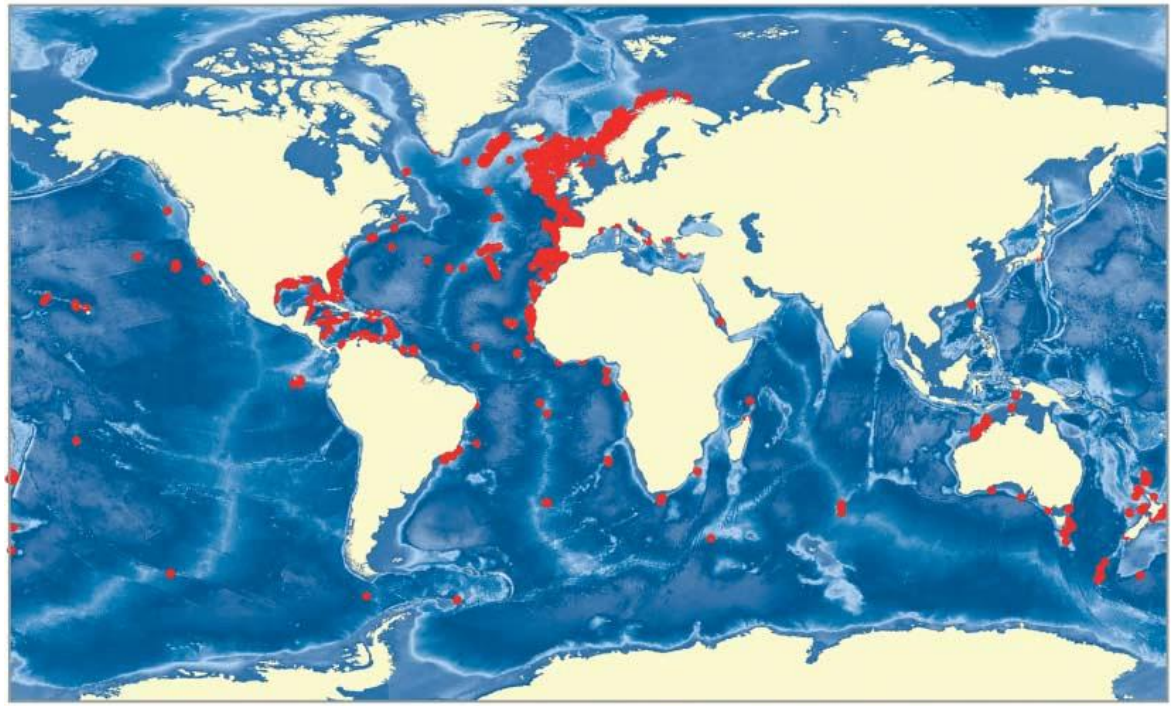
IsoEcol 2024 – Fredericton, Canada, 30/07/2024


COME TO THE
DARK SIDE
WE HAVE CORALS

Cold-water corals



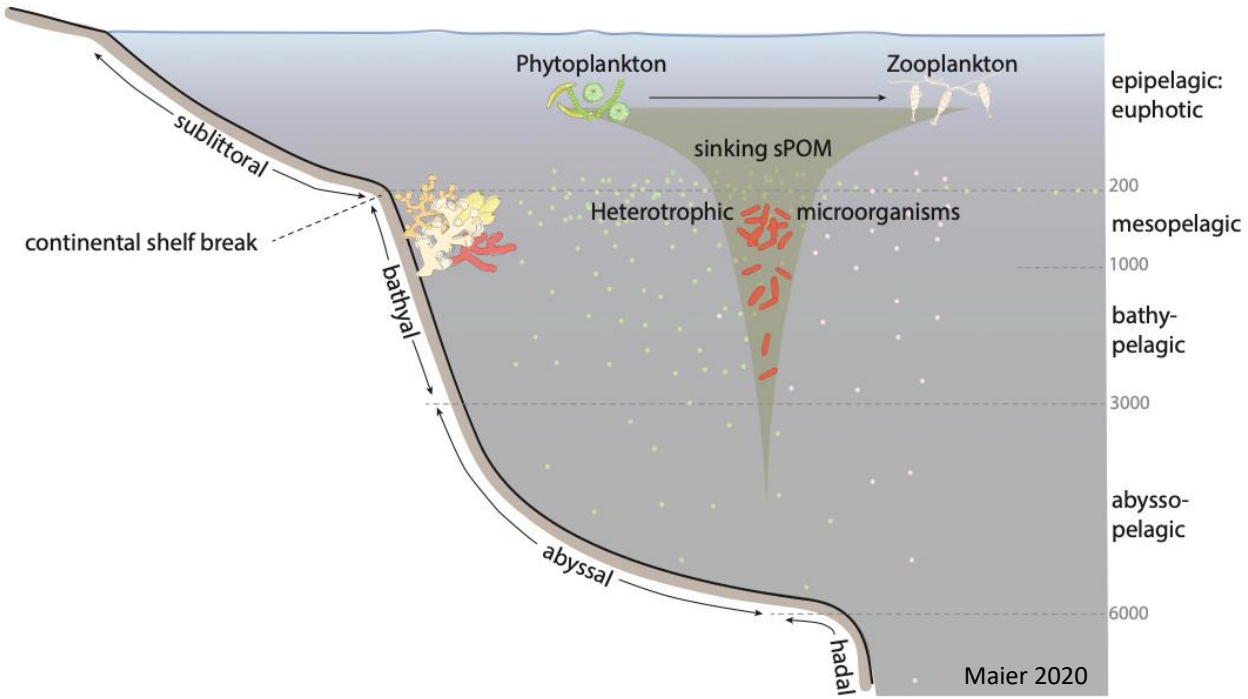
Roberts *et al.* 2006 Science 312: 543-547

Foundation species forming reefs with wordwide distribution, most commonly at depths ranging between 200 and 2000 meters

Those reefs provide habitats and/or nursery grounds for many other species: deep-sea biodiversity hotspots

Cold-water corals

Cold-water corals mostly depend on photosynthetic organic matter produced in the euphotic zone and exported through benthic-pelagic coupling



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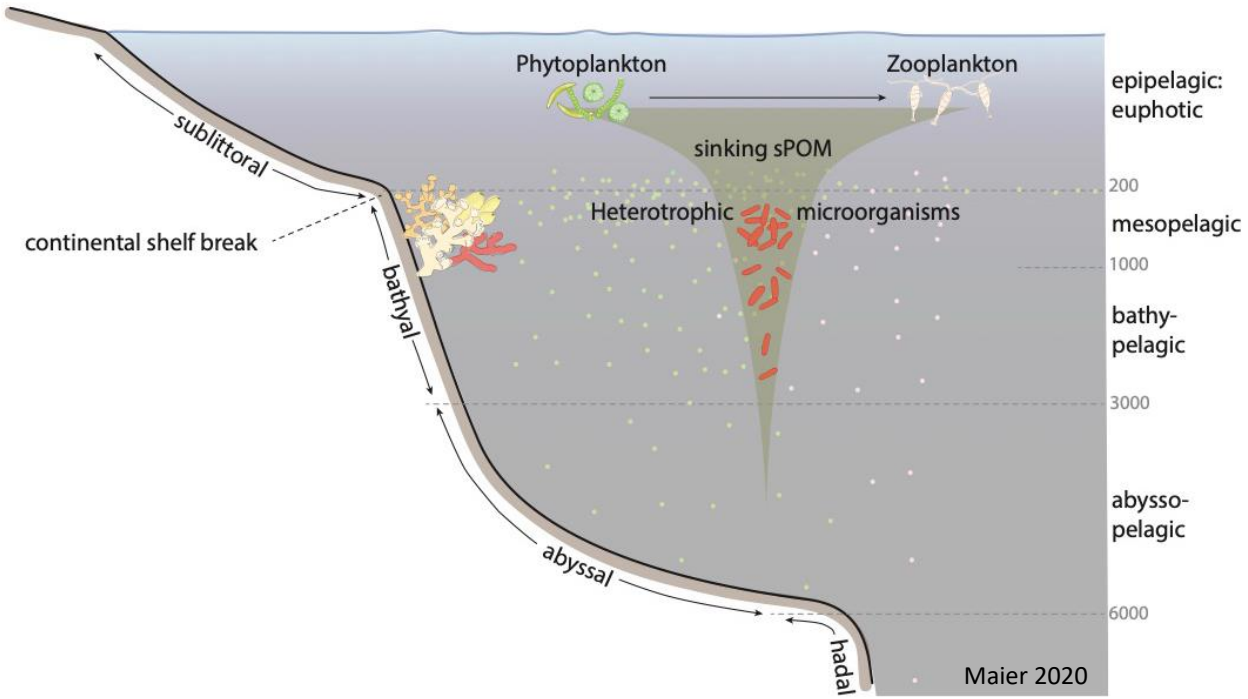
Food availability is therefore limited, and usually available through infrequent but massive resource pulses ("feast & famine" environment)



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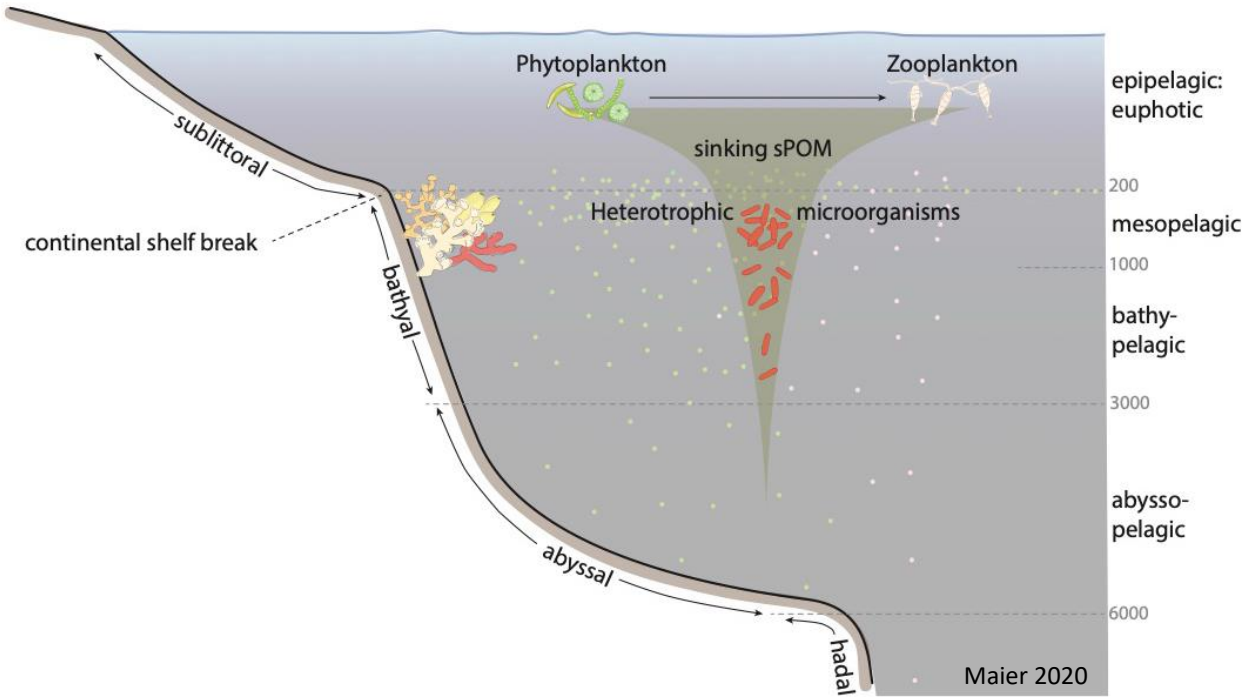
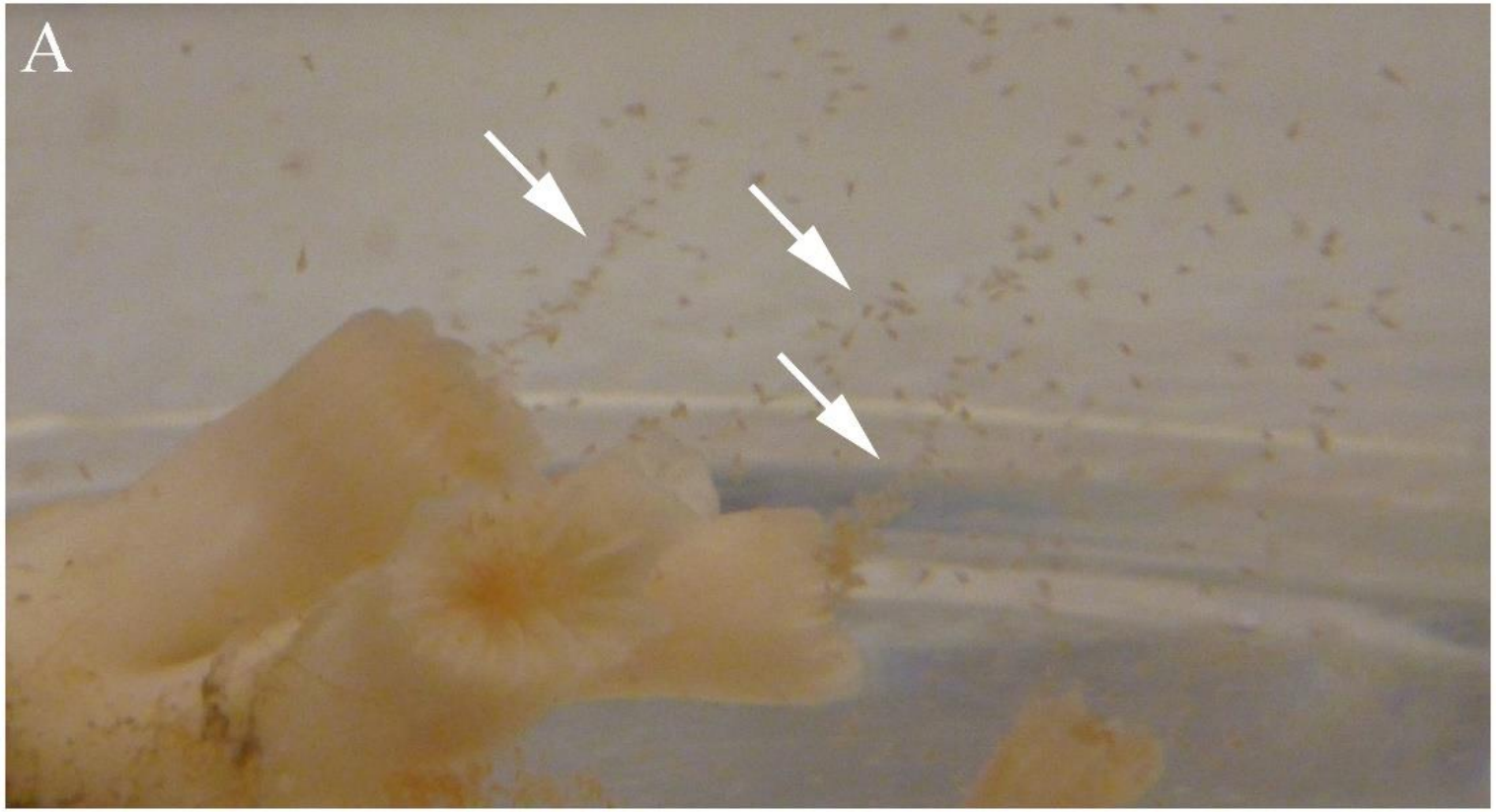
Energy acquisition is a major challenge for CWCs, and will likely become even more so in the next few decades as global change could shift coral energy budgets (ocean acidification)



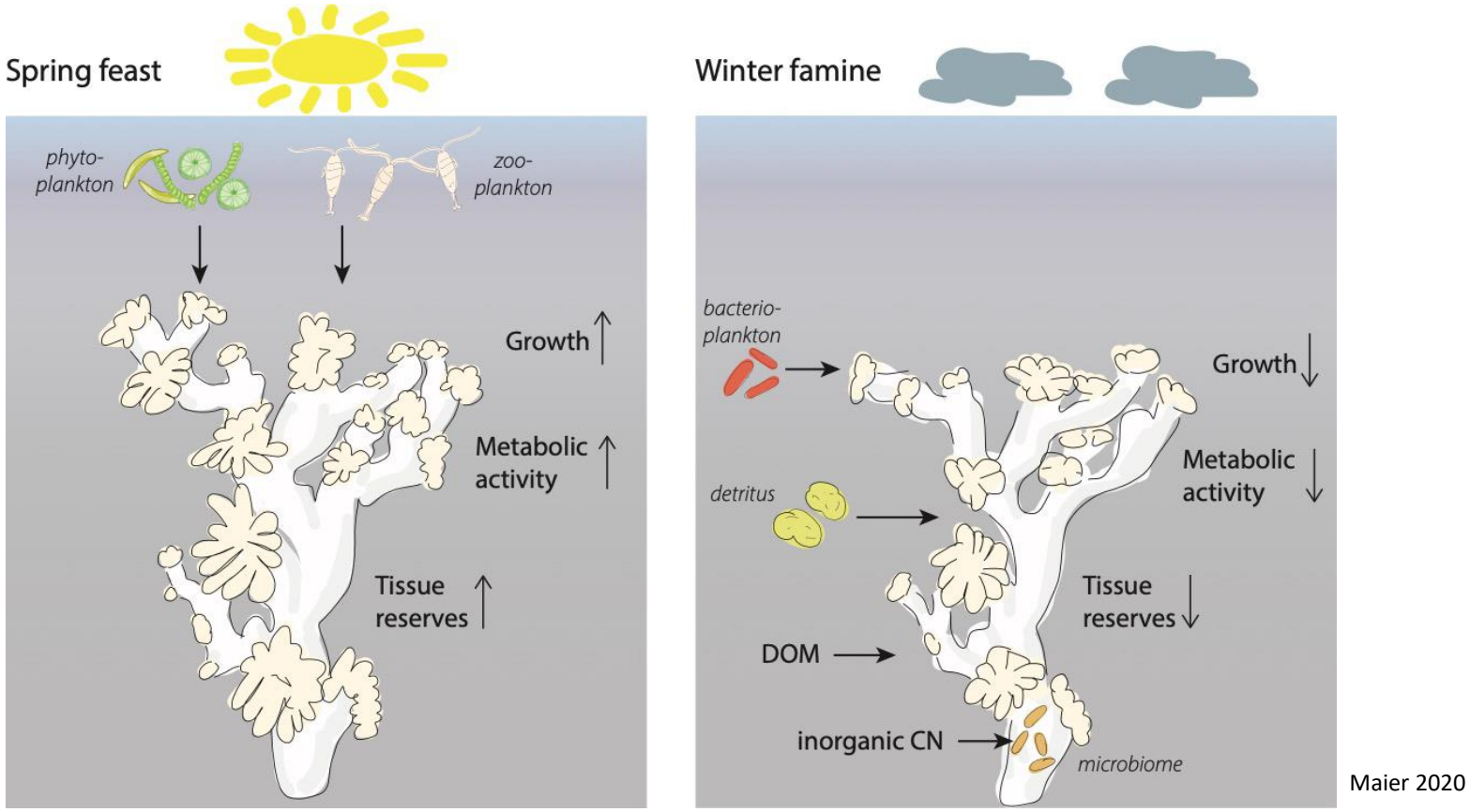
Adaptations to food-poor environments

Zetsche *et al.* 2016, PLoS ONE 11(2): e0146766



Passive suspension feeders: secret mucus to enhance particle/prey trapping

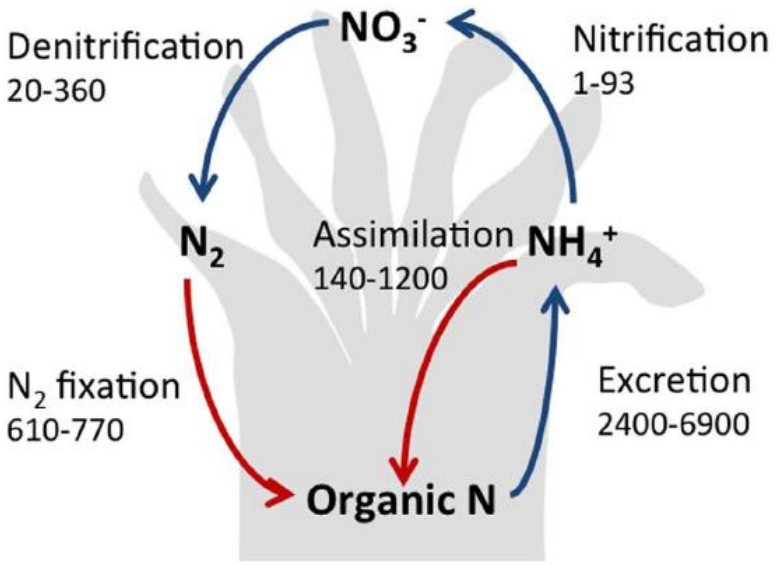
Adaptations to food-poor environments



Maier 2020

Selective suspension feeders: able to feed on multiple items (phytoplankton, zooplankton, bacterioplankton, detritus, etc.), with considerable ecological plasticity

Adaptations to food-poor environments



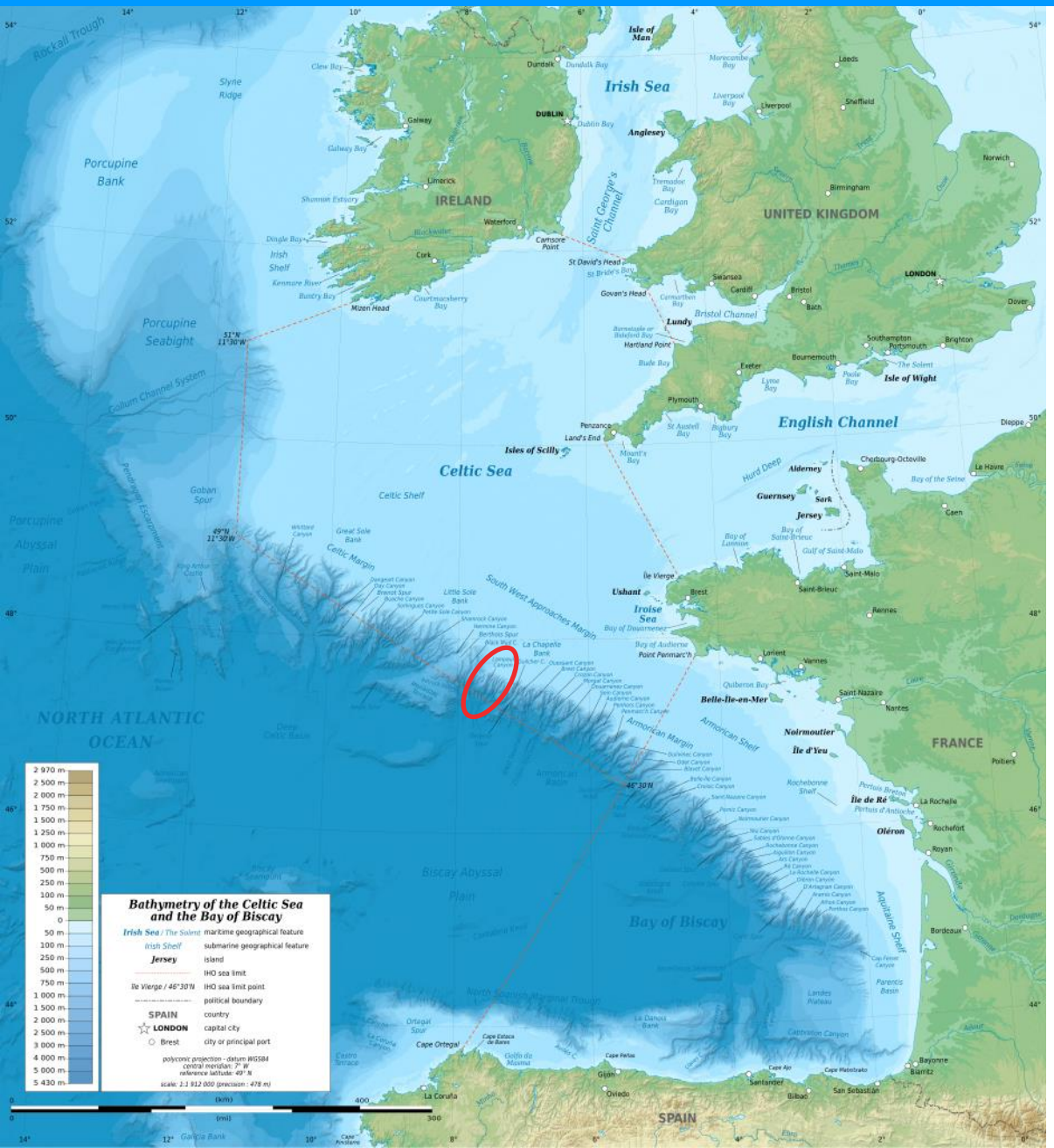
Middelburg *et al.* 2015, Sci. Rep. 5, 17962

No symbiosis with photosynthetic partners, but corals + micro-organisms colonizing them act as a holobiont

Micro-organisms : multiple metabolic activities

Desmophyllum pertusum derives nitrogen from bacterial metabolism and uses it to meet its nutritional requirements

Study site: Lampaul Canyon, Bay of Biscay

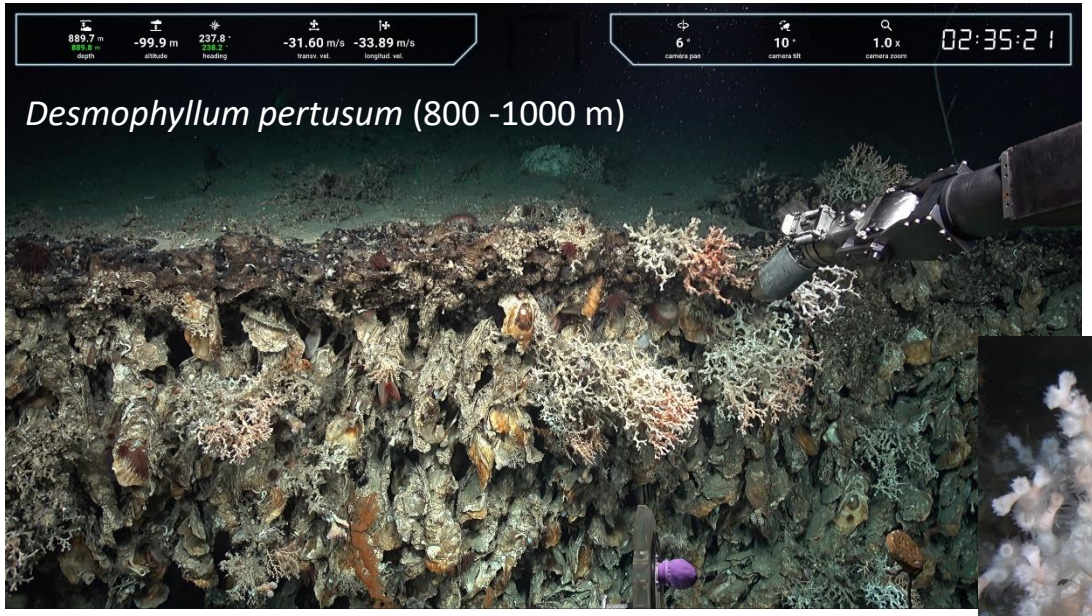


Extensive coral formations between 800 and 1600 m, built by three of the globally dominant cold-water reef building species

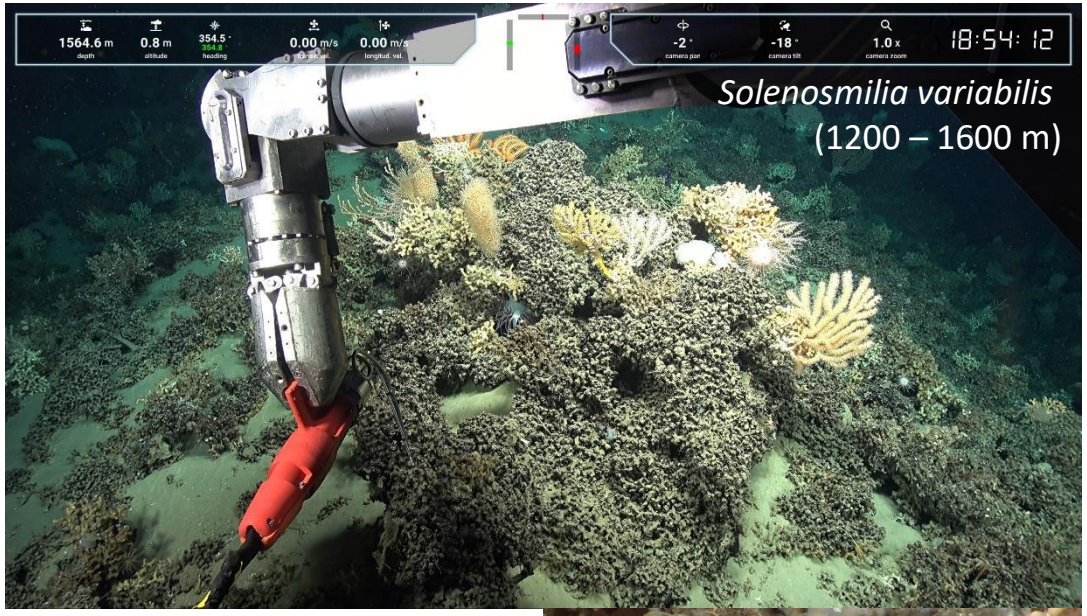
Image: Wikimedia Commons – Eric Gaba (Sting)

Study site: Lampaul Canyon, Bay of Biscay

Madrepora oculata (800 -1000 m)

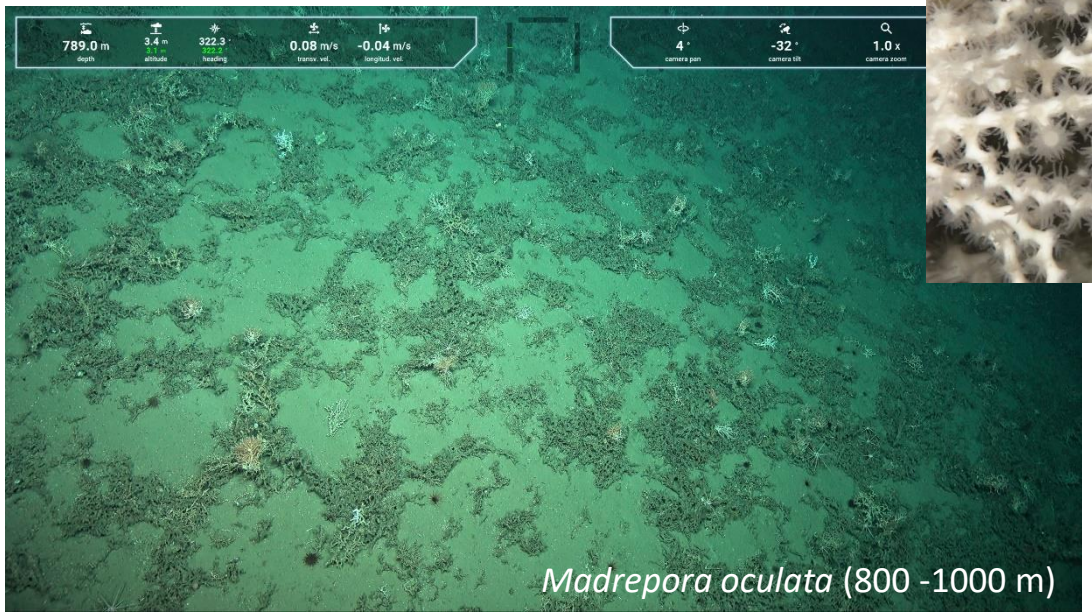


Desmophyllum pertusum (800 -1000 m)

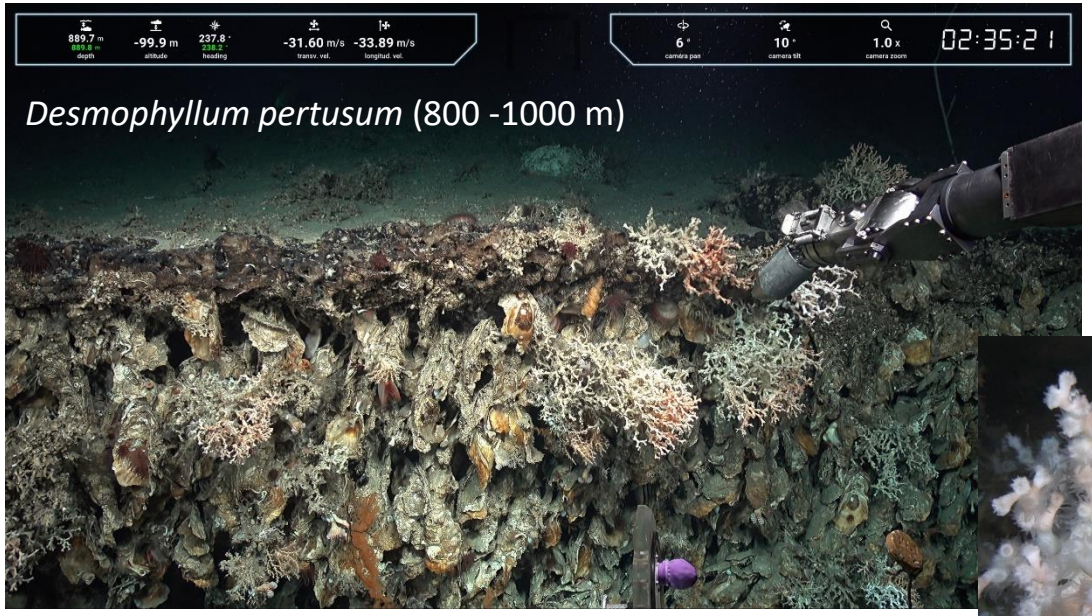


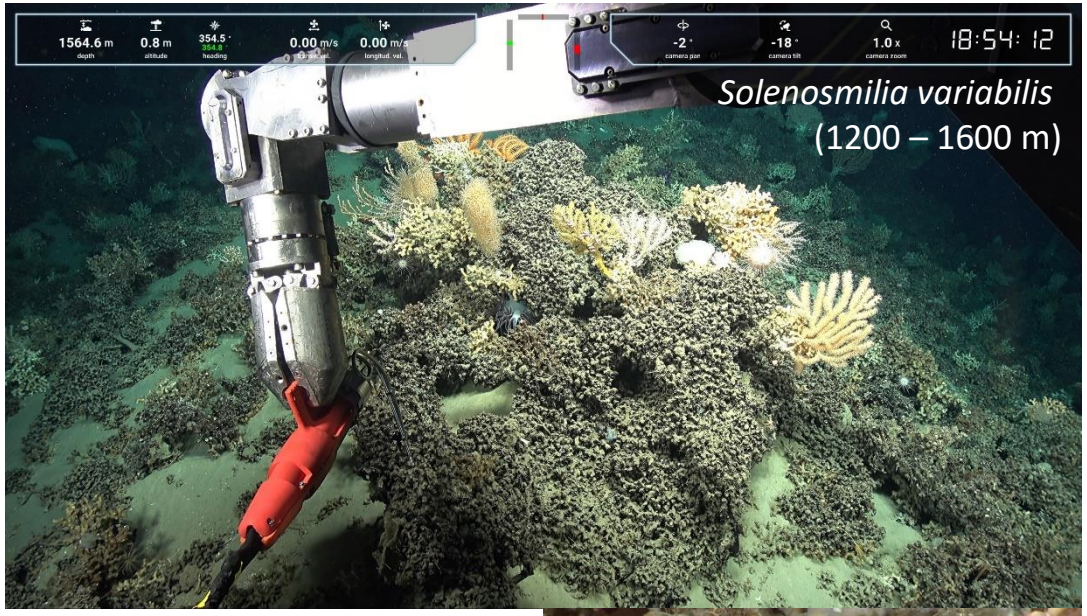
Solenosmilia variabilis
(1200 – 1600 m)

Study site: Lampaul Canyon, Bay of Biscay



M. oculata and *D. pertusum* are often found in syntopy (intertwined colonies)





Objectives

Cold-water corals have multiple feeding strategies (wide fundamental niches) and show considerable trophic plasticity.

Maier 2020

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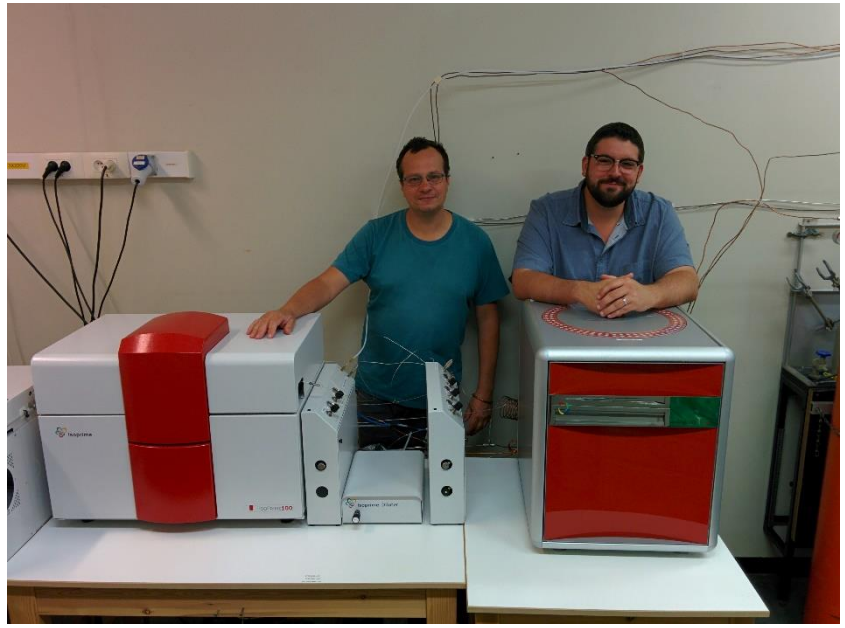
Cold-water corals have multiple feeding strategies (wide fundamental niches) and show considerable trophic plasticity.

In the Lampaul Canyon: what is the realized trophic niche of corals? How does each species acquire energy? Do they share dietary resources with each other or with associated fauna, particularly filter and/or suspension feeders?

Maier 2020

Methods



3 cruises onboard RV Thalassa (August/September 2021, 2022 and 2023)

Sampling of coral colonies and biomass-dominant associated fauna using either HROV Ariane or ROV Victor 6000

Dissection and extraction of relevant tissues

Measurement of stable isotope ratios of C, N and S using EA-IRMS

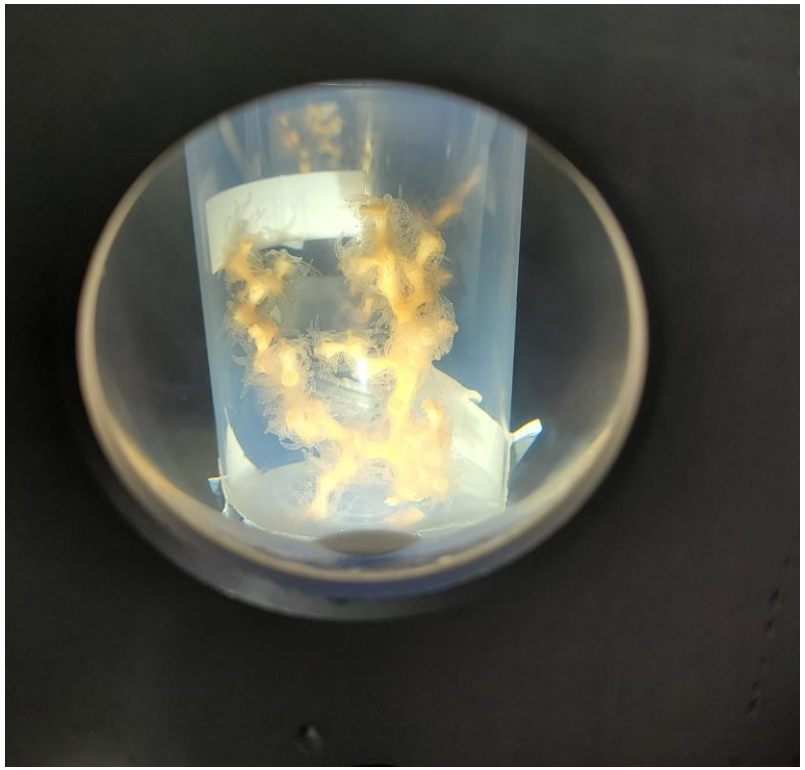
Analysis of fatty acid composition through GC-MS

Methods

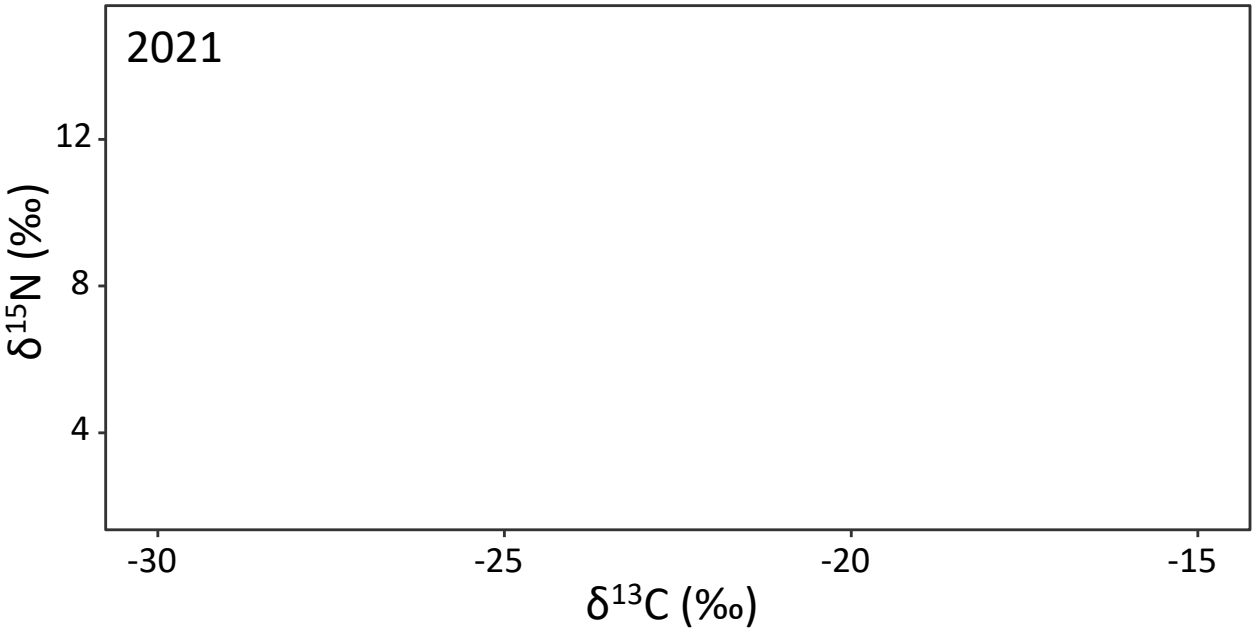
Ex situ labelling experiments onboard, during the cruises

Use of pressurized aquaria to recreate *in situ* pressure conditions

Addition of $^{15}NH_4Cl$ at environmental concentrations (3 µM), incubation for 24-72 hours, and quantification of inorganic nitrogen uptake by coral holobionts through stable isotope analysis



Isotopic niches of corals and associated fauna



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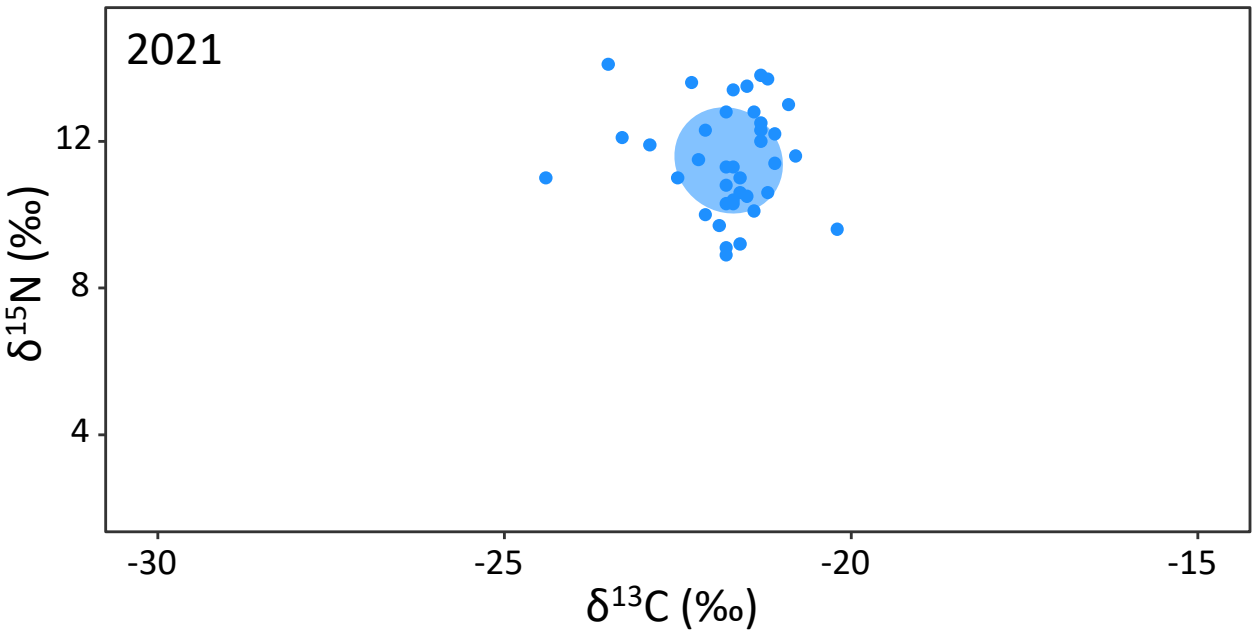

● Corals

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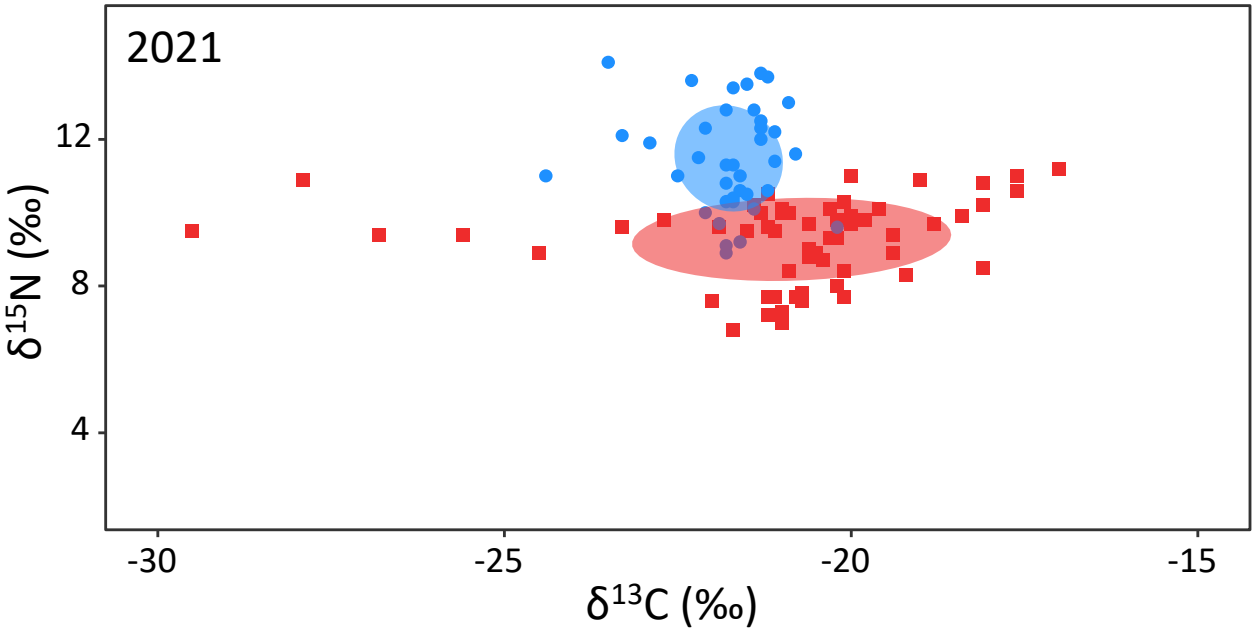

- Corals
- Associated filter and/or suspension feeders

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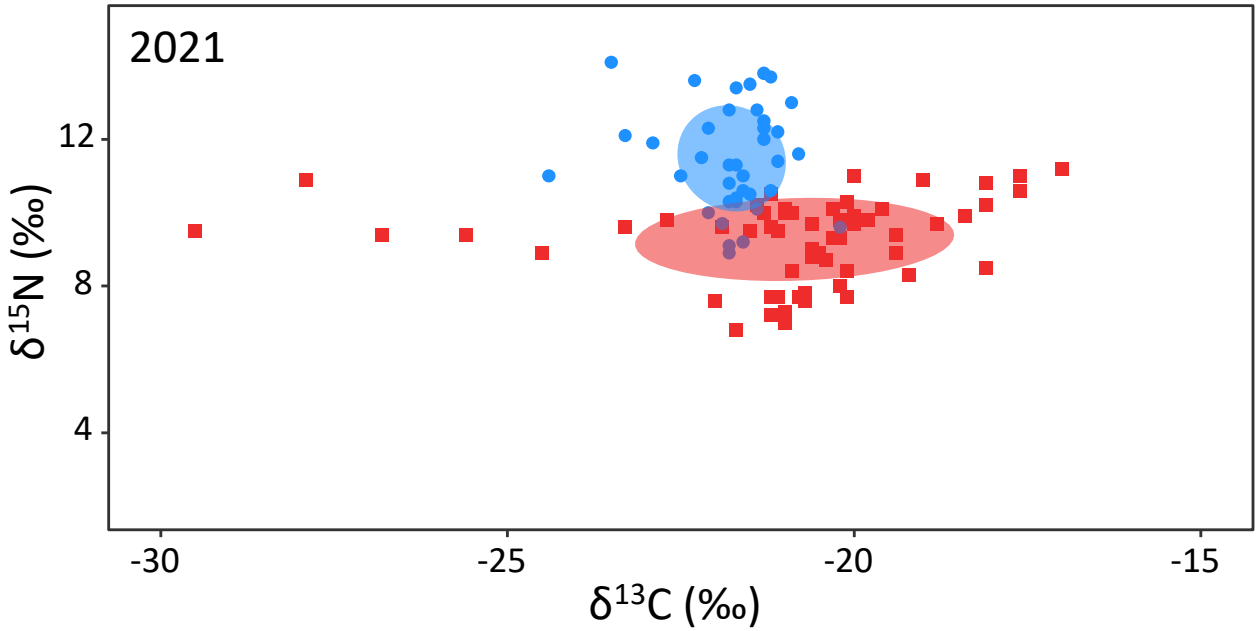

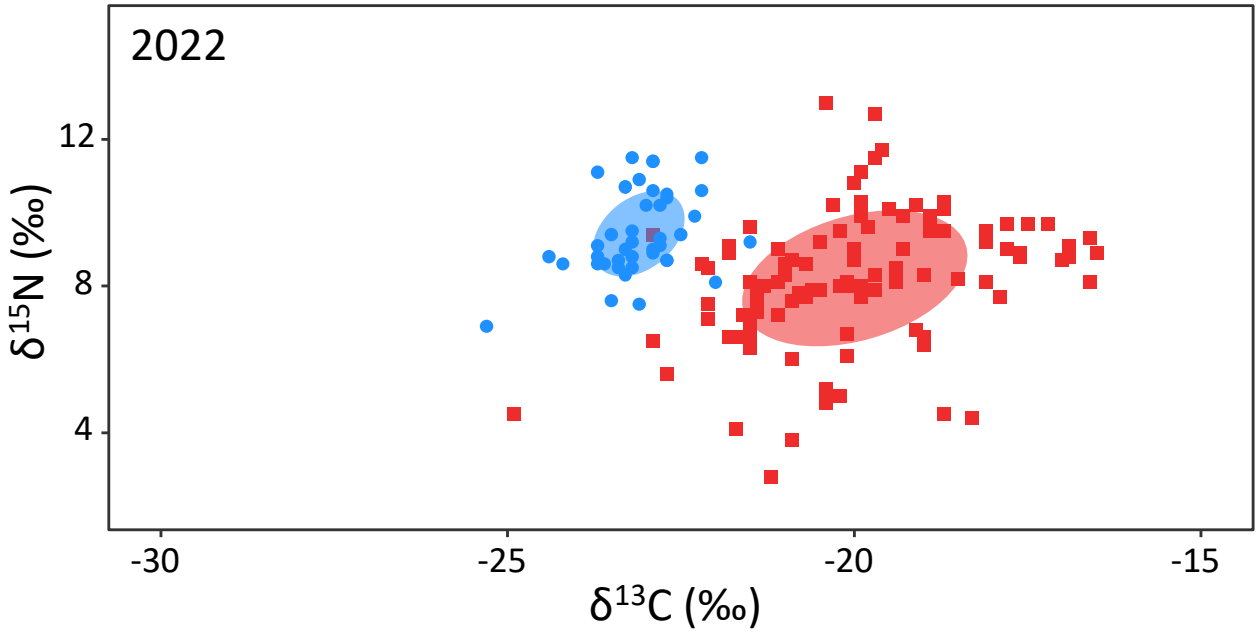

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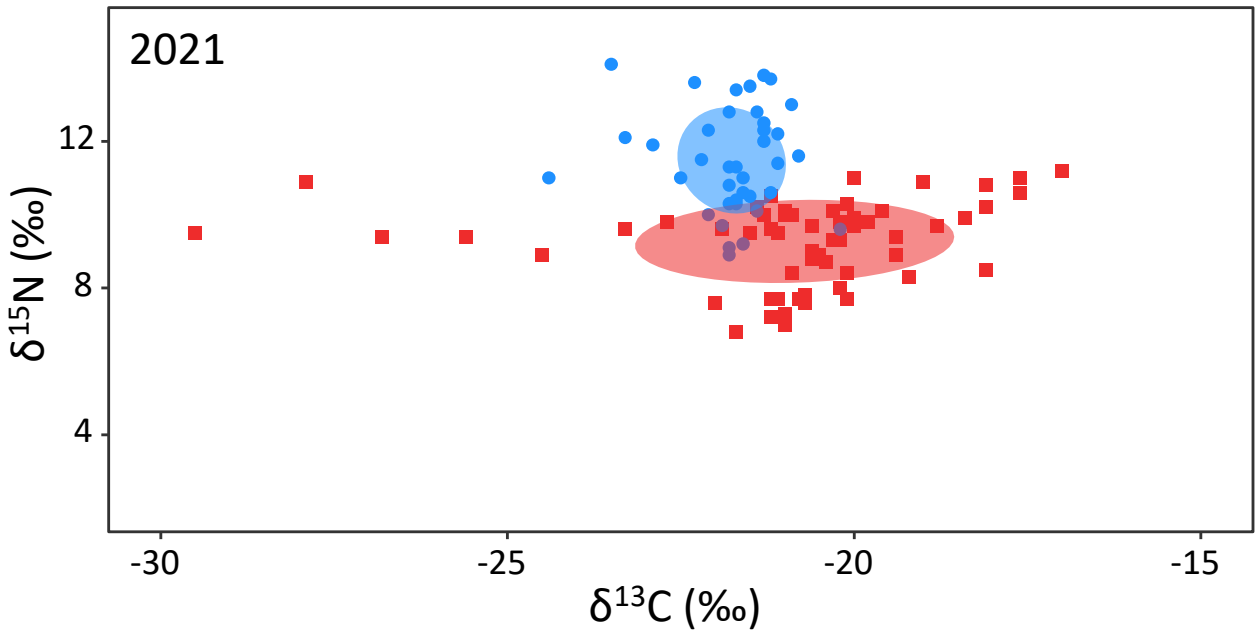

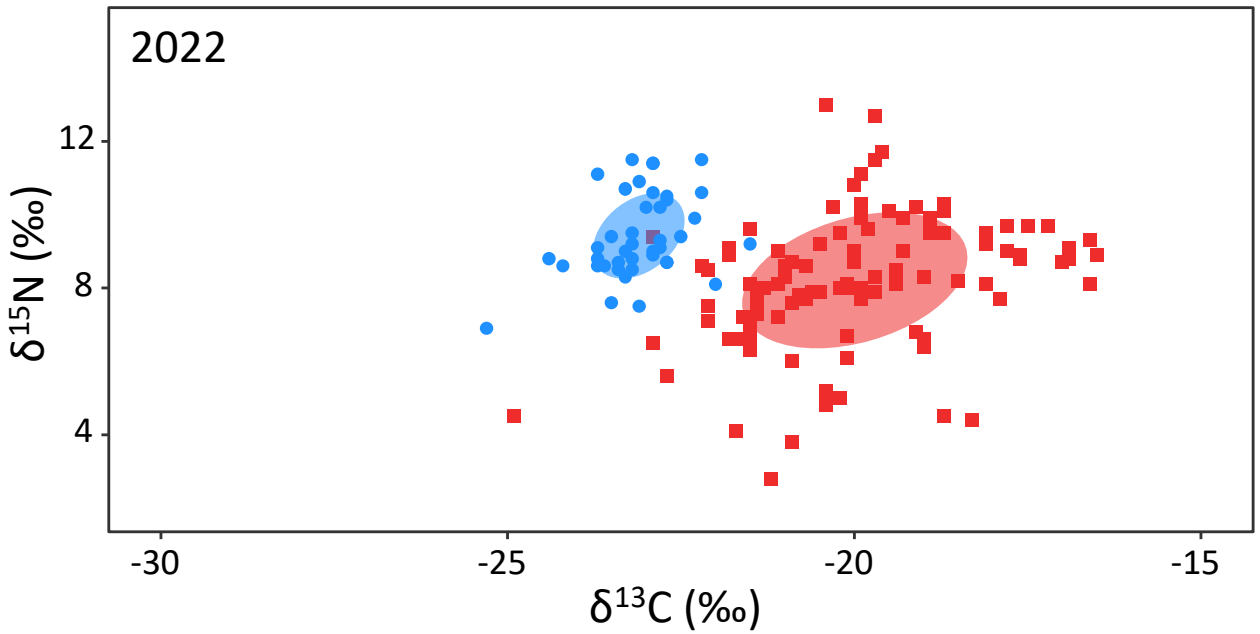

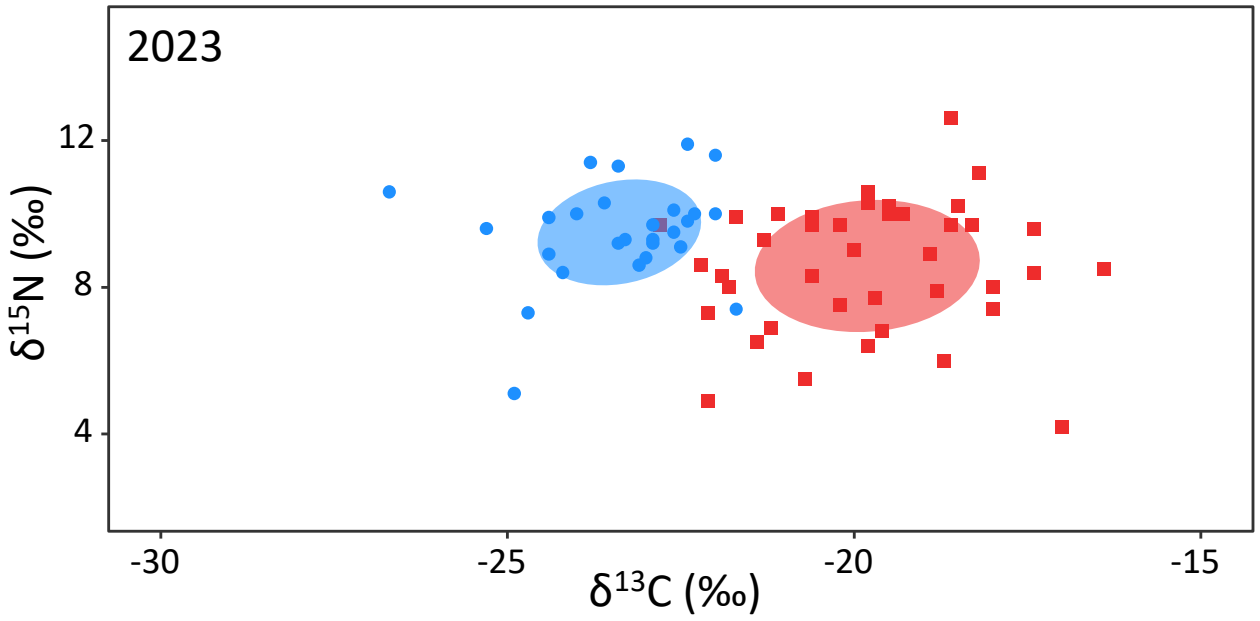

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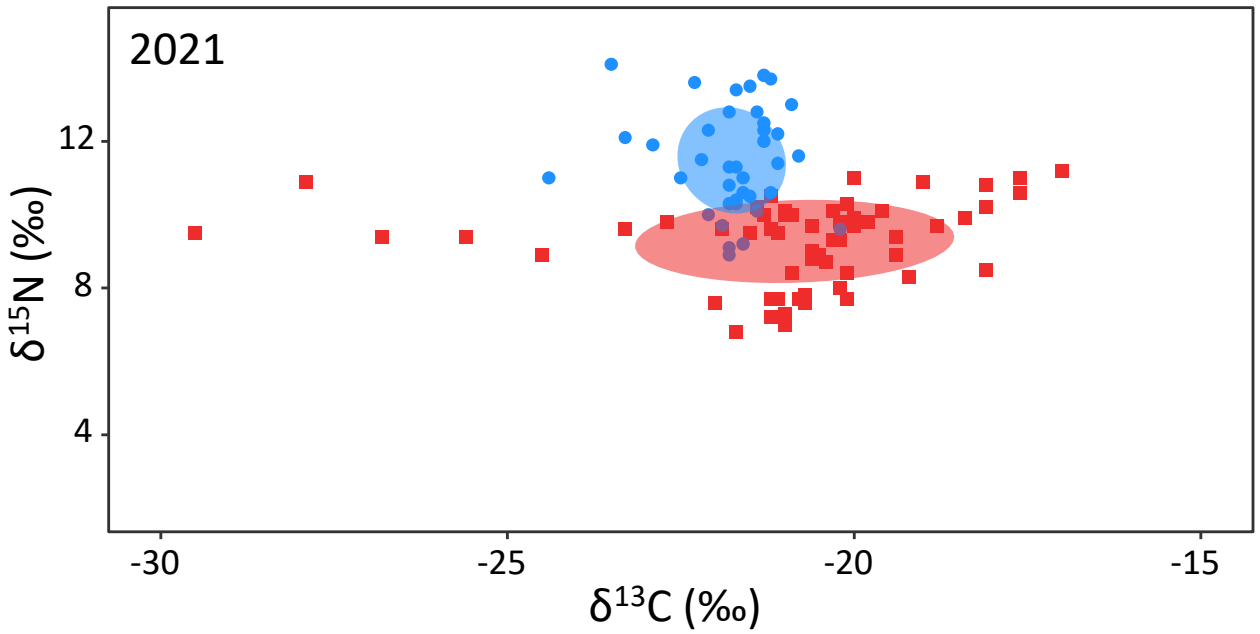

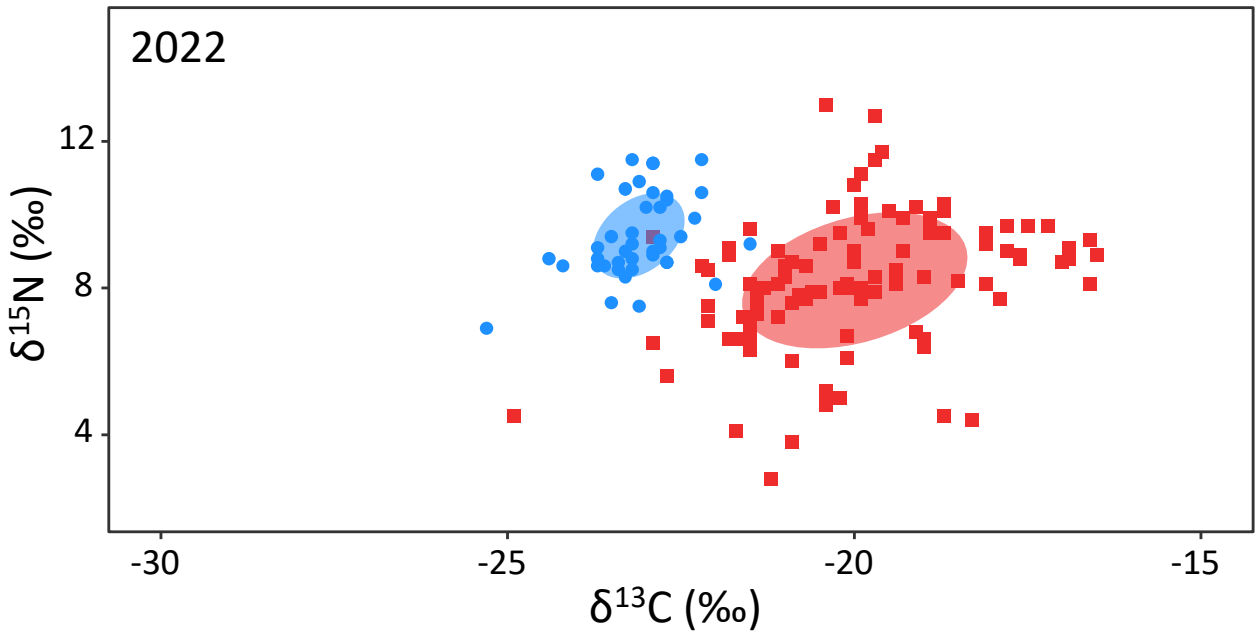

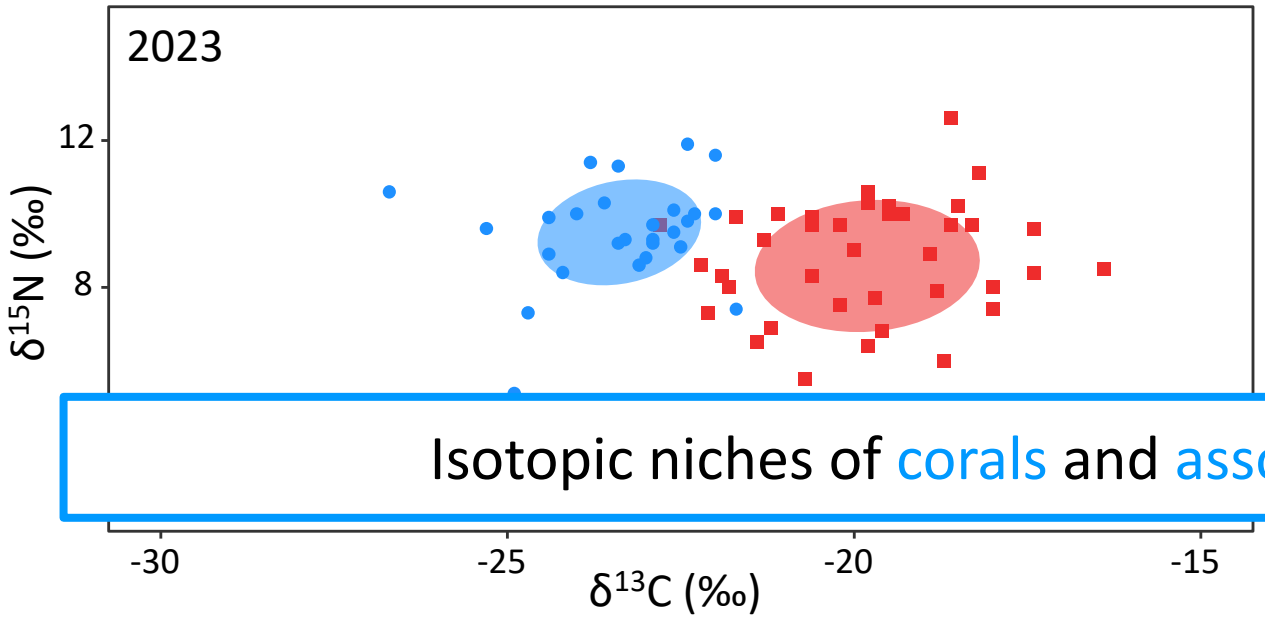

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Isotopic niches of corals and associated fauna: little to no overlap

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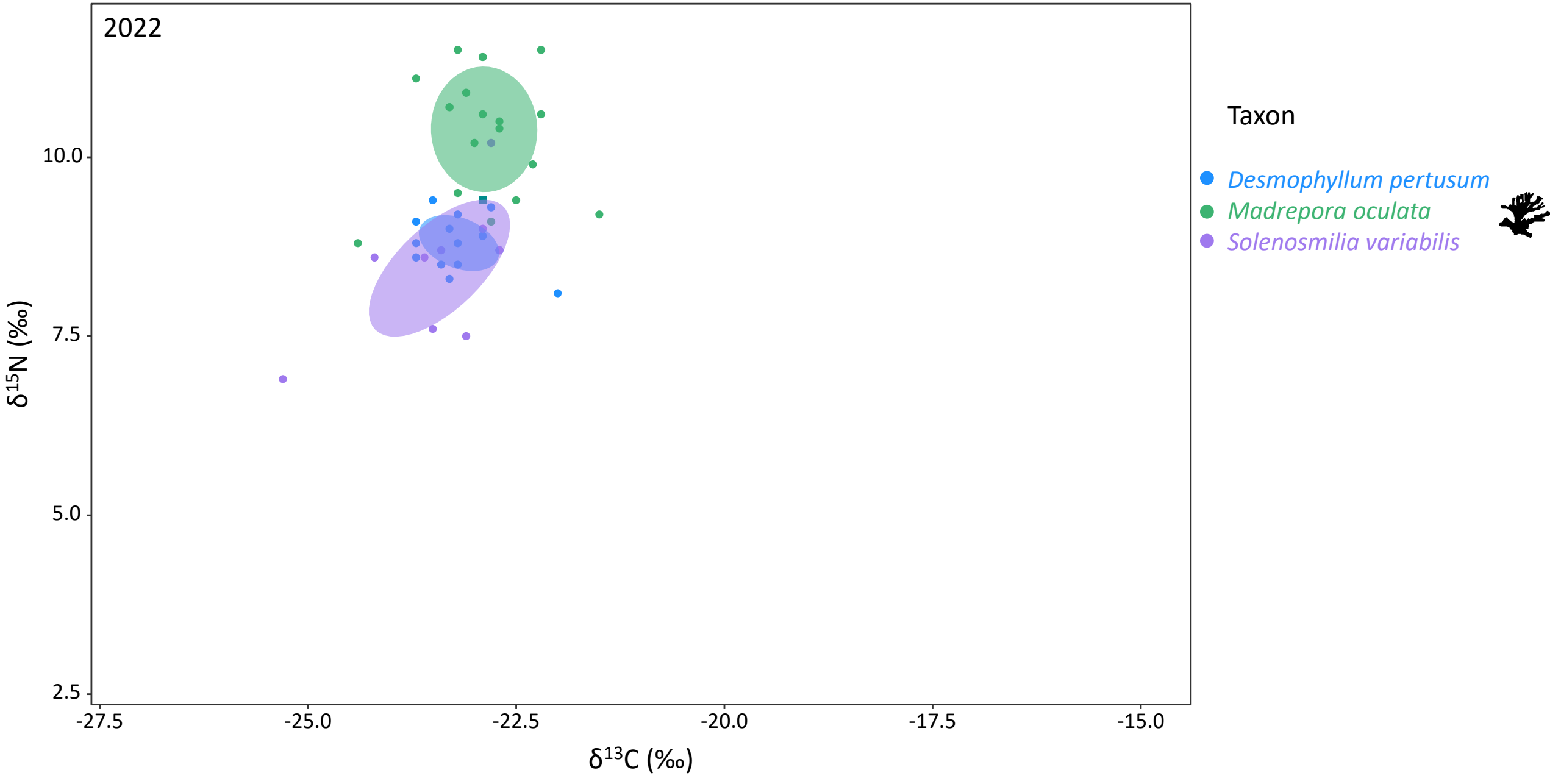

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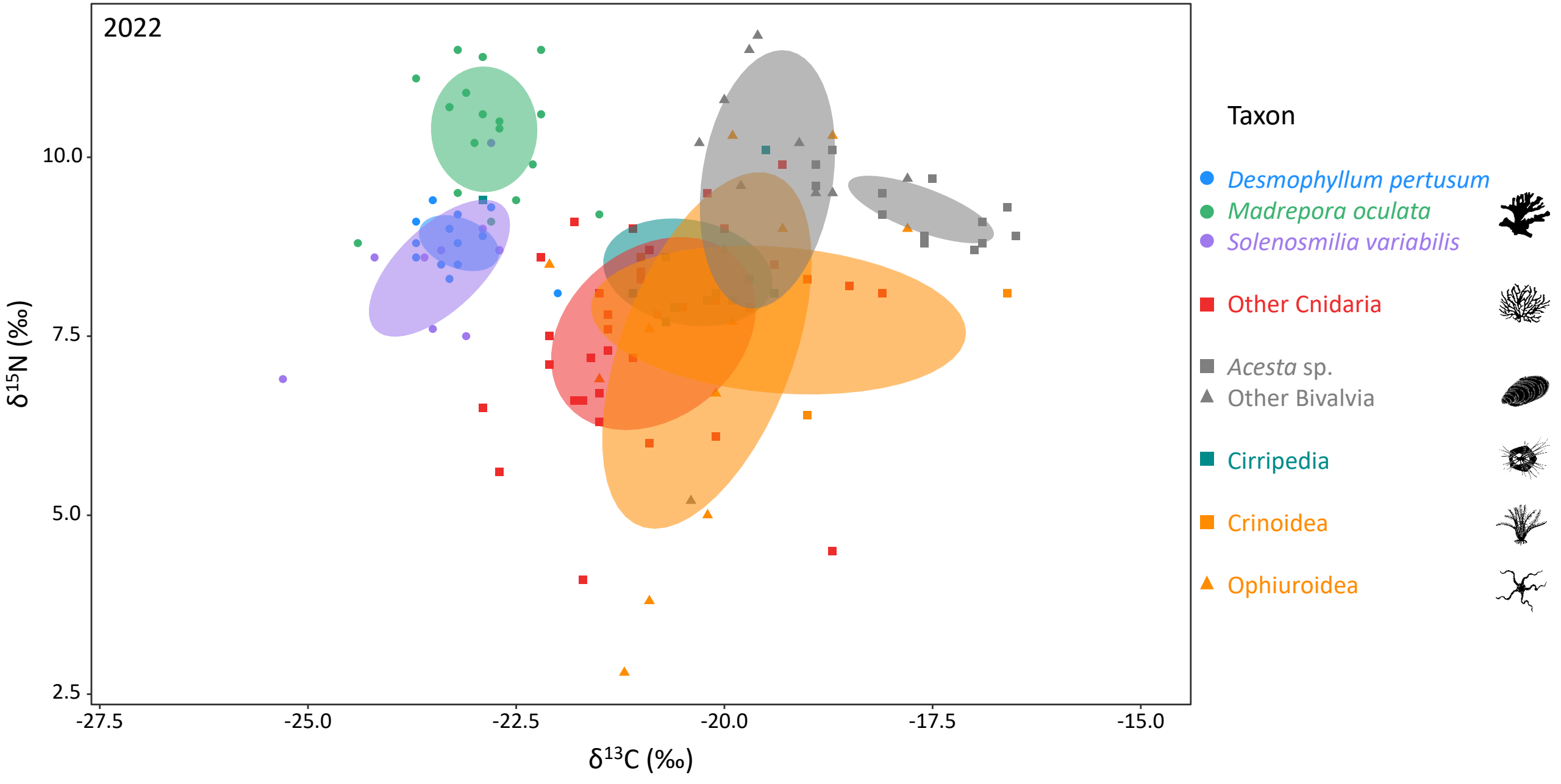

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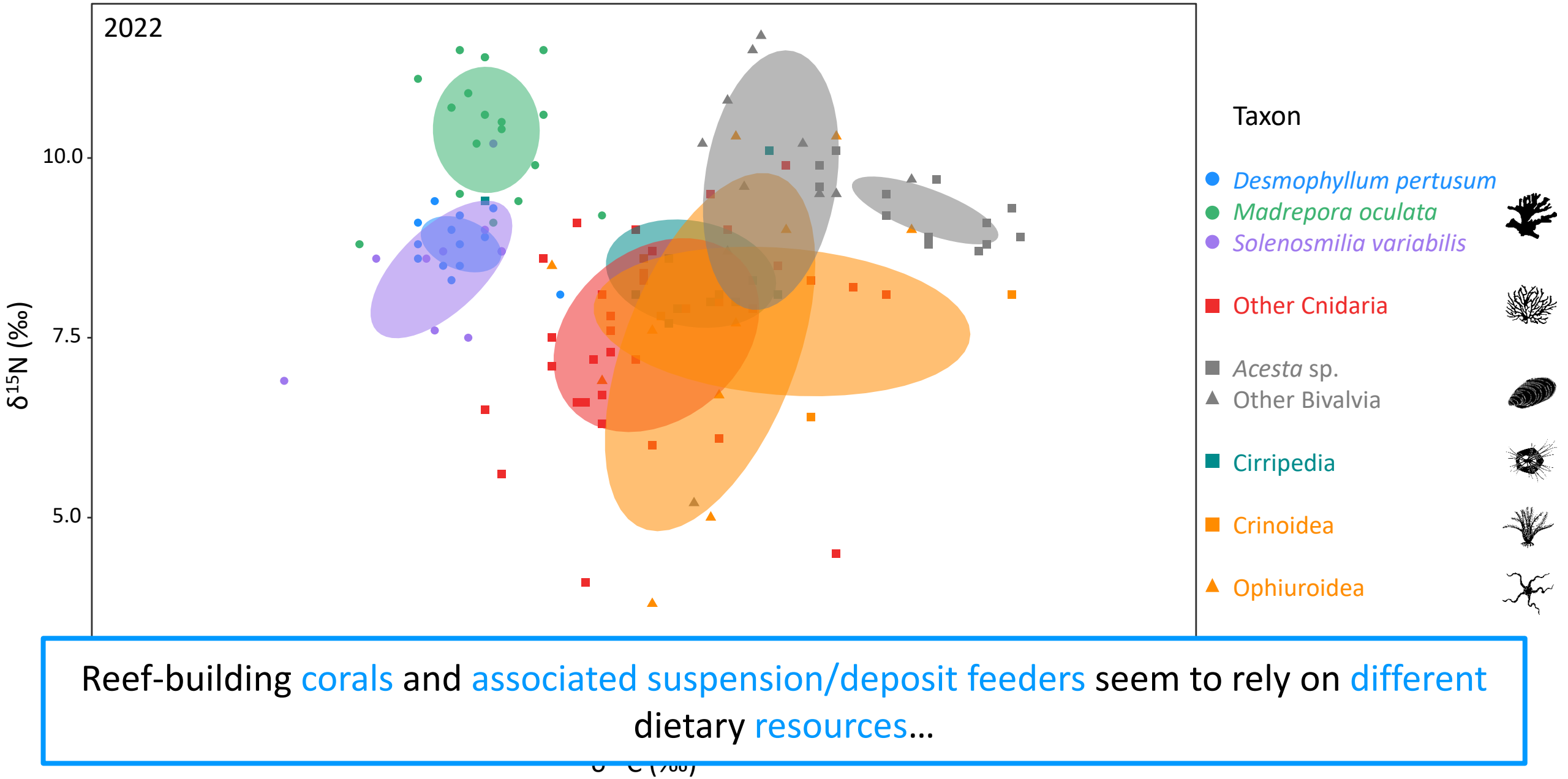
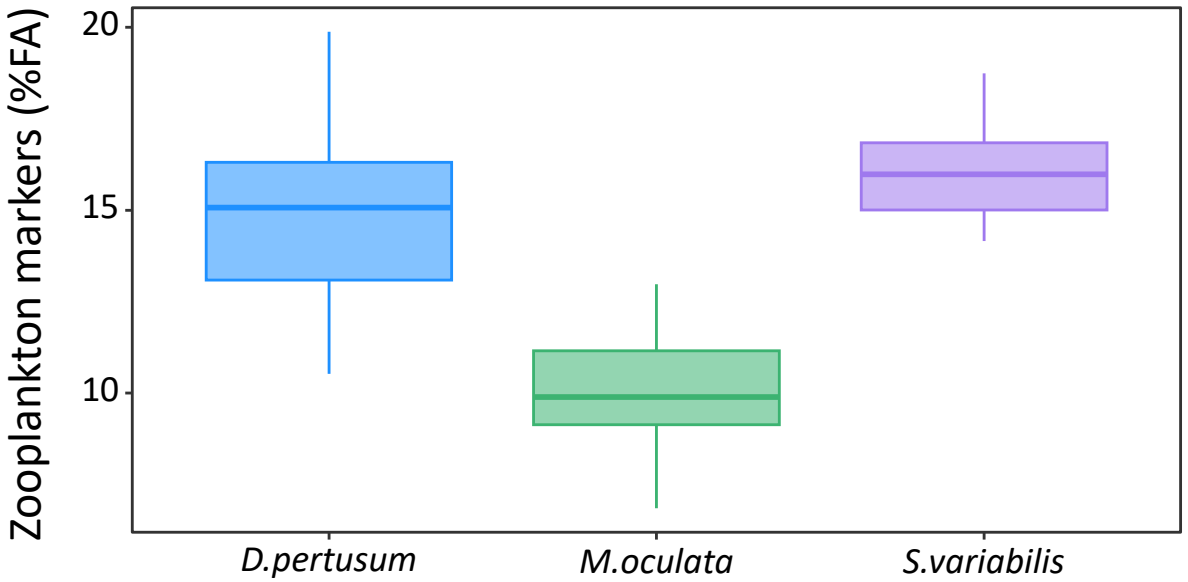


Reef-building corals and associated suspension/deposit feeders seem to rely on different dietary resources…

Fatty acid composition of reserve lipids



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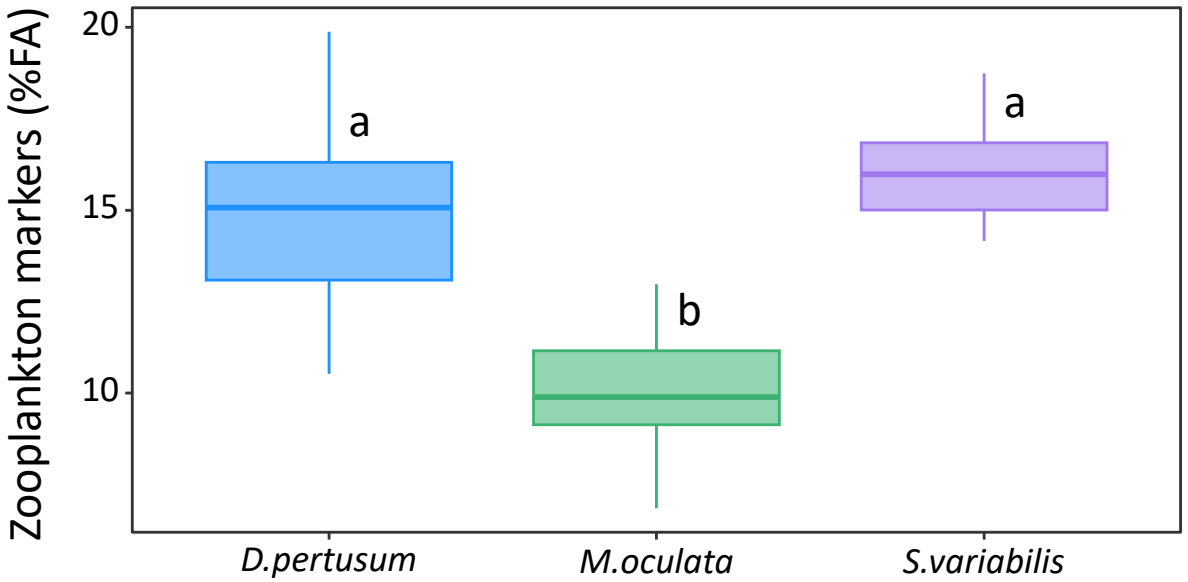

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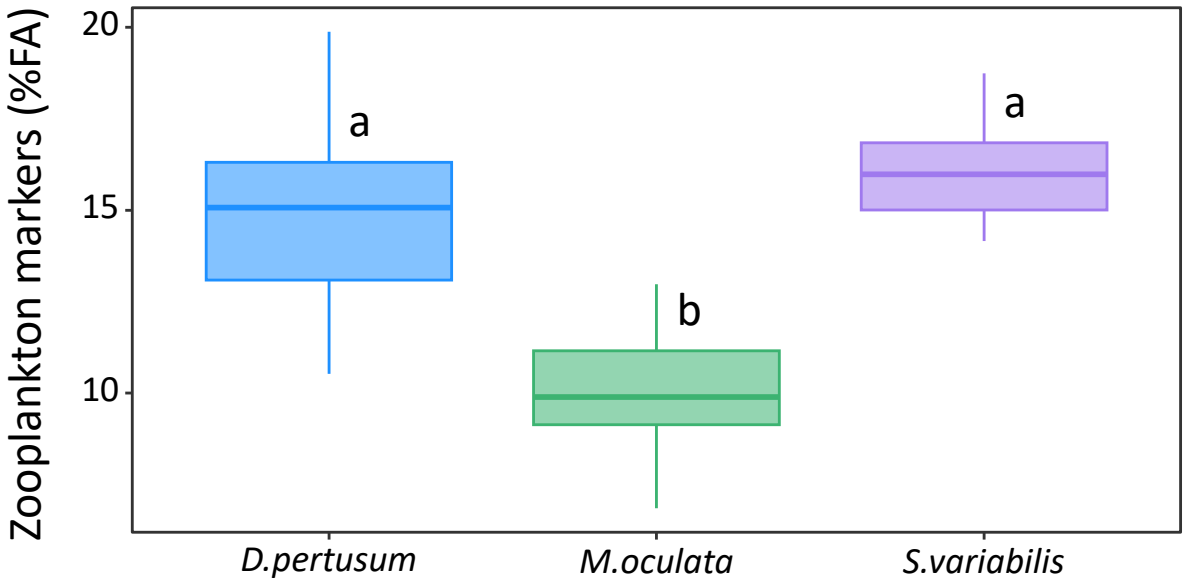

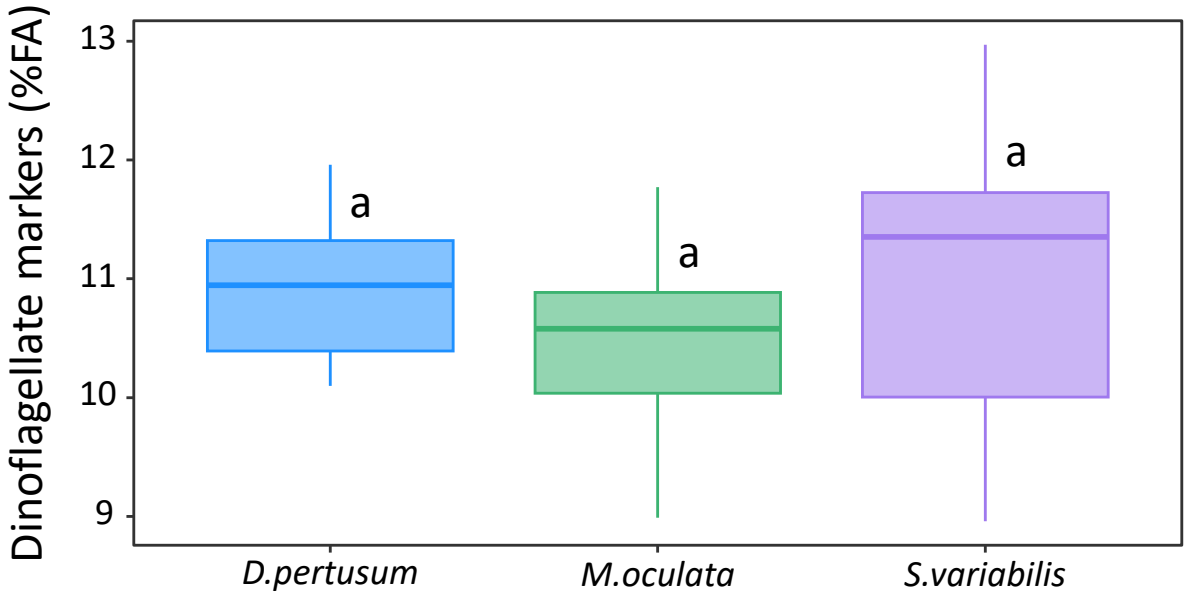

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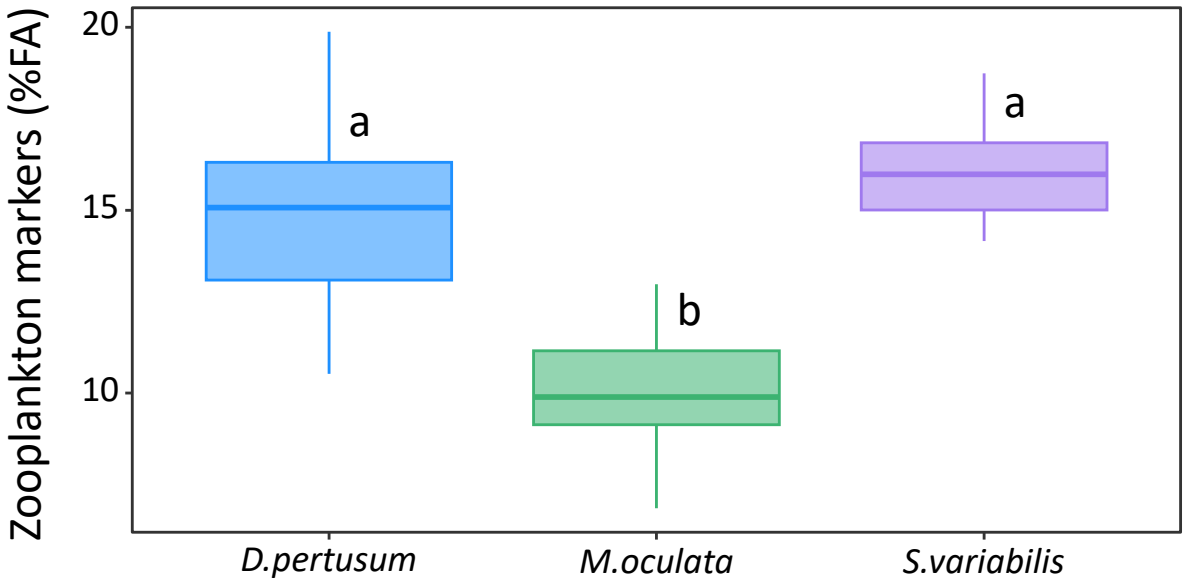

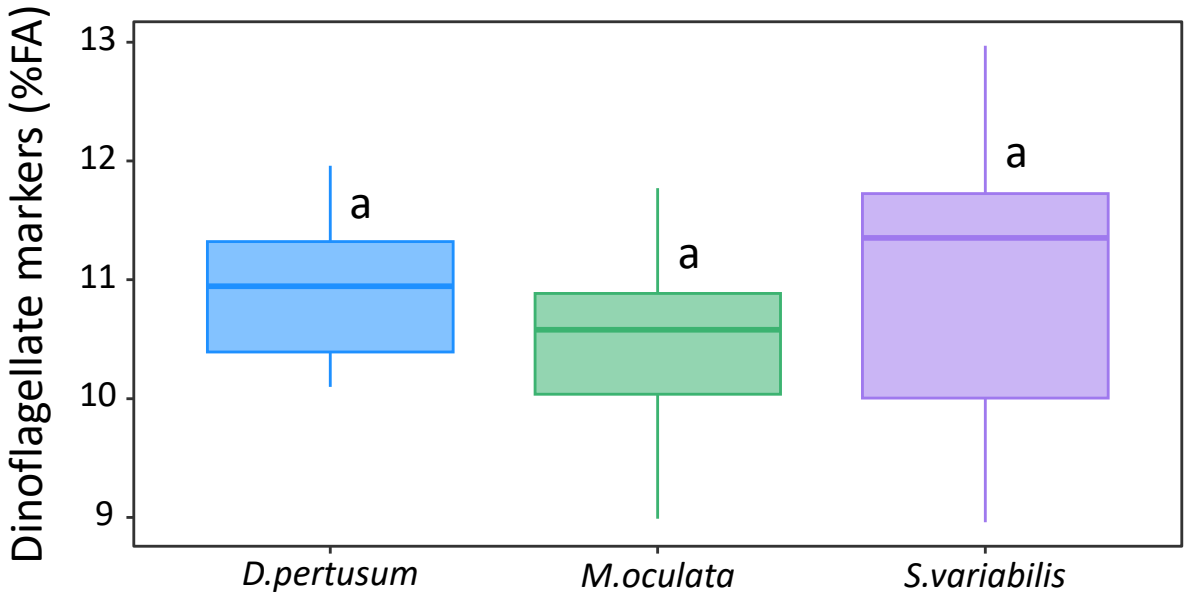

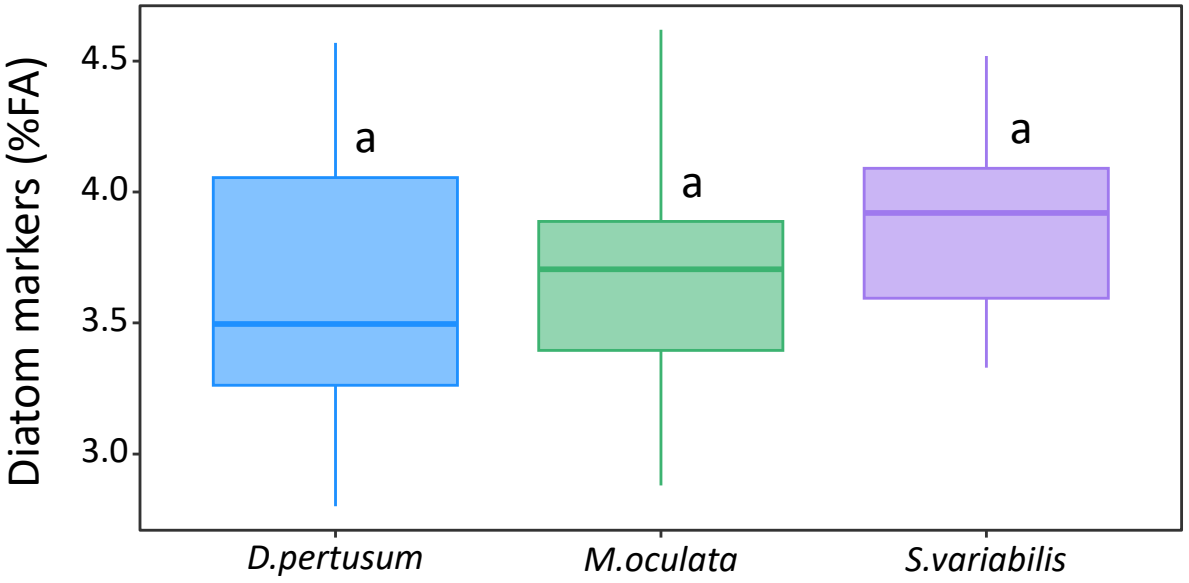

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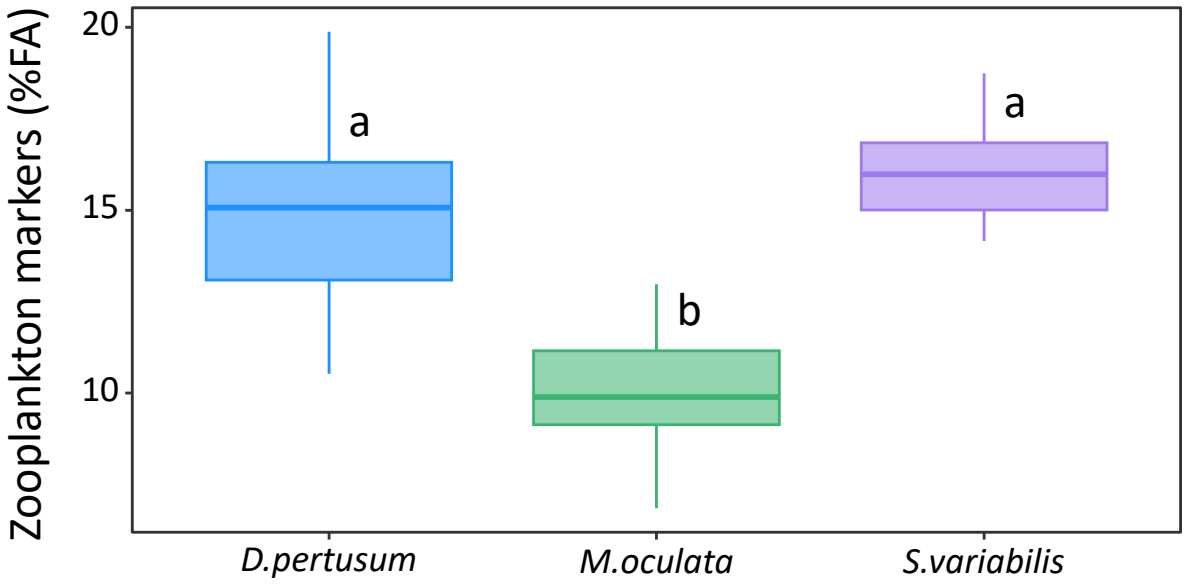

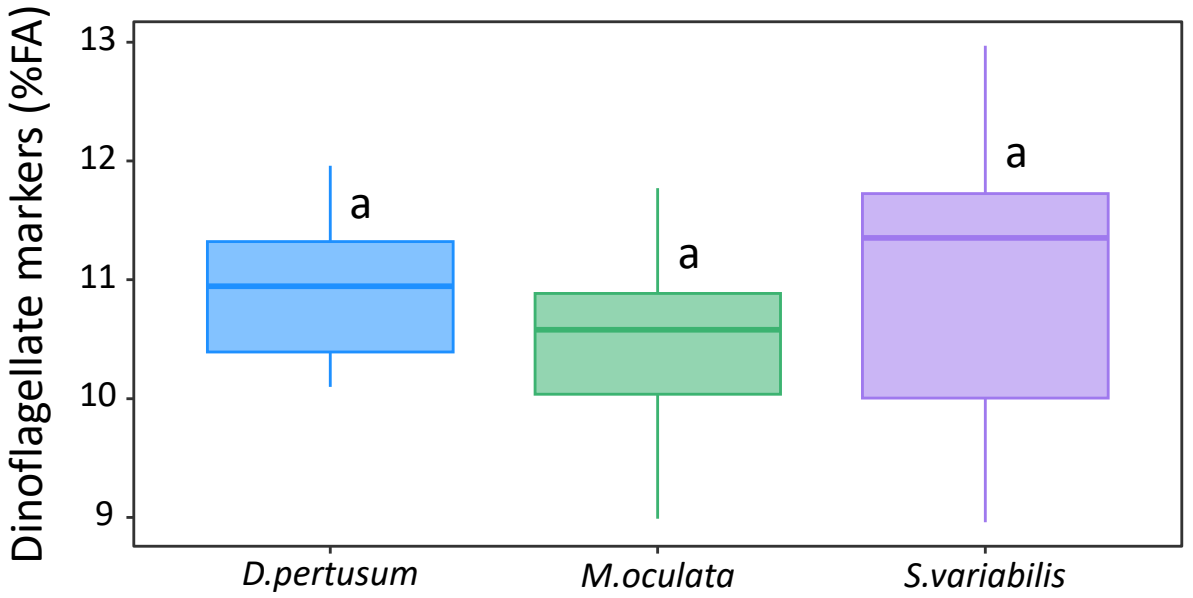

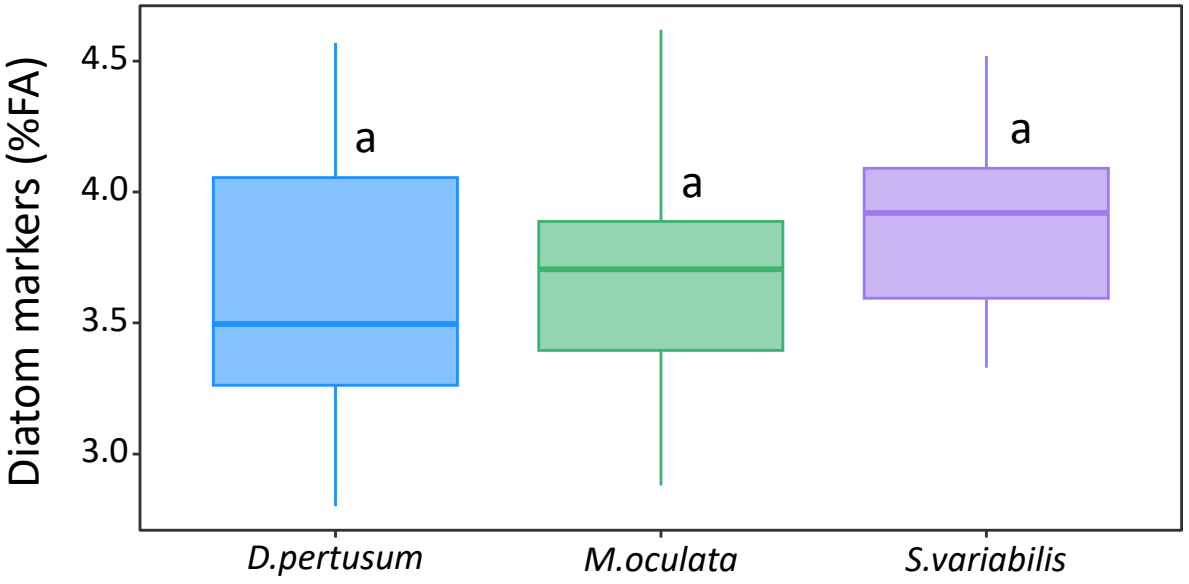

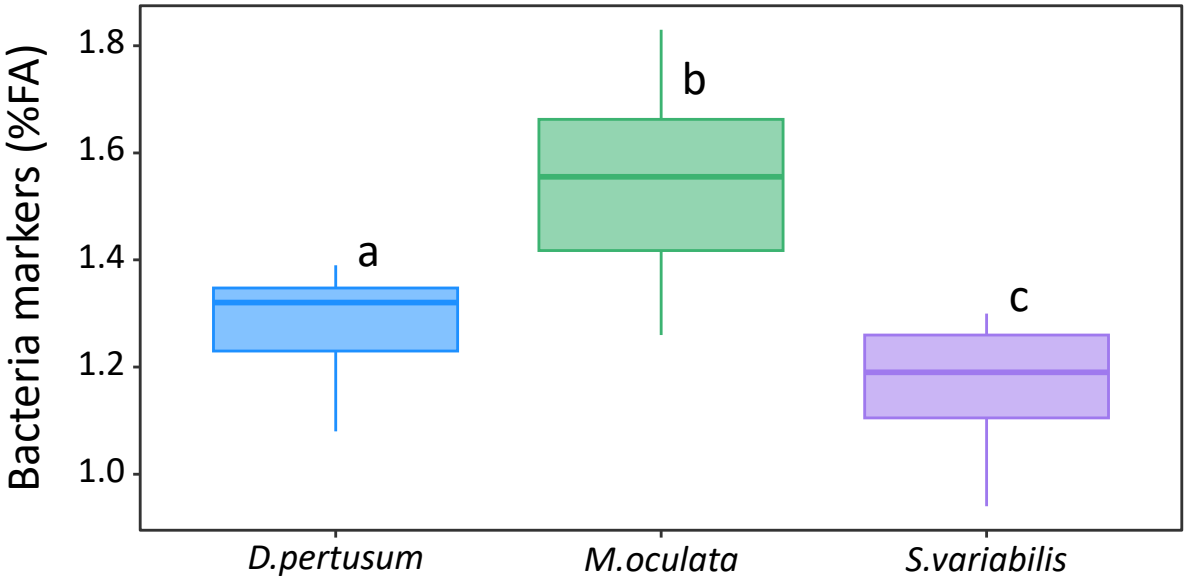

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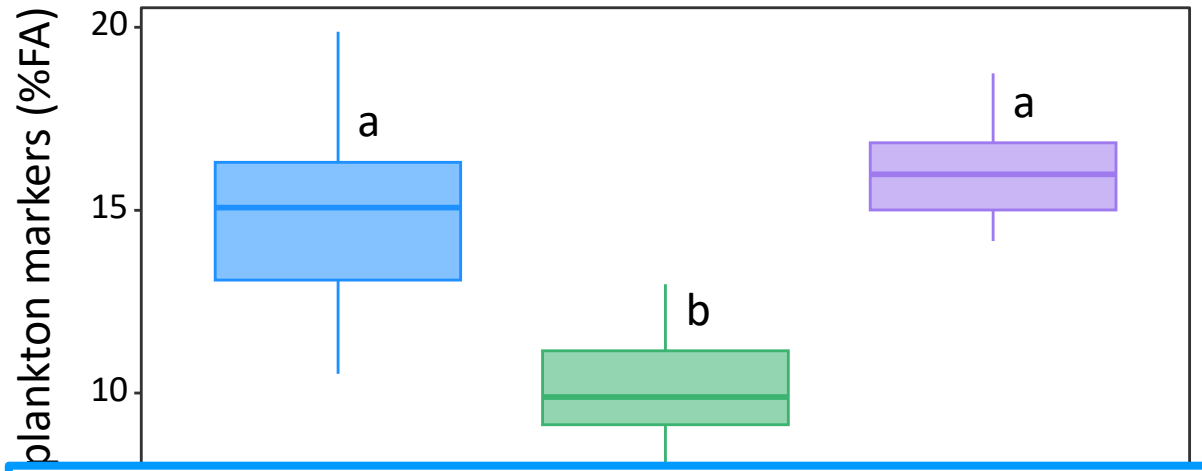
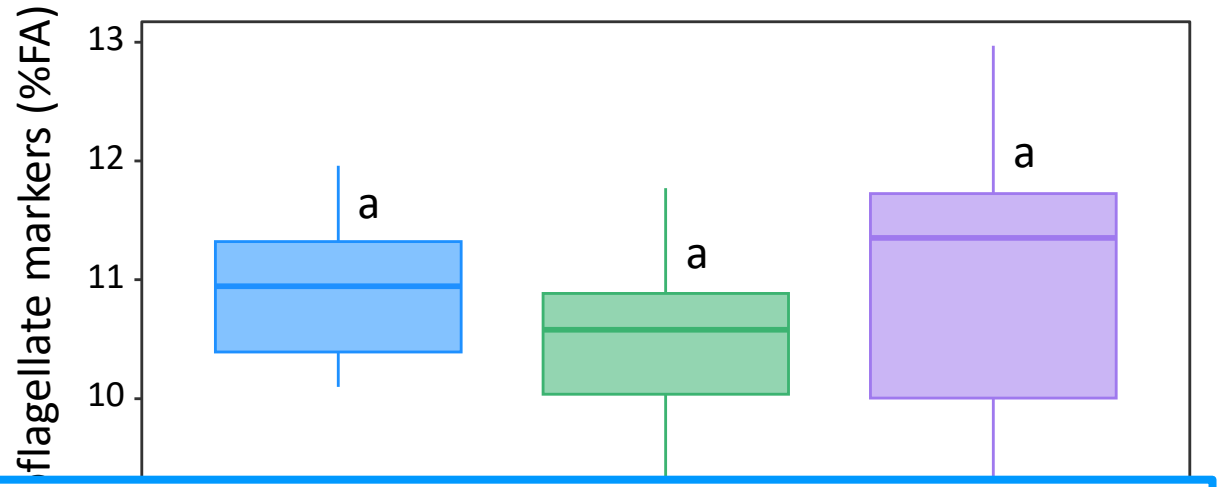
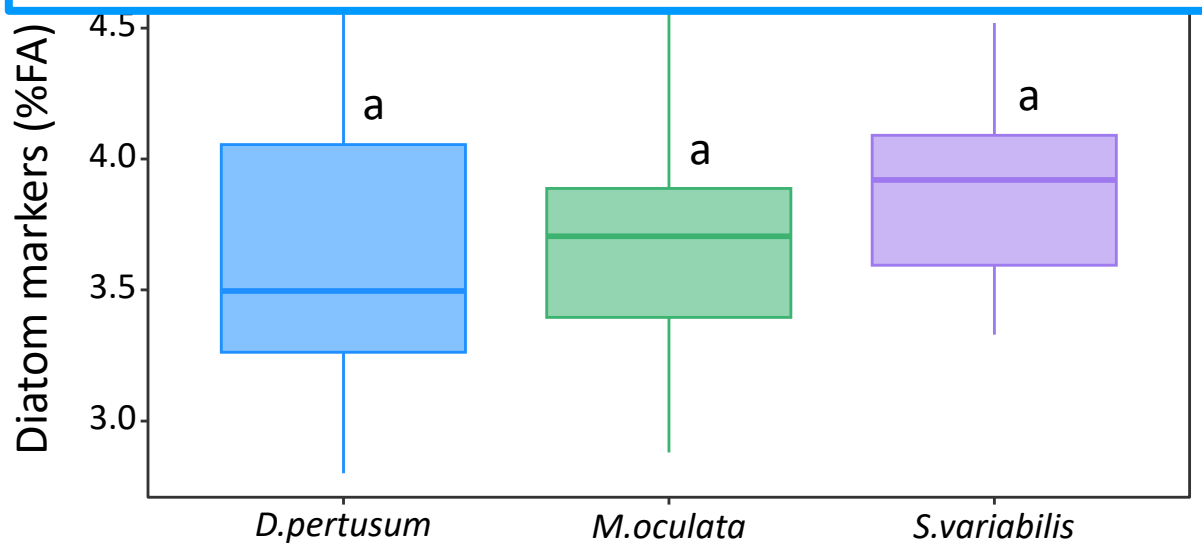
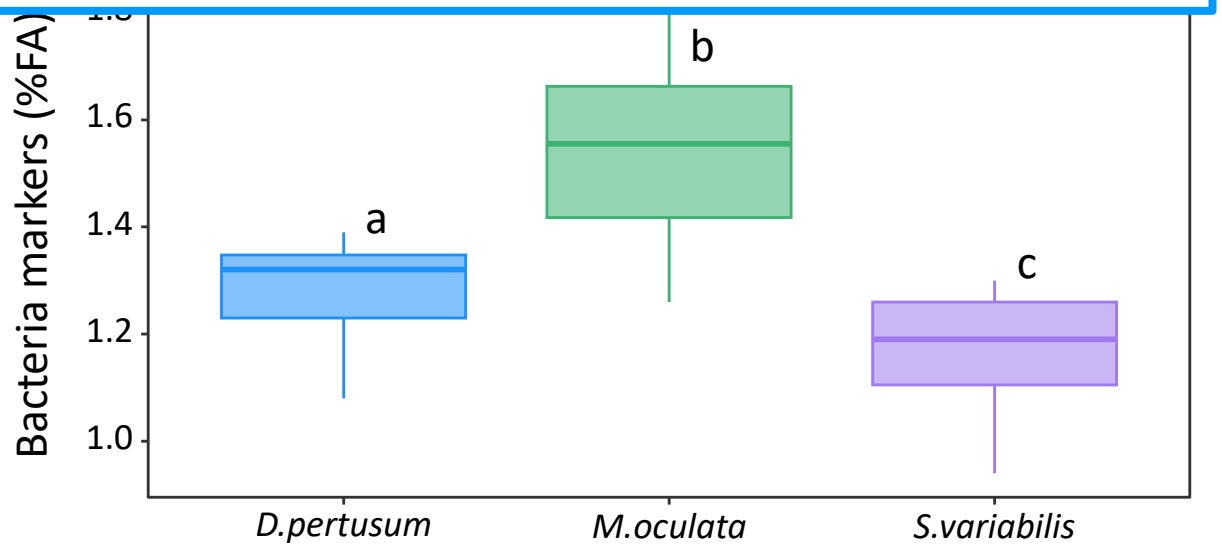


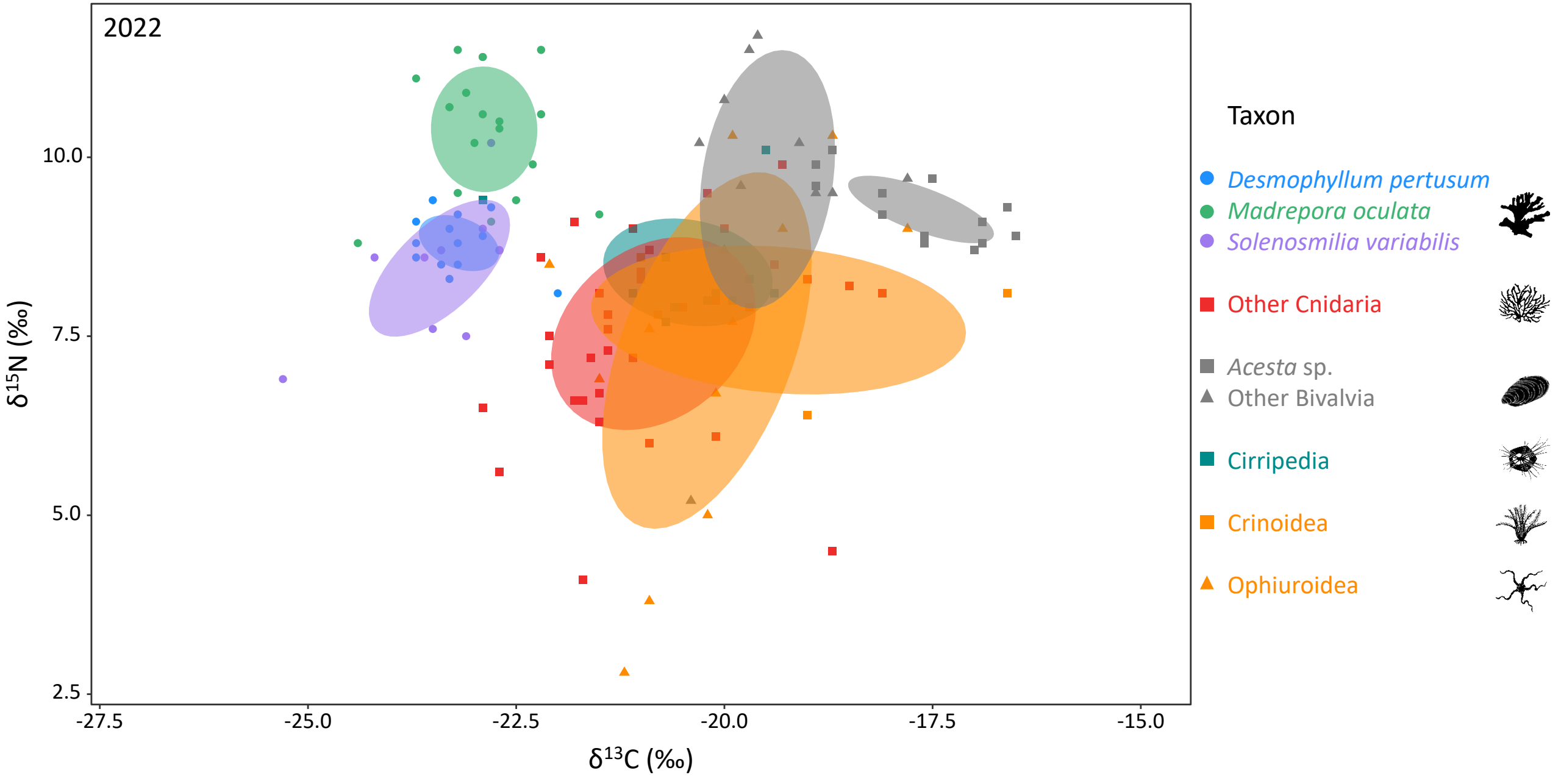


All three taxa seem to be selective plankton feeders, with species-specific differences for some resources likely increasing niche segregation

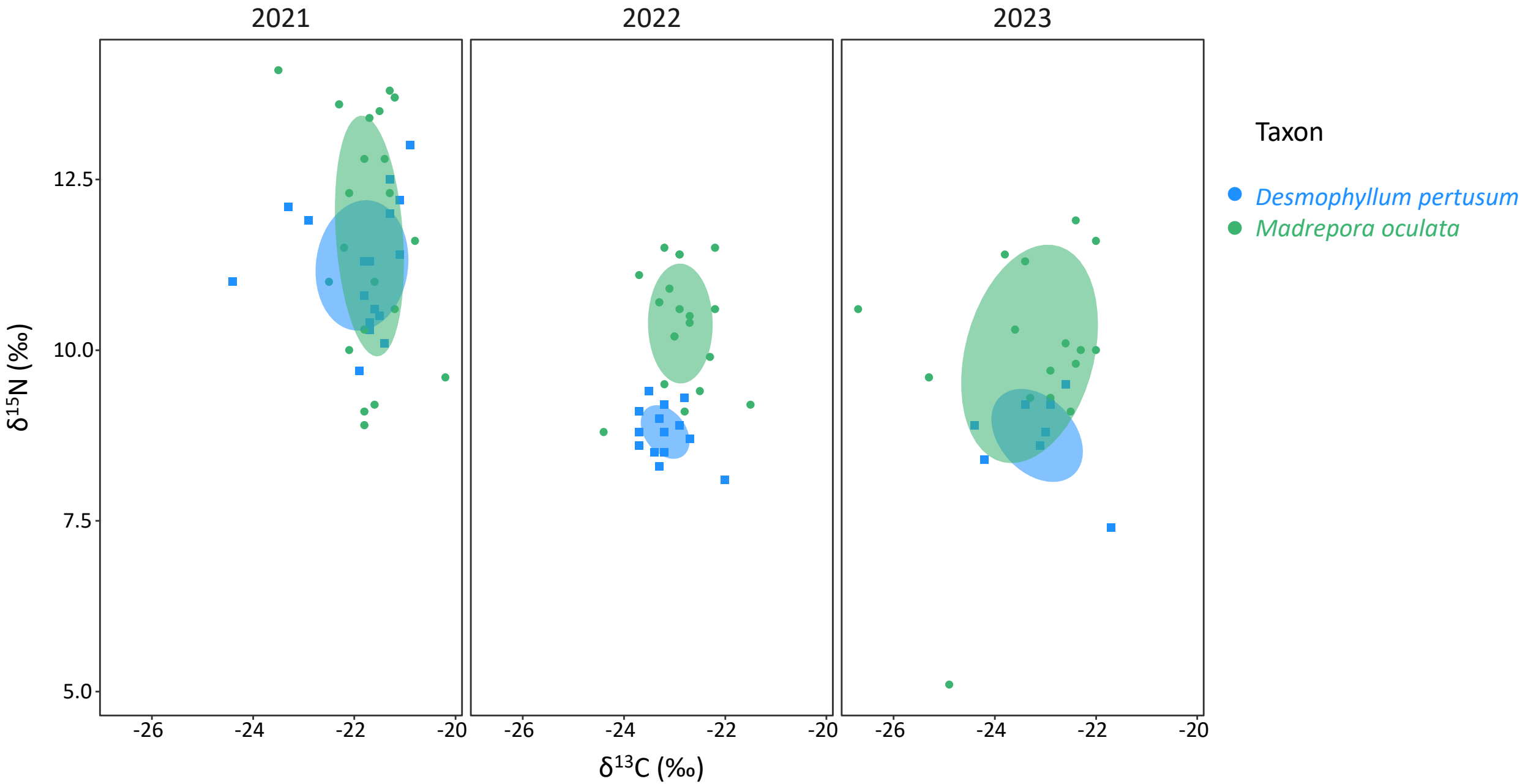




Isotopic niches of corals and associated fauna



Isotopic niches – dominant syntopic corals



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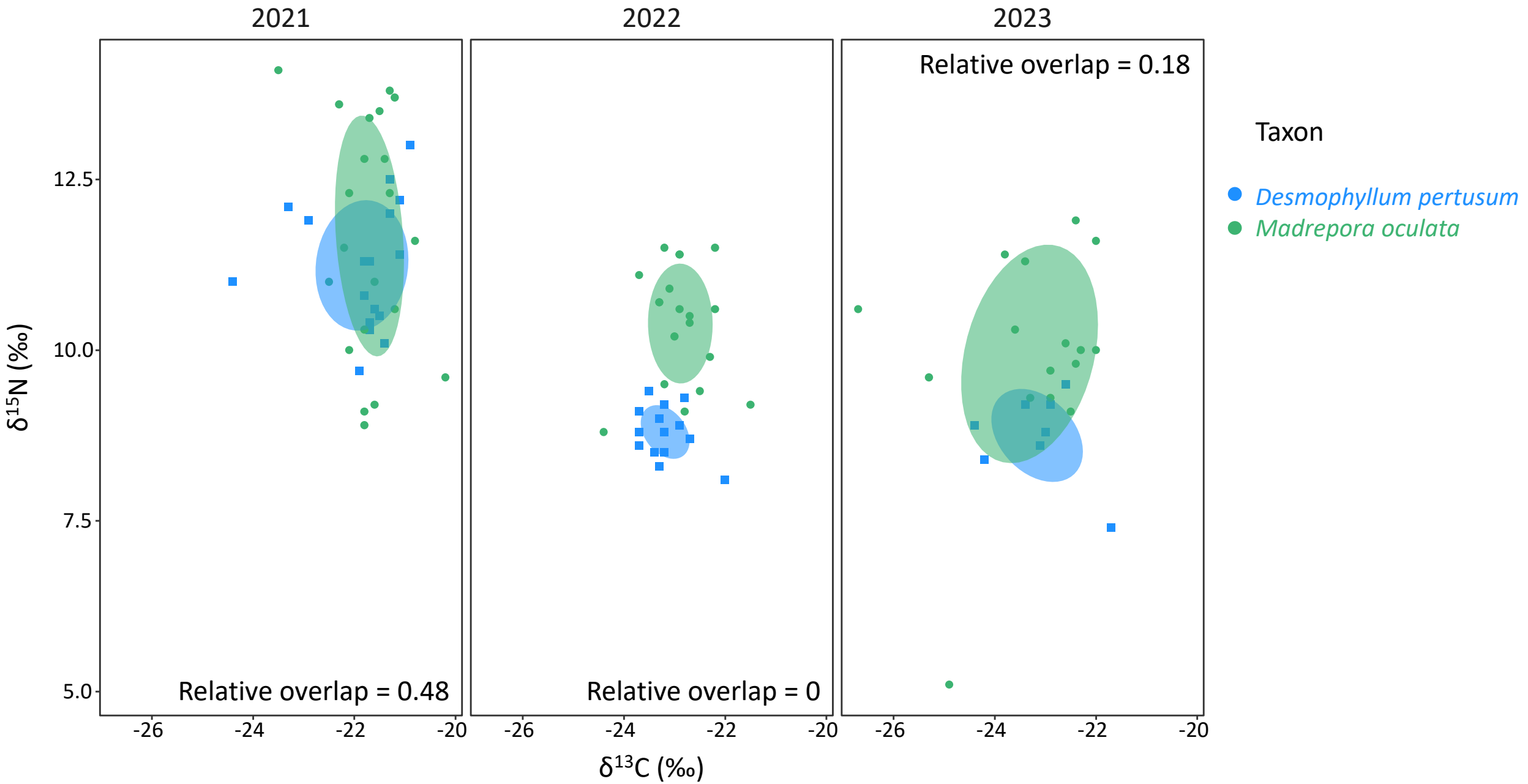

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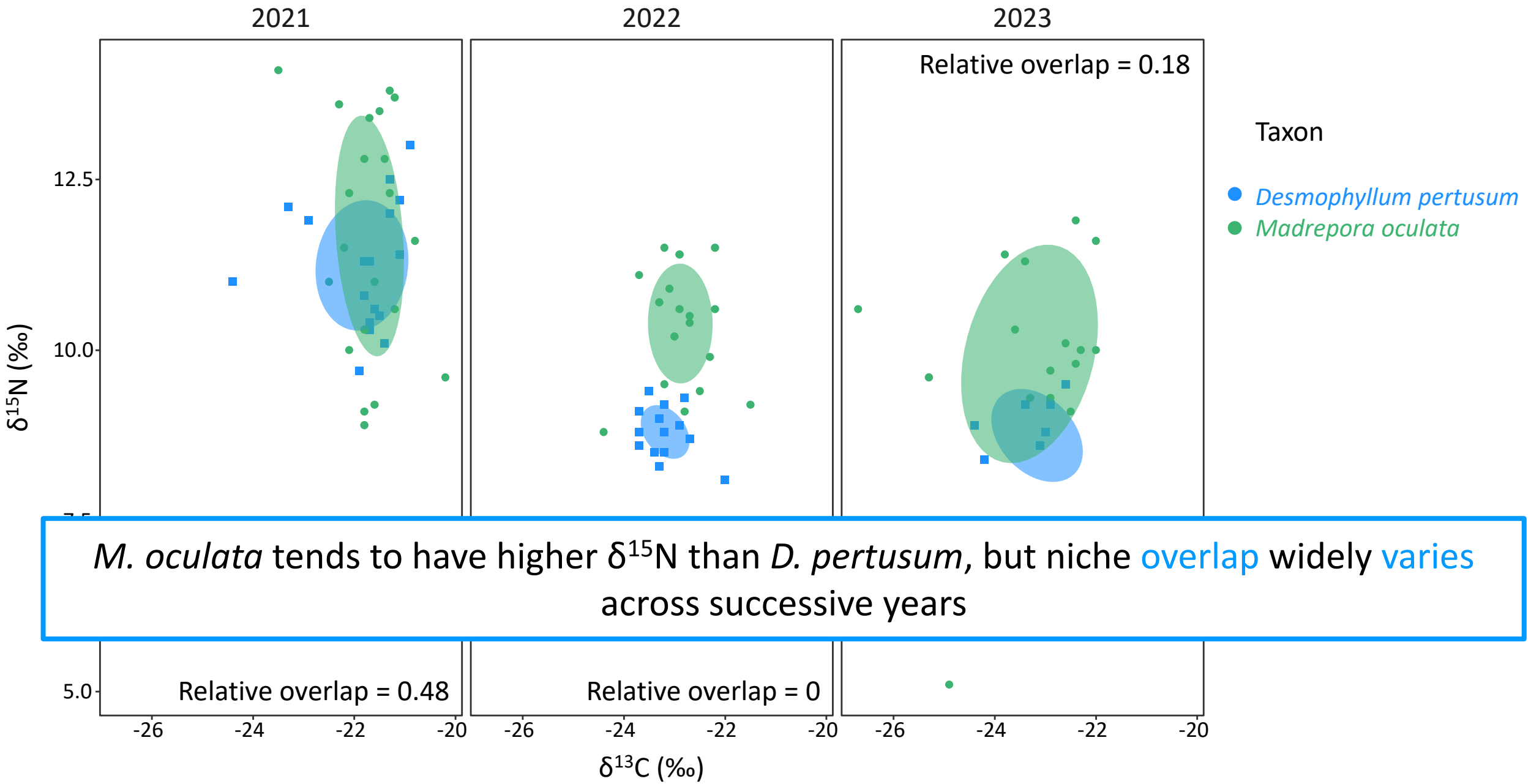

M. oculata tends to have higher $\delta^{15}N$ than *D. pertusum*, but niche overlap widely varies across successive years

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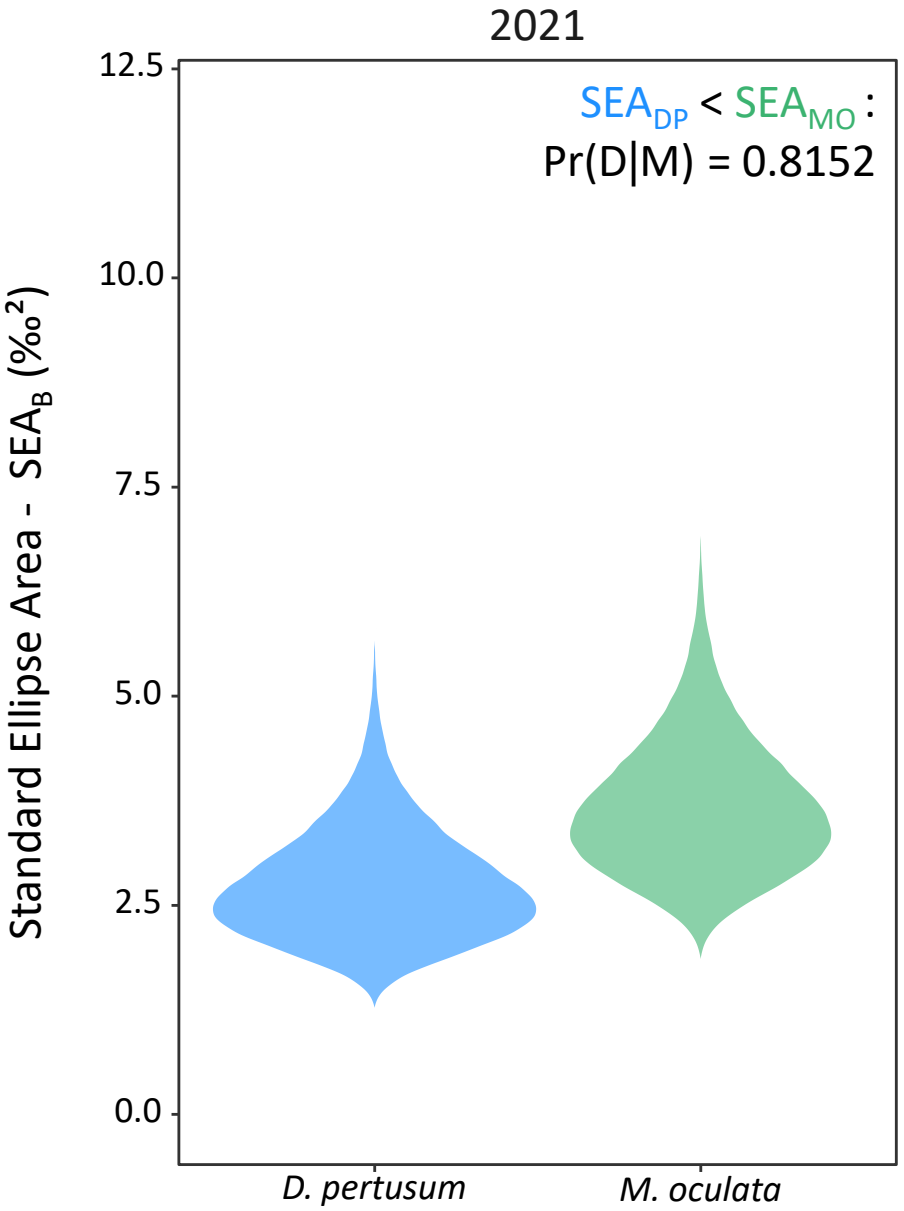

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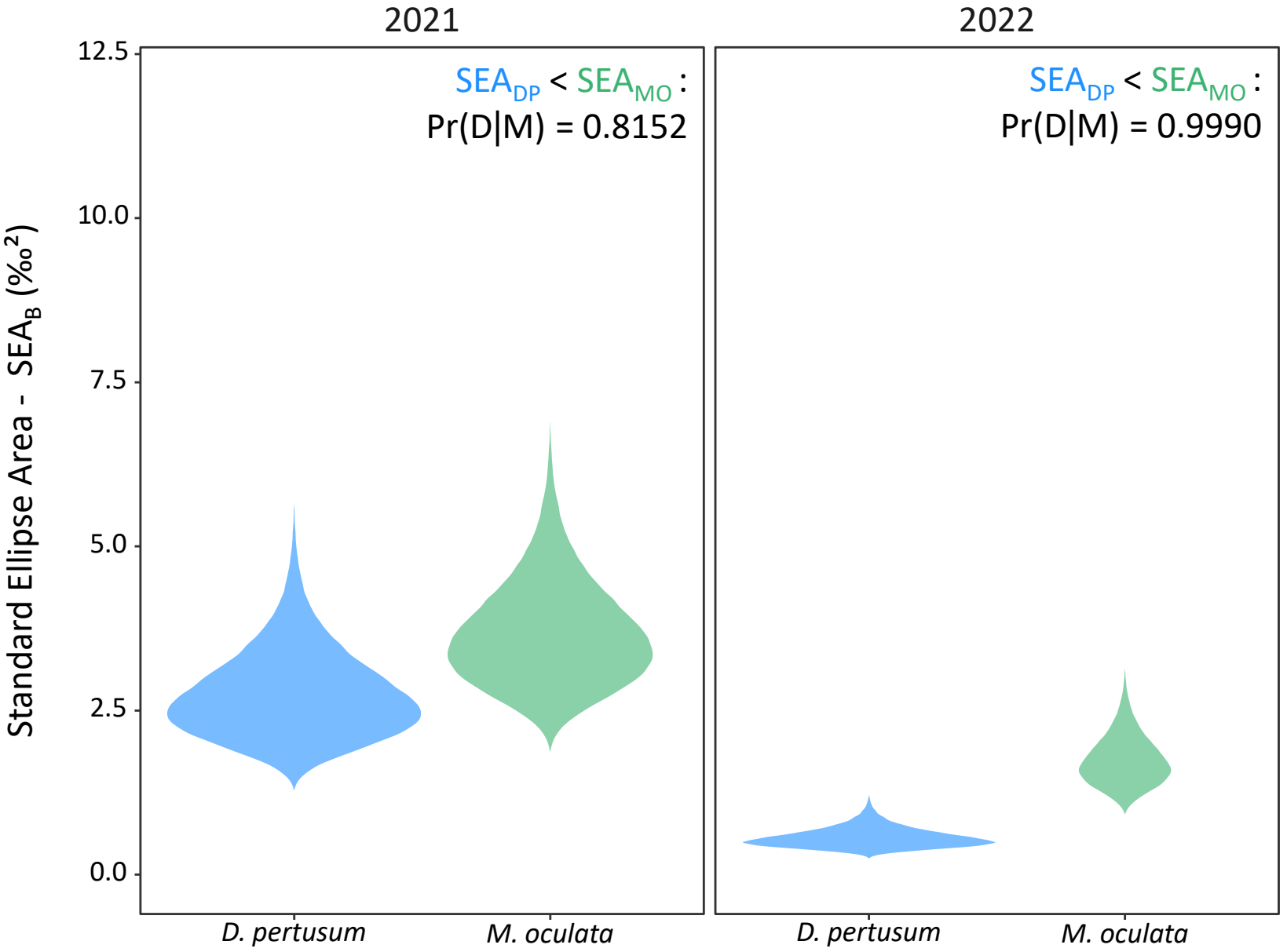

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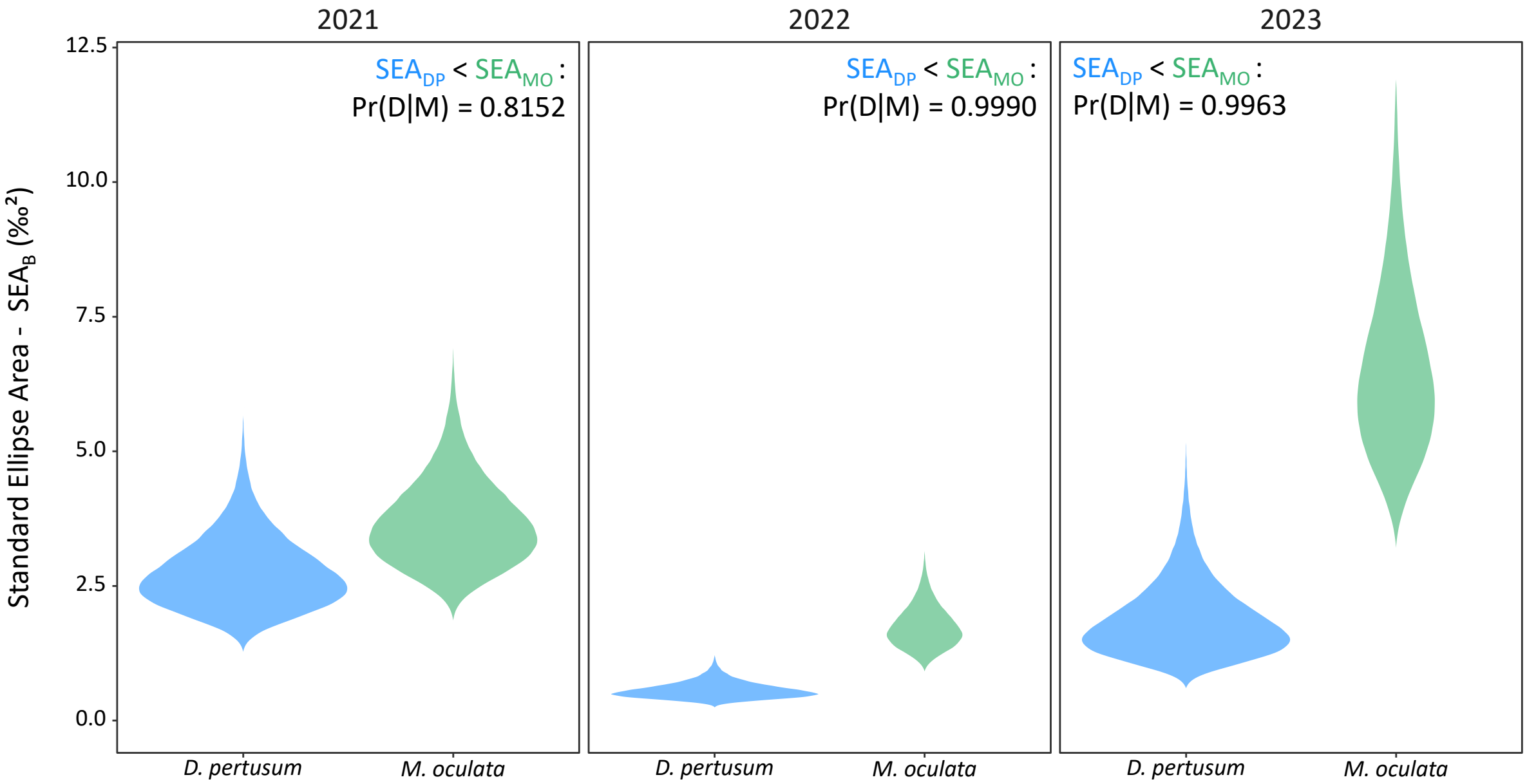
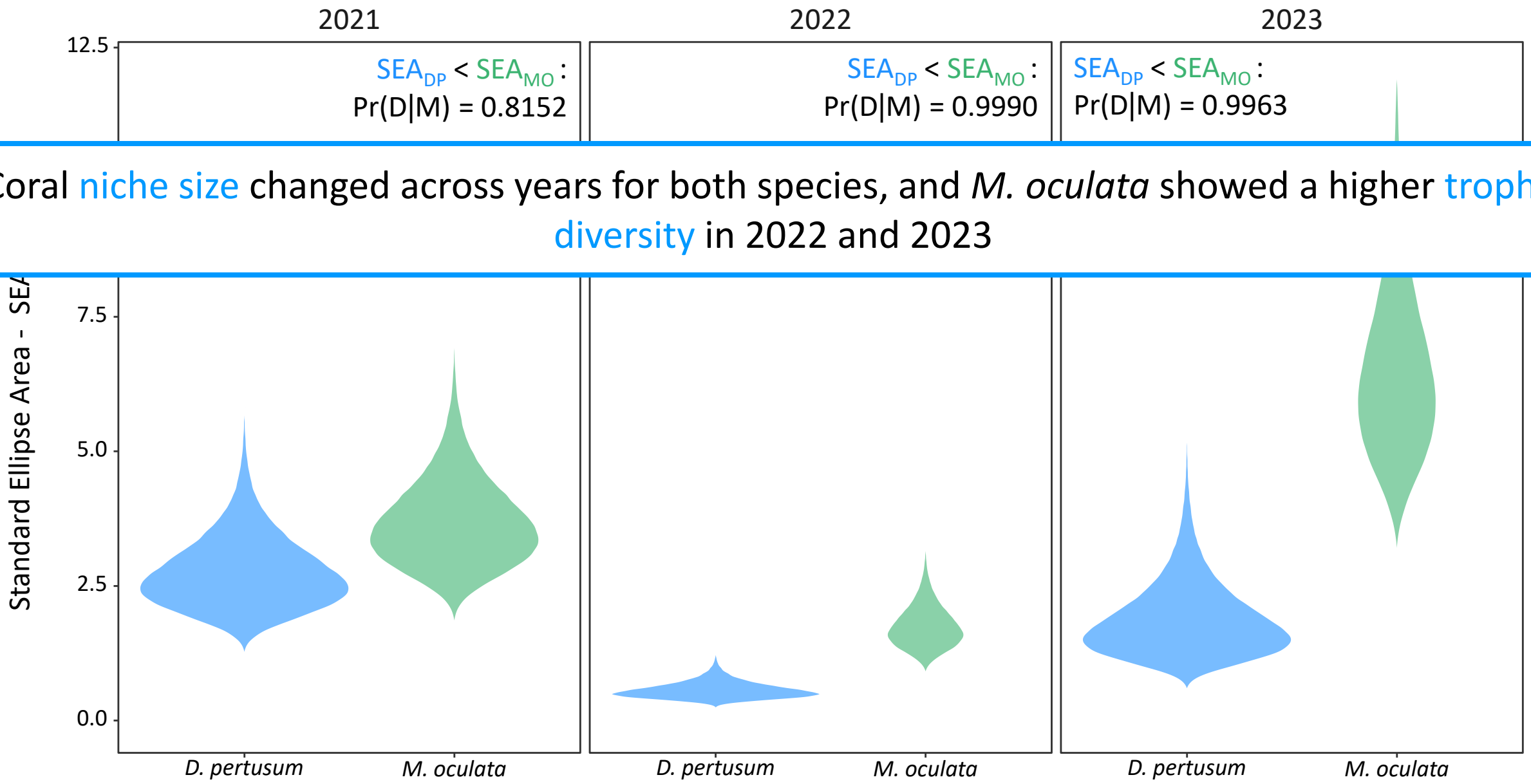
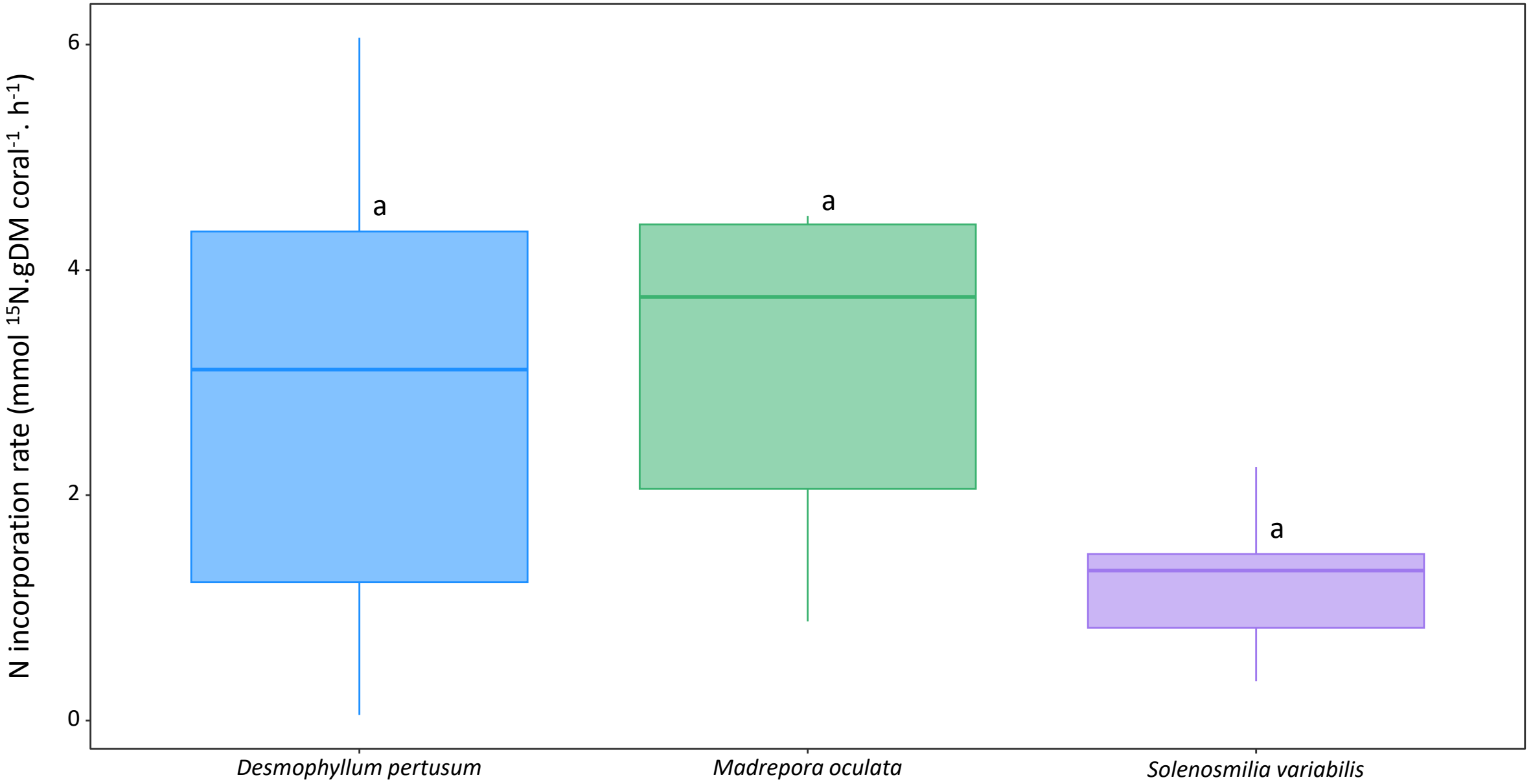


Isotopic niches – dominant syntopic corals



Coral niche size changed across years for both species, and *M. oculata* showed a higher trophic diversity in 2022 and 2023

Labelling experiments



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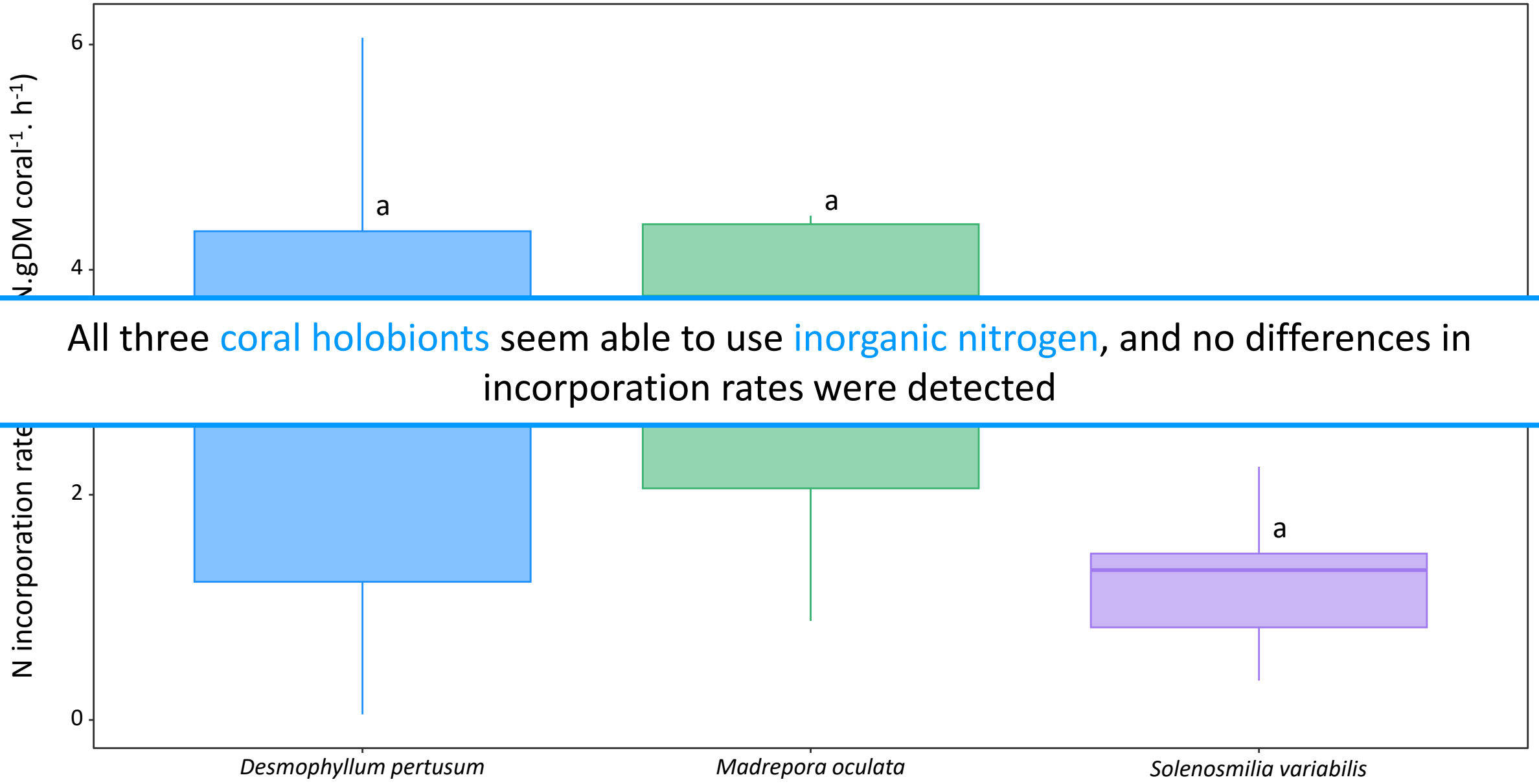

All three coral holobionts seem able to use inorganic nitrogen, and no differences in incorporation rates were detected

Inorganic nitrogen fixation: how?

Inorganic nitrogen fixation has previously been reported in *D. pertusum*, presumably through mutualistic bacteria. However, this symbiosis is currently poorly documented.

Use of NanoSIMS to elucidate incorporation patterns

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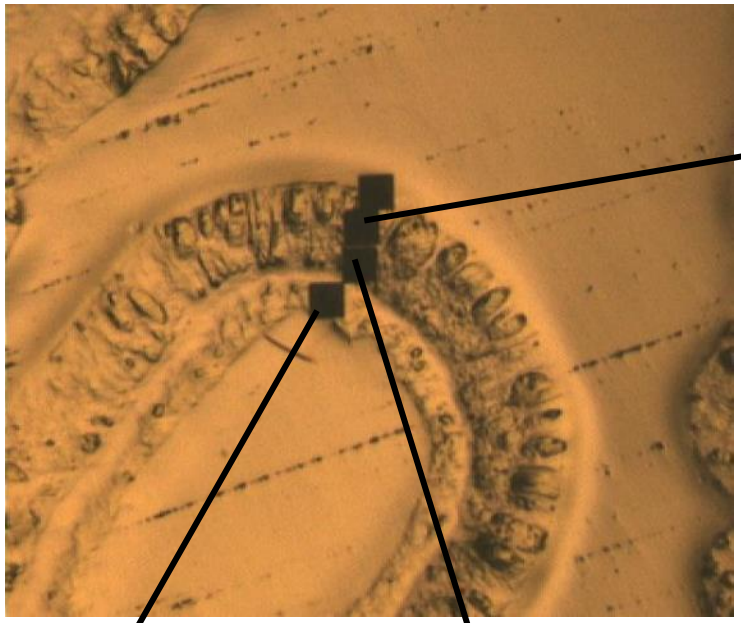
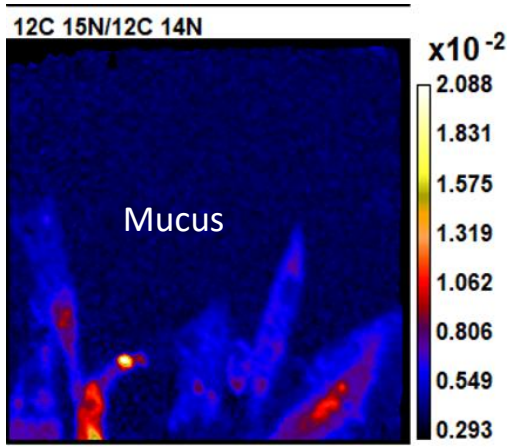
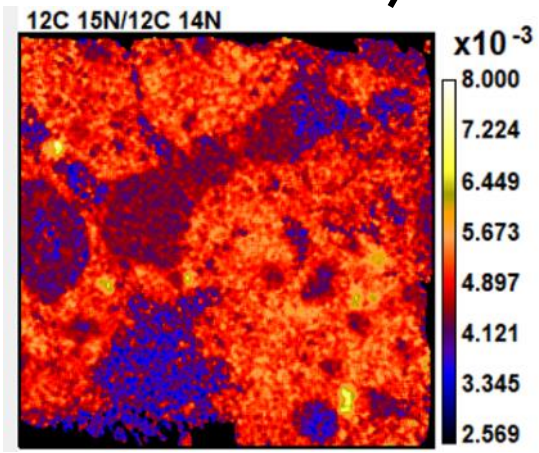
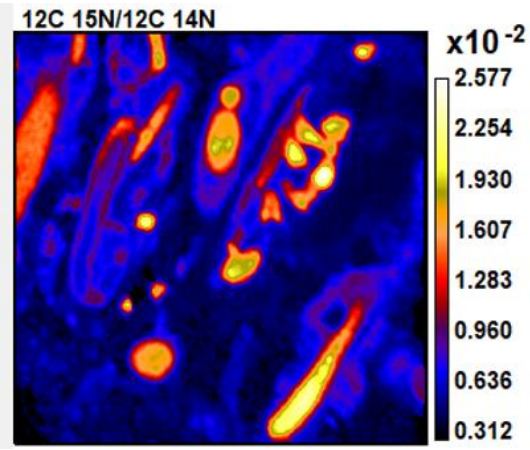




$\delta^{15}N$ (‰) = 3084.67 ± 30.35



$\delta^{15}N$ (‰) = 685.13 ± 12.10



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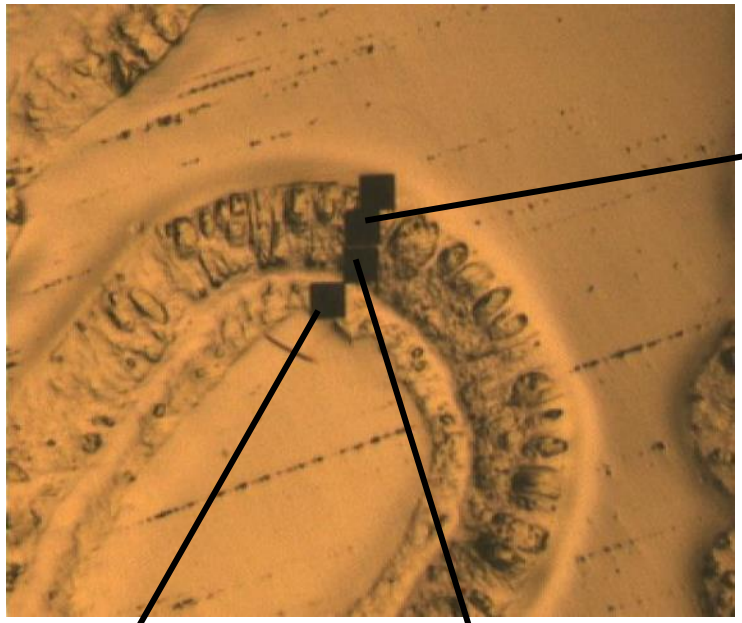
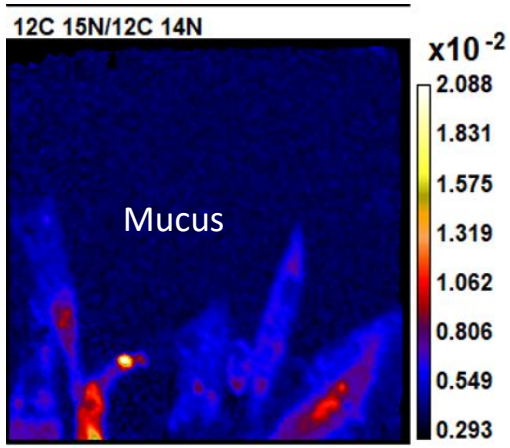
Stronger labelling in inner polyp structures (coelenteron and/or tentacle bases?)





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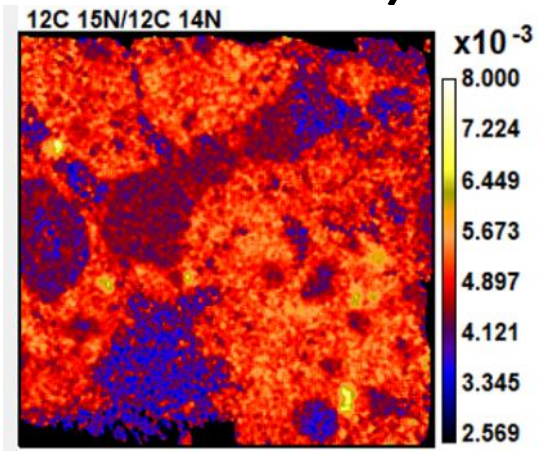

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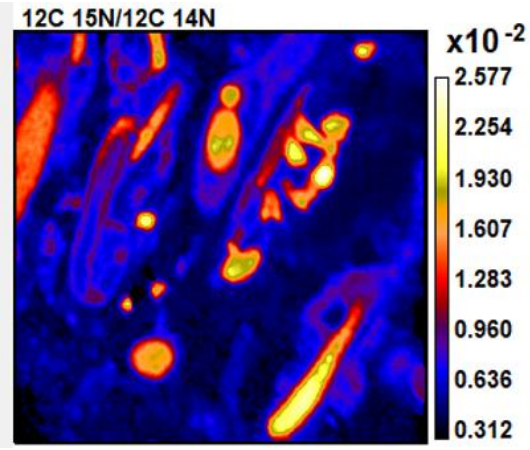

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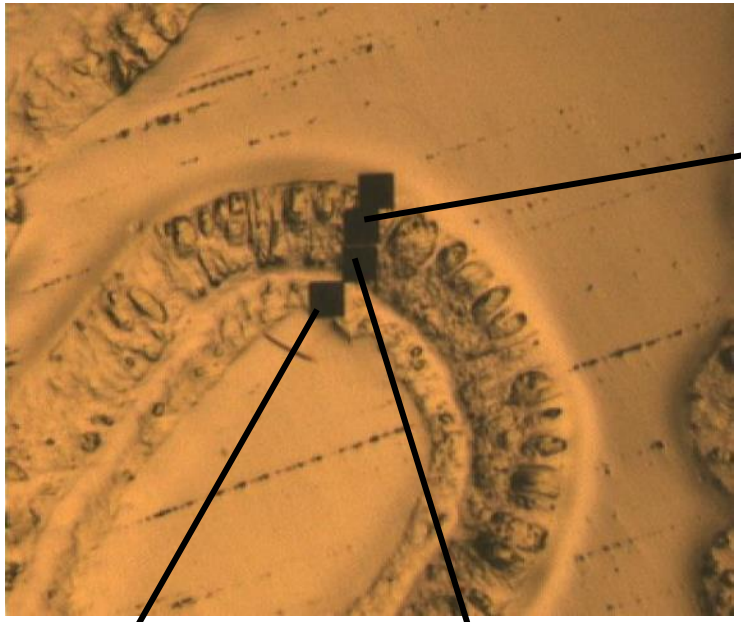

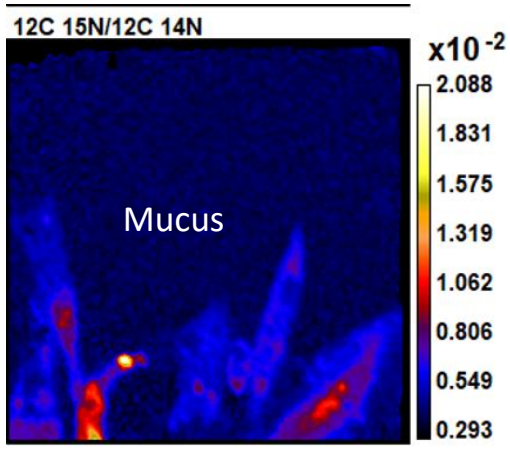

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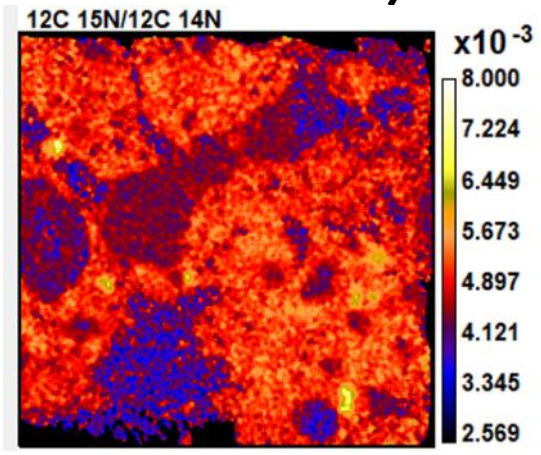

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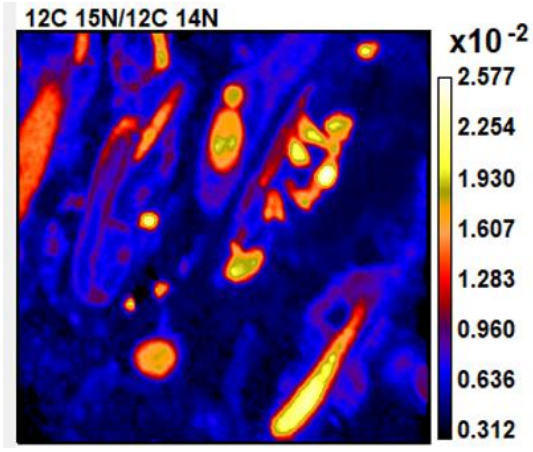

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In Norwegian fjords, *D. pertusum* might derive 10 – 30 % of its nitrogen from symbiote metabolism.

Spatial and temporal variability? Interspecific differences? Impacts of environmental changes (ocean warming and acidification)?

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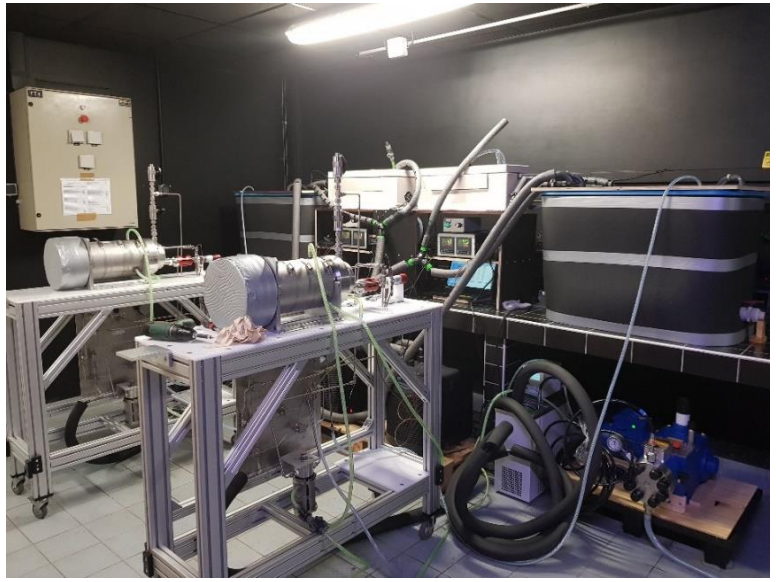

OcéanOpolis BREST

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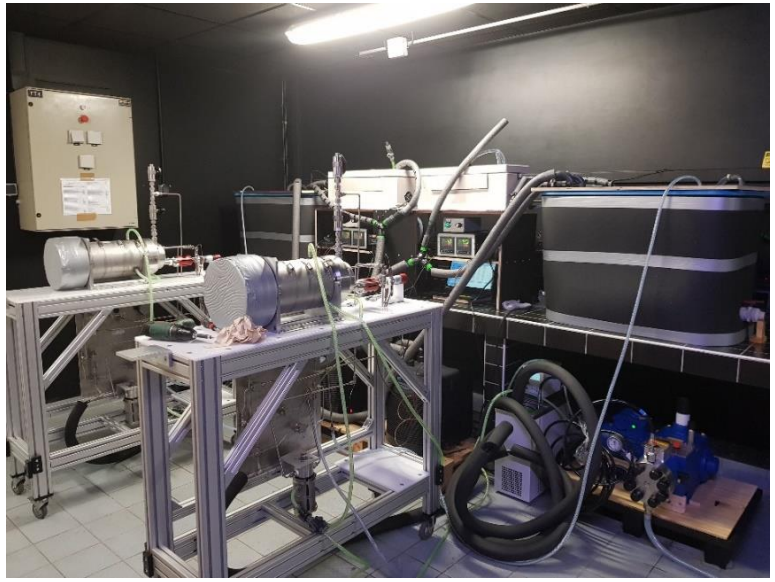
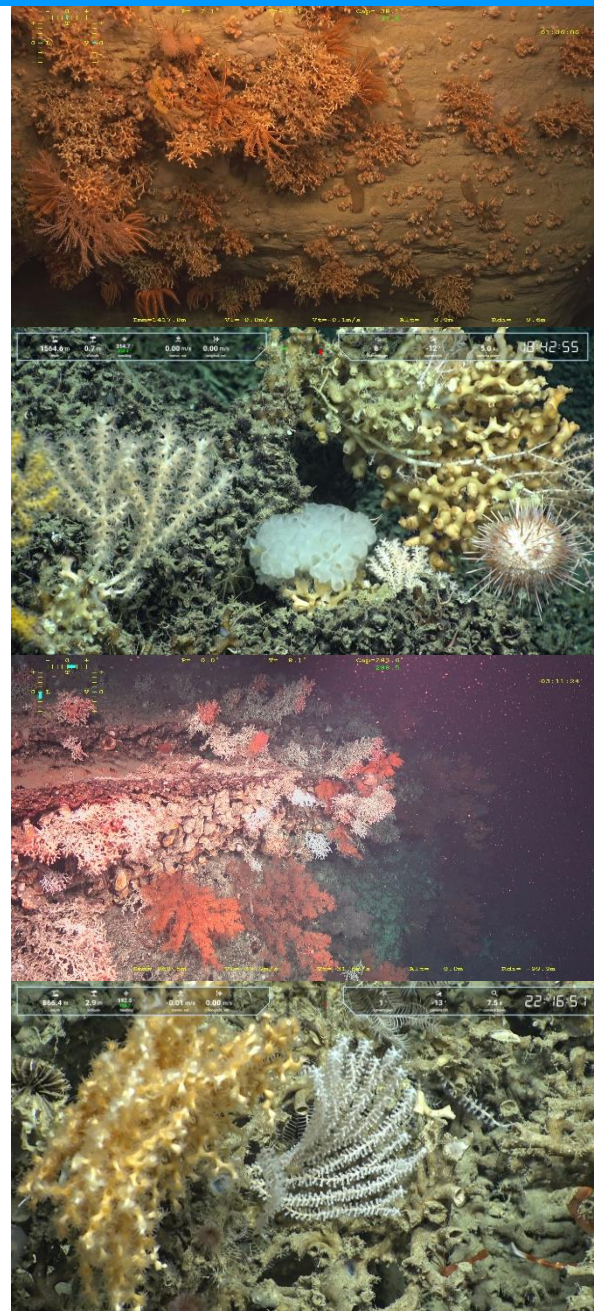


OcéanOpolis BREST

Take home message

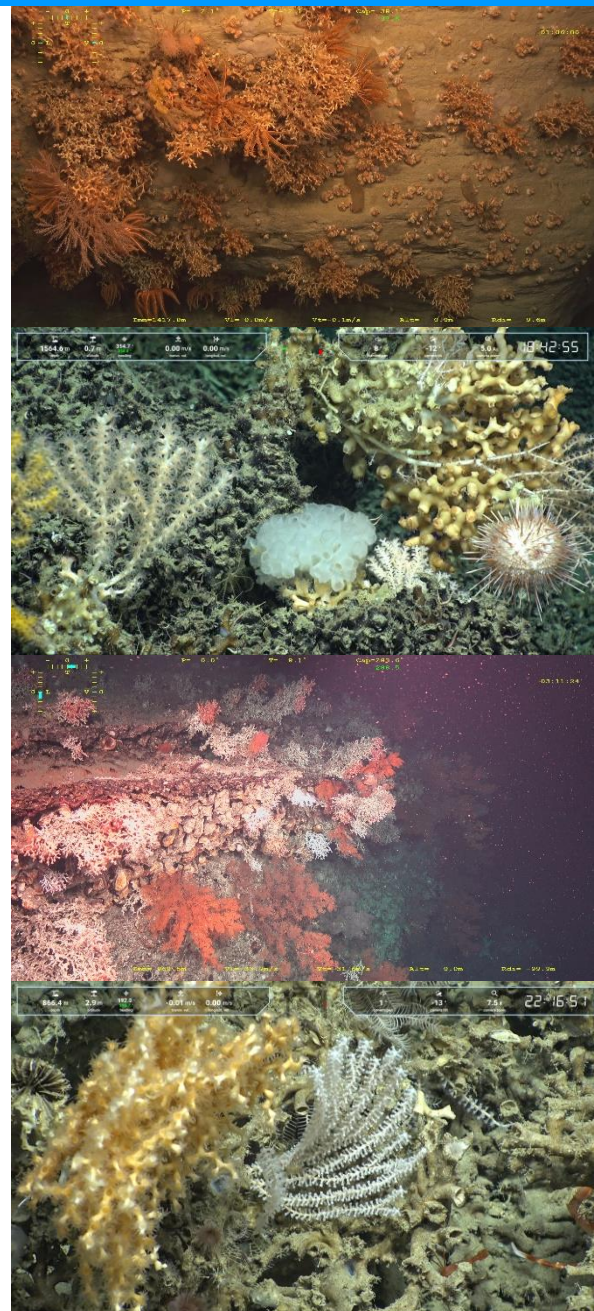
- In the Lampaul canyon, all 3 reef-building coral species exhibit marked resource segregation with associated fauna.



Take home message

- In the Lampaul canyon, all 3 reef-building coral species exhibit marked resource segregation with associated fauna.

- Corals are selective plankton feeders with species-specific preferences: *Desmophyllum pertusum* and *Solenosmilia variabilis* seem to favour zooplankton, while *Madrepora oculata* could have a more balanced diet comprising zooplankton, phytoplankton and bacterioplankton.



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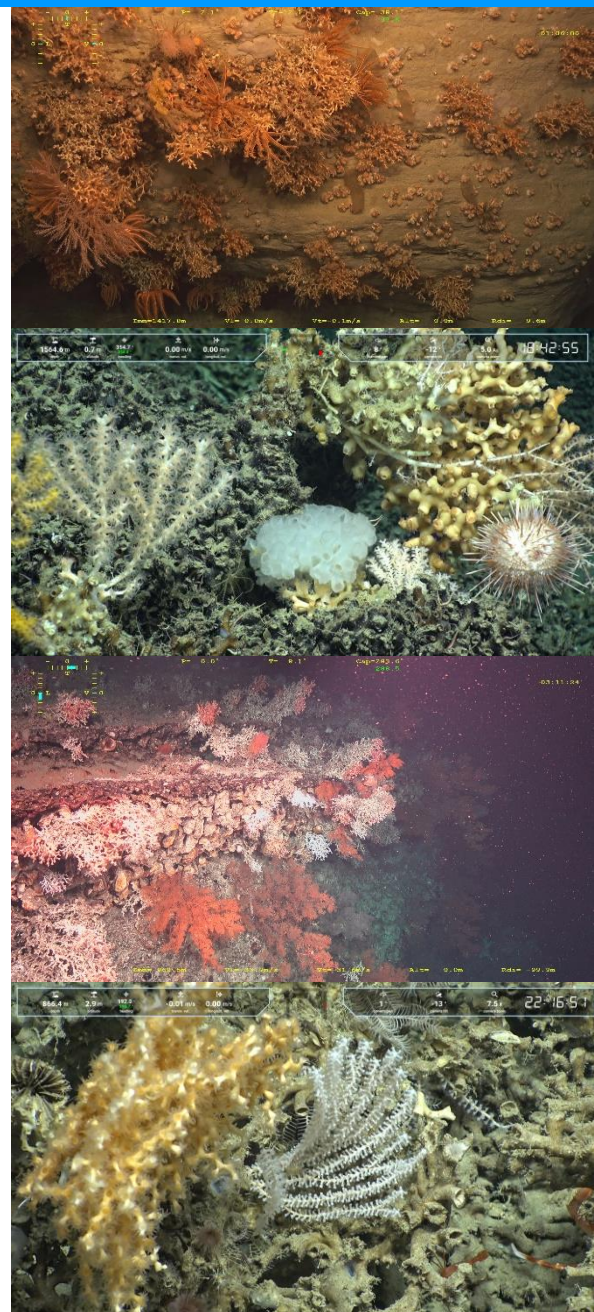

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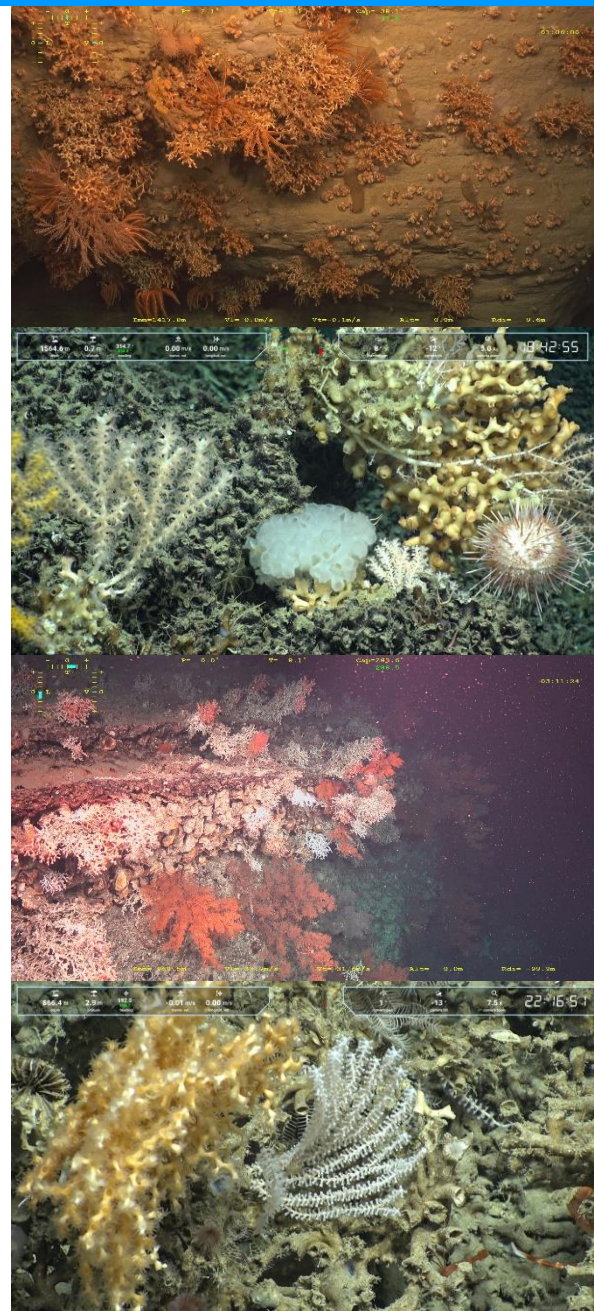
- The 3 species are able to supplement their dietary nutriment intake through inorganic nitrogen fixation. The relative importance of this mechanism for coral nitrogen budget is currently unclear.
- Joint use of stable isotopes and fatty acids suggest that although foraging strategies might differ, coral niches might partly overlap. Marked differences between successive sampling years hint towards highly dynamic trophic interactions.



Acknowledgements & funding

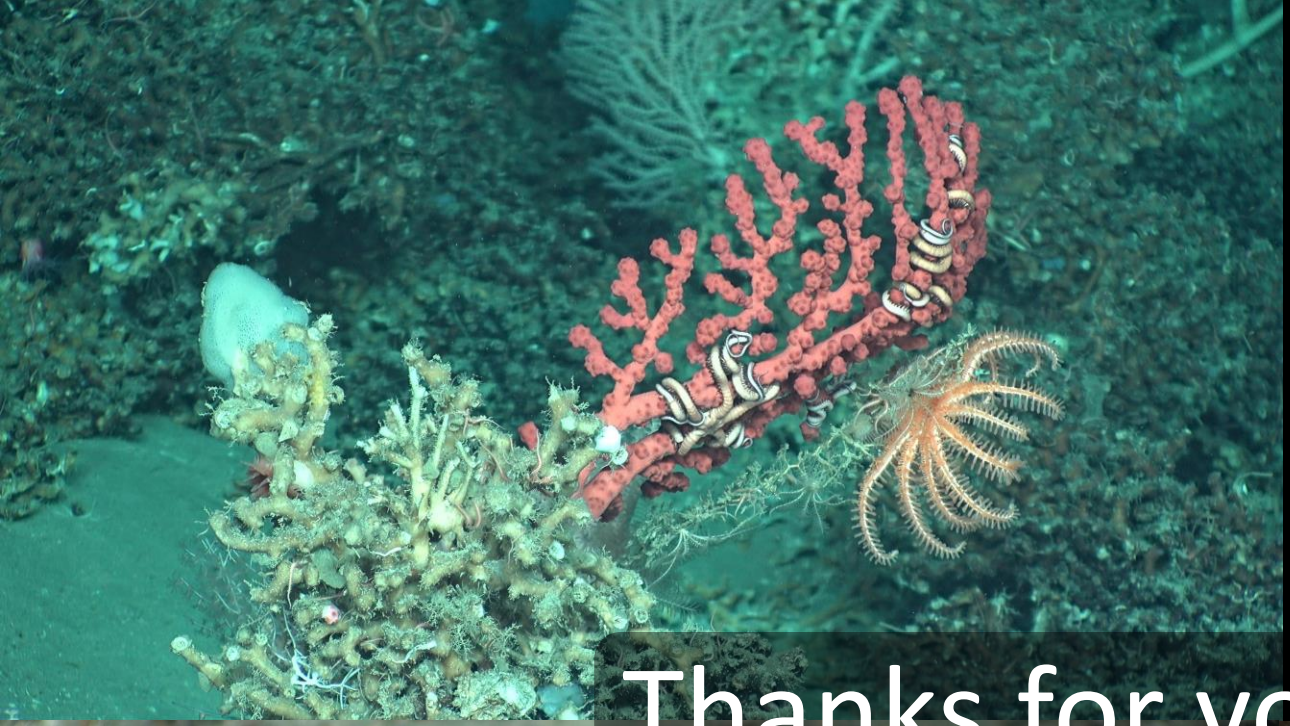
Many friends and colleagues involved in this project

CheReef (Characterization and Ecology of Cold-Water Coral Reefs) - French Oceanographic Fleet

ARDECO (Assessing Resilience of Deep-Water Corals to global change) - ANR (French National Research Agency)

iAtlantic (Integrated Assessment of Atlantic Marine Ecosystems in Space and Time) – European Union

Funded by the Horizon 2020 Framework Programme of the European Union



Thanks for your attention

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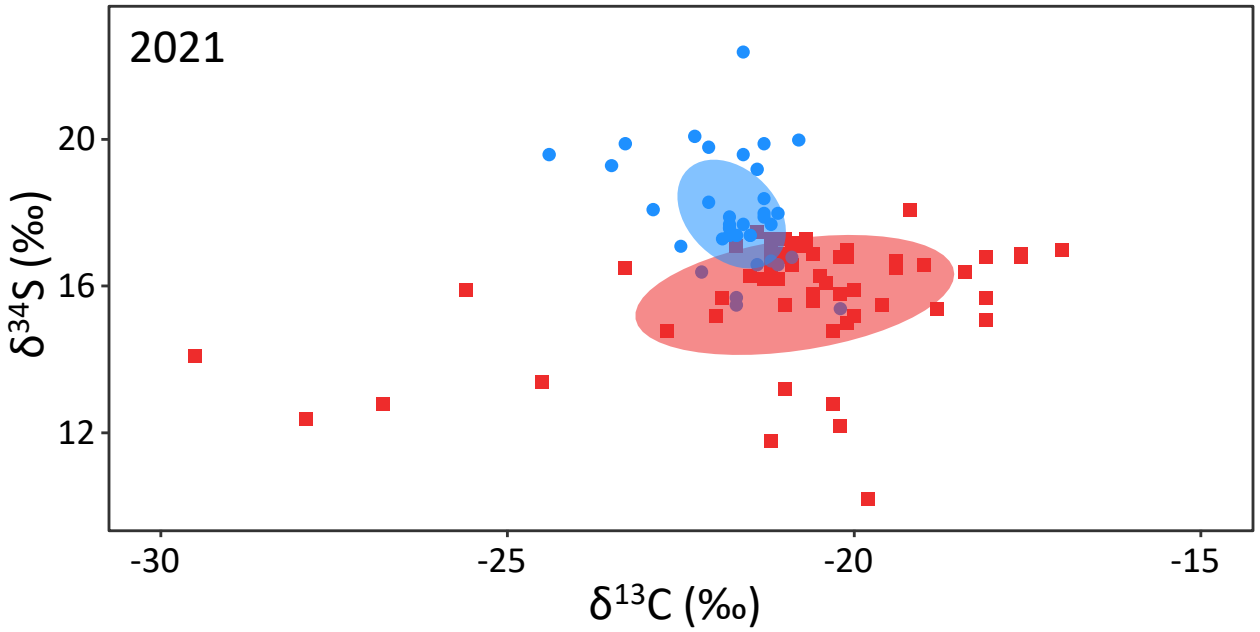
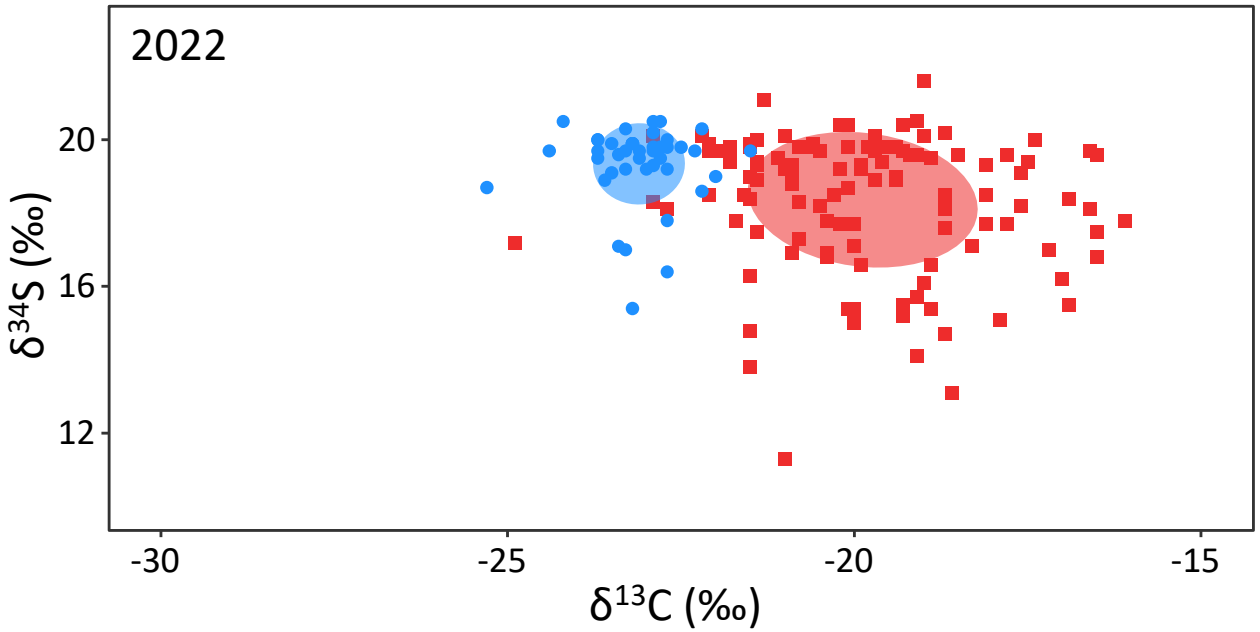
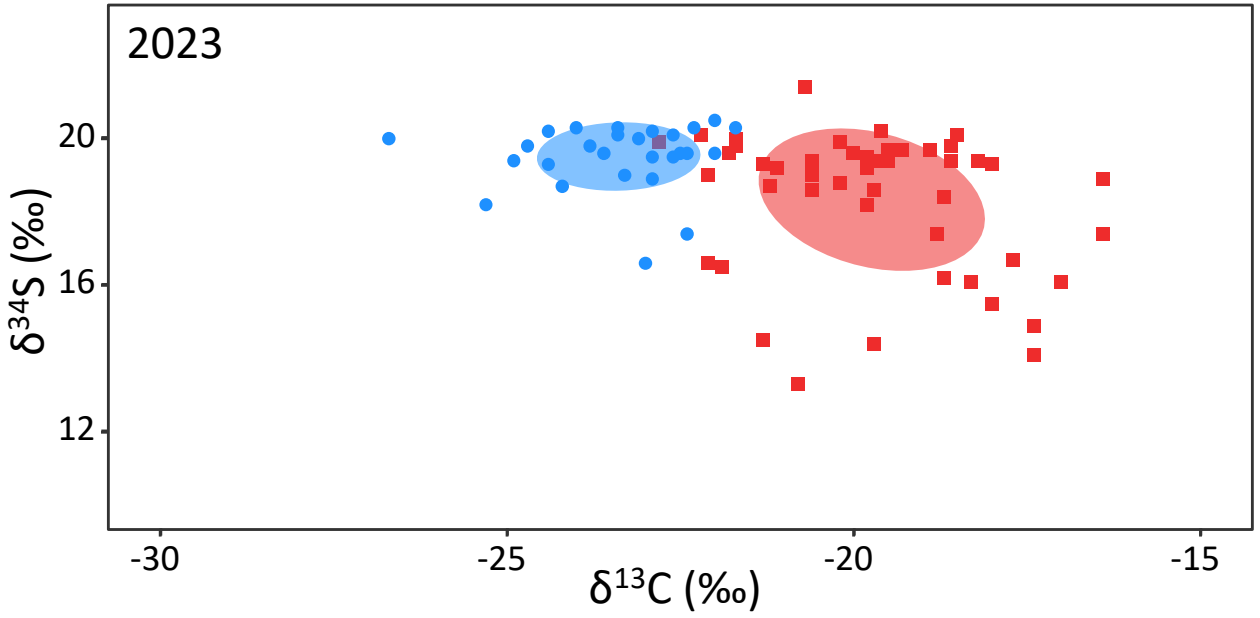
Trophic niches of corals and associated fauna







- Corals
- Associated filter and/or suspension feeders

Fatty acid composition of reserve lipids

Zooplankton trophic markers: Σ 20:1(n-9), 20:1(n-11), 22:1(n-11)

Dinoflagellate trophic markers: Σ 22:6(n-3), 18:4(n-3)

Diatom trophic markers: Σ 16:1(n-7), 16:2(n-4), 16:4(n-1)

Bacteria trophic markers: Σ 15:0, iso15:0, 17:0, iso17:0

Fatty acid composition of reserve lipids