

### III. Résultats

## *Chapitre 1 : Les ADAMTS2, 3 et 14 au cours du développement*

### 1. Introduction

Comme décrit en détail dans l'introduction générale de ce mémoire, les collagènes fibrillaires I, II, III et V sont sécrétés sous forme de procollagènes immatures. Ceux-ci sont formés d'une longue triple-hélice encadrée de part et d'autre de deux propeptides globulaires qui sont excisés par des enzymes spécifiques, la BMP1 pour le propeptide carboxyterminal et les aminoprocollagène peptidases (ADAMTS2, 3 et 14) pour le propeptide aminoterminal. L'inactivation de l'ADAMTS2, chez l'Homme (syndrome d'Ehlers-Danlos VIIC) et l'animal (dermatosparaxie), se marque par un syndrome sévère de fragilité cutanée sans affecter de façon significative les autres tissus conjonctifs riches en collagène de type I (tendon, os) ou de type II. Diverses études issues de notre laboratoire ont démontré une réduction drastique du clivage du propeptide aminoterminal du procollagène de type I au niveau du derme, alors que ce processus de maturation n'était que modérément altéré dans d'autres tissus et organes, ce qui était en parfaite adéquation avec les observations cliniques originales.

C'est dans ce cadre que nous avons démontré que l'ADAMTS3 est responsable du clivage de l'aminopropeptide du collagène de type II dans le cartilage de l'adulte et que l'ADAMTS14 possède une activité aminoprocollagène peptidase envers le procollagène de type I. Enfin, nous avons plus récemment démontré que les procollagènes de type III et V étaient également des substrats pour l'ADAMTS2.

Les ADAMTS2, 3 et 14 semblent donc constituer, dans la famille des ADAMTSs, un sous-groupe d'aminoprocollagène peptidases d'apparenté évolutionnaire. Nous avons participé, en collaboration avec le laboratoire de S. Apte (Lerner Research Institute, Cleveland Clinic Foundation) à l'analyse de l'expression spatiale et temporelle des ces trois

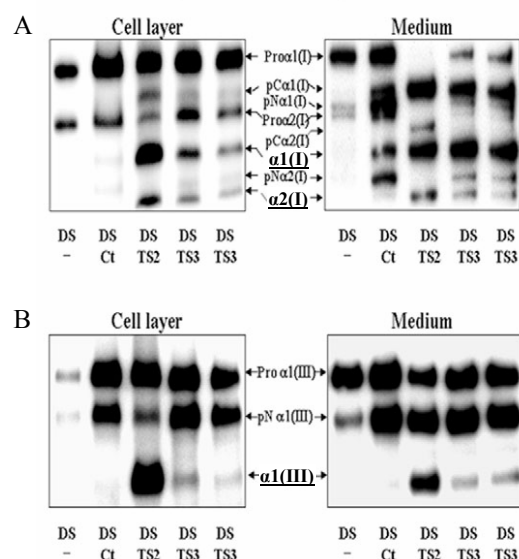
enzymes et de leurs substrats afin de préciser leur implication respective au cours de l'embryogenèse et du développement chez la souris.

## 2. Résumé des résultats

- La distribution des ARNm des collagènes fibrillaires, déjà connue, s'est vérifiée dans ce travail:
  - Collagène I: os, peau, périchondre, périoste, tendon, adventice des vaisseaux ;
  - Collagène II: cartilage ;
  - Collagène III: peau, paroi des vaisseaux, mésenchyme pulmonaire, péritoine, paroi des organes viscéraux.
  
- L'ARNm de l'ADAMTS3 est co-exprimé avec le transcrite de l' $\alpha 1(\text{II})$  dans tous les sites d'initiation et de développement de la chondrogenèse. Son expression est réduite dans les chondrocytes hypertrophiques. Il est également co-exprimé avec le transcrite de l' $\alpha 1(\text{I})$  durant l'ostéogenèse et le développement musculo-tendineux. Les ARNm de l'ADAMTS2 et de l'ADAMTS14 ne sont détectables dans ces diverses structures à aucun stade du développement. En accord avec ces données, nous avons démontré à la fois que les souris ADAMTS2<sup>-/-</sup> ne présentent pas d'anomalie macroscopiquement visible des cartilages, du squelette ou de la dentition, et que l'ADAMTS3 est capable de cliver l'aminoprocollagène de type I produit par les fibroblastes de derme dermatosparactique, déficients en ADAMTS2 (Fig. 22).

**Fig. 22.**

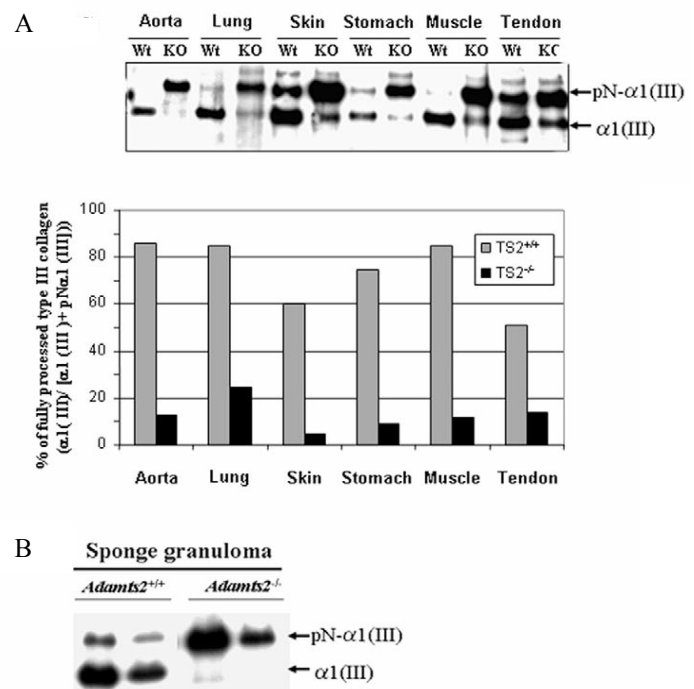
Les cellules HEK293 exprimant l'ADAMTS3 (TS3) ou l'ADAMTS2 (TS2) ou transfectées par le vecteur vide (Ct) sont co-cultivées avec des fibroblastes de derme de veau dermatosparactique (DS). Les polypeptides de collagène de type I (A) et de type III (B) sont détectés par western blotting dans la couche cellulaire et dans le milieu conditionné après 2 jours de culture. Le collagène produit par les fibroblastes DS seuls (DS/-) est détecté sous forme de précurseurs à différents stades de maturation (pro- $\alpha 1(\text{I})$ , pro- $\alpha 2(\text{I})$ , pro- $\alpha 1(\text{III})$ , pNa1(III), ...). A l'inverse, en présence d'ADAMTS2 et, dans une moindre mesure, d'ADAMTS3, on note l'apparition de chaînes  $\alpha 1(\text{I})$ ,  $\alpha 2(\text{I})$  et  $\alpha 1(\text{III})$  ayant subi une maturation complète.



- Chez l'embryon, l'ARNm de l'ADAMTS2 est co-exprimé avec le transcrite de  $\alpha 1(\text{III})$  à un niveau élevé dans le mésenchyme pulmonaire, ce qui se traduit chez les souris ADAMTS2<sup>-/-</sup> par une distension des alvéoles pulmonaires. Une co-expression de ces deux molécules est également observée dans la paroi des gros vaisseaux, la paroi intestinale, le péritoine, le mésentère et la paroi de la vessie. Les ADAMTS3 et 14 ne sont jamais co-exprimées de manière spécifique avec l' $\alpha 1(\text{III})$ . Dans la peau, les collagènes de type I et III sont tous deux exprimés. Seule l'ADAMTS2 est détectée dans le derme à 17,5 jours post-coït tandis que l'ADAMTS14 n'apparaît que 2 semaines après la naissance. Le rôle crucial de l'ADAMTS2 pour le clivage de l'aminopropeptide du collagène de type III est largement confirmé par des études biochimiques réalisées chez l'adulte montrant une réduction drastique de la maturation du collagène de type III dans divers organes des souris ADAMTS2<sup>-/-</sup> ainsi que dans le tissu fibrovasculaire développé au sein d'un granulome induit chez ces mêmes souris par implantation d'une éponge de PVA (Fig. 23).

**Fig. 23.**

Western-blotting illustrant la maturation de l'aminoprocollagène III (pNa1(III)) en collagène III ( $\alpha 1(\text{III})$ ). Celle-ci est largement déficitaire chez les souris ADAMTS2<sup>-/-</sup> (KO) par rapport aux souris sauvages (Wt) dans divers organes (A) ainsi que dans le tissu de granulation déposé au sein d'un granulome contre corps étranger (B).



- L'expression d'ADAMTS14 n'apparaît dans le derme que peu avant la naissance et perdure au cours de la vie adulte.

### 3. Conclusions

Une analyse récente de l'évolution des gènes des ADAMTSs montre que les clans à l'intérieur de cette famille se sont créés par duplication de leurs ancêtres primitifs. Les ADAMTS2, 3 et 14 sont hautement conservées et sont toutes trois douées d'activité aminoprocollagène peptidase dont les implications physiologiques respectives restaient à définir de manière plus précise.

L'évolution, au cours du développement embryonnaire, de l'expression de ces trois ADAMTSs et de certains de leurs substrats potentiels confirme que l'ADAMTS3 est bien l'enzyme principalement responsable du clivage du propeptide aminoterminal du procollagène de type II *in vivo*. Cette étude montre également que, de manière inattendue, l'ADAMTS3 est impliquée dans la maturation du collagène de type I au cours du développement musculo-squelettique. Ceci expliquerait que ces tissus ne sont que peu affectés chez les souris ADAMTS2<sup>-/-</sup> et dans la dermatosparaxie humaine et bovine. Par ailleurs, l'ADAMTS3 n'est pas exprimée dans le derme des souris et ne peut compenser l'absence d'ADAMTS2, ce qui serait à l'origine de l'atteinte préférentielle de ce tissu dans la dermatosparaxie. L'ADAMTS14 pourrait jouer ce rôle après la naissance mais son activité, plus réduite, ne permettrait pas l'élaboration de fibre normale dans le derme.

Ces études de co-localisation suggèrent également que, à l'inverse de la maturation du procollagène de type I qui peut être effectuée alternativement ou conjointement par les ADAMTS2 et 3, le clivage du propeptide aminoterminal du procollagène de type III est principalement le fait de l'ADAMTS2.

La réduction substantielle du clivage du pN-III chez les souris ADAMTS2<sup>-/-</sup> et dans la dermatosparaxie supporte cette fonction et contribue probablement à la fragilité cutanée caractéristique de cette pathologie.

Le déficit de maturation du procollagène de type III paraît avoir un impact plus dramatique chez la souris que chez l'Homme. Dans l'Ehlers-Danlos VIIC, les contusions et ecchymoses sont fréquentes et peut-être liées à la maturation réduite de l'aminoprocollagène de type III (pN-III) et/ou aux propriétés anti-angiogènes de l'ADAMTS2 (voir Chapitre 3). L'atteinte pulmonaire observée chez la souris n'a pas été rapportée dans la dermatosparaxie humaine. Le déficit presque complet du clivage de pN-III dans l'aorte chez la souris ne s'accompagne pas d'anomalies structurelles et fonctionnelles chez les souris jeunes mais pourrait se manifester avec l'âge ou suite à des atteintes diverses.

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# Regulation of procollagen amino-propeptide processing during mouse embryogenesis by specialization of homologous ADAMTS proteases: insights on collagen biosynthesis and dermatosparaxis

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Mutations in ADAMTS2, a procollagen amino-propeptidase, cause severe skin fragility, designated as dermatosparaxis in animals, and a subtype of the Ehlers-Danlos syndrome (dermatosparactic type or VIIC) in humans. Not all collagen-rich tissues are affected to the same degree, which suggests compensation by the ADAMTS2 homologs ADAMTS3 and ADAMTS14. In situ hybridization of *Adamts2*, *Adamts3* and *Adamts14*, and of the genes encoding the major fibrillar collagens, *Col1a1*, *Col2a1* and *Col3a1*, during mouse embryogenesis, demonstrated distinct tissue-specific, overlapping expression patterns of the protease and substrate genes. *Adamts3*, but not *Adamts2* or *Adamts14*, was co-expressed with *Col2a1* in cartilage throughout development, and with *Col1a1* in bone and musculotendinous tissues. ADAMTS3 induced procollagen I processing in dermatosparactic fibroblasts, suggesting a role in procollagen I processing during musculoskeletal development. *Adamts2*, but not *Adamts3* or *Adamts14*, was co-expressed with *Col3a1* in many tissues including the lungs and aorta, and *Adamts2*<sup>-/-</sup> mice showed widespread defects in procollagen III processing. *Adamts2*<sup>-/-</sup> mice had abnormal lungs, characterized by a decreased parenchymal density. However, the aorta and collagen fibrils in the aortic wall appeared normal. Although *Adamts14* lacked developmental tissue-specific expression, it was co-expressed with *Adamts2* in mature dermis, which possibly explains the presence of some processed skin procollagen in dermatosparaxis. The data show how evolutionarily related proteases with similar substrate preferences may have distinct biological roles owing to tissue-specific gene expression, and provide insights into collagen biosynthesis and the pathobiology of dermatosparaxis.

**KEY WORDS:** ADAMTS, Metalloprotease, Procollagen, Dermatosparaxis, Ehlers-Danlos syndrome

## INTRODUCTION

Collagens are triple-helical molecules that form the major structural elements of almost all tissues outside the nervous system. Collagen types I, II and III are evolutionarily related, and comprise the quantitatively major fibril-forming (fibrillar) collagens (Olsen, 1991). They form organotypic matrices by tissue-specific expression and by assembly of the collagen triple-helix into a variety of supramolecular aggregates (Birk et al., 1991). Collagen I is the major extracellular matrix (ECM) protein of bone, tendons, ligaments, and the dermis of the skin. Collagen II is the major collagen of cartilage, but it is also present in the vitreous of the eye and in the nucleus pulposus of the intervertebral disc (Zhu et al., 1999). Collagen III is a component of many stretchable tissues, such as the vascular wall, dermis, lung, heart and gastrointestinal tract (De Paepe and Malfait, 2004). It can co-polymerize with collagen I to form heterotypic fibrils and it is co-expressed in varying proportions with collagen I at many locations, with the exception of bone (Niederreither et al., 1995). Collagen I contains two  $\alpha 1(I)$  chains and one  $\alpha 2(I)$  chain, whereas collagens II and III are homotrimers of  $\alpha 1(II)$  and  $\alpha 1(III)$  chains, respectively (Olsen, 1991). Structural

mutations in collagen genes have serious consequences for tissues where single collagen types are major constituents. Collagen I mutations cause osteogenesis imperfecta (Rauch and Glorieux, 2004). Chondrodysplasias, sometimes with ocular anomalies, such as Stickler syndrome, are caused by collagen II mutations (Chan et al., 1995; Cole, 1997; Snead and Yates, 1999; Tiller et al., 1995), and collagen III mutations cause the vascular subtype of the Ehlers-Danlos syndrome (type IV), which is characterized by aortic aneurysms and skin fragility (De Paepe and Malfait, 2004).

Fibrillar collagens are made as a precursor (procollagen) and consist of a long triple-helical (collagenous) domain flanked at each end by globular non-collagenous propeptides (Olsen, 1991). The three major fibrillar procollagens share similar biosynthetic mechanisms. Following intracellular assembly of the triple helix, the procollagen molecules undergo proteolytic excision of each propeptide either prior to or following secretion (Canty et al., 2004; Olsen, 1991). The C-propeptide is excised by BMP1, an astacin-like protease (Kessler et al., 1996). ADAMTS2 (for A Disintegrin-like And Metalloprotease domain with Thrombospondin type 1 motifs 2) can excise the N-propeptide of all three major fibrillar collagens (Tuderman and Prockop, 1982; Wang et al., 2003). The processed collagen monomers assemble into fibrils and tissue-specific supramolecular aggregates. Because of a common biosynthetic pathway, it was expected that defects in ADAMTS2 would have broader effects than structural mutations involving individual procollagen chains. Instead, ADAMTS2 mutations lead to a limited clinical syndrome – dermatosparaxis (fragility of skin) in animals and the Ehlers-Danlos syndrome-type VIIC (EDS-VIIC or the dermatosparactic type) in humans (Colige et al., 2004; Colige et al.,

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1999; Lapiere and Nusgens, 1993; Nusgens et al., 1992; Petty et al., 1993; Smith et al., 1992; Wertelecki et al., 1992). Collagen fibrils in the dermis of dermatosparactic calves, humans and mice are thin, branched and 'hieroglyphic' on cross-section, instead of being round and of relatively uniform diameter (Li et al., 2001; Nusgens et al., 1992). Although this disorder is largely caused by anomalies in skin (dermal) collagen, there is only a modest effect on other collagen I-rich tissues, such as bone, tendon and muscle (Lapiere and Nusgens, 1993). Processing of procollagen II in the cartilage of dermatosparactic calves and mice is only minimally compromised, or not at all (Fernandes et al., 2001; Li et al., 2001).

It was hypothesized that enzymes closely related to ADAMTS2 may process procollagen I in tissues other than skin (Lapiere and Nusgens, 1993); this is supported by the observation that in bovine dermatosparaxis, the residual processed procollagen is cleaved at the expected site (Colige et al., 2002). We previously found that ADAMTS3, could process the amino-propeptide of procollagen II (Fernandes et al., 2001). Subsequently, it was shown that another homologous enzyme, ADAMTS14, could process procollagen I (Colige et al., 2002), and that ADAMTS2 could process procollagen III (Wang et al., 2003). Nevertheless, a comprehensive understanding of how these enzymes participate in collagen biosynthesis *in vivo*, particularly during embryogenesis, as the first collagenous matrices are laid down, has not been achieved.

ADAMTS2, ADAMTS3 and ADAMTS14 constitute an evolutionarily related cluster of ADAMTS proteases, the procollagen amino-propeptidases (PNPs). They have an identical domain structure and a unique modular composition not present in any other enzyme. The active site sequence in their catalytic domain (HETGHVVGMEHD) differs from that of the other ADAMTS proteases. This, and previous experimental evidence, suggested that their catalytic activities are very similar (Colige et al., 2002; Fernandes et al., 2001; Wang et al., 2003). Here, we demonstrate, through the temporal and spatial analysis of mRNA encoding ADAMTS2, ADAMTS3 and ADAMTS14, as well as of that encoding procollagen I, II and III, that there is highly coordinated developmental expression of two of these enzymes and their procollagen substrates during mouse embryogenesis. We demonstrate that ADAMTS3 can process procollagen I. Together with its prominent expression in some procollagen I rich tissues and in cartilage, ADAMTS3, and not ADAMTS2, may be the major procollagen I and II aminopropeptidase from a developmental perspective. ADAMTS2 is essential for procollagen III processing in mice, with possibly important implications for human dermatosparaxis.

## MATERIALS AND METHODS

### Riboprobes

cDNA clones suitable for *in vitro* transcription of mouse pro- $\alpha$ 1(I) collagen (*Col1a1*) and mouse pro- $\alpha$ 1(III) collagen (*Col3a1*) were provided by E. Vuorio (University of Turku, Finland) (Niederreither et al., 1995). The mouse pro- $\alpha$ 1(II) collagen (*Col2a1*) cDNA was provided by Dr Naomi Fukai (Harvard Medical School). The *Adams2* probe was generated by subcloning a 600 base pair (bp) *NotI-PstI* fragment from IMAGE clone AA832579 into pBluescript II (Stratagene, La Jolla, CA). The following primers were used to amplify *Adams3* and *Adams14* cDNA fragments by RT-PCR of mouse 17.5 days post-coitum (dpc) embryo cDNA.

*Adams3* (amplifying a 528 bp fragment corresponding to the unique C-terminal domain of mouse *Adams3*), 5'-CTGAAGAGCTGGTCTCAGTG-3' (forward) and 5'-TGGACTTGTACCCAAACATG-3' (reverse); and

*Adams14* (producing a PCR product of 310 bp that corresponds to the unique C-terminal domain), 5'-AGGTGGGTGACAGAAGAGTGG-3' (forward) and 5'-AGGCTGCAGATCTGCACCATG-3' (reverse).

The PCR products were cloned into pGEMT-Easy (Promega, Madison, WI). Radioactive sense and antisense cRNA probes were generated by *in vitro* transcription using T3, T7 or SP6 RNA polymerases in the presence of [<sup>35</sup>S]-UTP (Perkin-Elmer).

### In situ hybridization

*In situ* hybridization was performed on tissue sections of mouse embryos of gestational age 7.5 to 17.5 dpc. Embryos, including the surrounding uterus at 7.5 dpc and 9.5 dpc, were fixed overnight in 4% paraformaldehyde, embedded in paraffin wax, and sectioned at 6  $\mu$ m. The sections were hybridized to [<sup>35</sup>S]-UTP labeled riboprobes, as previously described (Somerville et al., 2003). Slides were dipped in NTB-2 emulsion (Kodak, Rochester, NY) and exposed at 4°C for 2 to 10 days. Nuclei were stained with Hoechst 33258 (Sigma, St Louis, MO). Dark-field (autoradiographic signal) and fluorescence microscopy (nuclear staining) images were captured on a Leica DMR upright microscope (Leica Microsystems, Wetzlar GmbH), with a Retiga Exi Fast 1394 cooled CCD camera with RGB liquid crystal color filter slider (Q Imaging, Burnaby, BC, Canada). Images were viewed with ImagePro Plus software (Media Cybernetics, Silver Spring, MD). Silver grains visualized on dark field microscopy were given a red pseudocolor and superimposed on nuclear fluorescence (blue).

### ADAMTS3 antibodies

A polyclonal antibody was generated in rabbits by immunization with a C-terminal peptide from human ADAMTS3 (CKKDGKIIDNRRPTRSSTLER-COOH), coupled to keyhole-limpet hemocyanin (Alpha Diagnostics International, Houston, TX). The antiserum was affinity-purified against the peptide coupled to NHS-sulfolink (Pierce).

### Cloning of cells expressing ADAMTS3 and co-culture with dermatosparactic fibroblasts

HEK 293F cells (Invitrogen, Carlsbad, CA) were maintained in growth medium (DMEM supplemented with antibiotics and 10% fetal bovine serum). Clones stably expressing the *ADAMTS3* expression plasmid pSHTS3 were isolated as previously described (Fernandes et al., 2001). Culture media were analyzed by western blotting with ADAMTS3 polyclonal antibody at a 1:1000 dilution, and clones expressing the highest levels of ADAMTS3 were used in subsequent studies. Peptide N-glycosidase F (PNGaseF, New England Biolabs, Beverly, MA) treatment of conditioned medium was carried out as previously described (Somerville et al., 2003). Dermatosparactic calf fibroblasts were cultured alone or co-cultured with equivalent numbers of HEK293 cells transfected with the empty plasmid vector (a negative control), or with the plasmid vector expressing either ADAMTS3 or ADAMTS2 (as a positive control) (Colige et al., 2002). Co-culture experiments were performed in DMEM supplemented with 2% FCS and ascorbic acid (50  $\mu$ g/ml) in the absence of G418. After 24 hours, the culture medium and the cell layer were collected separately and denatured in Laemmli sample buffer with or without 0.1 M DTT. The characterization of collagen polypeptides was performed by western blotting using type I or type III collagen-specific antiserum and ECL. These antisera were produced by repeated immunization of rabbit and guinea pig with bovine collagen I or collagen III, respectively. These antisera, used at a 1:2000 dilution, were highly specific and did not cross-react with other collagen types (data not shown).

### Histology, transmission electron microscopy (TEM) and mCT of *Adams2*-deficient (*Adams2*<sup>-/-</sup>) mice

*Adams2*<sup>-/-</sup> mice in the 129Sv strain were kindly provided by D. Prockop (Tulane University) and were used under approved institutional protocols. Genotype analysis was by PCR as previously described. Two-month-old mice were killed by CO<sub>2</sub> inhalation, and tissues and organs were isolated for histological analysis and for transmission electron microscopy. Lungs were fixed in 10% formalin under inflation as previously described (Oblander et al., 2005), followed by routine histology, including Hematoxylin and Eosin stain, Hart's stain for elastin and the Mallory trichrome stain. The skulls and hind limbs of the *Adams2*<sup>-/-</sup> mice and wild-type littermates were analyzed by micro-computerized tomography (mCT), followed by three-dimensional reconstruction of the images. Samples for TEM were prepared as previously described (Birk and Trelstad, 1986). Thick sections (1  $\mu$ m) were cut and



stained with methylene blue-azure B. Thin sections were cut using a Leica UCT ultramicrotome and a diamond knife. The lung was stained with 2.5% (w/v) ethanolic uranyl acetate followed by 0.04% (w/v) bismuth subnitrate. The aorta was stained with aqueous 2% (w/v) uranyl acetate followed by lead citrate. Sections were examined and photographed at 80 kV using a Tecnai 12 transmission electron microscope equipped with a Gatan US1000 2K Ultrascan digital camera.

#### Procollagen processing in *Adamts2*<sup>-/-</sup> mice

Individual organs were isolated from *Adamts2*<sup>-/-</sup> mice and wild-type littermates. These were either ground in liquid nitrogen and suspended in washing buffer [0.25 M sucrose, 20 mM EDTA, 2.5 mM NEM, 0.5 mM PMSF, 50 mM Tris (pH 7.5)] or directly solubilized in Laemmli sample buffer (aorta). After centrifugation (20000 g, 10 minutes), supernatants were discarded and pellets washed again as described above. Pellets were then suspended in 0.15 M extraction buffer [0.15 M NaCl, 50 mM Tris (pH 7.5)], rotated for 2 hours at 4°C and centrifuged (20000 g, 10 minutes). Extracts were collected and pellets sequentially extracted in 1 M extraction buffer [1 M NaCl, 50 mM Tris (pH 7.5)] and 0.1 M acetic acid. Adequate amounts of the various samples were then dialyzed at 4°C in 0.1 M acetic acid, lyophilized, denatured in Laemmli sample buffer with or without 0.1 DTT, and analyzed by SDS-PAGE. Sponge granulomas were induced by the subcutaneous insertion of circular PVA sponges (10 mm diameter, clinical PVA sponges grade 3, M-PACT Worldwide Management, Eudora, KS) rehydrated in PBS. After 4 weeks, mice were sacrificed and sponges were collected. Histology demonstrated the presence of abundant granulation tissue containing fibroblasts and blood vessels (data not shown). The sponges were ground, washed as described above, and then proteins were extracted in Laemmli sample buffer, separated by SDS-PAGE and western blotted with anti-collagen III as described above.

## RESULTS

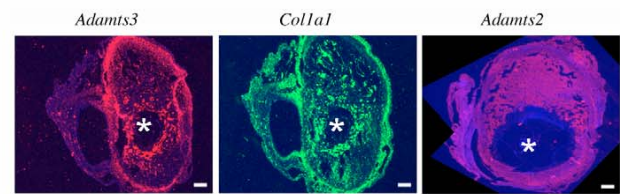
The distribution of the fibrillar collagen mRNA was consistent with previous reports (Niederreither et al., 1995). Thus, *Col1a1* was primarily expressed in bone, skin, perichondrium, periosteum, tendons and the adventitia (outermost layer) of blood vessels. *Col2a1* was expressed almost exclusively in cartilage. *Col3a1* expression was prominent in the vascular wall, lung mesenchyme, peritoneal lining and the wall of visceral organs. *Col3a1* was co-expressed with *Col1a1* in skin, but not in bone. None of the sense probes for any of the collagen or ADAMTS genes gave a signal above background levels.

#### *Adamts2* and *Adamts3* are expressed in the maternal tissues at day 7.5 of gestation

Northern analysis of commercial embryo RNA blots had shown the highest expression of both *Adamts2* and *Adamts3* at 7 dpc (Fernandes et al., 2001). However, in situ hybridization clarified that there was no expression in the embryo itself, whereas there was strong expression in maternal tissues, specifically in the decidual reaction and the myometrium of the uterus. Prominent *Col1a1* expression was seen in these tissues. *Col1a1* and *Adamts3* expression overlapped significantly in the decidual reaction (Fig. 1). The *Adamts2* expression pattern was different, but also overlapped with *Col1a1* (Fig. 1). The prominent co-expression of *Col1a1* with these biosynthetic enzymes reflects the intense remodeling of the uterus that occurs during pregnancy. These data suggest that the previously described northern analysis findings resulted from the inclusion of a significant amount of maternal tissue.

#### *Adamts3*, but not *Adamts2* or *Adamts14*, is co-expressed with *Col2a1* in cartilage

Most of the skeleton is formed from cartilage models that undergo endochondral ossification. Cartilage is first formed around 12 dpc in mesenchymal condensations that begin to express procollagen II and



**Fig. 1. *Adamts2* and *Adamts3* expression overlap with *Col1a1* in the 7.5 dpc uterus.** Note that *Adamts3* expression is similar to that of *Col1a1*, whereas *Adamts2* expression is different but has partial overlap. Asterisk indicates the embryo, which did not show expression. Scale bar: 100  $\mu$ m.

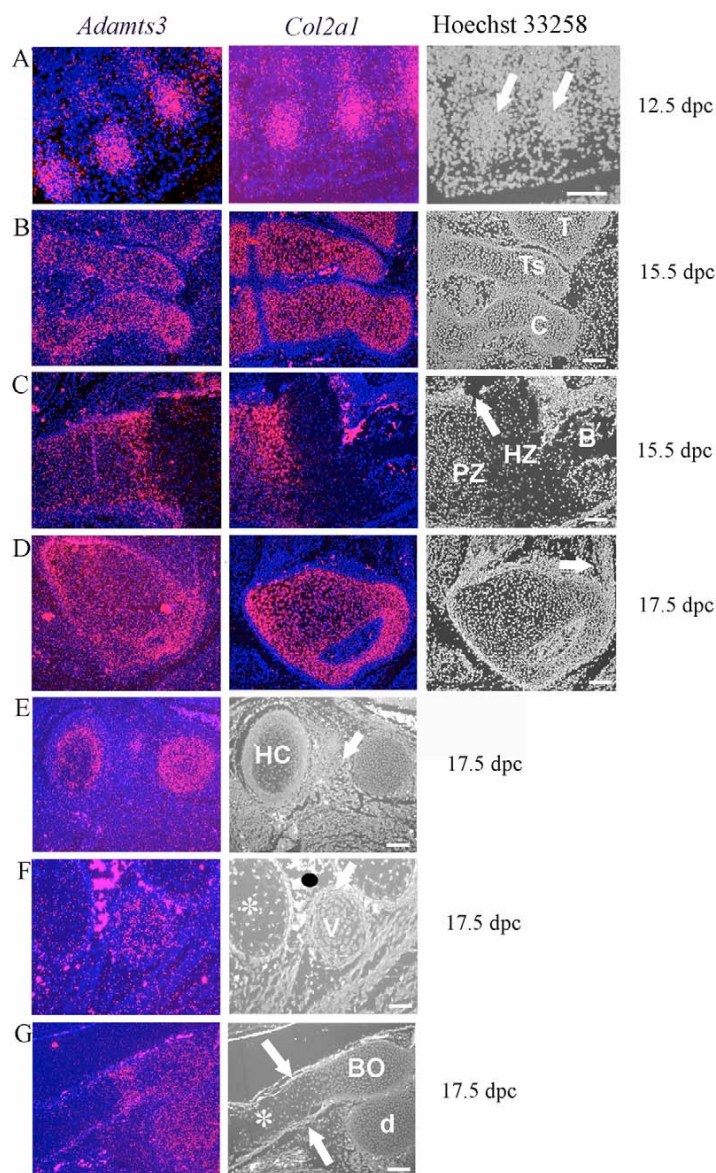
aggrecan. We have previously shown that ADAMTS3 processed procollagen II in vitro, and that ADAMTS3 mRNA was present at higher levels than ADAMTS2 mRNA in cultured chondrocytes by RT-PCR (Fernandes et al., 2001). However, in that study, the spatial and temporal association of *Adamts3* and *Col2a1* was not determined. *Adamts3* mRNA was associated with *Col2a1* mRNA expression from the initiation of chondrogenesis (Fig. 2A) and, subsequently, throughout skeletal development in all cartilage models, whether in the appendicular, axial or craniofacial skeleton (Fig. 2A-G). Chondrocytes in developing growth plates differentiate into hypertrophic chondrocytes and, as they do so, they suppress collagen II production and produce an abundance of a short-chain collagen, type X (LuValle et al., 1989), whose N-propeptides are not processed. Interestingly, *Adamts3* was downregulated as chondrocytes became hypertrophic (Fig. 2C,E-G). After 12 dpc, pre-cartilaginous mesenchymal tissue is located in the perichondrium around cartilage (Zhu et al., 1999); The *Adamts3* expression domain extended beyond the limits of the *Col2a1* expression domain into the perichondrium (Fig. 2B-G). Neither *Adamts2* nor *Adamts14* were detectable by in situ hybridization in cartilage at any developmental stage examined.

#### *Adamts3*, but not *Adamts2* or *Adamts14*, is co-expressed with *Col1a1* during bone and musculotendinous development

During endochondral ossification, as well as during intramembrane ossification, osteoblasts deposit collagen I in the osteoid, which is subsequently mineralized to form bone. During both types of osteogenic processes, *Adamts3* was co-expressed with *Col1a1* (Fig. 3A,B). In addition, *Adamts3* was co-expressed with *Col1a1* in perichondrium, including that around the hypertrophic zone of cartilage (Fig. 2C, Fig. 3A,C). Musculotendinous structures develop around the skeleton and synthesize abundant collagen I late in development; *Adamts3* was expressed in these structures at many locations in the skeleton at 17.5 dpc (Fig. 2D,E, Fig. 3B-D). Neither *Adamts2* nor *Adamts14* were detectable by in situ hybridization in bone or musculotendinous tissue during embryonic development.

***Adamts2*<sup>-/-</sup> mice have a normal skeleton and teeth**  
mCT analysis of two-month-old mice in whom skeletal maturity was attained, demonstrated no consistent anomalies in either the appendicular, axial or craniofacial skeleton. The overall morphology of the skull, as well as the surface features such as vascular foramina and the dimensions of its component bones, appeared no different from that of wild-type littermates (Fig. 4A). Severe abnormalities of the secondary dentition have been documented in EDS-VIIC patients (De Coster et al., 2003; Malfait et al., 2004). Incisors from





**Fig. 2. Co-expression of *Adamts3* and *Col2a1* in musculoskeletal tissues.** (A-G) In situ hybridization is used to show expression of *Adamts3* in developing cartilage from 12.5 dpc to 17.5 dpc. In A-D, images include hybridization of *Col2a1* to adjacent serial sections. Hoechst 33258 stained images are included on the far right for morphological correlation; the age of the embryo from which the section was taken is shown on the right. (A) Expression in initial sites of chondrogenesis in the developing ribs (arrows). (B) Expression in primordial cartilage of the ankle joint. T, tibia; Ts, Talus; C, calcaneus. (C) Expression in the proximal humerus growth plate. Note that expression is restricted to the proliferating zone (PZ). Hypertrophic zone chondrocytes (HZ) do not express *Adamts3*. *Col2a1* is not expressed in the bone of the primary ossification center (B). *Adamts3* is expressed in the perichondrium (arrow) and bone. (D) Proximal femur. The arrow indicates a tendon insertion site where *Adamts3*, but not *Col2a1* is expressed. (E-G) 17.5 dpc skeletal elements, demonstrating that *Adamts3* is not expressed by hypertrophic chondrocytes. (E) Absence of *Adamts3* expression in the hypertrophic chondrocytes (HC) of the manubrium sterni. Note *Adamts3* expression in developing intercostal muscle (arrow). (F) Ribs and vertebral lateral process. The rib elements contain hypertrophic chondrocytes (asterisk) that do not express *Adamts3*. By contrast, the lateral process of a thoracic vertebra (v) has small chondrocytes and these express *Adamts3*. The black oval indicates a spurious signal from erythrocytes. (G) Sagittal section through the basi-occiput (BO) and the dens (d) of the second cervical vertebra. Note the strong expression of *Adamts3* in regions containing small chondrocytes and absence from the hypertrophic zone (asterisk). Expression is seen in the perichondrium (arrows). Scale bar: 100  $\mu\text{m}$ .

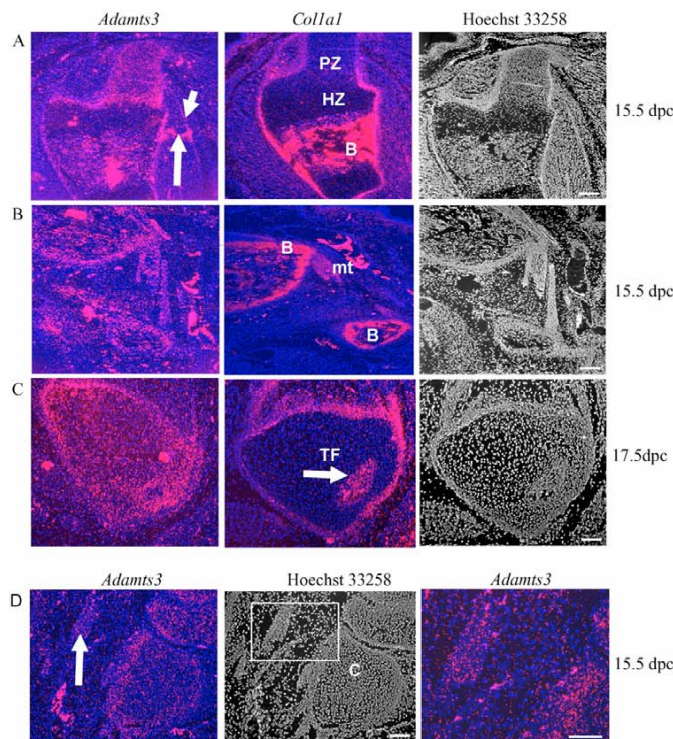
*Adamts2*<sup>-/-</sup> mice appeared normal, although molar teeth from *Adamts2*<sup>-/-</sup> mice showed a subtle loss of surface contour when compared with the wild-type littermates (Fig. 4A). Although *Adamts2* expression was not detectable during tooth development, *Adamts3*, but not *Adamts14*, was highly expressed in the dental papilla and at lower levels in ameloblasts (Fig. 4B).

#### ADAMTS3 rescues a procollagen I processing defect in dermatosaractic skin fibroblasts

HEK293F cells stably transfected with an ADAMTS3 expression plasmid, but not vector-transfected cells, produced an ~130-150 kDa protein that was reactive with the affinity-purified ADAMTS3 antibodies. The reactive band migrated faster after digestion with PNGase F as expected, since ADAMTS3 is predicted to have a N-linked carbohydrate (Fig. 4C). Type I collagen produced by dermatosaractic calf skin fibroblasts (DS) in culture consists of pro

$\alpha 1(\text{I})$  and pro  $\alpha 2(\text{I})$  in the cell layer, and pro  $\alpha 1(\text{I})$ , pro  $\alpha 2(\text{I})$  and pN $\alpha 1(\text{I})$  in the medium, demonstrating the absence of significant type I aminoprocollagen peptidase activity (Fig. 4D). In co-culture with control HEK293F transfected cells (Ct), pro  $\alpha 1(\text{I})$  and pro  $\alpha 2(\text{I})$  are also the most abundant products, but pC $\alpha 1(\text{I})$ , pN $\alpha 1(\text{I})$ ,  $\alpha 1(\text{I})$  and pN $\alpha 2(\text{I})$  are also detected, especially in the conditioned medium. These data illustrate the presence of low amino- and carboxy-procollagen peptidase activity. This is probably related to the synthesis of low levels of BMP1, ADAMTS3 and/or ADAMTS14 by DS, enzymes that may be more active in the high-confluence conditions observed in co-cultures than in monocultures. HEK293F cells expressing ADAMTS2 were used as a positive control. When co-cultured with DS, fully processed  $\alpha 1(\text{I})$  and  $\alpha 2(\text{I})$ , and a significant amount of pC $\alpha 1(\text{I})$ , were found associated with the cell layer. In the conditioned medium, only collagen molecules fully processed at the N terminus [ $\alpha 1(\text{I})$ ,  $\alpha 2(\text{I})$ , pC $\alpha 1(\text{I})$  and pC $\alpha 2(\text{I})$ ]

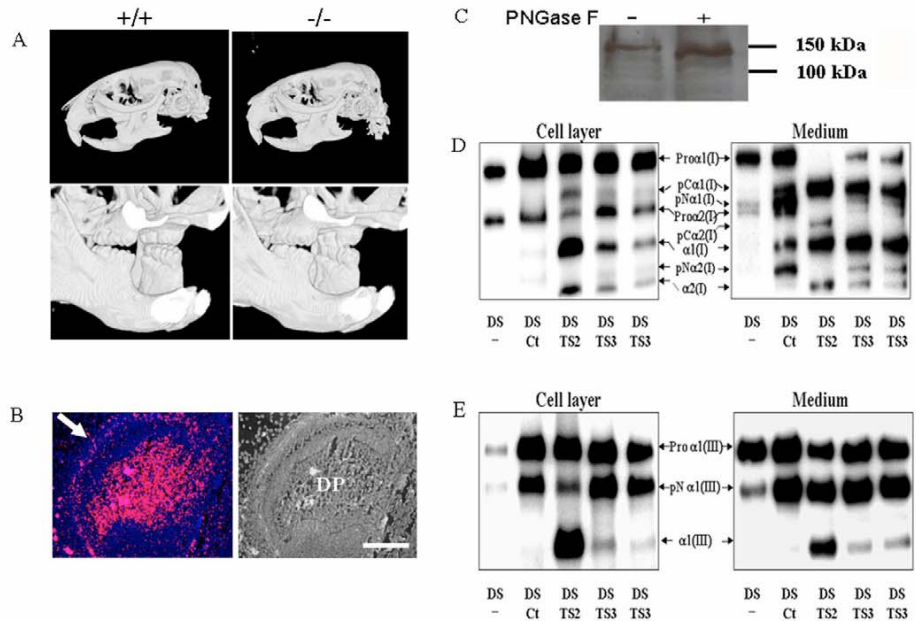




**Fig. 3. Co-expression of *Adamts3* and *Col1a1* in the musculoskeletal system.** All images are from 15.5 and 17.5 dpc embryos; Hoechst 33258-stained images are shown in the right-hand panel. (A) Expression in the appendicular skeleton (proximal humerus). Note that *Adamts3* expression overlaps with *Col1a1* in bone (B) and in the perichondrium around the hypertrophic zone and the periosteum (around bone). Chondrocytes in the proliferating zone (PZ) and hypertrophic zone (HZ) are indicated; arrows indicate mRNA in muscle. (B) Expression in the craniofacial skeleton. Note co-expression of *Col1a1* and *Adamts3* in bone (B) in the mandible and maxilla, as well as in developing masseter muscle (mt). (C) Expression in the proximal femur. The trochanteric fossa (TF), which contains the obturator tendons, is indicated by the arrow. The left panel in this figure is the same as that in Fig. 2D. (D) Expression in the developing Achilles tendon at 15.5 dpc. The image on the left shows *Adamts3* expression in the ankle cartilage (C) and tendon (arrow); that on the right shows the Achilles tendon at a higher magnification (boxed area in middle panel). Scale bar: 100 μm.

**Fig. 4. *Adamts3*, not *Adamts2*, has a role in procollagen I processing in the skeleton.**

(A) Normal craniofacial development, bone structure and minimal tooth involvement in *Adamts2*<sup>-/-</sup> mice. mCT analysis and three-dimensional reconstruction of skulls of *Adamts2*<sup>-/-</sup> mice and wild-type (+/+) littermates do not show differences in overall patterning, size, dimensions or organization of any craniofacial elements. The lower panels show tooth morphology from littermates. Note the poorly defined topography of molars from *Adamts2*<sup>-/-</sup> mice, although the overall morphology is similar. (B) *Adamts3*, but not *Adamts2* or *Adamts14*, was strongly expressed during tooth development in the dental papilla (DP), and was expressed less strongly in the ameloblasts (arrow). (C) Expression of ADAMTS3 in stably transfected 293 cells.



Conditioned medium from cells was analyzed by western blotting with a polyclonal antibody to ADAMTS3. Note that upon treatment with PNGase F, ADAMTS3 migrates faster. Untransfected cells showed no protein. Molecular mass markers are indicated on the right. (D) Rescue of the procollagen I processing defect by ADAMTS2 and ADAMTS3 in dermatosarptic calf skin fibroblasts (DS). DS were plated alone or mixed with an identical number of HEK-293 cells stably transfected with an empty expression vector (Ct), or with the same vector encoding ADAMTS2 (TS2) or ADAMTS3 (TS3). After two days in culture, western blotting of a non-reducing SDS-PAGE gel was used to detect type I collagen. Note that the antibody used detects both α1 and α2 chains, but reacts more strongly with the α1 chain. The identity of collagen polypeptides associated with the cell layer or recovered from the conditioned culture medium is indicated between the two panels. (E) Rescue of the procollagen III processing defect by ADAMTS2 and ADAMTS3 in DS. Samples used in D were also analyzed by western blotting using an anti-collagen III antiserum. The lower amount of collagen synthesized by DS cultured alone, as compared to co-cultures, was repeatedly observed and may be related to the lower level of confluence in the monoculture. Alternatively, HEK-293 cells may secrete factors activating the synthesis of procollagen. Scale bar: 100 μm.



were detected, illustrating a high aminoprocollagen peptidase activity. In DS/ADAMTS3-HEK293F cell co-culture, analysis of the collagen pattern demonstrates a high aminoprocollagen peptidase activity, although it is lower than that observed in presence of ADAMTS2.

The same co-culture samples were investigated for type III procollagen processing (Fig. 4E). A significant amount of fully processed  $\alpha 1(\text{III})$  was observed in the presence of ADAMTS2. Procollagen III processing was also detected when DS were co-cultured with cells expressing ADAMTS3.

#### **Adamts3 is expressed at extraskeletal locations**

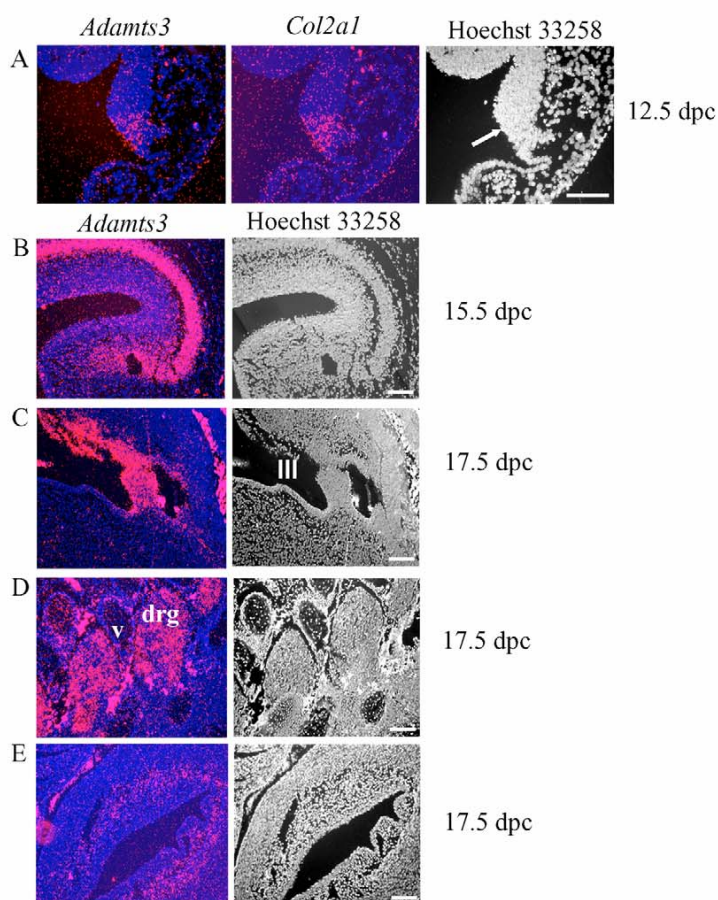
*Col2a1* expression has been previously reported at locations that are not associated with cartilage formation (Cheah et al., 1991). We found *Adamts3* to be precisely co-expressed with *Col2a1* at one such location in the hindbrain at 12.5 dpc (Fig. 5A). In addition, *Adamts3* was highly expressed at other locations where *Col2a1*, *Col1a1* or *Col3a1* gene expression was absent. At 15.5 dpc, *Adamts3* was highly expressed in the developing cerebral cortex (Fig. 5B), and at 17.5 dpc it was strongly expressed in the subcommissural organ, a specialized ependymal zone between the third and fourth ventricles (Fig. 5C), and in dorsal root ganglia (Fig. 5D). We have not investigated *Adamts3* expression in the vitreous humor, another extraskeletal site of *Col2a1* expression. *Adamts3* expression was seen in the wall of the urinary bladder (Fig. 5E).

#### **Adamts2, but not Adamts3 or Adamts14, is co-expressed with Col3a1**

*Adamts2* was co-expressed with *Col3a1* in the wall of large arteries, such as the aorta, carotid and pulmonary arteries, throughout mouse embryogenesis (Fig. 6A,B). By contrast, *Col1a1* was expressed only in the adventitial (outermost) layer of arteries (data not shown). Expression of both *Col3a1* and *Adamts2* was highest in the lung mesenchyme (Fig. 6B,C). *Adamts2* was also co-expressed with *Col3a1* at other sites, such as the palate (Fig. 6D), the intestinal wall, peritoneum and mesentery (Fig. 6E), the wall of the urinary bladder (Fig. 6F) and the skin (see Fig. 8A). *Adamts14* showed no co-expression with *Col3a1*, although expression of *Adamts3* in the wall of the urinary bladder partially overlapped with the expression of both *Adamts2* and *Col3a1* (Fig. 5E, Fig. 6F). With this exception, *Adamts3* was not otherwise co-expressed with *Col3a1*.

#### **Procollagen III processing is decreased in Adamts2<sup>-/-</sup> mice**

As only *Adamts2* was substantially co-expressed with *Col3a1*, and because ADAMTS2 processes procollagen III, we investigated whether procollagen III processing was affected in *Adamts2<sup>-/-</sup>* mice. Collagens were extracted from various mouse organs and western blotting was used to evaluate procollagen III processing. The different types of extraction gave essentially similar results. In all cases, a substantial proportion of procollagen III was



**Fig. 5. Expression of *Adamts3* outside the musculoskeletal system.** Hoechst 33258-stained images are shown in the right-hand panel. (A) Co-expression of *Adamts3* and *Col2a1* in the floor of the fourth ventricle in the hindbrain (arrow). (B) Expression of *Adamts3* in the cerebral cortex. (C) *Adamts3* expression in the ependyma of the subcommissural organ. III, third ventricle. (D) *Adamts3* in dorsal root ganglia (drg). Note the sparse expression in the vertebral cartilage (v) that comprises mainly hypertrophic chondrocytes. (E) Expression of *Adamts3* in the inner musculature of the urinary bladder. Note, in B-E, *Adamts3* was not co-expressed with *Col2a1*. Scale bar: 100  $\mu\text{m}$ .

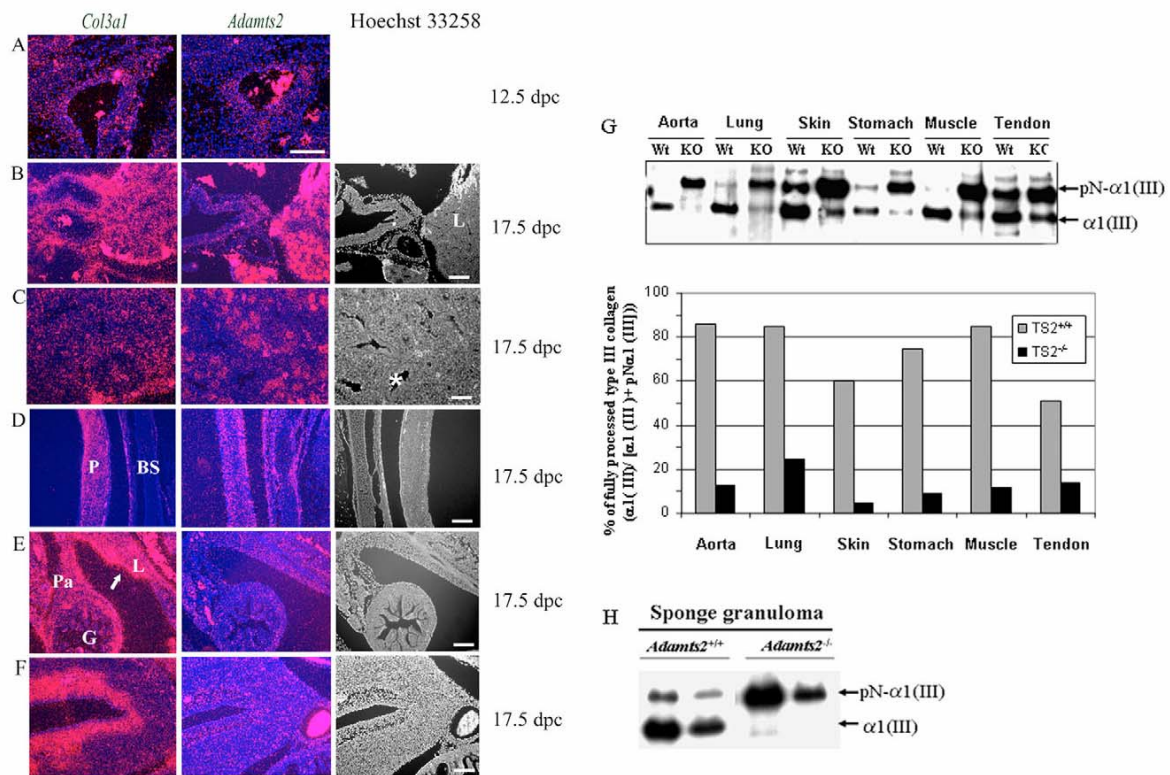
unprocessed in the *Adams2*<sup>-/-</sup> mice, whereas it was extensively or completely processed in wild-type mice (Fig. 6G). To investigate whether procollagen III processing by *Adams2* occurred in inflammation-induced fibrosis, proteins were analyzed from granulomas induced in *Adams2*<sup>-/-</sup> and wild-type mice by subcutaneous implantation of a PVA sponge. The analysis demonstrated that most of the newly synthesized procollagen III induced in the granulomas of wild-type mice was processed, whereas little or no procollagen III processing occurred in the granulomas induced in *Adams2*<sup>-/-</sup> mice (Fig. 6H).

***Adams2*<sup>-/-</sup> mice have distal airspace distension**

The lung was the most prominent site of developmental expression of *Adams2*, where its distribution corresponded with that of *Col3a1* (Fig. 6B,C). Lungs from two-month-old *Adams2*<sup>-/-</sup> mice and wild-type littermates were evaluated histologically after fixation under inflation to optimize parenchymal distension. *Adams2*<sup>-/-</sup> lungs had a consistent decrease in parenchymal density at both 2 months (Fig.

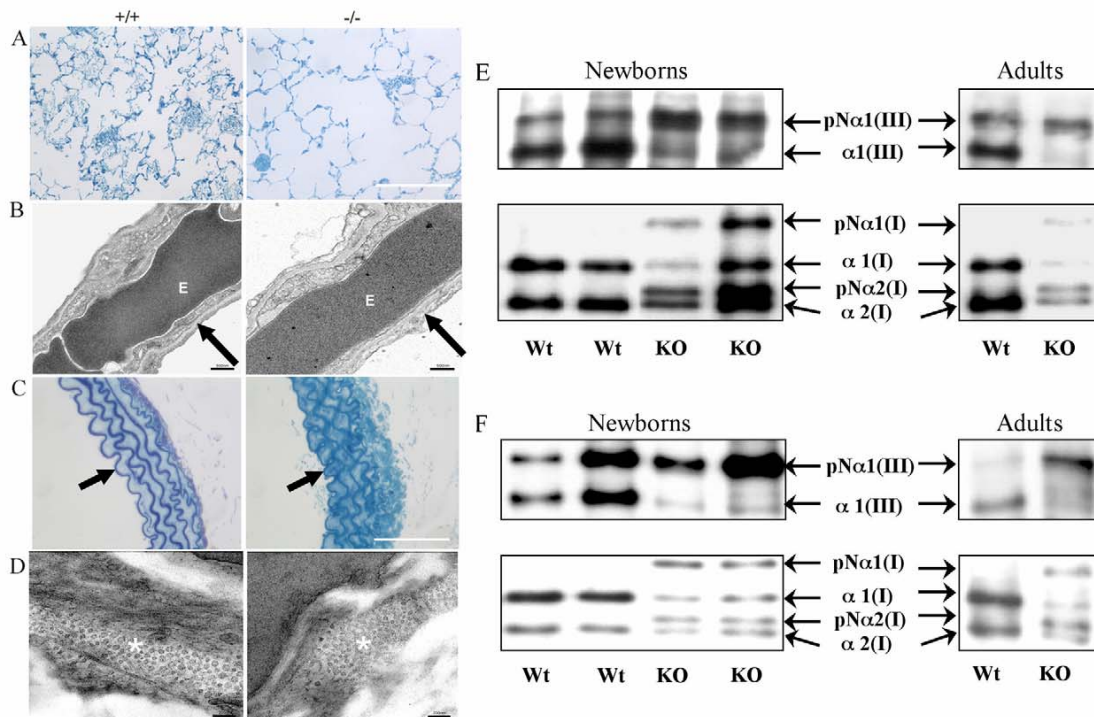
7A) and 2 weeks (data not shown). This emphysema-like appearance was unaccompanied by inflammatory cells or obvious fibrosis. Histochemical detection of elastin and collagen (data not shown) did not reveal any abnormalities and the ultrastructure showed the presence of normal basement membranes (Fig. 7B), elastic fibers and surfactant-producing pneumocytes (data not shown). By contrast, the histology (Fig. 7C) and electron microscopy of the aorta from *Adams2*<sup>-/-</sup> mice did not show consistent changes from that of the wild-type littermates. In both the lung and aorta (Fig. 7D) of *Adams2*<sup>-/-</sup> mice, the shape and diameter of collagen fibrils appeared to be unaffected.

We investigated whether the processing of both procollagen I and procollagen III were impaired in the aorta and lungs from *Adams2*<sup>-/-</sup> mice. Although it is not possible to comment from this data on the relative proportion of the two collagens in these tissues, it is clear that both newborn and adult *Adams2*<sup>-/-</sup> mice demonstrated significantly less processing of both collagens than did their wild-type littermates (Fig. 7E,F).



**Fig. 6. Co-expression of *Adams2* and *Col3a1*, and deficiency of procollagen III processing in *Adams2*<sup>-/-</sup> mice.** (A, B) *Adams2* is co-expressed with *Col3a1* in the arterial wall, as shown at two developmental stages: 12.5 dpc carotid artery (A) and 17.5 dpc pulmonary artery (B). L, lung. (C) Expression of *Col3a1* and *Adams2* in lung. Note the expression in lung mesenchyme but not in the epithelium of bronchial tubes (asterisk). (D) Co-expression of *Adams2* and *Col3a1* in the mesenchyme of the palate (P). *Adams2* is also expressed in the perichondrium of the basisphenoid (BS), at which location *Col3a1* is absent. (E) Co-expression of *Adams2* and *Col3a1* in the peritoneum lining the liver (L, arrow), and mesentery of the gut (G) and pancreas (Pa). (F) Co-expression of *Adams2* and *Col3a1* within the subepithelial layer of the developing urinary bladder. Scale bar: 100  $\mu$ m. In D-F, *Adams2* expression is weaker than in lung and arteries, and *Col3a1* expression is also significantly weaker and more dispersed. (G) Defective procollagen III processing in tissues from *Adams2*<sup>-/-</sup> mice. Western blot analysis of procollagen III isolated from mouse tissues with 0.15 M NaCl is shown. The tissue origin of the sample and the respective genotype is shown above each lane; procollagen III and collagen III molecular species are indicated on the left. Scanning densitometry of the different collagen III signals was used to calculate the proportion of fully processed molecules in the various tissues (lower panel). (H) Defective procollagen III processing in newly synthesized collagen in *Adams2*<sup>-/-</sup> mice. Protein was extracted from sponge granulomas and western blotting used to define the molecular species of procollagen III that were present. Sponge granulomas from two *Adams2*<sup>-/-</sup> mice and their wild-type littermates are shown. Scale bar: 100  $\mu$ m.





**Fig. 7. Histology and ultrastructure of lung and aorta in *Adamts2*<sup>-/-</sup> mice and wild-type (+/+) littermates.** (A) Comparative lung histology from *Adamts2*<sup>-/-</sup> and their wild-type (+/+) littermates. The distal air spaces are much larger in the mutant mice, but there is no inflammatory exudate. (B) Ultrastructure of an alveolar wall, demonstrating comparable basement membranes (arrows) in *Adamts2*<sup>-/-</sup> and wild-type mice. E, erythrocyte. (C) Methylene blue-azure B stain for elastin in *Adamts2*<sup>-/-</sup> and wild-type aortas demonstrates comparable aortic wall thickness and elastin lamellae (arrow). (D) Electron micrograph of collagen fibrils in cross section (in area marked by the asterisk) demonstrates comparable size and shape, and uniformity of fibril diameter. Scale bar: 100  $\mu$ m in A,C; 500 nm in B,D. (E) Western blot analysis of procollagen III (above) and I (below) processing in mouse lungs. Samples from wild-type and mutant newborn and adult mice are illustrated. (F) Western blot analysis of procollagen III (above) and I (below) processing in the mouse aorta. The various molecular species of collagen are indicated.

#### ***Adamts2* and *Adamts14* are co-expressed in mature mouse dermis**

In embryonic skin, low-level *Adamts2* expression was noted, but no expression of *Adamts3* or *Adamts14* was detected (Fig. 8A). However, in two-week old mice, both *Adamts2* and *Adamts14* were detected in the dermis, but not *Adamts3* (Fig. 8B).

#### **DISCUSSION**

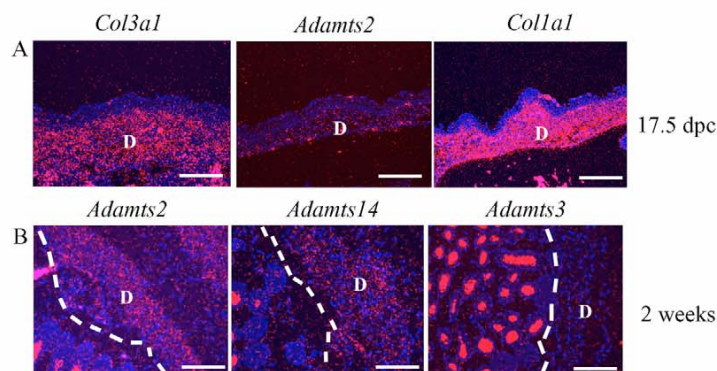
To rigorously establish the physiological significance of a protease-substrate interaction *in vivo* requires that the protease and substrate should be co-expressed, or that they should co-localize in tissues, and that in the absence of the protease, cleavage of the substrate should be reduced or absent. In addition, failure to cleave the substrate should have a significant impact on a biological system. We applied these criteria to exploring the roles of the procollagen N-proteinases during mouse embryogenesis. The results suggest that ADAMTS2 is the major procollagen III amino-propeptidase in mice, but that it appears to share the procollagen I processing function with ADAMTS3. Previous literature had suggested that a significant proportion of procollagen III in tissues was unprocessed and that the procollagen III-processing enzyme was not the same enzyme that processed procollagen I (Halila and Peltonen, 1986; Halila et al., 1986; Peltonen et al., 1985; Tuderman and Prockop, 1982). Wang et al. recently demonstrated that ADAMTS2 could process procollagen III *in vitro*, but the *in vivo* relevance was unclear

(Wang et al., 2003). The overlapping expression of *Adamts2* and *Col3a1*, and the substantial reduction in procollagen III processing in *Adamts2*<sup>-/-</sup> mice provides solid proof that ADAMTS2 is the major procollagen III-processing enzyme.

The role of ADAMTS2 in processing procollagen III may further contribute to the loss of mechanical integrity of dermosparactic skin, as collagen I and III are both present in the dermis. *In situ* hybridization revealed prominent *Col3a1* expression during lung development, in accordance with a previous study suggesting that, in mice, collagen III is a crucial structural component of the lung (Shiomi et al., 2003). The pseudo-emphysematous appearance of the lungs and the near-total lack of lung procollagen III processing in *Adamts2*<sup>-/-</sup> mice strongly suggests a vital requirement for collagen III in the lung. Lung abnormalities may result from a fragility of the pulmonary walls, similar to skin fragility in dermatosparaxis. Neither *Adamts3* nor *Adamts14* were expressed in the lung and, therefore, there was no compensating processing enzyme. However, in the aorta, which is severely affected by structural collagen III mutations, the near-total lack of procollagen III processing had no apparent effect on histology and collagen ultrastructure at 2 months of age. It is possible that changes may be seen in older mice, or by the induction of stress on the aortic wall in appropriate *in vivo* models.

The data suggest that ADAMTS3 probably makes a significant contribution to procollagen I processing during musculoskeletal development in mice. Expression of *Adamts3* in many *Col1a1*-





**Fig. 8. Procollagen N-propeptidase expression in skin.** D, dermis; the white dashed line indicates the line of demarcation of epidermis and dermis. **(A)** Co-expression of *Col3a1* and *Col1a1* in the dermis of the skin of a 17.5 dpc embryo. *Adamts2*, but not *Adamts14* or *Adamts3*, mRNA was detected in the dermis. **(B)** *Adamts2* and *Adamts14* mRNA, but not *Adamts3* mRNA, is detectable in the dermis of two-week-old skin. Scale bar: 100  $\mu\text{m}$ .

expressing tissues, such as bone and tendon, may explain why the most severe deficiencies in procollagen I processing in mouse dermatosparaxis are in the dermis. Together with our previous analysis of bovine dermatosparactic cartilage that demonstrated complete procollagen II processing, the data suggest that ADAMTS3 is probably the major procollagen II-processing enzyme, and predict that *Adamts3* mutations will lead to a chondrodysplasia or osteochondrodysplasia. Although *Col2a1* and *Col1a1* expression are generally mutually exclusive in tissues, *Adamts3* expression overlapped with that of both collagen genes. Although we detected *Adamts14* mRNA by RT-PCR of total embryo RNA (data not shown), in situ hybridization with three non-overlapping riboprobes showed no discernible developmental expression.

Is it possible that the PNPs have substrates other than the fibrillar procollagens? Although *Adamts2* expression was associated with procollagen gene expression, we noted strong expression of *Adamts3* in the nervous system at locations that lacked procollagen I, II or III mRNA. For instance, *Adamts3* was highly expressed in the sub-commisural organ (SCO), a specialized ependymal organ located at the entrance to the aqueduct of Sylvius. Its major secretory product is SCO-spondin, a glycoprotein that undergoes proteolytic processing prior to polymerization into the Reissner fiber of the spinal cord (Gobron et al., 1996). It is possible that ADAMTS3 might also be a SCO-spondin-processing enzyme.

Can the major fibrillar procollagen amino-propeptides be processed by other proteases? The highly conserved PNP modular organization (except for the variable C-terminal module) is different from all other proteases and is probably highly specialized for binding to triple-helical collagens. Heat-denatured collagen is not efficiently processed by ADAMTS2 (Tuderman et al., 1978), suggesting a need for spatially stringent interactions between the ancillary domain and the triple helix. Because residual procollagen processing in dermatosparactic skin occurs at the same cleavage site as in normal skin (Colige et al., 2002), it supports processing by an enzyme(s) closely related to ADAMTS2. It is clear from the *Adamts2*<sup>-/-</sup> mice that absence of ADAMTS2 leads to a significant loss of processing of procollagen III in many tissues. At these locations, neither *Adamts3* nor *Adamts14* are expressed, and, thus, do not compensate for its absence. Thus other enzymes do not appear to compensate significantly. A splice variant of collagen II, termed IIA is expressed in pre-cartilaginous mesenchyme and in perichondrium (Ryan and Sandell, 1990; Ryan et al., 1990). Ultrastructural and biochemical studies suggest its N-propeptide is not excised during development, suggesting perhaps that it is resistant to the action of these proteases (Zhu et al., 1999). By

contrast, matrix metalloproteinases (MMPs) have been shown to cleave the propeptide from collagen IIA at numerous sites other than the PNP bond (Fukui et al., 2002).

ADAMTS2 mutations in humans lead to characteristic craniofacial changes and decreased growth; features that are absent in *Adamts2*<sup>-/-</sup> mice. As individuals with EDS-VIIC age, the skin fragility is somewhat ameliorated and there is increasing joint laxity, which is not a prominent feature in early childhood. *Adamts2*<sup>-/-</sup> mice also have fragile skin, but they are not significantly growth retarded and have no craniofacial dysmorphism (Li et al., 2001). These differences between EDS-VIIC and mouse dermatosparaxis are compatible with the different deployment of functionally equivalent enzymes in humans and mice through the transcriptional regulation of their genes. The expression of ADAMTS3 and ADAMTS14 may change with age in humans, perhaps explaining how the properties of dermatosparactic skin change as affected individuals get older.

In addition to skin fragility, humans with EDS-VIIC manifest with a variety of other anomalies. Easy bruising, a universal sign in patients with EDS-VIIC, can be partly attributed to the ability of ADAMTS2 to process procollagen III, and to our observation, albeit in mice, that it is the only PNP expressed at significantly high levels in the walls of blood vessels. The procollagen III-processing defect in the absence of ADAMTS2 appears to be dramatic in most mouse tissues. We were particularly concerned that procollagen III processing in the mouse aorta was defective. Although histology and electron microscopy did not reveal specific anomalies in the aortic wall, and no aortic or other vascular aneurysms have been hitherto noted in EDS-VIIC patients, our findings suggest a need for the monitoring of EDS-VIIC patients as they age. Bladder rupture and intestinal perforations have been reported in individuals with EDS-VIIC. Our studies demonstrate co-expression of procollagen III and ADAMTS2 in the developing mouse at these locations. A recent clinical study emphasized the abnormal permanent dentition of individuals with EDS-VIIC, including hypodontia (De Coster et al., 2003). *Adamts2*<sup>-/-</sup> mice have mild dental changes and compensation by *Adamts3* may account for this.

Most of the 19 ADAMTS proteases can be assigned to phylogenetically distinct clades (Apte, 2004; Huxley-Jones et al., 2005; Nicholson et al., 2005). Recent analysis of ADAMTS gene evolution suggests that genes within mammalian ADAMTS clades arose by duplication of their primitive ancestors (Huxley-Jones et al., 2005; Nicholson et al., 2005). We speculated that, following duplication, closely related genes, such as ADAMTS2, ADAMTS3 and ADAMTS14, were highly conserved to provide subfunctionalization (retention of molecular function but with diversification of expression patterns) rather than

neofunctionalization (acquisition of new functions) (Huxley-Jones et al., 2005). The ability of all three PNP's to process procollagen I, but specialization in this regard as a result of differential gene regulation, supports this hypothesis. Other ADAMTS clades may exhibit the same phenomenon.

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