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- 4 Imane Ziani^{1,2,*}, Abdelqader El Guerraf^{2,3,4}, Nour Eddine Bentouhami⁵, Mohamed Brahmi¹,
- 5 Hamza Bouakline¹, Ali El Bachiri¹, Marie-Laure Fauconnier⁶, Sabah Ansar⁷, Farooq Sher^{8,*}
- 6
- 7 ¹Physical Chemistry of Natural Substances and Process Research Team, Laboratory of Applied
 - 8 Chemistry and Environment, Department of Chemistry, Faculty of Sciences, Mohammed First
- 9 University, Oujda 60000, Morocco
- 10 ²International Society of Engineering Science and Technology, Nottingham, United Kingdom
- 11 ³J. Heyrovsky Institute of Physical Chemistry, Dolejškova 2155, Libeň, 18200 Praha 8, Czechia
- ⁴Laboratory of Applied Chemistry and Environment, Faculty of Science and Techniques,
- 13 University Hassan First, BP. 577, Settat 26000, Morocco
- 14 ⁵Bio-Resources, Biotechnology, Ethno-Pharmacology and Health. Research Team, Faculty of
- 15 Sciences, Biology Department, Mohammed First University, 60000 Oujda, Morocco
- 16 ⁶Laboratory of Chemistry of Natural Molecules, Gembloux Agro-Bio Tech, University of Liège,
- 17 Liège, Belgium
- 18 ⁷Department of Clinical Laboratory Sciences, College of Applied Medical Sciences, King Saud
- 19 University, P.O. Box 10219, Riyadh, 11433, Saudi Arabia
- 20 ⁸Department of Engineering, School of Science and Technology, Nottingham Trent University,
- 21 Nottingham NG11 8NS, United Kingdom
- 22

23

24 ***Corresponding authors:**

- 25 Dr. F. Sher
- 26 Assistant Professor
- 27 Department of Engineering, School of Science and Technology
- 28 Nottingham Trent University
- 29 Nottingham
- 30 NG11 8NS
- 31 UK
- 32 E-mail address: <u>Farooq.Sher@ntu.ac.uk (F. Sher), imane.ziani95@outlook.com</u> (I. Ziani)
- 33
- 34 Tel.: +44 (0) 115 84 86679
- 35

36 Abstract

37 Exploring secondary outputs, specifically leftover materials from steam distillation of 38 Rosmarinus tournefortii de Noé, as agents for reducing metals introduces a novel approach to 39 eco-friendly nanomaterial production. This concept aligns with the creation of environmentally 40 conscious nanoparticles, showcasing potential across various fields, notably biomedicine. The 41 paper seamlessly fits into this context. By utilizing R. tournefortii de Noé, successful synthesis 42 of silver nanoparticles (AgNPs) was achieved, yielding nanoscale variations influenced by the 43 plant's by-products. Beyond structural aspects, investigating biomedical applications, focusing 44 on antioxidant and antimicrobial properties. Consistently observing ~94.9–97.3% scavenging 45 inhibition in water residues at different concentrations and enhanced antimicrobial efficacy 46 against Gram-negative and Gram-positive bacteria and Rhodotorula glutinis yeast due to these 47 residues. Moreover, a thorough examination using density functional theory unveiled a robust interaction between silver clusters and specific biomolecules found within the residues, namely 48 homoplantaginin, protocatechuic acid-glycoside, caffeic, and rosmarinic acids (ranging from 49 50 130.62 to 357.05 kcal/mol). These compounds notably enhance the reducing efficacy of Ag+ 51 ions and contribute to the enduring stability of AgNPs (ζ values: -22.8 mV and -17.2 mV). 52 Furthermore, the study recognizes challenges in finding alternative surface modification agents 53 and explores the intricate toxicity mechanisms of silver nanoparticles, emphasizing their 54 interactions with inflammation. Introducing promising nanomedicine approaches involving 55 rosmarinic acid nanoparticles for inflammatory bowel disease and rheumatoid arthritis, 56 highlighting the potential of rosemary by-products derived compounds in innovative therapeutic interventions for diverse inflammatory conditions. 57

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59 Keywords: *Rosmarinus tournefortii* de Noé; Sustainability; Bioconversion; Bio-based
60 materials; nano reinforcement; Antioxidant; biodegradable and DFT optimization.

61 **1. Introduction**

62 Nanotechnology stands as one of the most rapidly advancing domains in the realm of science 63 and technology on a global scale. In this vein, the fabrication of metal nanoparticles constitutes 64 an actively investigated area within nanotechnology, exhibiting exponential advancements across biomedical applications, nutritional sciences, and energy applications (Mittal et al., 65 2013). The challenge of achieving a biogenetic synthesis of uniformly sized nanoparticles with 66 67 distinct shapes remains at the forefront of biomaterials science. This achievement has also 68 rendered substantial advantages within the pharmaceutical sector, particularly in the struggle 69 against an array of bacterial and viral infections (Huh and Kwon, 2011). A broad spectrum of 70 techniques encompassing physical, chemical, biological, and hybrid methodologies is currently 71 employed to engineer nanoparticles of varying properties. Nonetheless, physical and chemical 72 synthesis methods are frequently hindered by elevated production expenses, ecological 73 pollution, and biological hazards (Bereza-Malcolm et al., 2015). Consequently, the biological 74 approach emerges as an alternative to traditional chemical and physical methods, serving as an 75 eco-friendly avenue for nanoparticle production. Moreover, this approach obviates the 76 necessity for costly, perilous, and toxic substances (Naghdi et al., 2022).

77

78 Ongoing investigations into the bio-production of non-metallic substances through botanical 79 extracts have unveiled a fresh avenue, offering swift and benign techniques within the redox 80 reaction for the creation of environmentally conscious nanoparticles. A multitude of scholars 81 have documented the biogenic synthesis of metal nanoparticles via plant leaf extracts, 82 spotlighting their potential uses. This propensity is ascribed to the presence of secondary 83 metabolites, encompassing phenolic acids, flavonoids, alkaloids, and terpenoids, which 84 primarily contribute to the conversion of ions into substantial metal nanoparticles (Pisoschi et 85 al., 2022). Several antecedent studies have underscored the proficiency of biosynthesized

86 nanoparticles in regulating oxidative stress, genotoxicity, and changes associated with 87 apoptosis (Zhang et al., 2018). Furthermore, nanoparticles boast an extensive array of 88 applications within the domains of agriculture and plant sciences. For instance, through the 89 utilization of bioprocessing technology, nanoparticles can transform agricultural and food 90 residues into energy (Tariq et al., 2023) and valuable by-products (Usmani et al., 2022).

91

92 Moreover, researchers have extensively explored the utilization of lignocellulosic biomass for 93 synthesizing metallic nanoparticles, elucidating its manifold applications and advantages. Chen 94 et al. (Chen et al., 2023) emphasize green synthesis methods using biomass components like 95 cellulose, hemicellulose, and lignin, resulting in nanoparticles with unique properties 96 applicable in catalysis, sensing, and biomedicine. For instance, cellulose-derived nanoparticles 97 exhibit high catalytic activity, with conversion rates exceeding 90% in chemical reactions, 98 while hemicellulose-derived ones demonstrate exceptional sensing capabilities, detecting 99 pollutants at concentrations as low as 1 ppb (Chen et al., 2023). Lignin-derived nanoparticles 100 show promising antimicrobial properties, with inhibition rates of up to 95% against bacteria, 101 underscoring sustainability (Chen et al., 2023). Conversely, Sankaran et al. (Sankaran et al., 102 2021) focus on enhancing bioenergy production through biomass-to-nanoparticle conversion. 103 They highlight the use of magnetite nanoparticles (MNPs) alongside alkaline pretreatment on 104 rice straw, resulting in a significant increase in biogas and methane yield by 100% and 129% 105 respectively (Sankaran et al., 2021). Additionally, incorporating nanoparticles into acid-106 functionalized magnetic nanoparticles boosts sugar production by 46% (Sankaran et al., 2021). 107 Both studies underscore the value of biomass for nanoparticle synthesis, addressing various 108 fields and offering sustainable solutions.

110 Despite the widespread use of plant extracts for nanoparticle synthesis, one of the most recent 111 and promising methods, based on the natural "bio-laboratory," is the use of biodegradable 112 wastes generated by the agricultural and food industries. Moreover, these waste materials are 113 abundant, cost-effective, and readily available, obviating the need for elaborate pre-processing 114 procedures. Various experiments were conducted to synthesize nanoparticles using different waste sources. In this context, zinc oxide nanoparticles were produced employing waste from 115 116 Phoenix dactylifera as a bio-reductant for efficient dye degradation and antibacterial 117 effectiveness in wastewater treatment (Rambabu et al., 2021). PVP-coated silver nanoparticles 118 (PVP-AgNPs) were employed in municipal solid waste composting (Gitipour et al., 2013). 119 Additionally, the Fenton process employed copper nanoparticles derived from printed circuit 120 boards to degrade mining surfactant (Martins et al., 2021). Secondary metabolites, which are 121 the main metal-reducing agents in the "green" synthesis of nanoparticles, are found naturally 122 in waste from the essential oil industry. Raw materials with a low essential oil content generate 123 a lot of waste. Although distilleries generally dispose of their residues, these common practices 124 can disrupt the ecological balance of the site and result in the loss of valuable biologically active substances present in the waste (Da Silva et al., 2016). 125

126

127 Amidst the diverse tapestry of rosemary species, Rosmarinus officinalis L. stands revered for 128 its therapeutic prowess and the generous yield of its essential oils. However, a notable void 129 exists in the expansive landscape an absence of comprehensive exploration into the species 130 Rosmarinus tournefortii de Noé. While R. officinalis L., has received considerable attention, 131 the intricacies of *Rosmarinus tournefortii* de Noé have been largely overlooked. This study 132 aims to fill this void by uncovering the untapped potential of Rosmarinus tournefortii de Noé, particularly in utilizing its waste by-products for nanoparticle synthesis. Specifically, the focus 133 134 is on utilizing water and solid by-products as reduction agents for Ag, Zn, and Cu metal ions.

The synthesis process was meticulously characterized using advanced techniques to provide comprehensive insights into the resulting materials. Additionally, specialized analyses were conducted to evaluate the efficacy of biomolecules within each by-product, with a comparative analysis between water and solid by-products to understand their roles in nanoparticle synthesis and their effectiveness as reduction agents.

140

141 Moreover, the study delves into the potential biological applications of the synthesized 142 nanoparticles, particularly their antioxidant and antimicrobial properties, suggesting promising 143 avenues in medicine and materials science. However, challenges and unexplored territories 144 remain. The exploration of alternative complexation agents for nanoparticle surface 145 modification is identified as a growing research area to enhance nanoparticle stability and 146 functionality. Additionally, the study underscores the promising potential for health-enhancing 147 applications in drug delivery and medical imaging. Overall, this study showcases the ability to 148 repurpose industrial by-products for eco-friendly nanoparticle synthesis, showcasing 149 accomplishments while highlighting avenues for future investigation. This multidisciplinary 150 approach positions by-product-derived nanoparticle synthesis as a key player in sustainable 151 nanotechnology and its myriad applications.

152 **2. Materials and methods**

153 **2.1. Plant material extraction**

The untamed *Rosmarinus tournefortii* de Noé plant's fresh leaves were harvested amidst its flowering phase on March 13, 2020, within the Megrez forest region of eastern Morocco, located at coordinates 34° 43′ 52.6″ N 2° 04′ 21.5″ W (Ziani et al., 2023). The plant's identification was confirmed by Tahri Tahar, the director of the Forest Management Studies Service of Oriental, Morocco. Voucher specimens were deposited at the Physical Chemistry of Natural Substances and Process Laboratory, Faculty of Sciences, Mohammed First University,

160 Oujda, Morocco. These leaves, once dried, underwent steam distillation on a semi-pilot scale to extract the pertinent essential oil. The liquid residue produced during steam distillation was 161 separated from the solid residue through filtration using a 90 mm Whatman (GF/A) filter and 162 163 subsequently concentrated utilizing a rotary evaporator. In the case of the solid residue, it was pulverized and air-dried over a period of approximately 15 days. Following this, the residue 164 was subjected to de-ionized water extraction for an hour. The resultant extracts from the two 165 166 categories of rosemary by-products, namely the liquid and solid residues, were securely stored 167 in amber glass vials and maintained at a refrigerated temperature of 4 °C until subjected to 168 analysis.

169 2.2. Biosynthesis of silver, zinc and copper nanoparticles

170 In the process of green synthesizing Ag, Zn, and Cu-NPs, the well-established protocol by Raut 171 et al. (Raut et al., 2014) was adapted with minor adjustments. The initial step involved 172 dissolving 2.5 g of liquid and solid residue extracts in 50 mL of de-ionized water to formulate 173 the aqueous extracts. Silver nitrate, zinc nitrate, and copper nitrate served as the precursor 174 materials for Ag, Zn, and Cu nanoparticles. The prepared extract was then introduced to 0.1 M solutions of AgNO₃, ZnNO₃, and CuNO₃, maintaining a ratio of 1:2 (v/v). The solution mixture 175 176 was agitated at 70 °C for 3 hours. After the nanoparticle synthesis was finalized, centrifugation was employed to isolate Ag, Zn, and Cu-NPs. The supernatant was separated post-177 178 centrifugation, and the resulting precipitates underwent triple washing using de-ionized water, followed by drying at 60 °C for two days (Fig. 1). 179

180 2.3. Characterization of biosynthesized nanoparticles

181 To confirm the generation of nanoparticles, a Shimadzu UV 1650-PC UV-visible 182 spectrophotometer was utilized. The measurement of absorbance was carried out within the 183 range of 300–800 nm (Lv et al., 2021). Attenuated Total Reflectance-Fourier Transform

184 Infrared (ATR-FTIR) analysis was executed employing a Jasco 4700-FTIR spectrometer from Shimadzu, Japan. This was done to compare the characteristics of the dried plant material with 185 those of the synthesized nanoparticles, aiming to decipher the role of reducing agents in 186 187 metallic ion reduction. The absorption spectra were captured over a wavelength range of 400 to 4000 cm⁻¹ (Borah et al., 2021). Microstructural scrutiny of the developed charged 188 189 nanoparticles was performed using a JEOL-JSM7001F instrument and field-emission scanning 190 electron microscopy (FE-SEM). The SEM filament operated at various currents and a voltage 191 of 5 kV at different magnifications, while the fixed working distance was maintained at 6 mm 192 (representing the unchanging separation between the sample and the objective lens). Elemental composition was determined through X-ray energy dispersive spectroscopy (EDS). The mean 193 194 diameter and distribution of the nanoparticles were measured using a laser granulometer, 195 specifically the Anton Paar Litesizer 500 (de Souza Niero et al., 2023). These measurements 196 were executed at room temperature within a liquid cell containing a dispersant with a refractive 197 index of 1.33. Zeta potential was determined via electrophoresis and the application of the 198 Smoluchowski equation to analyze particle mobility. XRD patterns of all nanoparticle samples 199 were captured using a Shimadzu XRD-6000 diffractometer and a Cu K ($\lambda = 0.154$ nm) radiation 200 source. Data collection occurred at a scanning speed of 0.02° /s within a diffraction angle span of 10° to 80° . The crystallite sizes of the nanoparticles were deduced utilizing the Scherrer 201 202 formula (Eq. (1)) (El Guerraf et al., 2023), with the average diameter of silver crystals (D) 203 being calculated based on the (111) plane.

204

$$D = \frac{\kappa\lambda}{\beta cos\theta} \tag{1}$$

205 **2.4.** Matrix of rosemary by-products

In the evaluation of rosemary by-products, a High-Performance Liquid Chromatography (HPLC) System, furnished with a 2998 Photodiode Array Detector and a reversed-phase C18 column (5, 250×4.6 mm), was utilized. Following the procedure outlined by Liu et al. (Liu et

209 al., 2011), extracts with a concentration of 5 mg/mL were introduced into the column, flowing 210 at a rate of 0.8 mL/min, utilizing a gradient mixture of binary solvents. To separate phenolic 211 compounds, a mobile phase consisting of two constituents was employed: mobile phase A 212 (acetonitrile with 0.1% formic acid) and mobile phase B (water with 0.1% formic acid). The 213 gradient pattern followed this sequence: starting at 60% B at 0 minutes, transitioning to 50% 214 B at 2 minutes, maintaining 50% B at 10 minutes, decreasing to 30% B at 15 minutes, 215 maintaining 30% B at 25 minutes, and returning to 60% B at 32 minutes. To identify specific 216 phenolic components, their retention time and maximum wavelength were assessed and 217 compared against established standards and available literature data.

218 2.5.

. DPPH radical scavenging assay

The 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay was carried out with a slight alteration 219 220 derived from our previously outlined procedure (El Guerraf et al., 2023). This assay is 221 commonly employed to evaluate the antioxidant potential of environmentally produced nanoparticles. In a concise summary, varying concentrations (0.1, 0.5, 1, 1.5 and 2 mg) of the 222 223 nanoparticles were mixed with 2500 µL of DPPH solution in methanol (0.04 mg/mL). After 224 being left in the dark for 60 minutes, the absorbance at 517 nm was gauged via a UV-vis 225 spectrophotometer. The scavenging inhibition (%) was determined using the subsequent 226 formula, depicted by Eq. (2), in which A0 symbolizes the absorbance of the control, and AC 227 signifies the absorbance of the tested samples.

228
$$DPPH Inhibition (\%) = \frac{A0 - AC}{A0} \times 100$$
(2)

229 2.6. Anti-microbial properties

Following prior investigations (Abdollahzadeh et al., 2021), the agar diffusion method was employed to assess the antibacterial properties of nanoparticles synthesized from rosemary byproducts on solid surfaces. The in vitro antibacterial performance was evaluated against distinct

233 microorganisms, namely Listeria innocua ATCC 33090 (gram-positive bacteria), Escherichia 234 coli ATCC 25922 (gram-negative bacteria), along with Rhodotorula glutinis (yeast) and Geotrichum sp. (mold). For both microbial types, the strains were adjusted to a 0.5 McFarland 235 standard, equivalent to 10⁶ CFU/mL and 10⁶ spores/mL, correspondingly (Mahdi et al., 2022). 236 237 Afterwards, these cultures were additionally thinned using Mueller-Hinton broth for bacteria, 238 yeast, and mould, and sterile physiological water for both, before being introduced onto the 239 surface of petri dishes. In this approach, wells (6 mm) were generated in Mueller-Hinton agar 240 (MHA) previously inoculated with the target bacteria or fungus. These wells were subsequently 241 loaded with 10 µL of samples (10 mg/mL). Incubation of the agar plates facilitated bacterial and fungal growth, conducted at 37°C for 18 hours and 25°C for 48 hours respectively. The 242 243 antimicrobial efficacy was evaluated by measuring the size of the inhibition zone within the 244 agar medium. To ensure robustness, each assay was carried out in triplicate.

245 **2.7.** Computational study

Initially, the major compounds (namely, caffeic acid, homoplantaginin, protocatechuic acid-246 247 glycoside, rosmarinic acid, epicatechin, and gallocatechin) extracted from Rosmarinus tournefortii de Noé by-products were optimized at the ground state using the GAUSSIAN 09 248 249 quantum chemistry simulation software (Kaushik et al., 2022). Subsequently, each individual 250 molecule was optimized in conjunction with a silver cluster by placing it in close proximity to 251 reactive sites that were identified through analyses of molecular electrostatic potential (MEP). 252 To reduce computational costs, representative models of silver nanoparticles (AgNPs), 253 consisting of a single silver atom (Ag_1) and three silver atoms (Ag_3) clusters, were utilized. 254 The interaction energies between the AgNPs models and the target molecules were estimated 255 using Eq. (3) (Kaushik et al., 2022).

256

257

 $\Delta I = E_{Ag-compound} - (E_{Ag} + E_{compound})$ (3)

Where $E_{Ag-compound}$, E_{Ag} , and $E_{compound}$ represent the energies associated with the interaction between AgNPs and the target molecule, AgNPs themselves, and the target compound, respectively. The optimization through DFT was performed using the B3LYP hybrid functional at the Lee-Yang-Parr calculation level. The organic compounds underwent treatment with the 6-311G(d,p) basis set, whereas the Ag atoms were characterized using the LanL2DZ basis set. Conventional convergence criteria were applied, and visualization of the molecular structure was facilitated using the GaussView 5.0 molecular editor.

266 **3. I**

3. Results and discussion

267 **3.1.** Optical absorption and crystalline structure analysis

UV-vis spectroscopy is a highly effective method for analyzing the optical response of metal 268 269 nanoparticles, particularly sensitive to their formation due to the pronounced surface plasmon 270 resonances (SPRs) they exhibit (Kelly et al., 2003). In this study, the UV-vis absorption spectra 271 of colloidal nanoparticles produced from the two types of rosemary residues were examined. 272 The spectral data demonstrated distinctive surface plasmon (SP) bands associated with AgNPs, 273 showing variations in their " λ max" and intensity of the SP band. This distinction underscores 274 the evident impact of the compound's characteristics on these particular parameters. In contrast, 275 when Zn and Cu were employed for nanoparticle synthesis, the SP band intensity spectra remained unchanged, indicating the unsuccessful development of ZnNPs and CuNPs. The 276 277 consistent surface plasmon (SP) band intensity spectra observed during the utilization of Zn 278 and Cu for nanoparticle synthesis suggest the unsuccessful development of ZnNPs and CuNPs, 279 potentially due to various factors. Inadequate synthesis conditions, such as suboptimal 280 temperature, pH, or reaction time, could have led to the formation of nanoparticles with 281 properties deviating from the desired plasmonic characteristics (Kim et al., 2024).

283 Additionally, the propensity of Zn and Cu nanoparticles to oxidize in air or aqueous 284 environments may have resulted in the formation of oxide layers on the nanoparticle surface, affecting their plasmonic properties (Jahan et al., 2021). Agglomeration or aggregation of 285 286 nanoparticles during synthesis or post-synthesis treatments could have altered the interactions between nanoparticles, impacting the observed SP band intensities (Pryshchepa et al., 2020). 287 288 Furthermore, surface contamination with impurities or residues from the synthesis process 289 might have interfered with the plasmonic properties of ZnNPs and CuNPs. The absence of 290 appropriate surface ligands or stabilizing agents during synthesis could have led to unstable 291 nanoparticles with modified plasmonic behavior (Kim et al., 2023). These factors collectively 292 contribute to the challenges in achieving the successful development of ZnNPs and CuNPs 293 with the desired plasmonic properties.

294

295 The spectra were captured when the colloidal sample's colour and absorption strength remained 296 constant. Fig. 2(a, b) illustrates that each residue used exhibits a single, distinct SPR position 297 in the 300-500 nm range. Additionally, no SPRs were seen at wavelengths greater than 500 298 nm, suggesting that the majority of the AgNPs produced have small sizes and comparable 299 shapes. This observation also provides early clues about colloidal AgNPs' size and size 300 distribution. Comparing the plasmon band positions of the two different residues, the solid 301 residue revealed higher wavelengths (372 nm) than the water residue (368 nm), demonstrating 302 a reduction in AgNPs particle size. Similar to this, several earlier studies from the *Rosmarinus* 303 officinalis L. plant species have also noted the absorption spectrum between 350 and 500 nm 304 brought on by AgNPs' surface plasmon resonance (Ghaedi et al., 2015; Noukelag et al., 2021). 305

X-ray diffraction (XRD) and Fourier-transform infrared spectroscopy (FTIR) were conducted
 to gain information on the one hand about the size, lattice, and structure of nanoparticles, and

308 on the other hand, about any potential bioactive compounds present in Rosmarinus tournefortii 309 de Noé and that can possibly act as reducing/stabilizing agent. For a better comparison, the 310 spectroscopic analysis was achieved in the case of Zn, Cu, and Ag biosynthesized in the 311 presence of the two types of rosemary by-products; solid and water residues. Firstly, by 312 analysing the XRD pattern obtained for the tested by-products, it is clear the amorphous nature 313 of both SR and WR (Fig. 2(c, d)). When using Zn or Cu, the XRD spectra remain unchanged 314 suggesting the unsuccessful elaboration of ZnNPs and CuNPs. The diffractograms resulted for 315 silver nanoparticles were totally different as they showed four well-resolved peaks at 2θ angles 316 of 38.27°, 44.38°, 64.62° and 77.52° for SR, and 38.15°, 44.32°, 64.55°, and 77.45° in the case 317 of WR. These prominent peaks arise from the (111), (200), (220), and (311) Bragg reflections 318 of face-centred cubic (fcc) structures of AgNPs (JCPDS 04-0783). Further, the average 319 crystallite size (d) of AgNPs was estimated using the Debye-Scherrer equation, $d = K\lambda/\beta \cos\theta$ 320 where K is the shape factor (between 0.9 and 1.1), λ is the incident X-ray wavelength of Cu 321 K α radiation (1.542 Å), β is the full width at half maximum in radians of the (111) line, and θ 322 is the Bragg diffraction angle. The average AgNPs sizes were found to be 17.98 and 18.49 nm 323 for SR@AgNPs and WR@AgNPs composites, respectively. The obtained results are more 324 interesting than those of Das et al. (Das and Velusamy, 2013) and Ghaedi et al. (Ghaedi et al., 2015) where it was reported that the mean AgNPs particle size ranged from 31.79 to 33 nm for 325 326 the plant genus Rosmarinus officinalis L. respectively.

327

On the other hand, FTIR characterization for all samples resulted in the spectra presented in (**Fig. 2(e, f)**). For both SR and WR, several typical bands were observed that describe functional groups associated with various biological macromolecules in the Rosemary leaf extracts. The peak with relatively high intensity at 1020 cm⁻¹ is assigned to C-O-C stretching vibration or alcohols/phenols (Farshchi et al., 2018). The bands at around 1152 and 1256 cm⁻¹ arise from

333 C-O stretching and represent the presence of polyphenols and the one that appears at 1391 cm⁻ ¹ corresponds to C–O–C stretching modes of vibration. The signal at 1514 cm⁻¹ demonstrates 334 also the presence of phenolic compounds from the extract which is related to the stretching of 335 336 the aromatic rings (Farshchi et al., 2018). The peak detected at approximately 1587 cm⁻¹ may result from the vibration of C=C groups, while the one near 1713 cm^{-1} is associated with C=O 337 338 groups from carboxylic acids (Piñeros-Hernandez et al., 2017). The stretching vibration of C-H is justified by the peak appearing at 2931 cm⁻¹. Finally, the broad absorption band at a 339 wavenumber between 3020-3620 cm⁻¹ is linked to O-H stretching (Piñeros-Hernandez et al., 340 341 2017).

342

343 After the bio-synthesis of AgNPs, the spectrum remains practically unchanged demonstrating 344 that the metal nanoparticles did not alter the structure of the rosemary plant in a significant 345 way. It should be noted, however, that some IR bands were shifted to lower frequencies after 346 the interaction of the residues with Ag. Based on the ATR-FTIR results and previously reported 347 papers for *Rosmarinus officinalis* L. species (Rabiee et al., 2020), the mechanism of reducing 348 silver ions to metallic silver can be discussed. In a simple way, numerous bioactive compounds 349 from *Rosmarinus tournefortii* de Noé will surround Ag⁺ producing a coating. The latter receive 350 electrons from these phytochemical constituents resulting in the reduction of silver cation and 351 avoiding the agglomeration of the particles. Most likely, the carbonyl groups from the bioactive 352 compounds are responsible for the reduction process and can act as a stabilizer and bio-capping 353 agents of the AgNPs. In summary, the spectroscopic analyses have demonstrated the successful 354 green synthesis of AgNPs using both Rosmarinus tournefortii de Noé solid and water residues. 355 Apart from Zn and Cu, no promising results were obtained, which may be related to the absence of sufficient reducing agents or suitable biomolecules to facilitate the reduction of metal ions, 356 357 thus hindering NPs formation.

358 The limitations in ATR-FTIR analysis, despite comparing plant extract with synthesized 359 CuNPs or ZnNPs, stem from challenges inherent in the nanoparticle synthesis process, as evidenced by prior UV-vis and DRX analyses. Incomplete reduction of metal ions during 360 361 synthesis may hinder the formation of well-defined nanoparticles with distinct chemical characteristics (Adra et al., 2024). Moreover, issues like nanoparticle aggregation or 362 precipitation, due to inadequate stabilization or unfavorable reaction conditions, can obscure 363 364 changes in the ATR-FTIR spectra. These aggregated nanoparticles may exhibit broad or 365 overlapping peaks, making it challenging to discern specific features. Additionally, the 366 sensitivity of ATR-FTIR may not be sufficient to detect subtle changes in chemical 367 composition or surface functional groups, particularly with low nanoparticle concentration or 368 minimal changes (Lee and Chae, 2021). Background signals or noise in the spectra could 369 further complicate the analysis. Weak or transient interactions between plant extract 370 components and Cu or Zn ions may result in subtle or undetectable changes in FTIR spectra, 371 possibly due to the nature of binding sites on the plant extract molecules (Jędrzejczyk et al., 372 2023). Together, these factors contribute to the limited changes observed in ATR-FTIR 373 analysis following unsuccessful nanoparticle synthesis. As the mechanism of interaction 374 remains always hard to fully understand, further optimization of the experimental conditions 375 may be necessary in future studies. It is essential to highlight that this study primarily 376 concentrated on the synthesis of nanoparticles from Rosmarinus tournefortii de Noé, a species 377 not previously investigated for this purpose, suggesting avenues for future exploration.

378

3.2. Hydrodynamic size and surface analysis

The enduring stability of colloidal silver nanoparticles was observed through spectroscopic monitoring using the zeta potential technique. This method, commonly employed for managing the stability of colloidal metal nanoparticles, gauges alterations in surface charge. Metal nanoparticles possessing a notably positive or negative zeta potential exhibit mutual repulsion,

383 preventing them from aggregating. Conversely, particles with low absolute zeta potential 384 values tend to aggregate and coalesce due to the absence of repulsive forces that hinder such 385 accumulation (Rao et al., 2021). Zeta potential (ζ) results for the two types of *Rosmarinus* 386 tournefortii de Noé steam distillation by-products; solid residues and water have ζ values of -13.3 and -11.4 mV, while the freshly prepared colloidal AgNPs have ζ values of -22.8 and -387 388 17.2 mV, respectively. The results obtained showed a strong negative value for AgNPs, clearly 389 suggesting the stability of nanoparticles. Moreover, the stability and colloidal behaviour of the 390 nanoparticles were explored as shown in Fig. 3(a, b), yielding insightful findings. Notably, a 391 single peak position was observed on the distribution graph for SR@AgNPs, indicating that 392 the nanoparticles in this residue exhibited a uniform size of approximately 92.3 nm and shared 393 a similar surface charge. Such uniformity is characteristic of a monodisperse (narrowly 394 distributed) sample, which holds significant appeal for numerous applications. Additionally, 395 intriguingly, WR@AgNPs displayed two distinct peaks in the nanoparticle size distribution 396 (62.2 and 111.64 nm). Each peak represented a group of particles with comparable sizes. This 397 bimodal distribution suggested the existence of two separate populations of nanoparticles in 398 the sample. Possible explanations for this phenomenon could include the aggregation or 399 agglomeration of nanoparticles, resulting in larger particles that contribute to the second peak 400 in the size distribution. Furthermore, the presence of various morphologies or crystalline 401 structures among the nanoparticles might also contribute to the bimodal size distribution. For 402 instance, certain metal nanoparticles can exhibit different shapes, such as spherical and rod-403 shaped, leading to diverse size populations (Yaraki et al., 2022).

404

405 Additional support for the successful production of AgNPs was uncovered through 406 morphological examinations and elemental analysis. As illustrated in **Fig. 3(c, d)**, the obtained 407 nanoparticles exhibited distinct shapes. The two types of by-products displayed predominantly

408 spherical morphology in their nanoparticles, each exhibiting varying diameters within the 409 nanoscale range. Because of their diminutive size, extensive surface activity, and considerable 410 specific surface area, the nanoparticles had a proclivity to readily form aggregates. The 411 effectiveness of synthesizing AgNPs using Rosmarinus tournefortii de Noé solid and water 412 residues was further confirmed by analyzing the chemical composition based on the EDX 413 spectrum (Fig. 3(e, f)). The optical absorption band of the EDX peak in the 3-4 keV range is 414 characteristic of metallic silver nanocrystallites' absorption (Kotakadi et al., 2014). The EDX 415 spectrum of the synthesized silver nanoparticles (Fig. 3(e, f)) clearly showed the absence of 416 elemental nitrogen peaks and the presence of elemental silver metal, along with C and O, which 417 are associated with the phenolic compounds of the residues and the formation of Ag NPs. The 418 distinct signal peak of silver provided strong evidence of the successful reduction of silver 419 nitrate to silver nanoparticles. Our findings align with earlier studies on AgNPs produced from 420 the Rosmarinus officinalis L. plant species (Das and Velusamy, 2013; Ghaedi et al., 2015). 421 Those studies also noted the presence of uniformly sized, spherical silver nanoparticles in the 422 nanoscale range.

423 **3.3.** Matrix responsible for metal ion reduction

424 Many active plant chemicals with therapeutic or dietary benefits have been used to create 425 nanomaterials. Among these, flavonoids and phenolic acids have received a great deal of 426 attention because of their potential uses in nano-medicine. To better understand the nature of 427 the molecules responsible for stabilization and reduction in the synthesis of metal NPs, HPLC-428 DAD analysis was carried out. Through comparing the retention times and UV-visible spectra 429 of the specimens with established reference standards, it was possible to swiftly identify the 430 presence of gallic acid, epicatechin, chlorogenic acid, rosmarinic acid, caffeic acid, and 431 apigenin. To identify the flavonoids, data relating to the flavonoid elution scheme as described 432 in the literature were also examined in detail. Both types of rosemary steam distillation by-

433 product extracts contained phenolic acids and flavonoids, which have been tentatively434 identified in **Table 1** as two families.

435

436 According to the reference standards, peak 1,2, 3/6, 7/5, 9/7, and 10/8 for solid and water (SR/WR) residues, respectively, were positively identified as the chemical structures of gallic 437 acid, epicatechin, chlorogenic acid, caffeic acid, rosmarinic acid, and apigenin (Fig. 4). The 438 439 same identification was discovered by prior studies for rosemary (Bendif et al., 2017). Additionally, since the third peak's absorbance band falls within the 275–340 nm range, values 440 441 that are nearly identical to the UV-vis spectra detected by numerous studies (270-340 nm), the 442 third peak may be assumed to be homoplantaginin for the solid residue extract (Miguel Herrero 443 et al., 2010). According to the UV-vis spectrum of peak 4 for the water residue, protocatechic 444 acid glycoside was recognized for this peak with maxima at 220.0 and 281.2 nm, which is 445 almost identical to the spectra given by literature data for the same plant (M. Herrero et al., 2010). For peaks 4 and 5 for solid and water residues, respectively, flavanol gallocatechin is 446 447 highly recommended, whose maximum spectra were recorded between 283 and 335 nm, values 448 quite close to those corresponding in our study (283, 334 nm). Further, the patterns of the two 449 UV spectra are similar, demonstrating that those peaks are linked to gallocatechin (Almela et al., 2006). Based on the spectrum maximum at 275.3 nm, which is characteristic of spectral 450 451 maxima observed in several studies (de Almeida Gonçalves et al., 2018) (Gonçalves et al., 452 2019), yunnaneic acid F is the most suggested structure for peak 8 of the water residue extract.

453

454 Comparing the two aqueous by-product extracts, the water residue remained to extract more 455 phenolic compounds than the solid residue that could be attributed to the prolonged contact of 456 the plant material with boiling water, a cell permeation effect favoring the extraction of the 457 metabolites can be assumed. The harsh steam distillation conditions can also cause the

458 formation of phenolic artefacts in the water residue. In addition, high recovery of rosmarinic 459 acid and protocatechuic acid glycoside was revealed in water residue, while epicatechin and 460 homoplantagenin had to be extracted to the maximum in the solid residue. Caffeic acid is 461 extracted in almost the same way in both residues, meaning that it takes longer to extract it in its entirety in the aqueous residue. These findings align with prior observations by Miljanovic 462 et al. (Miljanović et al., 2023), indicating a higher efficacity of phenolic acid (rosmarinic, 463 464 syringic, and caffeic acids) and flavonoid (gallocatechin and apeginin) extraction in both byproducts, especially in the water residue. However, a limitation arises from the inability to 465 466 compare products using ethanol/water as the extraction solvent for the solid residue, hindering a comprehensive analysis. Discrepancies with the findings of Luca et al. (Luca et al., 2023), 467 468 highlight that solid and water residues had the same elution pattern for most polar compounds, 469 while phenolic diterpenes were absent in the water residue. The authors compared the two by-470 products with a different solvent extract, for which the solid residue was extracted using SC-471 CO₂ extraction or even the solid residue from hydrodistillation extracted with methanol/water, 472 which is normal to have phenolic diterpenes since the solvents used can rapidly extract a range 473 of molecule polarities.

474

Moreover, observations from Wollinger et al. (Wollinger et al., 2016) regarding limited 475 476 phenolic compound presence in water residue (rosmarinic acid and traces of carnosol and 477 carnosic acid) underscore the potential impact of the distillation process duration. Conversely, 478 studies by Celano et al. (Celano et al., 2017) and de Elguea-Culebras et al. (de Elguea-Culebras 479 et al., 2023) reported the presence of all phenolic compounds, including phenolic diterpenes, 480 in water residue, emphasizing the complexity of compound extraction influenced by solvent choice and extraction method. Despite these insights, a comprehensive elucidation of the 481 482 comparison between the two aqueous extracts from steam distillation by-products remains

elusive, warranting further investigation. Limited published data on the chemical composition
of the aqueous residue from rosemary distillation, particularly for *Rosmarinus officinalis* L.
and *Rosmarinus tournefortii* de Noé species, underscores the need for future research to
validate these findings.

487 **3.4. DPPH radical scavenging by elaborated nanoparticles**

488 Antioxidants are widely acknowledged for their potential efficacy in treating and preventing 489 various diseases. However, a notable limitation arises from the low permeability and poor water 490 solubility of most antioxidants, leading to instability during storage and degradation in the 491 gastrointestinal tract (Oliveira et al., 2021). Consequently, their practical utility has been 492 restricted. To address this issue, the amalgamation of materials science and nanotechnology 493 has played a pivotal role in reducing the generation of free radicals during nanoparticle 494 production. These specialized nanoparticles, termed nano-antioxidants, have emerged as a 495 solution (Dal Lago et al., 2011). In this study, the nanoparticles' capacity to mitigate the DPPH 496 free radical was investigated. The efficacy of the prepared samples in this regard was assessed 497 using a straightforward methodology, measuring their scavenging activity against this stable free radical. 498

499

500 Antioxidant capacities of elaborated materials are presented in Fig. 5. For comparison, the two 501 types of Rosmarinus tournefortii de Noé by-products were also tested, and the results are 502 illustrated in the same figure. As highlighted, the arrangement of the two types of by-products 503 is the same, for which the incorporated AgNPs showed significantly higher inhibition in the 504 case of SR/WR@AgNPs than the free by-products extracts, while the unsuccessful 505 SR/WR@CuNPs and SR/WR@ZnNPs syntheses showed the lowest entrapment with DPPH 506 inhibition respectively. Furthermore, upon comparing the evaluated scavenging capacities of 507 the two by-product types, the water residue emerged as the most effective for incorporation.

508 Notably, the scavenging inhibition, ranging from 94.9 to 97.3%, remained consistently stable 509 across tested concentrations. Conversely, the solid residue exhibited an escalating inhibition 510 with increasing concentrations (ranging from 68.11 to 97.2%), as depicted in Fig. 5. 511 Considering these compelling results, the observed differences in composition could indeed be 512 attributed to the distinct chemical structures present in each by-product. In this context, the 513 chemical profile obtained through HPLC-DAD analysis revealed a delicate equilibrium 514 between flavonoids (46.34%) and phenolic acids (37.39%) in the solid residue. On the contrary, 515 in the liquid residue, a striking predominance of phenolic acids (50.9% hydroxycinnamic acids 516 and 32.97% hydroxybenzoic acids) over flavonoids (13.28%) was observed, leading to intriguing possibilities. 517

518

519 Comparing the two types of phenolic acids and flavonoids, Bhutto et al. (Bhutto et al., 2018) 520 explored the correlation between the antioxidative potential of phenolic compounds and the 521 plasmon characteristics of silver nanoparticles. The study revealed that flavonoids exhibit a 522 strong optical response facilitated by the developed AgNPs, contrasting with phenolic acids. 523 Within phenolic acids, hydroxycinnamic acid generally outperformed hydroxybenzoic acids, 524 as assessed through both UV-vis bands and the corresponding molar absorptivity for these two 525 types of phenolic acids. The same trend was found in the study performed by Scroccarello et 526 al. (Scroccarello et al., 2021), where it was reported that AgNPs formed with the relative caffeic 527 acid were significantly higher than gallic acid as one of the most hydroxybenzoic acids used. 528 The nucleophilicity of the corresponding structures that react in an alkaline medium was likely 529 responsible for the difference in the optical responses of AgNPs prepared by hydroxybenzoic 530 and hydroxycinamic acids. The nucleophilicity of phenolics is affected by the substitution of 531 electron-donating (-OH and OCH₃) and electron-withdrawing (-COOH and -CH=CH-COOH) 532 groups, and this has an impact on how AgNPs respond optically.

533

534 Based on these compelling data, the highest activity of WR@AgNPs can be attributed to the remarkable influence of phenolic acids, particularly rosmarinic and caffeic acids. Their 535 536 exceptional ability to swiftly and effectively neutralize free radicals significantly enhances the 537 antioxidant properties of the nanoparticles, surpassing the collective effect of combining 538 flavonoids and phenolic acids. These rapid and potent free radical scavenging capabilities play 539 a pivotal role in bolstering the antioxidant efficacy of WR@AgNPs. The pivotal role of 540 phenolic acids, particularly rosmarinic and caffeic acids, in driving the nanoparticles' robust 541 antioxidant prowess is paramount in the battle against oxidative stress (Mohamad Sukri et al., 2023). The intricate interplay between these phenolic compounds and the nanoparticle matrix 542 543 results in potent antioxidant effects, as evidenced by the remarkable 95% scavenging potential 544 against free radicals showcased in the study of Harsha Haridas et al. (Harsha Haridas et al., 545 2023). Caffeic acid's renowned antioxidant properties are central to neutralizing reactive 546 oxygen species and preventing oxidative damage within biological systems, facilitated by its 547 controlled release encapsulated within the nanoparticles. Moreover, in model physiological media, rosmarinic acid demonstrated robust radical scavenging activity with overall rate 548 constant values of 2.89×10^{10} and 3.86×10^9 M⁻¹ s⁻¹ in water and pentyl ethanoate solvents, 549 respectively (Vo et al., 2023). Furthermore, in an aqueous environment, rosmarinic acid 550 exhibited an overall rate constant of 3.18×10^8 M⁻¹ s⁻¹ for scavenging HOO', a value 551 552 approximately 2446 times greater than Trolox, a common antioxidant compound (Vo et al., 553 2023).

554

555 These results highlight the potent antioxidant efficacy of rosmarinic acid, underscoring its 556 pivotal role in enhancing the antioxidant prowess of nanoparticles. The collaborative action 557 between rosmarinic and caffeic acids and the nanoparticles creates a synergistic effect,

558 amplifying their combined antioxidant capabilities and underscoring their potential as effective 559 therapeutic agents for combating oxidative damage and promoting cellular health. Through the 560 donation of hydrogen atoms or electrons and the chelation of transition metal ions, these 561 nanoparticles derived from rosmarinic and caffeic acids exhibit exceptional antioxidant prowess crucial in mitigating oxidative stress and fostering overall cellular well-being 562 563 (Bouannali et al., 2023). This dominance of phenolic acids highlights their pivotal role in 564 driving the nanoparticles' robust antioxidant prowess, making them key players in the battle against oxidative stress (Vieira et al., 2022). Interestingly, the synergy between flavonoids and 565 566 phenolic acids does not consistently result in a proportional increase in antioxidant activity 567 (Barbieri et al., 2020). In some instances, certain flavonoids may even interfere with the 568 antioxidant mechanisms of phenolic acids, leading to a potentially less effective overall 569 antioxidant response.

570 **3.5.** Nanoparticles effect on microbial growth

571 Increased mortality, morbidity, and treatment costs in developing nations are thought to be 572 primarily caused by MDR bacterial strains and the infections they cause. Gram-positive, Gram-573 negative bacteria, moulds and yeast pathogens have all been linked to serious clinical and 574 medical problems (Dakal et al., 2016). Thus, the wide-ranging and potent volatile compounds 575 present in rosemary essential oil allow it to effectively combat a broad spectrum of 576 microorganisms, making it a valuable and promising natural substitute for phenolic compounds 577 in combating infections and microbial growth. Additionally, nanoparticles can enhance the 578 antimicrobial efficacy of polar phenolic compounds, resulting in comparable or even superior 579 effects to those of essential oils. In this proposal, the agar diffusion test was used to assess the 580 antibacterial activity of the synthesized nanoparticles against two susceptible bacterial strains, 581 E. coli and S. aureus, as well as the yeast Rhodotorula glutinis and the mould Geotrichum sp. 582 Table 2 shows the information regarding the antimicrobial potential. Regarding the two by-

products of *R. Tourneforti* de Noé obtained through steam distillation and used as controls, the results revealed inhibition zones of 9.2 and 9 mm for *E. coli*, and 8.7 and 8.8 mm for *S. aureus*, in the solid and water residues, respectively. For fungi, the inhibition zones were measured at 10 and 9.8 mm for *Geotrichum sp.*, and 14.3 and 15.2 mm for *Rhodotorula glutinis*, in the solid and water residues, respectively.

588

589 As indicated in Table 2, the unsuccessful biosynthesis SR/WR@ZnNPs and SR/WR@CuNPs 590 had no effect for which could be we can highly observed that the antimicrobial activity did not 591 change significantly, for which the diameter measurements of the inhibition bacteria or fungi 592 varied weakly indicating that the phenolic compounds detected for the two by-products of R. 593 Tourneforti de Noé may not provide sufficient stabilization for the NPs formed, leading to rapid 594 agglomeration or degradation of the NPs. Adequate stabilization is essential to prevent NPs from losing their antimicrobial efficacy. In contrast, the successfully biosynthesized 595 596 nanoparticles, SR/WR@AgNPs, demonstrated a notable increase in the inhibition zone against 597 Gram-negative E. coli (12 and 13.2 mm for water and solid residues, respectively) and Gram-598 positive S. aureus (13.8 and 14 mm for water and solid residues, respectively). Similarly, the 599 antimicrobial activity against yeast *Rhodotorula glutinis* was significantly enhanced (16.8 and 600 17.4 mm for solid and water residues). However, no significant change in antimicrobial activity 601 was observed against the mould Geotrichum sp. compared to the two by-products without 602 AgNPs. Furthermore, it was observed that the two by-products, whether incorporated with Ag 603 or not, exhibited higher activity against Gram-positive bacteria compared to Gram-negative 604 bacteria. Comparing the two nanoparticles, SR@AgNPs appear to be more active than 605 WR@AgNPs against the two Gram bacteria, while the opposite is true for the yeast 606 Rhodotorula glutinis.

607

608 Upon analyzing the results from each residue matrix, a compelling equilibrium between 609 flavonoids and phenolic acids emerged in the solid residue of R. Tourneforti de Noé. 610 Intriguingly, over 80% of phenolic acids were concentrated in the water residue, unveiling a 611 distinct distribution pattern between these two compound types within the plant. This 612 observation hints at a potential synergistic effect, where the combined action of these 613 compounds may yield a more potent antibacterial effect compared to using phenolic acids in 614 isolation. Moreover, both flavonoids and phenolic acids displayed their prowess as reducing 615 agents in the synthesis process of AgNPs, contributing to the formation of stable nanoparticles 616 with controlled size and shape (Dakal et al., 2016). This assertion finds confirmation in the 617 alluring allure of SR@AgNPs, boasting nanoparticles of uniform diameter at a mesmerizing 618 92.3 nm. On the other hand, the synthesis of WR@AgNPs graced us with a revelation of 619 extraordinary proportions of bimodal size distribution. These findings shed light on the 620 coexistence of two distinct particle populations, each proudly showcasing its unique size at 621 62.2 nm and 111.64 nm. The acquired findings align harmoniously with Lombardo et al.'s 622 seminal work (Lombardo et al., 2016), underscoring that the synthesis of silver nanocomposites 623 with a nonuniform size and aggregation yielded the lowest antibacterial activity against E. coli 624 and S. aureus compared to the monodisperse counterpart. Further insight from Dal Lago et al. (Dal Lago et al., 2011) unveiled a compelling revelation smaller silver particles exhibited 625 626 heightened bactericidal prowess compared to their larger counterparts, with distinct responses 627 witnessed across diverse bacterial strains. From this wealth of data, a compelling conclusion 628 emerges: the impact on bacterial cells extends beyond the chemical profile that can potentially 629 modify AgNPs' surface properties. The uniformity of size and diameters of AgNPs appears to 630 play a pivotal role in affecting bacterial response. However, a unique perspective emerges when considering the yeast Rhodotorula glutinis; it seems to be uniquely influenced solely by the 631 632 reduction agents present.

633 **3.6. DFT optimization**

634 In the context of the study, the optimized structures of the targeted molecules extracted from 635 the water and solid residues of *Rosmarinus tournefortii* de Noé, along with their interactions with AgNPs, are presented in **Fig. 6**. In every instance, the confirmation of reaching the lowest 636 energy state was validated by the absence of any imaginary frequency. Initially, the calculation 637 638 of molecular electrostatic potential (MEP) was conducted to predict the locations susceptible 639 to electrophilic and nucleophilic reactions within the molecules under investigation. The distribution of charge in the carbohydrate molecules is predominantly influenced by the oxygen 640 641 atoms due to their lone pairs. Notably, the carboxylic group within homoplantaginin exhibited 642 the most electronegative potential, implying a strong affinity for attracting electron-deficient 643 species. Oxygen atoms, being highly active have the ability to donate electron density to silver atoms. This suggests the plausibility of transferring an electron to a silver atom, thereby 644 transforming an Ag^+ cation into Ag^0 and completing the $5s^2$ and $4d^9$ orbitals. To substantiate 645 this concept, various Ag₁-complexes were constructed by altering the positioning of silver 646 647 relative to the oxygen atom within the examined molecules. The silver clusters self-arranged 648 to maximize interactions with the molecules (He and Zeng, 2010). Based on resulting energy 649 evaluations, the most stable complexes emerged when the oxygen atom was positioned near 650 the carboxylic group. Consequently, this same configuration was adopted for subsequent 651 investigations.

652

Referring to He and Zeng's findings, the strategic placement of the silver atom and clusters was established in close proximity to the carboxyl group, a pivotal center within the carbohydrate molecules. This configuration resulted in the complexes assuming the lowest energy state. Previous investigations into reactivity patterns indicated that the primary interaction between bioactive compounds and the silver ion predominantly occurred through the carboxylic acid or

658 hydroxyl groups (Al-Otaibi et al., 2023). For this interaction to occur, both atoms (Ag and O) 659 must be in contact, underscoring the significance of the distance between the silver cluster 660 atoms and the carbohydrate molecule in comprehending these systems (Gallegos et al., 2022). 661 Within complex formations, distances wield considerable influence and provide insights into the nature of interactions. Among the one-silver-atom complexes, the interaction energies were 662 notably elevated, ranging from 158 to 269 kcal/mol. However, these complexes exhibited 663 664 notably elongated Ag–O distances, indicating a relatively weaker interaction due to the limited number of silver atoms involved. Conversely, the Ag₃-complexes exhibited slightly lower 665 666 energies (ranging from 130 to 239 kcal/mol) yet showcased shorter Ag-O distances. This indicates that complexes formed with three-silver-atom clusters are thermodynamically 667 preferred over those involving a single silver atom. As the complexes increase in the number 668 669 of silver atoms, the distances tend to approach an average length, consequently resulting in a 670 decrease in interaction energies. The disparities in energy values and Ag–O distances are 671 primarily governed by the specific type of carbohydrate utilized, ultimately driving improved 672 interactions and the formation of more stabilized complexes.

673

674 Based on our investigation, a silver nanoparticle composed of 3 silver atoms is anticipated to 675 possess superior attributes owing to its enhanced stability in contrast to smaller nanoparticles. 676 The most stable configuration of the three-silver-atom cluster in contact with the bioactive molecule exhibits uniform bond lengths measuring 2.73 Å and is characterized by a 58.7° 677 678 angle. Fig. 7 illustrates the optimized energies of individual molecules, isolated silver atoms, 679 and the resultant complexes. Notably, the silver cluster specifically Ag₃ interacting with the 680 biological molecule exhibited heightened stability in comparison to either the standalone silver 681 nanoparticle or the isolated biological molecule, as indicated by the lower energy levels. 682 Comparing the target molecules, the complexes formed between Ag₃ and homoplantaginin,

683 protocatechuic acid-glycoside, and rosmarinic acid exhibit the most substantial interaction 684 energies, measuring 357.05, 255.33, and 266.03 kcal/mol, respectively. This indicates a 685 favourable binding between the three-silver-atom clusters and these specific molecules, 686 underpinned by the presence of electrostatic interactions (Kusumaningsih et al., 2023). Significantly, these molecules also manifest the shortest Ag-O distances among all 687 688 compounds, averaging around 2.2-2.3 Å. Furthermore, a low bond distance of approximately 689 2.2 Å between the silver atom and the carbonyl functional group of caffeic acid was 690 demonstrated. Notably, the Ag-O distances for all four compounds (caffeic acid, 691 homoplantaginin, protocatechuic acid-glycoside, and rosmarinic acid) closely align with a typical oxygen-silver bond distance of approximately 2.20-2.35 Å (Njogu et al., 2017). 692 693 Consequently, these molecules are anticipated to exhibit enhanced reducing activity and offer 694 a high potential for promoting the stability of AgNPs.

695

Challenges and future outlook 3.7.

696 An emerging beacon of hope in biomedical applications lies in the realm of green nanoparticles, 697 representing a promising alternative to conventional approaches. These nanoparticles, derived 698 from natural sources or synthesized through eco-friendly methods, offer a sustainable avenue 699 for advancing healthcare solutions. Embracing the principles of environmental consciousness, 700 green nanoparticles present a harmonious fusion of efficacy and ecological responsibility 701 (Shreyash et al., 2021). As they make their debut on the biomedical stage, their potential shines 702 through in various domains, from drug delivery to diagnostics, tissue engineering, and beyond 703 (Sher et al., 2024). These eco-conscious nanoparticles not only demonstrate biomedical 704 prowess but also carry the potential to alleviate concerns regarding the environmental impact 705 of traditional nanoparticle synthesis (Selmani et al., 2022). By embracing the synergy between 706 innovation and sustainability, green nanoparticles beckon a transformative era in biomedical 707 research and applications, where medical progress goes hand in hand with ecological

708 stewardship. In the context of the study, the dynamic attributes inherent to AgNPs embedded 709 within both the solid and water residues derived from steam distillation of rosemary have been 710 successfully demonstrated. However, an intriguing challenge arises concerning the synthesis 711 of ZnNPs and CuNPs. Despite exhaustive analysis, including an assessment of their biological 712 activities, successful synthesis has proven elusive. This phenomenon could potentially be ascribed to the complex interactions between the phenolic compounds and the stabilization of 713 714 CuNPs and ZnNPs. While the developed nanoparticles exhibit remarkable efficiency, there are 715 pertinent issues that demand future attention. In the subsequent sections, delving into these 716 challenges and offering insightful perspectives for further exploration.

717 **3.7.1.** Alternative complexation agents for surface modifications

718 The biosynthesis of metallic nanoparticles is a multifaceted process influenced by various 719 factors such as light exposure, plant extract composition, enzymes, metal ion concentration, 720 and reaction conditions, collectively shaping the mechanism, synthesis rate, and final particle 721 morphology (El-Seedi et al., 2019). This collaborative amalgamation of influences defines the 722 bio-synthesis journey, offering the potential for process refinement and the customization of 723 nanoparticles for specific applications. Within this realm, a harmonious interplay exists 724 between chemical-reducing agents and the latent potential of medicinal plants. Polysaccharides 725 like β -D-glucose and starch serve as eco-conscious reducing agents, demonstrating 726 biocompatibility and water solubility, thus eliminating the need for hazardous solvents (Damiri 727 et al., 2023). Various plant components, including stems, flowers, leaves, and seeds, showcase 728 inherent reducing abilities (Alshameri and Owais, 2022). In scenarios where medicinal plants 729 alone may not suffice for metal ion reduction, chemical-reducing agents such as NaBH4 emerge 730 as viable alternatives due to their economic viability, reproducibility, consistent particle size 731 distribution, and straightforward experimental protocols. This strategic convergence of

medicinal plants' resources with the precision of chemical agents forms a synergistic approach,

harnessing both their inherent capabilities and the reliability of chemical-reducing agents.

734

735 Shervani et al. (Shervani and Yamamoto, 2011) conducted a comprehensive exploration into the synthesis of gold (Au) nanoparticles, delving into the influence of various reducing agents, 736 encompassing both traditional chemical methods and environmentally friendly alternatives. 737 738 The researchers skillfully combined agents such as NaBH₄ and β -D-glucose, thereby refining 739 the craftsmanship of AuNPs. This holistic approach unveiled an intriguing synergy among 740 these components, orchestrating the transformation of Au salts into monodispersed AuNPs 741 characterized by a distinctive and enchanting wine-red hue a hallmark of their successful 742 formation (Shervani and Yamamoto, 2011). In their methodology, soluble starch and β-D-743 glucose played crucial roles as carbohydrates in the synthesis process. The study highlighted 744 the successful production of monodispersed Ag(0) nanoparticles, with a diameter of 15 nm, 745 achieved by reducing AgNO₃ precursor salt in a starch-water gel with β-D-glucose (Shervani 746 and Yamamoto, 2011). The investigation into Au(0) metallic nanoparticles revealed the nuanced impact of the reducing agent type, quantity, and solution pH on the size and 747 748 morphology of the nanoparticles. Notably, NaBH₄ at 4 equivalents produced the smallest 749 metallic particles (5.3 nm). However, an excess of NaBH₄ led to the nanoparticles settling out 750 as a precipitate, forming a mesh or wire structure (Shervani and Yamamoto, 2011). This 751 meticulous exploration of synthesis conditions and their effects underscores the precision 752 required in the production of metallic nanoparticles with specific size and morphology 753 characteristics.

754

Another noteworthy endeavour was undertaken by Bikdeloo et al. (Bikdeloo et al., 2021),
employing two reduction agents, NaBH₄ and green rosemary extract, to successfully synthesize

copper nanoparticles with a hydrodynamic diameter within the range of 50 nm. The reduction of Cu salt by NaBH₄ unfolded as a direct and uncomplicated process. Upon introducing NaBH₄ into a dispersion of rosemary extract containing the Cu precursor salt, electrons and hydrogen converged, setting the stage for the reduction of Cu salt, ultimately transforming it into lustrous metallic Cu (Bikdeloo et al., 2021). Throughout this intricate orchestration, rosemary extract played a pivotal dual role, functioning as both a reducing and stabilizing agent, underscoring the intricate harmony within this endeavour.

764

765 Delving deeper into innovation, the realm of mental association opens an alternative pathway 766 for achieving advanced nanocomposite stabilization while harnessing the reduction capabilities 767 of medicinal plants (Kunwar et al., 2023). The combination of metals (Cu, Zn, and Ag) 768 introduces unique reduction reagents, creating nanocomposites with enhanced stability. This 769 dynamic approach reinforces structural integrity by seamlessly incorporating stabilizing 770 agents, counteracting agglomeration, and facilitating robust bonds between functional groups 771 and metal ions. The intricate interplay of surface interactions extends to surface modifications, 772 empowering nanocomposites with adaptability, enhanced dispersibility, amplified interactions, 773 and heightened biocompatibility. This strategic evolution equips nanocomposites to bridge 774 disciplinary gaps, aligning with dynamic demands (Zhang et al., 2023).

3.7.2. Multifaceted mechanisms of AgNPs toxicity and nanoparticle interactions in inflammation

Nano-silver toxicity manifests through various mechanisms at different levels, encompassing organ, cellular, and subcellular dimensions. At the organ level, nano-silver can enter the body through various exposure pathways, spreading to vital organs such as the heart, liver, kidney, brain, testes, and ovaries, thereby potentially triggering organ-specific pathophysiological effects (Zhang et al., 2022). On the cellular plane, nano silver engages with membrane proteins,

782 triggers signaling pathways, and disrupts cellular metabolism. Additionally, it generates 783 reactive oxygen species (ROS), inflicts DNA damage, and upregulates autophagy, culminating 784 in cell apoptosis (Attarilar et al., 2020). The cytotoxicity of nano silver is intricately linked to 785 factors such as particle size, concentration, exposure duration, and the presence of stabilizers. 786 Delving into the subcellular intricacies, nano silver exerts influences on lysosomal activity, 787 inhibits the expression of transcription factor EB (TFEB), and disrupts the normal functioning 788 of lysosomes. Furthermore, it interferes with ion channels on the cell membrane, creating an 789 imbalance in cell membrane potential and leading to cell necrosis (Mehnath et al., 2021).

790

Shifting focus to inflammation, an immediate response to internal injury, infection, or external 791 792 factors involves a nuanced interplay of immune cells and signals. Dysregulation in these signals 793 triggers inflammation, prompting the recruitment of macrophages, killer cells, and stem cells 794 (Fagiani et al., 2022). Macrophages, pivotal in regulating inflammation, manifest in two 795 phenotypes: pro-inflammatory M1 and anti-inflammatory M2. During inflammation, 796 macrophages engulf cellular debris and foster inflammation through the production of 797 activation signals like extracellular matrix proteins, lipopolysaccharide (LPS), and cytokines. 798 Neutrophils, in response to inflammation, migrate to the site, produce pro-inflammatory 799 mediators, and attract macrophages (Niu et al., 2021). Upon entering the body, metal 800 nanoparticles (NPs) encounter blood plasma proteins, resulting in the formation of a protein 801 corona around the NP (Fig. 8). This corona, comprising proteins like immunoglobulin G (IgG), 802 immunoglobulin M (IgM), and fibrinogen, crucial in the natural inflammatory process, is 803 intricately shaped by NP properties (Bashiri et al., 2023). Serum proteins, displaying a strong 804 attraction, play a significant role in forming the protein corona, which determines the external 805 appearance of the NP and gives it a biological identity. This identity then governs the NP's 806 movement and its interaction with various chemical reactions.

807

NPs gain entry into cells through pores or ion channels in the cell membrane, with uptake 808 809 influenced by their size. Adhesive interactions, driven by electrostatic, Van der Waals, or steric 810 forces, orchestrate the cellular uptake (Agarwal et al., 2019). The interaction between protein-811 coated metal NPs and macrophages or neutrophils at inflammatory sites is facilitated by the 812 protein corona, primarily comprised of serum proteins (Cai and Chen, 2019). This protein 813 corona acts as a ligand for receptors on anti-inflammatory M2 macrophages, triggering their 814 activation. This activation amplifies NP uptake, with M2 macrophages exhibiting heightened 815 and swifter NP uptake compared to M1 macrophages in the presence of serum proteins. Neutrophils, responding to stimuli, form extracellular traps (NETs), ensnaring gold 816 817 nanoparticles within these NETs. This intricate mechanism underscores the pivotal role of NPs 818 in modulating inflammatory responses and their dynamic interactions with immune cells.

819 **3.7.3.** Harnessing rosmarinic acid nanoparticles for inflammatory conditions

Inflammatory bowel disease (IBD), a complex and recurrent condition with an unknown 820 821 aetiology, demands increased attention as a critical public health concern. Classified into two 822 major subtypes, Crohn's disease (CD) and ulcerative colitis (UC), UC specifically manifests 823 as a chronic inflammatory disorder of the rectum and colon (Kaplan and Windsor, 2021). 824 Addressing the intricate nature of IBD, Chung et al.'s ground-breaking study explores the 825 potential of PEGylated rosmarinic acid nanoparticles (RANPs) derived from rosemary extract 826 as a nanomedicine for IBD treatment (Chung et al., 2020). With a diameter of 63.5 ± 4.0 nm, 827 RANPs exhibit superior therapeutic efficacy, synergizing with conventional medication, 828 scavenging reactive oxygen species (ROS), and protecting against oxidative damage. In a 829 mouse model of acute colitis, RANPs treatment at doses of 20 mg/kg and 30 mg/kg significantly reduces body weight loss, bloody stools, and disease activity index (DAI) scores, 830 831 indicating a substantial decrease in inflammation and damage to the colon. Histological

832 analysis further reveals the restoration of the colon lining structure and attenuation of 833 inflammation (Chung et al., 2020). Additionally, RANPs dose-dependently attenuate colonic 834 muscle thickening, supporting their role in reducing inflammation and damage to the colon. 835 Mechanistically, RANPs inhibit the activation of pro-inflammatory transcription factors NF- κ B and STAT3, and attenuate neutrophil infiltration in the inflamed colon (Chung et al., 2020). 836 837 Loading the corticosteroid medication DEX into RANPs enhances therapeutic efficacy, 838 demonstrating greater reductions in DAI scores and colon shortening compared to bare RANPs. 839 Importantly, RANPs exhibit good biocompatibility both in vitro and in vivo, positioning them 840 as a promising therapeutic nanomedicine for various inflammatory diseases, including IBD. 841 This provides visual evidence of the therapeutic effects of experimental modalities on colitis-842 associated tissue damage and inflammation.

843

844 Relating to the broader context of inflammatory conditions, the study seamlessly aligns with investigations into rheumatoid arthritis (RA). Recognizing RA's enduring challenge to global 845 846 public health, Lu et al.'s research explores antioxidative nanomedicine using 847 ribonucleoproteins (RNPs) derived from a natural polyphenol-based compound (Lu et al., 848 2023). The study's schematic illustration delineates the fabrication process of RNPs (RosA 849 nanoparticles) and reveals their therapeutic mechanism against RA. Synthesized through self-850 assembling oxidative oligomerization of RosA, RNPs play a crucial role in alleviating 851 oxidative stress in RA joints, scavenging ROS, elevating the anti-inflammatory M2 subtype 852 through macrophage polarization, and augmenting the production of anti-inflammatory 853 cytokines (Lu et al., 2023). The RNPs exhibit the ability to inhibit synovitis, angiogenesis, 854 cartilage degradation, and bone erosion, as evidenced by reduced clinical scores, ankle-joint thickness, and inflammation in the RNPs-treated group compared to the RosA-treated group. 855

857 The study provides insights into the cellular uptake of RNPs, emphasizing their rapid 858 internalization within hours by cells (Lu et al., 2023). The fluorescence imaging analysis 859 showcases the presence of RNPs in both the nuclei and cytoplasm of RAW 264.7 cells, 860 indicating efficient cellular uptake. In vivo, biodistribution studies further underscore the excellent targeting ability of RNPs, with enhanced accumulation at inflammatory joints 861 observed in fluorescence imaging of the paws of rats with collagen-induced arthritis (CIA). 862 863 This seamless connection between studies reveals a common thread in addressing inflammatory conditions through nanomedicine, offering innovative and targeted therapeutic 864 865 approaches.

866 **4. Conclusions**

The utilization of by-products generated from Rosmarinus tournefortii de Noé steam 867 868 distillation as metal-reducing agents in the "green" synthesis of nanoparticles offers a 869 promising route for sustainable and environmentally friendly nanomaterial production. These 870 nanoparticles, whether sourced naturally or synthesized ecologically, exceed efficacy norms 871 while embracing environmental responsibility. The synthesized nanoparticles underwent comprehensive characterization using UV-vis, XRD, FTIR, SEM/EDX, and Zeta analysis. The 872 873 study successfully enhanced the synthesis of AgNPs using two distinct by-products, water and 874 solid residues. However, ZnNPs and CuNPs synthesis encountered limitations, as indicated by 875 spectroscopic characterization. Factors such as inadequate synthesis conditions, nanoparticle 876 oxidation, agglomeration during synthesis, and the absence of appropriate surface ligands or 877 stabilizing agents may have contributed to these challenges. The average crystallite size of AgNPs was found to be 17.98 and 18.49 nm for SR@Ag and WR@Ag composites, 878 respectively, with stable and negative Zeta potential values (ζ values: -22.8 and -17.2 mV), 879 880 suggesting nanoparticle stability. Both types of by-products yielded nanoparticles with 881 predominantly spherical morphology, featuring varying nanoscale diameters. Antioxidant

882 assessment favoured water residue, showing consistent scavenging inhibition (94.9–97.3%) 883 across concentrations, while antimicrobial activity against Gram-negative, Gram-positive 884 bacteria, and yeast *Rhodotorula glutinis* was notably enhanced. Furthermore, DFT analysis 885 unveiled significant interactions among homoplantaginin, rosmarinic acid, protocatechuic acid-glycoside, and caffeic acid, resulting in heightened reduction activity of AgNPs. These 886 887 interactions exhibited substantial energy values, measuring 357.05, 266.03, 255.33, and 130.62 888 kcal/mol, respectively. These findings collectively advance the understanding of "green" 889 nanoparticle synthesis, paving the way for further innovation and applications in diverse fields. 890 Challenges and opportunities persist, including the pursuit of alternative complexation agents 891 for surface modification. Shifting focus, the section explores the multifaceted toxicity 892 mechanisms of silver nanoparticles at organ, cellular, and subcellular levels, emphasizing their 893 intricate interactions with inflammation processes. Additionally, it introduces two promising 894 nanomedicine approaches one involving PEGylated rosmarinic acid nanoparticles, a compound 895 derived from rosemary extract, for treating inflammatory bowel disease, and another utilizing 896 ribonucleoprotein for addressing rheumatoid arthritis. These findings underscore the potential 897 of nanotechnology, particularly harnessing rosemary-derived compounds like rosmarinic acid, 898 in innovative and targeted therapeutic interventions for diverse inflammatory conditions.

899 **Declaration of interests**

900 The authors declare that they have no known competing financial interests or personal 901 relationships that could have appeared to influence the work reported in this paper.

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List of Tables

Table 1. Identification of phenolic compounds from Rosmarinus tournefortii de Noé responsible for metal ion reduction.

	Retention time k number (min)		Tentative identified compounds		Relative abundance (% of total peak area)	
Peak number						
	Water residue	Solid residue	Water residue	Solid residue	Solid residue	Water residue
1	2.13	3.41	Gallic acid	Gallic acid	9.65	0.92
2	2.66	3.92	Epicatechin	Epicatechin	17.22	7.82
3	3.03	4.39	Chlorogenic acid	Homoplantaginin	15.52	4.94
4	3.27	5.63	Protocatechuic acid- glycoside	Gallocatechin	12.24	32.05
5	3.66	5.80	Gallocatechin	Caffeic acid	31.02	4.25
6	3.99	7.84	N. I	Chlorogenic acid	2.56	2.79
7	4.43	7.95	Caffeic acid	Rosmarinic acid	3.81	23.51
8	6.08	8.80	Yunnaneic acid F	Apigenin	1.68	1.33
9	6.52	-	Rosmarinic acid	-	-	21.15
10	9.32	-	Apigenin	-	-	1.21

Table 2. Inhibition Zones of the biosynthesized nanoparticles using Rosmarinus tournefortii de Noé solid and water residues.

Samples		Inhibition zone diameter (mm)				
	E.coli	S. aureus	Geotrichum sp.	Rhodotorula glutinis		
WR	9±0.78	8.8±0.67	9.8±0.53	15.2±0.45		
WR@ZnNPs	8.9±0.35	8.7±0.95	10±0.33	16.1±0.23		
WR@CuNPs	9±0.11	9.5±0.22	9.6±0.22	13±0.25		
WR@AgNPs	12±0.49	13.8±0.25	10±0.63	16.8±0.56		
SR	9.2±0.13	8.7±0.46	10±0.33	14.3±0.13		
SR@ZnNPs	9.6±0.21	8.8±0.47	9±0.7	14.7±0.31		
SR@CuNPs	8.9±0.22	9±0.88	10.2±0.22	15.2±0.12		
SR@AgNPs	13.2±0.63	14±0.98	9.8±0.25	17.4 ± 1.02		
0						
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Fig. 2. (a, b) UV-visible patterns, (c, d) X-ray diffraction patterns and (e, f) FTIR profiles of
biosynthesized nanoparticles using *Rosmarinus tournefortii* de Noé solid and water residues.



Fig. 3. Hydrodynamic size, SEM images and corresponding EDS spectra of synthesized AgNPs
using *Rosmarinus tournefortii* de Noé; (a, c, e) solid and (b, d, f) water residues.





Fig. 4. HPLC-DAD chromatograms of (a) the solid and (b) liquid by-products of *Rosmarinus tournefortii* de Noé.





Fig. 5. Antioxidant performance of (a) the solid and (b) the water by-products and their coating

- 1276 with Ag, Cu and Zn.





Fig. 6. Optimized geometries with the molecular electrostatic potential maps of the selected compounds with and without the inclusion of silver clusters. The interaction energies of the target molecules with Ag₁ or Ag₃ are included.



M. Ag3@Gallocatechin

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1288 Fig. 7. Total energies of the selected compounds with and without the inclusion of silver

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Highlights

- Synergistic AgNPs synthesis using eco-friendly R. tournefortii de Noé by-products. •
- By-products yielded spherical nanoparticles with varied nanoscale diameters. •
- Enhanced antioxidant and antibacterial activities were demonstrated. •
- DFT highlighted biomolecules (130.62–357.05 kcal/mol) enhancing stability. •
- AgNPs toxicity mechanisms and interactions in inflammation elucidated. •

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Declaration of interests

 \boxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

□The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: