

1 **Dynamic changes of key metabolites in Longjing green tea during processing**
2 **revealed by widely targeted metabolomic profiling and sensory experiments**

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24 **Abstract:**

25 In this study, widely targeted metabolomics and chemometrics were utilized to
26 comprehensively analyse the formation of taste substances in Longjing green tea. A
27 total of 580 non-volatile metabolites were identified using ultra-performance liquid
28 chromatography-electrospray ionization-tandem mass spectrometry, and alterations in
29 three metabolic pathways were investigated. Notably, the fixation process reduced
30 phosphatidic acid levels, resulting in the formation of lyso-phosphatidylcholine and
31 lyso-phosphatidylethanolamine, as well as the release of esterified polyunsaturated
32 fatty acids. Baiye No.1 had high levels of L-glutamic acid and L-glutamate, while
33 Longjing 43 showed elevated levels of flavones. Correlation analysis and sensory
34 verification indicated that an appropriate concentration of L-aspartic acid increased
35 the stringency of the tea. These findings advance our understanding of Longjing green
36 tea quality improvement and cultivar development.

37 **Keywords:** Longjing tea; Widely targeted metabolomics; Processing; Taste; Cultivar

38 1. Introduction

39 Green tea, a non-fermented tea, is the most produced and consumed tea in China.
40 It is renowned for its health-enhancing properties, including antioxidant, anti-
41 inflammatory, and anticancer effects (Musial et al., 2020). Manufactured from the
42 fresh new shoots of tea plant (*Camellia sinensis*), the typical green tea process
43 involves picking, fixation, rolling, and drying. Sensory evaluations of green tea reveal
44 a variety of aromas attributes, including faint, floral, chestnut-like, and other
45 categories. The tea infusion exhibits a green and bright colour and the taste is usually
46 composed of bitterness, astringency, sweetness, and umami. Importantly, taste is a
47 key determinant of consumer preference and acceptance, making it a critical aspect of
48 the sensory characteristics of green tea. The quality of tea depends on the secondary
49 metabolites it contains. For instance, the astringency of tea is primarily due to the
50 presence of polyphenols, alkaloids, and catechins. Anthocyanins contribute
51 significantly to its bitter taste (Ye et al., 2022). Amino acids constitute approximately
52 70% of the umami taste intensity of green tea (Nakagawa, 1975), while soluble sugars
53 are the primary source of sweetness (Yue et al., 2017).

54 Taste compounds in tea leaves are influenced by the quality of fresh tea leaves
55 (varieties, growing conditions, and picking tenderness), processing technique, and
56 storage (Zeng et al., 2022). Firstly, the quality of tea is closely associated with the tea
57 cultivar (Wang et al., 2021). For instance, Longjing 43 is considered the most suitable
58 cultivar for producing Longjing tea (Dragon well tea), primarily due to its chlorophyll
59 *b* content that contributes to the formation of a brown-beige colour in dry tea (Wang

60 & Ruan, 2010). The fresh and mellow taste of Anji bai tea processed from Baiye No.1
61 is related to its high L-theanine content and low tea polyphenol levels (Zeng, Lin, Liu
62 & Liu, 2019). Secondly, tea processing technologies substantially impact metabolites
63 in the final tea product. Many bioactive compounds, which are present in the final tea
64 product and contribute to its quality or functional properties, are produced during the
65 tea manufacturing process (Liao, Zhou, & Zeng, 2020). In the fixation or roasting
66 process, amino acids can react with carbonyl compounds to form Strecker aldehydes
67 that contribute to the formation of the tea aroma (Rizzi, 2008). The fixation stage is
68 primarily associated with chlorophyll decomposition, phosphatidic acids reduction
69 and glycolipids degradation (Li et al., 2021). Finally, the storage year is one of the
70 quality evaluation criteria for tea, especially dark tea and white tea (Zhou., et al 2023).
71 However, green tea is usually recommended to be consumed without long-term
72 storage because its taste and aroma deteriorate quickly. Therefore, comprehensive
73 studies are necessary to explore the dynamic changes of green tea used fresh tea
74 leaves from different cultivars as raw materials during the entire manufacturing
75 process.

76 In recent years, there has been significant progress in the development and
77 refinement of precision instruments, enabling the application of accurate and powerful
78 techniques for food analysis. Metabolomics studies in the field of tea research have
79 utilized various approaches, including targeted, untargeted, and widely targeted
80 metabolomics. These methodologies have been employed in investigating different
81 aspects of tea, such as evaluating the grade of *Tieguanyin* tea (Zeng et al., 2023),

82 assessing the impact of processing on green tea (Shi et al., 2022), and discriminating
83 tea cultivars in oolong tea (Zeng et al., 2022). Targeted metabolomic investigations
84 have traditionally involved the identification and quantification of a predefined set of
85 analytes, enabling the monitoring of metabolic changes over time. This approach has
86 been extensively utilized for several decades in the field of metabolism research. In
87 contrast, untargeted metabolomics represents a comprehensive and unbiased
88 analytical approach that aims to detect metabolic disturbances without relying on a
89 predefined list of analytes. However, the identification of metabolites in untargeted
90 metabolomics can be complex and time-consuming (Hertzog et al., 2022). A recent
91 advancement in metabolomics is the widely targeted approach, which combines the
92 advantages of untargeted and targeted methods. Widely targeted metabolomics
93 integrates the generality of untargeted metabolomics with the accuracy of targeted
94 metabolomics, offering a valuable tool for comprehensive detection of metabolites
95 (Zhou et al., 2022). Therefore, in this study, we employed the widely targeted
96 metabolomics approach to analyse the metabolites.

97 The objective of this research is to investigate the influence of cultivars and
98 processing on the metabolite profile of Longjing green tea, utilizing ultra-performance
99 liquid chromatography-electrospray ionization-tandem mass spectrometry (UPLC-
100 ESI-MS/MS) and chemometrics. Specifically, three tea cultivars were selected, which
101 were the albino tea cultivar "Baiye No.1," the traditional Longjing cultivars "Longjing
102 43" and the variety "Quantizhong." Fresh tea leaves from these cultivars underwent
103 the same manufacturing process to become Longjing green tea samples. Our primary

104 goal is to elucidate the dynamic changes in metabolites that occur during processing
105 and establish a sensory evaluation based on these metabolite variations. The findings
106 of this study are expected to contribute to the theoretical understanding of Longjing
107 green tea quality improvements and the development of high-quality tea varieties.

108 **2. Materials and methods**

109 *2.1. Chemicals*

110 Pure water was from Hangzhou Wahaha Group Co., Ltd. (Hangzhou, China).
111 Acetonitrile (ACN, HPLC grade), methanol (MeOH, HPLC grade), and glacial acetic
112 acid (HPLC, $\geq 99.9\%$) were from Merck (Darmstadt, Germany). Formic acid (FA), L-
113 aspartic acid, epigallocatechin (EGC), dihydromyricetin, and L-leucine were from
114 Aladdin (Shanghai, China). Ethyl caprate (99%) and ethanol (99%) were from
115 Sinopharm Group Chemical Reagent Co., Ltd. (Shanghai, China).

116 *2.2. Tea samples*

117 In 2020, young shoots consisting of one bud and two leaves from *Camellia*
118 *sinensis* cultivars 'Longjing 43 (LJ)', 'Longjing Quntizhong (QTZ)', and 'Baiye No.1
119 (BY)' were collected from Pang'an County in Zhejiang Province, China. These leaves
120 were utilized in Longjing tea processing experiments. The tea processing procedure
121 involved sequential steps, starting with the picking of fresh leaves and followed by
122 natural withering at temperatures ranging from 18 to 24 °C for a duration of 8 hours.
123 Subsequently, fixation was carried out using a roller-hot air fixation machine at
124 temperatures between 180 and 195 °C for a period of 1 hour, with a rotational speed
125 of 20 rpm and a leaf load of 80-100g. After fixation, the roasting process was

126 conducted for 30 min in the same roller-hot air fixation machine with a temperature of
127 90 ° C. Finally, the leaves were dried at a temperature of 200 ° C for 1 min. Samples
128 were collected and preserved for analysis during the processing stages, including fresh
129 tea leaves (FTL), withering (Wi), fixation (Fix), roasting (R), and the final tea product
130 (T). The samples were carefully obtained and stored to ensure their integrity before
131 being subjected to further analysis.

132 2.3. *Sensory evaluation*

133 Each tea infusion was prepared by brewing 3.0 g of dried tea leaves with 150 mL
134 of boiling water for a duration of 4 min at room temperature (RT, 25 ± 2 ° C),
135 according to the standard method for Longjing green tea brewing outlined in Chinese
136 standard GB/T 23776-2018. The evaluation and scoring of the tea infusion were
137 conducted by a trained panel of experts from the Tea Research Institute, Chinese
138 Academy of Agricultural Sciences. The panellists, aged between 25 and 48 years,
139 possessed certifications for tea quality evaluation issued by the Tea Scientific Society
140 of China. Prior to the evaluation, all panellists signed a consent form and willingly
141 underwent comprehensive training to develop their ability to discern various taste
142 attributes, including bitterness, astringency, umami, and total score. Each member of
143 the panel assigned scores to the different taste attributes, using a 0-10 scale to indicate
144 the intensity. The scoring scale ranged from 0-2 (very weak/just perceptible) to 8-10
145 (very strong intensity). The mean values of the scores were calculated and reported
146 (Zeng et al., 2023).

147 2.4. *Sample preparation and extraction*

148 *2.4.1. Dry sample extraction*

149 Biological samples were subjected to vacuum freeze-drying using a Scientz-
150 100F lyophilizer. Subsequently, the dried samples were ground to a powder form
151 using a Retsch grinder (MM 400) operating at a frequency of 30 Hz for 1.5 min. For
152 further analysis, 50 mg of the powdered sample was weighed using an MS105DM
153 electronic balance. Subsequently, 1200 μ L of a 70% methanolic aqueous internal
154 standard extract, pre-cooled to -20° C, was added to the sample. The sample and
155 extract were vortexed once every 30 min for a duration of 30 seconds, repeating this
156 process six times. Following centrifugation at a rotation speed of 12000 rpm for 3
157 minutes, th

158 e supernatant was aspirated, and the sample was filtered using a microporous
159 membrane with a pore size of 0.22 μ m. The filtered sample was then stored in an
160 injection vial for subsequent UPLC-MS/MS analysis.

161 *2.4.2. UPLC Conditions*

162 The sample extracts were subjected to analysis using a UPLC-ESI-MS/MS
163 system (UPLC, ExionLC™ AD) coupled with tandem mass spectrometry. The
164 analytical conditions included an Agilent SB-C18 UPLC column (1.8 μ m particle
165 size, 2.1 mm \times 100 mm); the mobile phase consisted of solvent A, which was
166 composed of pure water with 0.1% formic acid, and solvent B, which was composed
167 of acetonitrile with 0.1% formic acid. Sample measurements were carried out using a
168 gradient program. Initially, the composition was 95% A and 5% B. Within 9 minutes,
169 a linear gradient to 5% A and 95% B was applied, and this composition was

170 maintained for 1 minute. Subsequently, within 1.1 minutes, the composition was
171 adjusted to 95% A and 5.0% B, and this composition was maintained for 2.9 minutes.
172 The flow velocity was set at 0.35 mL per minute, and the column oven temperature
173 was maintained at 40 ° C. The injection volume was 2 µL. The effluent from the
174 UPLC system was directed to an ESI-triple quadrupole-linear ion trap (QTRAP)-MS
175 for analysis.

176 *2.4.3. ESI-Q TRAP-MS/MS*

177 The electrospray ionization source operation parameters were set as follows: the
178 source temperature was maintained at 500 ° C and the ion spray voltage (IS) was set to
179 5500 V for positive ion mode and -4500 V for negative ion mode. The ion source
180 gases I (GSI) and II (GSII) and the curtain gas (CUR) were set at 50, 60, and 25 psi,
181 respectively. The collision-activated dissociation (CAD) was set to high. Quantitative
182 multiple reaction monitoring (MRM) scans were acquired using a triple quadrupole
183 mass spectrometer, with the collision gas (nitrogen) set to medium. The declustering
184 potential (DP) and collision energy (CE) for each MRM transition were optimized
185 through further DP and CE optimization. A specific set of MRM transitions was
186 monitored for each period based on the eluted metabolites within that period.

187 *2.4.4. Principles of metabolite qualitative and quantitative analysis*

188 In our study, we employed the self-built Metware Database (MWDB) for
189 substance qualification, utilizing secondary spectral information. During the analysis,
190 we implemented a filtering process to eliminate duplicate signals originating from
191 isotopes, as well as ions such as K⁺, Na⁺, NH₄⁺, and fragment ions that are inherent

192 components of larger molecular weight substances. For metabolite quantification, we
193 utilized the MRM mode of a triple quadrupole mass spectrometer. In this mode, the
194 quadrupole initially screened the precursor ions (parent ions) specific to the target
195 substance, thereby excluding ions corresponding to other molecular weight
196 compounds, and effectively minimizing interference. Subsequently, the precursor ions
197 were induced to undergo ionization within the collision chamber, resulting in
198 fragmentation into multiple ion fragments. These fragment ions were further filtered
199 through the triple quadrupole to select a characteristic fragment ion, thereby
200 eliminating non-target ion interferences. This approach significantly enhanced the
201 accuracy and reproducibility of quantification.

202 Upon acquisition of mass spectrometry data for metabolomic analysis from
203 diverse samples, peak area integration was performed for all chromatographic peaks
204 corresponding to the compounds of interest. Subsequently, integration correction was
205 applied to the mass spectrometry peaks of the same metabolite across different
206 samples. Metabolites exhibiting a matching score of 0.7 or higher for retention time
207 and spectral fragmentation ions in the database were selected and retained for further
208 analysis.

209 *2.5. Sensory verification experiment*

210 We prepared 450 mL of tea according to the method described in section 2.3 of
211 the study. We divided the tea into 10 equal parts, with each part containing 40 mL,
212 while reserving 50 mL as the control sample (CK). Design different concentration
213 gradients for the sensory verification experiment based on the desired concentrations

214 of L-aspartic acid, L-phenylalanine, EGC, dihydromyricetin, and L-leucine in the tea.

215 The additive amount for each compound should be as follows:

216 L-aspartic acid: 1.2mg and 2.8mg

217 L-phenylalanine: 8mg and 24mg

218 EGC: 8mg and 24mg

219 Dihydromyricetin: 200mg and 400mg

220 L-leucine: 12mg and 36mg

221 Add the appropriate amount of each compound, according to the designed
222 concentration gradients, to the respective 40 mL portions of tea. Ensure thorough
223 mixing to achieve homogeneity. Ask panellists to evaluate the corresponding taste
224 attributes of the tea samples. Each member will receive a sample with a specific
225 concentration of the compound(s) for evaluation. The taste attributes to be assessed
226 include bitterness, astringency, umami, and the overall flavour score. The CK should
227 serve as a reference for comparison. Collect the evaluation scores and feedback from
228 panellists for further analysis and interpretation.

229 *2.6. Data processing and statistical analysis*

230 Partial least squares discriminant analysis (PLS-DA) modelling was conducted
231 using SIMCA 13.0 software (Umetrics, Sweden). Heat map analysis was performed
232 using TBtools v2.003 software (China). Hierarchical cluster analysis and radar map
233 visualization were conducted using Origin 2023b software (USA). Pearson correlation
234 analysis between taste attributes and chemical compounds was performed using SPSS
235 software (version 20.0), and the resulting network diagram was generated using

236 Cytoscape software (version 3.9.1). K-means clustering analysis was performed using
237 R software (version 3.5.1).

238 **3. Results and discussion**

239 *3.1. Overall determination of non-volatile metabolites in three varying varieties*

240 A total of 2616 ion features were obtained after peak-picking and alignment in
241 the analysis. Subsequently, 580 non-volatile metabolites were screened out based on
242 their retention time and spectral fragmentation ions. These metabolites comprised
243 various classes, including 36 alkaloids, 64 amino acids and derivatives, 138
244 flavonoids, 29 lignans and coumarins, 78 lipids, 27 nucleotides and derivatives, 33
245 organic acids, 70 phenolic acids, 22 tannins, 19 terpenoids, and 64 others (Figure 1A).

246 To gain a better understanding of the changes in non-volatile metabolites during
247 the processing of Longjing tea, a hierarchical cluster was performed on three different
248 varieties (Figure 1B). The analyses revealed significant differences among the three
249 varieties, indicating distinct metabolic profiles. When comparing variations in
250 processing within the same variety, it was observed that samples taken before and
251 after fixation showed noticeable discrepancies. Additionally, by examining the
252 processing stages, four different change tendencies were identified for these
253 metabolites (Figure 1C). For instance, 184 metabolites exhibited an initial increase in
254 the withering stage followed by a subsequent decrease in their levels.

255 *3.2. Alteration of metabolites by fixation*

256 The hierarchical cluster analysis results demonstrated significant disparities in
257 non-volatile metabolites subsequent to fixation. By implementing chemometrics

258 techniques to isolate essential metabolites, a PLS-DA model was established, with
259 results depicted in [Figure 2A and 2B](#). The findings indicate improved isolation of tea
260 samples post-fixation, with variable importance in projection (VIP) values exceeding
261 2. Thirty metabolites were identified, as visualized in [Figure 2C](#), 12 of which
262 decreased post-fixation, while the rest increased. These key metabolites contained 10
263 lipid compounds, indicating that the lipids changed greatly during the process of
264 fixation.

265 Lipids contribute not only to energy provision, texture, and mouthfeel but also
266 significantly influence the odour and flavour formation of food ([Shahidi & Hossain,
267 2022](#)). Thermal treatment is a routine used to moderate lipid oxidation, thereby
268 enhancing the palatability of food ([Zhang et al., 2022](#)). For instance, non-volatile
269 lipids are converted to volatile metabolites such as aldehydes and alcohols, which
270 contribute to the green and fresh notes in green tea ([Matsui, Kurishita, Hisamitsu, &
271 Kajiwara, 2000](#)). Further examination of lipid metabolite alterations following
272 fixation revealed that the majority of glycerol esters displayed a downward tendency,
273 while lyso-phosphatidylcholine (LPC) and lyso-phosphatidylethanolamine (LPE)
274 increased, as illustrated in [Figure 2D](#). The presence of LPC and LPE in peanuts and
275 green tea were also found to decrease post-thermal treatment ([Zhang et al., 2023;](#)
276 [Wang et al., 2021](#)) and a previous study showed that LPC levels (16:0, 18:1, 18:2, and
277 18:3) notably increased after the rolling and fermentation stages in black tea
278 processing ([Liu et al., 2023](#)). A previous study revealed that phosphatidic acids were
279 the most significantly reduced lipids during green tea manufacturing ([Li et al., 2021](#)).

280 Additionally, carbonyl compounds from lipid oxidative products can interact with
281 amino acids to form lipid-derivatived aroma compounds (Shahidi & Hossain, 2022).
282 Combined with the lipid metabolic pathway (Figure 2D), thermally-induced processes
283 potentially decreased the content of phosphatidic acids, forming LPC and LPE and
284 releasing the esterified form of polyunsaturated fatty acids.

285 3.3. Differences in sensory and metabolites among three varieties of final tea

286 Tea plants from different varieties contain varying biochemical compositions,
287 such as large-leaf varieties with a high level of polyphenols, and albino cultivars with
288 a high content of amino acids (Zhao et al., 2022). In this experiment, Longjing teas
289 made from three varieties were analysed and well discriminated, as shown in Figure
290 3A, B. Using a criterion of a VIP value greater than 2, key differential metabolites
291 were identified and visualized in a heat map (Figure 3C), with a total of 35
292 metabolites identified. The BY variety exhibited higher levels of epigallocatechin, 3-
293 methylellagic acid, and dihydromyricetin. It was found that the intensity of the sweet
294 aftertaste increased with the molar concentration of epigallocatechin (Zhang et al.,
295 2020). Ellagic acid is a polyphenol that results from the dimerization of gallic acid,
296 and it is known to contribute to the characteristic astringent taste due to its stable
297 cross-links with proteins (Bakkalbasi et al., 2009). Additionally, dihydromyricetin, the
298 main bioactive component in vine tea, has been extensively studied for its potential
299 health benefits (Carneiro et al., 2021).

300 In the LJ variety, isoorientin-7-O-glucoside, orientin-2''-O-galactoside, 3-
301 isopropylmalic acid, 2-propylmalic acid, epicatechin gallate, cryptochlorogenic acid

302 (4-*O*-caffeoylquinic acid), morin-3-arabinoside, and quercetin-3-*O*-arabinoside were
303 found to be more abundant compared to the other two varieties. The QTZ variety
304 exhibited higher levels of naringenin-7-*O*-glucoside (flavanones), kaempferol-3-
305 *O*(2''-galloyl) galactoside, and epicatechin-3-(3''-*O*-methyl) gallate. Previous studies
306 have reported that the flavonol glycoside profiles of dry teas were able to discriminate
307 tea plant cultivars, rather than the manufacturing procedure (Zhang et al., 2018). In
308 this study, it was also observed that the key differential compounds were
309 predominantly flavonol glycosides.

310 To quantify specific sensory factors attributes (i.e., bitterness, astringency, and
311 umami), an organoleptic test was conducted. As shown in Figure 3D, the BY variety
312 achieved the highest total and umami scores, while the LJ variety scored highest in
313 bitterness. On the other hand, the QTZ variety exhibited a pronounced astringency.
314 Combining these results with the metabolite analysis, it was inferred that the high
315 content of epigallocatechin in the BY variety might contribute to its high total and
316 umami scores. Additionally, previous studies have reported that the superior
317 performance of the BY variety was attributed to its high level of amino acids and low
318 levels of catechins and caffeine, which reduced astringency and bitterness while
319 enhancing the umami taste (Feng et al., 2014). This observation was further supported
320 by the dynamic changes in 12 amino acids (Figure 3E), where the content of L-
321 glutamic acid and L-glutamate were highest in the BY variety. These compounds are
322 known to be the primary contributors to the umami taste in green tea. Therefore, the
323 L-glutamic acid and L-glutamate content may be the reason for the intense umami

324 taste of the BY variety. Additionally, previous studies have identified caffeoyl- or
325 feruloyl-substituted quinides as contributors to the bitter taste in roasted coffee (Frank
326 et al., 2006). Hence, it is possible that cryptochlorogenic acid (4-*O*-caffeoylquinic
327 acid) contributes to the strong bitterness of the LJ variety. Similarly, compounds
328 found enriched in the QTZ variety, including naringenin-7-*O*-glucoside (a flavanone),
329 kaempferol-3-*O*-(2''-galloyl) galactoside, and epicatechin-3-(3''-*O*-methyl) gallate
330 (polyphenols), are known to present bitter flavours. However, these results were only
331 tentative and based on previous studies. To further demonstrate these findings,
332 correlation analyses between sensory results and these key metabolites, as well as
333 sensory verification experiment, were conducted.

334 3.4. Correlation analysis and the sensory verification experiment

335 The data analysis workflow, illustrated in Figure 4A, involved the utilization of a
336 PLS-DA model to identify 35 metabolites exhibiting differential expression (VIP >
337 2). Subsequently, these 35 metabolites were assessed for correlation with the sensory
338 evaluation results. The significance was set as the absolute value of the correlation
339 coefficients surpassing 0.8. The findings revealed significant correlations solely
340 between these metabolites and the sensory attributes of umami and astringency,
341 whereas no correlations were observed with bitterness. Astringency, a critical sensory
342 characteristic of green tea, is primarily influenced by hydrolysable and condensed
343 tannins (Granato et al., 2014). Among the 8 metabolites showing a strong correlation
344 with astringency (Figure 4B), compounds such as N-(sulfonyl) phenylalanine,
345 jaceosidin-7-*O*-galactoside, epicatechin-3-(3''-*O*-methyl) gallate, 4-hydroxy-3-

346 methoxyphenyl 1-*O*- β -D-(6'-*O*-galloyol)-glucopyranoside, and L-aspartic acid
347 displayed a significantly positive correlation with the astringency score. Conversely,
348 dihydromyricetin (ampelopsin), epigallocatechin, and L-phenylalanine exhibited a
349 notably negative correlation with the astringency score.

350 To further validate their actual taste contributions, epigallocatechin, L-
351 phenylalanine, L-aspartic acid, and dihydromyricetin were selected for additional
352 experiments (Figure 4C). The results indicated that L-aspartic acid and
353 dihydromyricetin augmented the astringency score, while L-phenylalanine reduced it.
354 Epigallocatechin displayed varying effects depending on its concentration. In green
355 tea, L-glutamic acid, L-theanine, and L-aspartic acid are considered as main
356 contributors to the umami flavour (Yu et al., 2014). Previous research demonstrated
357 that L-aspartic acid significantly inhibited the astringency of EGCG at low
358 concentrations (0.02 mg/mL), but significantly intensified the astringency at medium
359 to high concentration (0.17-1.39 mg/mL) (Liu et al., 2023). This finding suggests that
360 L-aspartic acid bidirectionally modulates astringency at different concentrations,
361 which possibly the reason for the positive correlation with astringency observed in
362 this study. Despite exhibiting a negative correlation in the correlation analysis,
363 dihydromyricetin was found to enhance the astringency score. On the other hand, the
364 bitterness of L-phenylalanine, which was proven to be a major contributor to
365 bitterness in bamboo shoots (Gao et al., 2019), reached an equivalent level to
366 berberine hydrochloride at a concentration of 0.5 mg/mL. However, the interaction
367 between L-phenylalanine and astringency remains unknown and requires further

368 investigation to better understand their relationship.

369 Shifting focus to the umami taste, although 8 metabolites exhibited strong
370 correlations with umami taste, L-leucine demonstrated a negative correlation (Figure
371 4D). To validate this finding, we conducted a sensory verification experiment (Figure
372 4E), and the results indicated that the umami score decreased upon the addition of L-
373 leucine. L-leucine itself exhibits a bitter taste, but previous research suggested that
374 changes in L-leucine concentration were difficult for the human body to perceive.
375 (Scharbert & Hofmann, 2005). Based on the results, L-aspartic acid, L-phenylalanine,
376 and L-leucine were identified as essential taste metabolites in Longjin tea.

377 Furthermore, an analysis of the changes in L-aspartic acid, L-phenylalanine, and
378 L-leucine during processing was conducted using a heat map (Figure 4F), revealing
379 distinct variations in the levels of these metabolites among the three cultivars.
380 Specifically, QTZ exhibited the highest content of L-aspartic acid and L-leucine,
381 whereas BY had the highest level of L-phenylalanine. Notably, all three amino acids
382 displayed an increasing trend across the three varieties during processing. The
383 increase in amino acids was attributed to the high temperature employed during
384 processing, which promoted protein hydrolysis. Similar observations have been
385 reported in other studies on the processing of green tea (Wang et al., 2021).

386 3.5. *Dynamic changes in the main taste metabolites of three different varieties of* 387 *Longjing tea during processing*

388 The primary taste constituents of tea are amino acids (umami), flavonoid

389 (bitterness and astringency), and alkaloids (bitterness) (Zhang et al., 2020). In order to
390 gain a more comprehensive understanding of the transformations of the non-volatile
391 compounds during Longjing tea processing, we focused on three salient metabolic
392 pathways: the flavonoid pathway, the amino acid pathway, and the alkaloid pathway.
393 The modifications in these metabolic pathways are discussed in detail below.

394 3.5.1 Modifications in the flavonoid metabolic pathway

395 The major components of the flavonoid metabolic pathway in tea include
396 flavones and flavone glycosides, flavonol glycosides, and flavanols, as depicted in
397 Figure 5. Flavones and flavonols are predominantly present as *O*-glycosides, with a
398 glycoside moiety attached to the C-3 position of the aglycones. These compounds
399 play a crucial role in contributing to the bitter taste of tea (Fang et al., 2019). During
400 processing, the concentrations of flavones (such as vitexin, apigenin, and isovitexin)
401 and most apigenin glycosides generally exhibited an upward trend, particularly after
402 the fixation process. However, exceptions were observed for apigenin-4'-*O*-glucoside,
403 apigenin-7-*O*-glucoside (cosmosiin), and isovitexin-7-*O*-glucoside (saponarin).
404 Although the abundance of flavones and flavone glycosides differed among the three
405 tea varieties, the overall tendency during processing was similar.

406 The concentration of quercetin significantly increased after the fixation process.
407 Quercetin is known to contribute to the green colour of tea infusions (Wang et al.,
408 2004), which may explain intensified tea infusion colour observed after fixation. The
409 detected flavonol glycosides were categorized as kaempferol glycosides, quercetin
410 glycosides, and myricetin glycosides. The abundance of quercetin glycosides

411 primarily displayed a notable decrease during processing, particularly after fixation,
412 whereas the trends for kaempferol glycosides and myricetin glycosides varied.

413 Flavanols, which are the most characteristic and abundant metabolites in tea, are
414 considered the primary contributors to the astringency and bitterness taste (Zhang et
415 al., 2020). In this study, different types of flavanols exhibited diverse changes.
416 Epigallocatechin gallate and catechin gallate showed an increase during processing.
417 Previous research has indicated that catechins undergo various transformations,
418 including isomerization, optical isomerization, hydrolysis, thermal polymerization,
419 and pyrolysis, during the processing of green tea (Huang et al., 2005).

420 *3.5.2 Modifications in the amino acid metabolic pathway*

421 Dynamic alterations in amino acids during processing are illustrated in Figure
422 6A. Throughout the tea manufacturing process, significant changes occurred in the
423 abundance of various amino acids. Initially, fresh tea leaves contained high
424 concentrations of L-theanine and L-glutamic acid, which gradually diminished during
425 processing. L-theanine, a unique amino acid found in different tea types, plays a
426 prominent role in contributing to the umami taste of tea (Zhang et al., 2020). It has
427 been suggested that the decrease in L-theanine content is attributed to the Maillard
428 reaction between theanine and glucose, resulting in the formation of Amadori
429 rearrangement products, methylpyrazine and 2,5-dimethylpyrazine, which are found
430 in various teas (Guo et al., 2018; Han et al., 2022).

431 Following withering, the concentrations of L-valine, L-aspartic acid, L-tyrosine,

432 and L-tryptophan significantly increased. The rise in proteinaceous amino acids is
433 attributed to protein degradation facilitated by the hydrolytic activity of endogenous
434 peptidases during withering, particularly in black tea (Chen et al., 2020). Notably, L-
435 arginine contributes to sweetness, while tryptophan and phenylalanine contribute to
436 the astringency and bitter taste of tea (Zhu et al., 2020). These changes in amino acid
437 composition likely represent differentially expressed metabolites responsible for the
438 distinctive taste profile of Longjing green tea.

439 The dynamic changes observed in L-valine, L-aspartic acid, L-theanine, L-
440 glutamic acid, L-tyrosine, and L-tryptophan were similar across the three tea varieties
441 studied. However, there were variations in the abundance of L-arginine during
442 processing. In the case of QTZ, L-arginine content appeared to keep increasing, while
443 BY and LJ exhibited a tendency to decrease followed by an increase. The variability
444 in L-arginine levels could be attributed to varietal specificity.

445 *3.5.3 Modifications in the alkaloid metabolic pathway.*

446 Caffeine and theobromine are the primary alkaloids found in tea and are
447 responsible for contributing to the characteristic bitter taste of tea infusions. As
448 depicted in Figure 6B, the processing of tea leaves had a significant impact on the
449 levels of caffeine and theobromine. Specifically, the content of theobromine
450 consistently decreased throughout the processing stages, while caffeine initially
451 increased, decreased after roasting, and then increased again. This observed
452 phenomenon can be attributed to the conversion of theobromine, an intermediate
453 compound in the biosynthesis of caffeine, into caffeine during the processing steps

454 (Xia et al., 2017). Furthermore, the fluctuation in caffeine content throughout the
455 processing stages can potentially be attributed to sublimation of caffeine caused by
456 exposure to high temperatures. It is worth noting that when comparing LJ and QTZ
457 with BY, the content of both theobromine and caffeine was found to be the lowest in
458 BY. These findings highlight the dynamic changes in caffeine and theobromine levels
459 during tea processing and demonstrate the influence of processing on the composition
460 and taste characteristics of tea.

461 **4. Conclusion**

462 This study combined widely targeted metabolomics and chemometrics to
463 investigate the effects of cultivars and processing on Longjing green tea's metabolite
464 profile. A total of 580 non-volatile metabolites were identified, highlighting
465 alterations in flavonoid, amino acid, and alkaloid metabolic pathways. The fixation
466 process potentially reduced phosphatidic acid levels, leading to the formation of LPC,
467 LPE, and the release of esterified polyunsaturated fatty acids. Distinct metabolites and
468 taste profiles were observed for the three cultivars, with L-glutamic acid and L-
469 glutamate predominant in BY, and accumulate flavones predominant in LJ.
470 Correlation analysis linked taste attributes with metabolites, revealing that a certain
471 concentration of L-aspartic acid increased tea astringency. Changing trends of key
472 metabolites during processing were similar among cultivars except for L-arginine in
473 QTZ. These findings advance our understanding of Longjing green tea quality
474 improvement and contribute to the development of high-quality tea varieties. Future
475 research will focus on volatile metabolite dynamics in Longjing green tea.

476 **Ethical statement**

477 All the participants (healthy and nonsmokers from TRICAAS) were conducted in
478 accordance with the principle set forth in the Declaration of Helsinki and informed
479 written consent was obtained. This study was approved by the Zhejiang Gongshang
480 University Human Ethics Committee.

481 **CRedit authorship contribution statement**

482 **Lin Zeng:** Writing – original draft, Methodology, Investigation, Data curation,
483 Formal analysis, Visualization. **Yan-Qing Fu:** Writing – review & editing, Resources,
484 Data curation, Methodology, Supervision. **Ying Gao:** Writing – review & editing,
485 Resources, Software, Visualization. **Fang Wang:** Writing – review & editing. **Jun-**
486 **Feng Yin:** Writing – review & editing. **Marie-Laure Fauconnier:** Writing – review
487 & editing. **Lijing Ke:** Investigation, Data curation. **Yong-Quan Xu:**
488 Conceptualization, Writing – review & editing, Data curation, Formal analysis,
489 Investigation, Project administration.

490 **Conflicts of Interest**

491 The authors declare no conflicts of interest regarding the publication of this paper.

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