

Article

Application of DNA Barcoding for Monitoring Madagascar Fish Biodiversity in Coastal Areas

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Abstract: Madagascar is a marine biodiversity hotspot. A recent checklist recorded 1689 marine or transitional water fish species, 2.5% being endemic. To date, studies in this country were mostly focused on adult fishes using morphological-based identification. The early life stages of fishes remain largely understudied. The present work aimed to improve knowledge of fish biodiversity in Madagascar by focusing on post-larval reef fishes and settled juveniles in seagrass meadows of southwest Madagascar by using either species identification keys or DNA barcoding. Up to 119,500 individuals were collected, and 1096 individuals were successfully barcoded. We identified 387 species—85 through their morphology (with 58 unsuccessfully sequenced) and 302 by using CO1 barcoding corresponding to 302 barcode index numbers (BINs). This study added 27 new BINs for the BOLD database, 120 new for Madagascar, but only 159 were assigned a precise species name. By referring to the updated checklist of Madagascar fishes, 10 new species were detected for Madagascar. This number will probably increase when all the barcoded specimens become assigned to precise species names. These preliminary findings stress our poor knowledge of marine fish biodiversity in Madagascar and demonstrate the relevance of DNA barcoding in improving this knowledge.

Keywords: molecular tool; fish post-larvae; fish juvenile; coastal habitat; marine biodiversity



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1. Introduction

Madagascar is recognized for its terrestrial and marine biodiversity [1], including fishes. The country has about 1800 fish species with about 1700 marine fish, including anadromous and catadromous species [2]. Among marine fish species, up to 43 are endemic to Madagascar according to Fricke et al. [2]. The knowledge about Madagascar marine fish biodiversity mainly focuses on morphology-based identification which is usually not useful for some families or for early life stages. However, surveying the early life stages of fishes could probably improve this knowledge as demonstrated by [3] in the waters of Reunion Island.

Specimen identification at the species level constitutes a major issue when working on post-larvae or juvenile fish. Indeed, no specialized identification guide exists for reef fish juveniles, and the available identification guides for fish larvae rarely go beyond the genus level [4]. Several more precise identification guides exist, but they have been developed for specific areas and generally concern only a few species [5,6]. Obtaining the precise

species richness in early life stages of fish remains, therefore, challenging [7]. For this reason, most previous studies describing larval fish diversity remain limited to the family level in temperate and tropical areas [8–13].

DNA barcoding based on mitochondrial cytochrome-c oxidase 1 sequence (CO1) is one of the most widely used tools for species identification [3,7,14,15]. According to [16], this approach appears more effective for identifying fish larvae than techniques based on individual morphology and meristic character. Moreover, Meredith [17] reported the potential of DNA barcoding in revealing the misidentification of species when using traditional identification keys. Despite the potential ability of DNA barcoding to assign a species name to a given specimen, this tool presents, however, some limitations, such as its low capacities in identifying recently diverged or hybrid species [15]. These limitations allowed these authors to suggest an approach integrating both morphological characteristics and DNA barcoding to improve the efficiency of species identification. This identification-based approach was adopted by [3] through the combination of DNA barcoding with high-definition photos for identifying fish larvae from Reunion Island waters. Another limitation of molecular-based identification is the capacity of reference libraries to associate CO1 sequences with precise species names. Indeed, these libraries are currently confronted with problems related to misidentifications as reported by [18].

The present work focused on surveying fish larvae and juveniles by using the approach combining DNA barcoding and high-definition photos developed by [3]. The ultimate goal was to enhance our knowledge of Malagasy fish biodiversity, and to pursue the building of a CO1 sequence library for Madagascar fishes. More specifically, the present work aimed at: (i) characterizing the diversity of post-larval fish in coral reef habitats and juvenile fishes in seagrass beds, and (ii) discovering eventual new records for Madagascar.

2. Materials and Methods

2.1. Study Sites and Sampling

This study was carried out in coral reef habitats and seagrass beds of the southwestern coast of Madagascar (Figure 1). The post-larval fish survey was conducted in (i) a flat coral reef surrounding Nosy Ve Island, and (ii) in the northern part of the Toliara Great Barrier Reef. These sites were selected because previous scientific information on habitat morphology and the associated resources were available [10,19]. A juvenile fish survey was conducted in the seagrass beds off Ankilibe, 15 km to the south of Toliara. This area was chosen because of the presence of small-scale fishermen engaged in juvenile fishing using mosquito seine nets [20].

Post-larvae sampling took place over three six-month periods during the warm season: November to April of 2014–2015, 2016–2017, and 2017–2018. The warm season was chosen as the peaks of abundance and species richness always occur during this period [10]. For each warm season, three stations per site were sampled by using light traps called “SLEEP” (acronym of “Système Lumineux Electronique d’Echantillonnage des Post-larves”) developed by OCEA Consult’ on Reunion Island [3]. Each monthly sampling consisted of three consecutive nights centered on the new moon period, as the fish larval supply on the reef environment mainly occurs around this lunar phase [21]. Although light traps are selective and their efficiency is influenced by the water-current conditions or by water mass turbidity [22,23], they were used because they allow one to catch fish larvae before they settle onto benthic habitats, at a stage called “post-larvae” [24,25]. Six light traps per night were deployed, with three traps per site (i.e., one trap per station). Light traps were set around sunset and retrieved around the crepuscule. The individuals caught by each trap were placed into separate containers filled with seawater for keeping them alive.

For seagrass fishes, the catches of two small-scale fishermen using mosquito seine nets were sampled during the warm seasons of 2016–2017 and 2017–2018 (i.e., November to April). This period corresponds to the highest landings in southwestern Madagascar [26]. Although several small-scale fishing gears are commonly used in southwest Madagascar, fishermen using mosquito seine nets were chosen as they capture many juveniles [27].

The catches were collected for three days at spring tide, a period more practical for the deployment of mosquito seine net needing a depth of less than 1.3 m and thus more favorable for fishing. Sampled catches were put into a cooler with ice and transported to the IH.SM laboratory for the identification process.

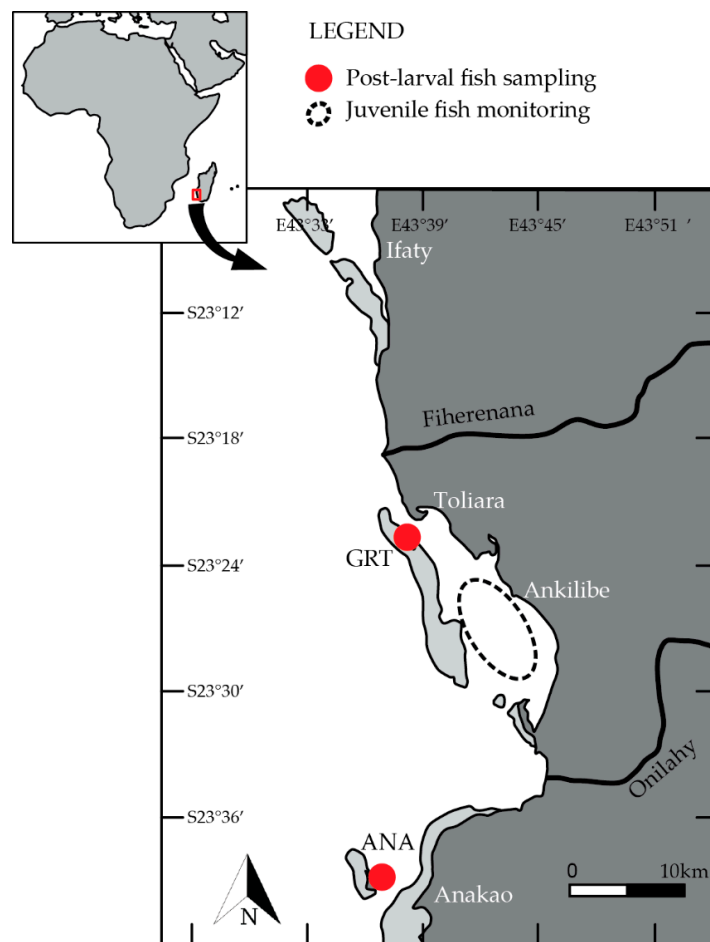


Figure 1. Locations of the sampling sites for post-larvae (red circles) and juvenile fish (dotted circle). With ANA = Anakao reef, and GRT = Great Barrier Reef of Toliara.

2.2. Identification Process

In the laboratory, living individuals from light traps, and dead fishes from mosquito nets, were sorted by morphospecies, i.e., individuals presenting a similar morphology. Each morphospecies was identified at the lowest possible taxonomical level by using published keys [28–30], the identification of some specimens remaining to family or order level. One specimen per morphospecies and per sample was randomly selected and photographed with a camera Nikon model D90 equipped with a Sigma 105 mm macro lens. Post-larvae were euthanized with an overdose of clove oil. The camera was connected directly to a computer equipped with the Adobe Lightroom 5.7[®] software—created by Adobe Systems Inc. in San Jose California USA—used for managing the photos and all the information related to each specimen. A tissue fragment from each photographed specimen was preserved in 90% ethanol and stored at $-20\text{ }^{\circ}\text{C}$ until total DNA extraction. Several tissue fragments per morphospecies were brought for barcoding at the genotyping and sequencing facility, Institut des Sciences de l'Evolution, CEMEB, University of Montpellier, France. Tissue fragments were then rinsed with distilled water to remove the alcohol and then dried in 2 mL individual tubes. An automated purification of genomic DNA from dried tissues was performed by using a Macherey–Nagel NucleoMag[®] 96 Tissue kit [31]. About 650 bp were amplified from cytochrome oxidase 1 (CO1) mitochondrial

gene by using the cocktail of primers FishF1-5'TCAACCAACCACAAAGACATTGGCAC3' and FishF2-5'TCGACTAATCATAAAGATATCGGCAC3' [BOLD Primer code: C_FishF1] in combination with FishR1-5'TAGACTTCTGGGTGGCCAAAGAATCA3' [BOLD Primer code: VR1] [32]. A complete description of the sequencing process can be found in [3]. The CO1 sequences were manually edited by using Chromas 2.6.4 (DNA Sequencing software, available online: <http://technelysium.com.au/wp/chromas/>, accessed on 15 May 2017). CO1 sequences with their corresponding specimen images and sampling details (e.g., location, time, taxonomy) were uploaded into the Barcode of Life Data System database [dx.doi.org/10.5883/DS-PHDJAO]. In BOLD, each sequence was assigned a barcode index number (BIN) by delineating sequence clusters through single linkage analysis by using a 2.2% sequence divergence for obtaining the operational taxonomic units [33]. The resulting OTUs were refined by using Markov clustering and the optimal partition was selected through silhouette criterion ([33]). For assigning a species name to each BIN, we adopted an approach consisting of several steps. First, if the BIN corresponded to only one species in BOLD, and this species was observed in this BIN only, the specimen was identified as "Genus + species". Second, if the BIN corresponded to different species from the same genus, or if the species corresponding to this BIN was also assigned to different BINs in BOLD, the specimen was identified as "Genus + BIN". Third, if the BIN corresponded to species from different genera, but belonged to the same family, the specimen was identified as "Family + BIN". Fourth, when the BIN was new for BOLD (i.e., corresponding to specimens that had never been barcoded before), identification was based on the combination of the lowest taxonomical level based on morphological character and the BIN (i.e., Genus + BIN and/or Family + BIN). Note that identification such as "Genus + BIN", or "Family + BIN", do correspond to identifications at the species level, as each BIN corresponds to an OTU, and thus to a putative species [33].

2.3. Data Analyses

Data analyses were implemented with R programming software R3.5.1 created by R Core Team in Vienna-Austria [34]. For providing the expected observed fish species richness and assessing the additional sampling benefits, species accumulation curves of the population from mosquito seine nets and light-trap catches were plotted by using the "vegan" package (Version 2.5.4, [35]). Bootstrap estimators were demonstrated as better estimators of total species richness than jackknife and Chao's estimator [36]. The "wqid" package (Version 0.2.2, [37]) using richboot function was used to obtain the bootstrapped estimation of species richness [38].

The number of species corresponding to the initial names obtained from morphological-based identifications was compared to the resulting OTU from BOLD to highlight the strength of molecular-based identification approach. For assessing the effort of fish DNA barcoding in Madagascar, the existing BINs in BOLD—associated with fish species names from this country—were extracted from the Public Data Portal on 4 May 2022 by using the keywords "Actinopterygii Madagascar". The species names that occurred most were assigned to each of the extracted BINs and compared to the list of fish species reported by [2]. To obtain the contribution of the present study in increasing DNA barcoding effort for Madagascar fishes, these extracted BINs were compared with the BINs obtained in this study.

3. Results

3.1. Species Richness: Morphological vs. Molecular-Based Identification

In total, 364 samples were collected: 286 from light traps and 78 from mosquito seine nets. 119,500 individuals were collected, 50,342 from light traps and 69,158 from mosquito seine nets. Using morphological characteristics only, we identified 656 morphospecies among post-larvae and juvenile fish. A total of 1096 individuals from 571 morphospecies were successfully barcoded (with 1 to 17 barcoded specimens for each morphospecies; see Supplementary Material Table S1). Finally, 387 species from 66 families were obtained by the combination of molecular-based identification and morphology approach (see Supplementary Material Table

S2). Ten families were the most speciose: Apogonidae with 37 species, Pomacentridae (26), Syngnathidae (25), Lethrinidae (22), Labridae (22), Mullidae (20), Chaetodontidae (18), Gobiidae (16), Acanthuridae (15), and Carangidae (14). Among the 387 species, 238 were observed as post-larvae and 232 as seagrass fishes, 83 species being observed in both sampling gears. Based on the species accumulation curves, the maximum values of species richness were not reached with light traps (Figure 2a) nor with mosquito seine nets (Figure 2b). The bootstrap estimator of the total species richness estimated that light traps and mosquito seine nets should catch up to approximately 275 and 264 species, respectively.

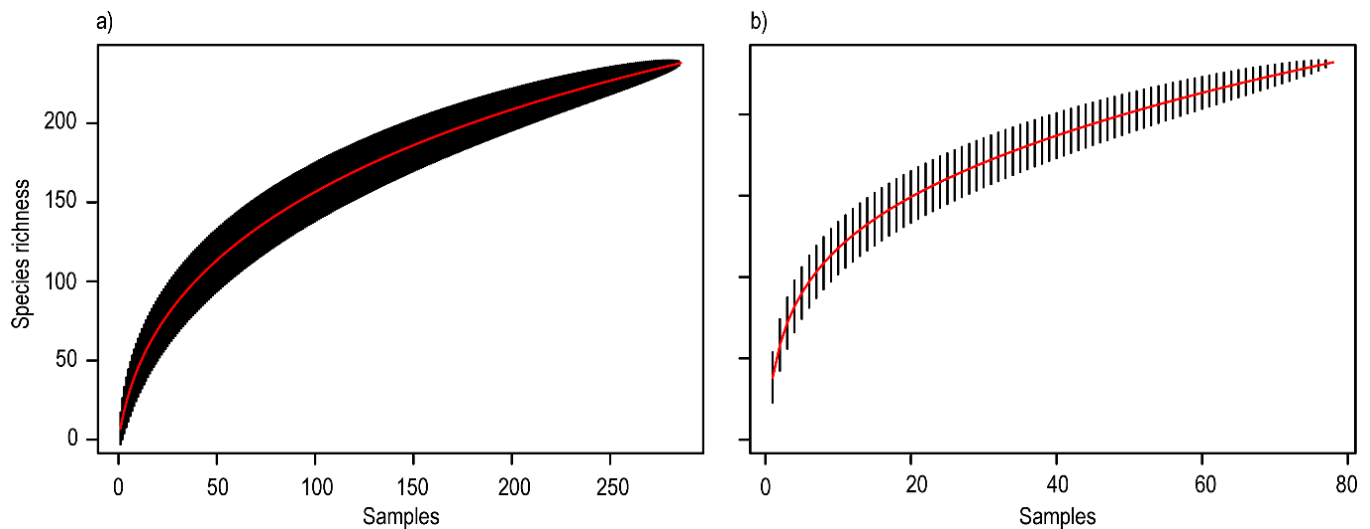


Figure 2. Species accumulation curves for post-larval (a) and juvenile fishes (b). One sample corresponds to the catch of each light trap per night (a) or each mosquito seine net per day (b). With black bars: interval of confidence and Red line: accumulation curve.

Among the 387 observed species, 85 (22%) were identified by their morphology only (27 at the species level, 58 as morphospecies which were unsuccessfully sequenced) and 302 (78%) by using CO1 barcoding corresponding to 302 BINs (see Supplementary Material Table S2). Among the 302 obtained BINs, 159 were unambiguously associated with a genus + species name, 115 presented ambiguities in species' name assignment, and 27 BINs had never been recorded before in the BOLD database. The 115 BINs presenting ambiguities in species' name assignment included 104 identified as "Genus + BIN" (e.g., *Hippocampus* [BOLD:ACE6993]), 11 as "Family + BIN" (e.g., Labridae [BOLD:ACT0048]) and one as "Subclass + BIN" (Actinopterygii [BOLD:ADN1799]) (Figure 3). Among the 27 new BINs, 14 were identified as "Genus + BIN" and 13 as "Family + BIN" (see Supplementary Material Table S2). More than half of these 27 new BINs for BOLD belonged to Apogonidae (four BINs), Gobiidae (three BINs), Labridae (four BINs), Lethrinidae (four BINs), and Syngnathidae (three BINs).

3.2. Discordances in Species Assignment

Among the 141 BINs presenting ambiguities in the species' name assignment, 70, including the 27 new ones, were simply not assigned to species name in the BOLD database. The remaining 71 BINs presented ambiguities in species assignment. These ambiguities came either from BINs associated with several species names in BOLD, or from BINs assigned to one species name to which two or more BIN(s) were associated. The first type of ambiguity concerned three BINs: BOLD:AAA9764 associated with *Myripristis hexagona* and *M. murdjan*, BOLD:AAD1777 associated with *Pempheris adusta* and *P. nesogallica*, and BOLD:AAD5600 associated with *Apogon erythrinus* and *Ostorhinchus aureus*. The second type of ambiguity corresponded to 68 BINs.

Identification levels

Total of obtained morphospecies = 656		CO1 & Morphology (387 species)	
Morphology-based identification	Species Level (27)	Morphology-based identification (85)	
Failed identification (based on CO1 barcoding)	Morphospecies level (58)		
Successfully barcoded (using CO1)	Morphospecies (571)	Identification based CO1 (302)	
		Genus + species (158)	Genus + BIN (118)
		Family + BIN (25)	Subclass + BIN (1)

Figure 3. Identification levels. The numbers in brackets correspond to the number of species for each category.

3.3. Geographical Distribution of Ambiguous Species

The species' geographical distribution is important for understanding whether the ambiguities in species name assignment could be linked to the different regions where they occur. The geographical distribution of the 71 BINs presenting ambiguities in the species' name assignment could be categorized into two groups. The first group comprised 44 BINs for which the associated species name was also associated with another BIN(s) observed in different regions (Table 1). For the second group of 27 BINs, no clear geographical distribution was observed (Table 2).

Table 1. Ambiguously identified species with clearly separated origins of the specimens of each BIN, with IO: Indian Ocean, PO: Pacific Ocean, and AO: Atlantic Ocean.

Species Name in BOLD	Current Work		Other Projects in BOLD Database	
	BINs	Distribution in BOLD	Other BINs	Distribution in BOLD
<i>Amblygobius phalaena</i>	BOLD: AAB8727	West IO & PO	BOLD: AAB8728	Center PO
<i>Antennarius striatus</i>	BOLD: AAO6018	East AO & IO, South PO	BOLD: AAF2509	West AO, North PO
<i>Apogon semiornatus</i>	BOLD: AAD2206	West IO	BOLD: AAD2207	West PO
<i>Asterropteryx semipunctata</i>	BOLD: AAC0108	West IO	BOLD: AAC0105/BOLD: AAC0106/ BOLD: AAC0107/BOLD: AAC0109	South & North PO, North AO
<i>Cheilinus chlorourus</i>	BOLD: AAB4186	West IO	BOLD: AAB4185/BOLD: ACE8465	PO
<i>Corythoichthys flavofasciatus</i>	BOLD: AAI8860	West IO	BOLD: AAE9785/BOLD: ACR0421	North PO
<i>Dendrochirus brachypterus</i>	BOLD: AAC9564	West IO	BOLD: AAC9563	East IO, West PO
<i>Diagramma pictum</i>	BOLD: AAD4477	West IO	BOLD: AAB9940	North & East IO, West PO
<i>Epinephelus rivulatus</i>	BOLD: ACZ9919	West IO	BOLD: AAF0433	East IO, West PO
<i>Fowleria variegata</i>	BOLD: AAD8726	West IO	BOLD: AAD8727/BOLD: ACC5212	West PO
<i>Gazza minuta</i>	BOLD: AAB7096	West IO	BOLD: AAA9823	North IO & Northwest PO
<i>Gerres longirostris</i>	BOLD: AAE6359	West IO	BOLD: AAE6360	Souther PO
<i>Halichoeres nebulosus</i>	BOLD: AAC7896	West IO	BOLD: AAC7897/BOLD: AAD5811	East IO, North PO
<i>Halichoeres zeylonicus</i>	BOLD: AAF7654	West IO	BOLD: AAF7655	North & East IO
<i>Hippichthys spicifer</i>	BOLD: AAE5342	West IO	BOLD: ADA9516	North PO
<i>Hippocampus histrix</i>	BOLD: AAE5356	Southwest & North IO	BOLD: AAE5355	North PO
<i>Lethrinus harak</i>	BOLD: AAC1521	West IO	BOLD: AAC1520	East IO, West PO
<i>Lethrinus lentjan</i>	BOLD: AAB0511	West IO	BOLD: AAB0508	West PO
<i>Lethrinus miniatus</i>	BOLD: AAC8078	southwest & north IO	BOLD: AAC8077	East IO

Table 1. Cont.

Species Name in BOLD	Current Work		Other Projects in BOLD Database	
	BINs	Distribution in BOLD	Other BINs	Distribution in BOLD
<i>Lethrinus nebulosus</i>	BOLD: ABY6363	West IO	BOLD: ACE7416/BOLD: AAE3737/ BOLD: AAB2308	Northeast IO, West PO
<i>Lutjanus lutjanus</i>	BOLD: AAA8168	West, North & east IO	BOLD: AAA8169	North PO
<i>Nectamia fusca</i>	BOLD: AAL9262	West IO	BOLD: AAF8240/BOLD: ABZ1276	West PO
<i>Nectamia savayensis</i>	BOLD: AAD9453	West IO	BOLD: AAD6832/BOLD: AAD9454/ BOLD: AAF1913/BOLD: ABY4865	North & East IO, West PO
<i>Neoniphon sammara</i>	BOLD: AAC8278	West IO	BOLD: AAC6632	East IO, West PO
<i>Ostorhinchus aureus</i>	BOLD: ACE9301	West IO	BOLD: AAD5599	East IO, West PO
<i>Ostorhinchus cookii</i>	BOLD: AAC2084	West & North IO	BOLD: AAC2085/BOLD: AAC2086 BOLD: AAC2086	West PO
<i>Ostorhinchus taeniophorus</i> ^a	BOLD: AAD8453 BOLD: AAD8454	West & North IO	BOLD: ACP7798	West & Center PO
<i>Oxycheilinus bimaculatus</i>	BOLD: AAC3195	West IO	BOLD: AAC7530/BOLD: AAC7531	East IO, West PO
<i>Petroscirtes mitratus</i>	BOLD: AAE6131	West IO	BOLD: AAE3283/BOLD: ADK6555	North IO, West PO
<i>Plectroglyphidodon lacrymatus</i>	BOLD: AAB6988	West IO	BOLD: AAB6989	West PO
<i>Plesiops coeruleolineatus</i>	BOLD: AAE4785	West IO	BOLD: AAE4786/BOLD: AAE4787	South PO
<i>Pomacentrus caeruleus</i>	BOLD: AAB9539	West & East IO	BOLD: ADW3804/BOLD: ADW3805	North IO
<i>Pristiapogon fraenatus</i>	BOLD: AAJ1264	West IO	BOLD: AAD6833/BOLD: ADC1718	East IO, West PO
<i>Pristiapogon kallopterus</i>	BOLD: AAB4082	West IO & PO	BOLD: ABZ7996	Center PO
<i>Pseudalutarius nasicornis</i>	BOLD: AAI4531	West IO	BOLD: AAE8257	South PO
<i>Sargocentron diadema</i>	BOLD: AAB3424	West, North & East IO	BOLD: AAB3425	PO
<i>Scarus ghobban</i>	BOLD: ABY4451	West IO	BOLD: AAB3544/BOLD: AAB3545	East IO, West & East PO
<i>Sphyraena flavicauda</i>	BOLD: AAF8900	West IO	BOLD: AAG2679	East IO
<i>Stolephorus indicus</i>	BOLD: AAG4825	West IO	BOLD: AAB7978/BOLD: AAB7978	North IO & North PO
<i>Synanceia verrucosa</i>	BOLD: AAE2821	West IO, south & center PO	BOLD: AAE2820	North PO
<i>Tylosurus crocodilus</i>	BOLD: AAC4148	IO, North & center PO	BOLD: AAB9821/BOLD: ABY6772	West AO, East PO
<i>Valenciennea puellaris</i>	BOLD: AAC4124	West & North IO	BOLD: AAC4125	West PO
<i>Vanderhorstia ornatissima</i>	BOLD: AAF0288	West & North IO	BOLD: ACC5236	Southern PO

^a Species complex.

Table 2. Ambiguously identified species without separated origins of the specimens of each BIN, with IO: Indian Ocean, PO: Pacific Ocean, and AO: Atlantic Ocean.

Species Name in BOLD Database	Current Work		Other Projects in BOLD Database	
	BINs	Distribution in BOLD	BINs	Distribution in BOLD
Species Assigned to More Than Two BINs				
<i>Trachinocephalus myops</i> ^a	BOLD: AAA9578 BOLD: ABX6347	IO, PO	BOLD: AAA9580/BOLD: AAA9581 BOLD: AAA9582/BOLD: ACF3856	AO, IO, North PO
<i>Selar crumenophthalmus</i>	BOLD: AAB0871	West IO, West PO	BOLD: ACF3856 BOLD: AAB0870	West AO, West IO, West PO
<i>Parupeneus heptacanthus</i>	BOLD: AAB2590	West & East IO	BOLD: AAB2589/BOLD: AAI4268	East IO, West PO
<i>Caesio caeruleaurea</i>	BOLD: AAB4822	West IO	BOLD: AAB4823/BOLD: ABZ2197	West IO, East IO, West PO
<i>Synodus variegatus</i>	BOLD: AAB5069	West & East IO, West PO	BOLD: AAB5072	East IO

Table 2. Cont.

Species Name in BOLD Database	Current Work		Other Projects in BOLD Database	
	BINs	Distribution in BOLD	BINs	Distribution in BOLD
<i>Dactyloptena orientalis</i>	BOLD: AAB5966	West & East IO, West PO	BOLD: AAB5965	East IO, West PO
<i>Parupeneus barberinus</i> ^a	BOLD: AAB5978 BOLD: AAB5980	IO, West PO	BOLD: AAB5979	IO, West PO
<i>Gerres filamentosus</i>	BOLD: AAC0377	West & North IO	BOLD: AAC0379/BOLD: AAC0380/ BOLD: AAC0382/BOLD: AAC0383/ BOLD: AAL5737/BOLD: AAY1477/ BOLD: ADC2070	North & East IO, West PO
<i>Hemiramphus far</i>	BOLD: AAC0565	West IO	BOLD: AAC0563/BOLD: AAX1441	West, North & East IO, West PO
<i>Gerres oyena</i>	BOLD: AAC1291	West IO	BOLD: AAC1288/BOLD: AAC1289/ BOLD: AAC1290/BOLD: AAC1290	West, North & East IO, West PO
<i>Sebastapistes strongia</i>	BOLD: AAC4542 BOLD: AAC4543	South West IO, West PO	BOLD: AAC4545/BOLD: ACD1673	West PO
<i>Lagocephalus scleratus</i>	BOLD: AAC5565	IO, Mediterranean Sea, West PO	BOLD: AAC5566/BOLD: AAC5567	East IO, West PO
<i>Epinephelus flavocaeruleus</i>	BOLD: AAD1767	West & East IO, West PO	BOLD: ACE3678/BOLD: ACE8264	West IO
<i>Carangoides coeruleopinnatus</i>	BOLD: AAD2297	West & East IO, West PO	BOLD: AAB3475 BOLD: AAD2298	West & East IO, West PO
<i>Stethojulis albobittata</i>	BOLD: AAD4824	West IO	BOLD: ACF2947	West IO
<i>Lactoria fornasini</i>	BOLD: AAF2668	West IO, West PO	BOLD: AAF2667	West IO
<i>Zebrasoma desjardini</i>	BOLD: AAF6311	West IO	BOLD: ACV8450	West IO
<i>Apogon coccineus</i>	BOLD: AAU1535	West IO	BOLD: AAJ8751	West IO
<i>Chromis opercularis</i>	BOLD: ACF0042	West IO	BOLD: AAJ0279	West IO
<i>Upeneus vittatus</i>	BOLD: ACV4665	West & East IO	BOLD: ABZ7416	West & East IO
<i>Terapon jarbua</i>	BOLD: ACV8977	West IO	BOLD: AAA9351/BOLD: AAA9352/ BOLD: AAA9354/BOLD: ADR6412	West, North & East IO, West PO
Species Assigned to a BIN				
<i>Myripristis hexagona & Myripristis murdjan</i>	BOLD: AAA9764	West IO, PO		West IO, PO
<i>Pempheris adusta & Pempheris nesogallica</i>	BOLD: AAD1777	IO, Mediterranean Sea		IO, Mediterranean Sea
<i>Apogon erythrinus & Ostorhinchus aureus</i>	BOLD: AAD5600	West & East IO, North PO		North PO

^a Species complex.

3.4. DNA Barcoding Effort for Madagascar Fishes

Among the 1689 marine or transitional water fish species recorded in Madagascar EEZ [2], only 419 (i.e., about 24.8%) had already been barcoded and assigned to a BIN in the BOLD database in May 2022 (Table S3). Based on the comparison of these 419 BINs with the obtained BINs from this study, the present work added 120 new BINs for Madagascar (see Supplementary Material Table S2) including the 27 new BINs that were never recorded before for the BOLD database. These 120 new BINs belonged to 41 families, half of them corresponding to species of Pomacentridae (16 species), Apogonidae (14), Chaetodontidae (eight), Labridae (eight), Mullidae (eight), and Lethrinidae (six).

This study allowed us to identify 10 new records for Madagascar, i.e. species that had never been observed in the coastal waters of this country. These new species for Madagascar were among the 159 BINs that were unambiguously associated to a species name. These species were *Chaetodon ulietensis* Cuvier 1831, *Diagramma labiosum* Macleay

1883, *Dipterygonotus balteatus* Valenciennes 1830, *Equulites klunzingeri* Steindachner 1898, *Foa fo* Jordan & Seale 1905, *Neamia octospina* Smith & Radcliffe 1912, *Ostorhinchus gularis* Fraser & Lachner 1984, *Paramonacanthus frenatus* Peters 1855, *Pempheris ibo* Randall & Victor 2015, and *Scarus fuscopurpureus* Klunzinger 1871.

4. Discussion

The present work highlighted the usefulness of DNA barcoding as a tool for inventorying biodiversity based on early life stages of fish. We observed an over-evaluation of species richness when using only morphological characters, 571 morphospecies vs. 387 species when using DNA barcoding combined with a morphological approach. Inversely in other studies, using only a morphological approach has also led to an underestimation of the number of species as observed by [13] in the southwest tropical Atlantic Coast. These authors identified 76 post-larval reef fish morphospecies from 465 samples (i.e., the catches of one light trap for one night) over five years by using morphological and meristic characteristics. Even though these authors did not use DNA barcoding, this richness appeared to be low compared to the 733 reef fish species known in this part of the Atlantic Coast [39]. For instance, in [13] only one morphospecies assigned to “Genus spp. or Family spp.” was observed for each of the families Scorpaenidae, Gobiidae, and Synodontidae because meristic characters largely overlap between species. This highlights the inconsistency of traditional identification techniques due to the inefficiency of morphological characteristics for separating some species as already reported by [16].

The present work—using DNA-based identification—observed up to 265 putative species (i.e., 265 BINs) caught as post-larvae from 286 samples only. Previous work using similar light traps at the same sites obtained 128 post-larval reef fish morphospecies from 145 samples, 79 only being identified at the species level ([11]. Based on the graph presented in Figure 2a, 145 samples would correspond to approximately 180 species, i.e., ~40% more species than observed without using DNA barcoding. These findings confirm the DNA barcoding usefulness for identifying the early life stage of fish species as reported by [17], and support its higher efficiency compared to traditional techniques.

DNA barcoding can also contribute to knowledge improvement of fish biodiversity at local or global scales. As revealed by [40], DNA barcoding is useful for enhancing fish biodiversity knowledge as they detected up to 90 new species for South Africa during their ten-year-long post-larval survey. Similarly—in waters of Reunion Island—Collet et al. [3] demonstrated the importance of post-larval surveys in improving fish biodiversity knowledge using DNA barcoding. These authors found ten new species that had never been recorded in the Reunion Exclusive Economic Zone over six months of sampling during a single warm season (about 108 samples). These findings appear to be in line with our results as 10 species that had never been recorded in Madagascar were observed during three warm seasons (364 samples). By referring to the Catalog of Fishes, four of these new species for Madagascar (*Dipterygonotus balteatus*, *Foa fo*, *Neamia octospina*, *Paramonacanthus frenatus*, and *Pempheris ibo*) have already been observed in the Western Indian Ocean region. On the other hand, five of the new species (*Chaetodon ulietensis*, *Diagramma labiosum*, *Equulites klunzingeri*, *Ostorhinchus gularis*, and *Scarus fuscopurpureus*) appeared to be new records for this region. This result may be related to their misidentification during previous works using morphological identification only. This could also correspond to an extension of their geographical distribution linked to their larvae dispersal during tropical storms, which has already been demonstrated for *Epinephelus marginatus* whose larvae were suggested to be transported by a cyclone from South Africa—which holds the nearest known population—to Reunion Island [41]. Moreover, the present work also contributed to the enhancement of fish biodiversity knowledge at a global scale as 27 BINs new for the BOLD database were obtained. These new BINs may either correspond to fish species that have never been described due to the rarity of molecular-based studies in the region. This can also correspond to species that have never been barcoded at the adult stage [42] as most barcoding studies are still based on the commercial value of the species. The lack of

CO1 sequences in BOLD may also be linked to the way specimens are usually selected for DNA analyses. Most of the time, specimens are first grouped by similar morphologies (i.e., morphospecies) and patterns of colors. As the number of barcoded specimens per morphospecies is often limited, some species that look similar based on their morphology may be ignored. In the present work, several specimens were barcoded within each morphospecies. This was particularly the case for Lethrinids post-larvae and juveniles as cryptic species are often encountered in this family [43] and morphological keys are unable to distinguish the species [44]. In the present study, BOLD assigned the CO1 sequences of 138 Lethrinids specimens to 14 BINs, four of them being new for BOLD. This finding not only confirms that DNA barcoding can deal with species that are hard to differentiate [15] but also demonstrate that the way individuals are selected for DNA analyses is important for increasing fish biodiversity knowledge.

Although DNA barcoding appeared to be a promising tool for identifying the early life stage of fish, some barcoded individuals were not assigned to a species name. Wibowo et al. [45] were also unable to assign to species names a large proportion of larval fish morphospecies (68%) they had collected in Indonesia tropical swamps. According to these authors, the main problem was the lack of reference CO1 sequences. In the present work, 141 BINs were not clearly assigned to a species name. Of these 141 BINs, 43 (excluding the 27 new BINs for BOLD) were not associated with a precise species name. For the remaining 71 BINs, the species name corresponding to each of these BINs was also assigned to other BINs in BOLD. Ideally, there should be one BIN only for each species and vice versa [33]. Interestingly, for 44 of these 71 BINs (i.e., 62%), the species name associated with each BIN was also associated with another BIN corresponding to specimens caught in a different geographical location (Table 2). For example, among the two BINs corresponding to *Plectroglyphidodon lacrymatus*, BIN BOLD:AAI8860, observed during the present study, is solely observed in the Western Indian Ocean, whereas BIN BOLD:AAB6989 is observed in the Western Pacific Ocean. These findings confirm the DNA barcoding ability for tracing species origin [15]. It is, however, difficult to determine if these BINs correspond to sister species or only to one species presenting a high phylogeographic structure (i.e., an intraspecific divergence without speciation process) as observed in the groupers [46]. In order to not increase noise in BOLD, the specimens caught during the current study were, thus, identified as *Plectroglyphidodon* [BOLD:AAI8860]. For the remaining 27 BINs out of these 71 BINs, no clear spatial distribution in the BINs corresponding to each species name was observed (Table 2). For example *Zebrasoma desjardini* is associated either with the BIN BOLD:AAF6311 or with BOLD:ACV8450, both of them being observed in the Western Indian Ocean. This kind of ambiguous case may be associated with specimen voucher misidentifications that often occur in laboratories [47]. Indeed, *Zebrasoma desjardini* is difficult to distinguish morphologically from *Zebrasoma veliferum* [48]. Such misidentifications cause serious problems in reference libraries such as in BOLD and has already been stressed by [18] for the Indo-Pacific fish larvae. This highlights the important need for revising the information present in reference libraries.

An increase in the DNA barcoding effort, not only for adult fishes but also for larval and juvenile fishes, remains mandatory for a better knowledge of fish biodiversity. For example, 1919 fish species are known from South Africa's exclusive economic zone, but only 1006 (52.4%) have been barcoded [40]. Nevertheless, these authors demonstrated that the DNA barcoding effort added 90 new species caught as larvae and 139 as adults. For Madagascar, among the 1689 marine or transitional water fish species reported by [2], only 419 (~25%) had been barcoded before the present work. The current study sequenced up to 120 fish species of Madagascar that were never barcoded, and added 27 new BINs in BOLD, and recorded at least 10 new species for Madagascar. Thus, the number of marine, or transitional water, fish species in Madagascar rose to 1710, 530 of them (~36%) being barcoded. In Madagascar, the effort in terms of DNA barcoding is still in an early stage of development compared to other countries in the Western Indian Ocean such as South Africa. Increasing this DNA barcoding effort by conducting surveys at broader geographic

and temporal scales, using all life stages from post-larvae to adults, will thus improve the knowledge of Malagasy fish biodiversity. This will allow updating the checklist of Malagasy fishes reported by [2].

5. Conclusions

In conclusion, this study clearly stressed the usefulness of DNA barcoding as a promising tool for enhancing the knowledge of the Malagasy marine fish biodiversity. This work—which added a large number of new sequences to BOLD—represents an important step for establishing a national sequence reference library for marine fish of Madagascar. The large number of fish species that remain to be barcoded demonstrates that Madagascar is still in the early stages of development in terms of molecular-based identification of marine fish. Increasing research efforts based on molecular-based identification by conducting a study on a broader scale will contribute to a significant effect on fish biodiversity. Although the present work was carried out in a restricted area only, nine new records for Madagascar were detected. This number will probably increase when all the barcoded specimens—which presented ambiguities—are assigned to a precise species name. However, even if some specimens were not assigned to a species name, BIN can be used for biodiversity or ecological surveys [49]. Therefore, accurately identifying species based on their corresponding BIN will permit linking the early life stages to adults and investigating the response of these different life stages to environmental conditions.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/d14050377/s1>. Table S1: Dataset of individuals successfully barcoded during the present work. Table S2: List of species captured by light traps and mosquito seine nets during austral warm seasons. The species are organized according to Nelson (2006). Total abundances are reported for each sampling gear as well as the number of successfully barcoded specimens. Table S3: List of barcoded fish species from Madagascar in BOLD in 4 May 2022.

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Institutional Review Board Statement: Ethical review and approval were waived for this study as we work on dead animals retrieved from small-scale fisher catches for juvenile monitoring. For the collected living fish post-larvae from light trap, only one individual per taxa were collected and the remaining we returned to the sea.

Data Availability Statement: The data presented in this study are openly available in the Barcode of Life Data System database at dx.doi.org/10.5883/DS-PHDJAO and in GENBank with Reference accession number OL409189-OL410277 (this will publicly accessible by October 2022).

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References

1. MEF. *Fourth National Report on Biological Diversity Convention Madagascar*; Ministry of Environment and Forests: Antananarivo, Madagascar, 2009; p. 144.
2. Fricke, R.; Mahafina, J.; Behivoke, F.; Jaonalison, H.; Léopold, M.; Ponton, D. Annotated Checklist of the Fishes of Madagascar, Southwestern Indian Ocean, with 158 New Records. *FishTaxa* **2018**, *3*, 432.
3. Collet, A.; Durand, J.-D.; Desmarais, E.; Cerqueira, F.; Cantinelli, T.; Valade, P.; Ponton, D. DNA Barcoding Post-Larvae Can Improve the Knowledge about Fish Biodiversity: An Example from La Reunion, SW Indian Ocean. *Mitochondrial DNA Part A* **2018**, *29*, 905–918. [[CrossRef](#)] [[PubMed](#)]
4. Leis, J.M.; Carson-Ewart, B.M. *The Larvae of Indo-Pacific Coastal Fishes: An Identification Guide to Marine Fish Larvae*, 2nd ed.; Leiden; Brill: Boston, FL, USA, 2004; ISBN 978-90-04-11577-4.
5. Maamaatuaiahutapu, M.; Remoissenet, G.; Galzin, R. *Guide d'identification Des Larves de Poissons Récifaux de Polynésie Française*; Coral Reef Initiative for the South Pacific; Téthys: Nouméa, New Calédonie, 2006; 108p.
6. Juncker, M. *Young Coral Reef Fish of Wallis and the Central Pacific: Identification Guide*; Wallis and Futuna-Environment: Noumea, New Caledonia, 2007; 170p.
7. Frantini-Silva, W.; Sofia, S.H.; Orsi, M.L.; Almeida, F.S. DNA Barcoding of Freshwater Ichthyoplankton in the Neotropics as a Tool for Ecological Monitoring. *Mol. Ecol. Resour.* **2015**, *15*, 1226–1237. [[CrossRef](#)] [[PubMed](#)]
8. Brodeur, R.; Peterson, W.; Auth, T.; Soulen, H.; Parnel, M.; Emerson, A. Abundance and Diversity of Coastal Fish Larvae as Indicators of Recent Changes in Ocean and Climate Conditions in the Oregon Upwelling Zone. *Mar. Ecol. Prog. Ser.* **2008**, *366*, 187–202. [[CrossRef](#)]
9. Carassou, L.; Ponton, D.; Mellin, C.; Galzin, R. Predicting the Structure of Larval Fish Assemblages by a Hierarchical Classification of Meteorological and Water Column Forcing Factors. *Coral Reefs* **2008**, *27*, 867–880. [[CrossRef](#)]
10. Mahafina, J. Perception et Comportement Des Pêcheurs Pour Une Gestion Durable de La Biodiversité et de La Pêche Récifale Du Sud-Ouest de Madagascar. Ph.D. Thesis, Université de La Réunion et Université de Toliara, Toliara, Madagascar, 2011.
11. Jaonalison, H.; Mahafina, J.; Ponton, D. Fish Post-Larvae Assemblages at Two Contrasted Coral Reef Habitats in Southwest Madagascar. *Reg. Stud. Mar. Sci.* **2016**, *6*, 62–74. [[CrossRef](#)]
12. Chen, L.-C.; Lan, K.-W.; Chang, Y.; Chen, W.-Y. Summer Assemblages and Biodiversity of Larval Fish Associated with Hydrography in the Northern South China Sea. *Mar. Coast. Fish.* **2018**, *10*, 467–480. [[CrossRef](#)]
13. Grande, H.; Rezende, S.d.M.; Simon, T.E.; Félix-Hackradt, F.C.; García-Charton, J.A.; Maida, M.; Gaspar, A.L.B.; Francini-Filho, R.B.; Fredou, T.; Ferreira, B.P. Diversity of Settlement-stage Reef Fishes Captured by Light-trap in a Tropical South-west Atlantic Ocean Coastal Reef System. *J. Fish Biol.* **2018**, *94*, 210–222. [[CrossRef](#)]
14. Pegg, G.G.; Sinclair, B.; Briskey, L.; Aspden, W.J. MtDNA Barcode Identification of Fish Larvae in the Southern Great Barrier Reef—Australia. *Sci. Mar.* **2006**, *70*, 7–12. [[CrossRef](#)]
15. Trivedi, S.; Aloufi, A.A.; Ansari, A.A.; Ghosh, S.K. Role of DNA Barcoding in Marine Biodiversity Assessment and Conservation: An Update. *Saudi J. Biol. Sci.* **2016**, *23*, 161–171. [[CrossRef](#)]
16. Ko, H.-L.; Wang, Y.-T.; Chiu, T.-S.; Lee, M.-A.; Leu, M.-Y.; Chang, K.-Z.; Chen, W.-Y.; Shao, K.-T. Evaluating the Accuracy of Morphological Identification of Larval Fishes by Applying DNA Barcoding. *PLoS ONE* **2013**, *8*, e53451. [[CrossRef](#)] [[PubMed](#)]
17. Meredith, C.; Hoffman, J.; Trebitz, A.; Peterson, G.; Lietz, J.; Hatzembuhler, C.; Pilgrim, E.; Okum, S.; Martinson, J. *Development of DNA-Based Identification Methods to Track the Species Composition of Fish Larvae within Nearshore Areas of the Great Lakes*; Minnesota Chapter of the American Fisheries Society: St. Cloud, MN, USA, 2017.
18. Leis, J.M. Taxonomy and Systematics of Larval Indo-Pacific Fishes: A Review of Progress since 1981. *Ichthyol. Res.* **2015**, *62*, 9–28. [[CrossRef](#)]
19. Bruggemann, J.H.; Rodier, M.; Guillaume, M.M.M.; Andréfouët, S.; Arfi, R.; Cinner, J.E.; Pichon, M.; Ramahatratra, F.; Rasoamandriana, F.; Zinke, J.; et al. Wicked Social-Ecological Problems Forcing Unprecedented Change on the Latitudinal Margins of Coral Reefs: The Case of Southwest Madagascar. *Ecol. Soc.* **2012**, *17*, 47. [[CrossRef](#)]
20. Ranaivomanana, L.N.J. Identification Des Conditions d'appropriation de La Gestion Durable Des Ressources Naturelles et Des Écosystèmes: Cas Du Grand Récif de Toliara. Ph.D. Thesis, Institut Haliéutique et des Sciences Marines (IH.SM) de l'Université de Toliara, Toliara, Madagascar, 2006.
21. Dufour, V. Colonisation Des Recifs Coralliens Par Les Larves de Poissons. Ph.D. Thesis, Université Pierre & Marie Curie, Paris, France, 1992.
22. Marchetti, M.P.; Esteban, E.; Limm, M.; Kurth, R. Evaluating Aspects of Larval Light Trap Bias and Specificity in the Northern Sacramento River System: Do Size and Color Matter? *Am. Fish. Soc. Symp.* **2004**, *39*, 269–279.
23. Lindquist, D.C.; Shaw, R.F. Effects of Current Speed and Turbidity on Stationary Light-Trap Catches of Larval and Juvenile Fishes. *Fish. Bull.* **2005**, *103*, 438–444.
24. Wilson, D. Patterns of Replenishment of Coral-Reef Fishes in the Nearshore Waters of the San Blas Archipelago, Caribbean Panama. *Mar. Biol.* **2001**, *139*, 735–753. [[CrossRef](#)]
25. Anderson, T.W.; Bartels, C.T.; Hixon, M.A.; Bartels, E.; Carr, M.H.; Shenker, J.M. Current Velocity and Catch Efficiency in Sampling Settlement-Stage Larvae of Coral-Reef Fishes. *Fish. Bull.* **2002**, *100*, 404–413.
26. Laroche, J.; Ramanarivo, N. A Preliminary Survey of the Artisanal Fishery on Coral Reefs of the Tulear Region (Southwest Madagascar). *Coral Reefs* **1995**, *14*, 193–200. [[CrossRef](#)]

27. Raharinaivo, L.R.; Jaonalison, H.; Mahafina, J.; Ponton, D. How to Efficiently Determine the Size at Maturity of Small-sized Tropical Fishes: A Case Study Based on 144 Species Identified via DNA Barcoding from Southwestern Madagascar. *J. Appl. Ichthyol.* **2020**, *36*, 402–413. [[CrossRef](#)]
28. Bauchot, M.L.; Bianchi, G. *Fiches FAO d'identification Des Espèces Pour Les Besoins de La Pêche. Guide Des Poissons Commerciaux de Madagascar (Espèces Marines et d'eaux Saumâtres), Avec Le Support Du Programme Des Nations Unies Pour Le Développement (Projet RAF/79/065)*; FAO: Rome, Italy, 1984; 135p.
29. Smith, M.M.; Heemstra, P.C. (Eds.) *Smiths' Sea Fishes*; Springer: Berlin/Heidelberg, Germany, 1986; ISBN 978-3-642-82860-7.
30. Collet, A.; Bosc, P.; Pothin, K.; Chabanet, P. *Guide d'identification Des Post-Larves de Poissons Des Récifs Coralliens*; ARDA et OCEA Consult': La Réunion, France, 2013; 176p.
31. Werner, F.; Fröndt, R. Automated Purification of Genomic DNA from Tissue with the Macherey-Nagel NucleoMag®Tissue Kit on the EpMotion® 5075. *Appl. Note* **2015**, *365*, 1–7.
32. Ward, R.D.; Zemlak, T.S.; Innes, B.H.; Last, P.R.; Hebert, P.D.N. DNA Barcoding Australia's Fish Species. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2005**, *360*, 1847–1857. [[CrossRef](#)] [[PubMed](#)]
33. Ratnasingham, S.; Hebert, P.D.N. A DNA-Based Registry for All Animal Species: The Barcode Index Number (BIN) System. *PLoS ONE* **2013**, *8*, e66213. [[CrossRef](#)] [[PubMed](#)]
34. R Core Team. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2018.
35. Oksanen, J.; Blanchet, F.G.; Friendly, M.; Kindt, R.; Legendre, P.; McGlinn, D.; Minchin, P.R.; O'Hara, R.B.; Simpson, G.L.; Solymos, P.; et al. *Vegan: Community Ecology Package*. 2019. Available online: [https://www.scirp.org/\(S\(351jmbntvnsjt1aadkposzje\)\)/reference/ReferencesPapers.aspx?ReferenceID=1778707](https://www.scirp.org/(S(351jmbntvnsjt1aadkposzje))/reference/ReferencesPapers.aspx?ReferenceID=1778707) (accessed on 12 October 2021).
36. Poulin, R. Comparison of Three Estimators of Species Richness in Parasite Component Communities. *J. Parasitol.* **1998**, *84*, 485–490. [[CrossRef](#)] [[PubMed](#)]
37. Meredith, M. Quick and Dirty Estimates for Wildlife Populations. 2019. Available online: <http://www.r-project.org/> (accessed on 12 October 2021).
38. Smith, E.P.; Van Belle, G. Nonparametric Estimation of Species Richness. *Biometrics* **1984**, *40*, 119–129. [[CrossRef](#)]
39. Pinheiro, H.T.; Rocha, L.A.; Macieira, R.M.; Carvalho-Filho, A.; Anderson, A.B.; Bender, M.G.; Di Dario, F.; Ferreira, C.E.L.; Figueiredo-Filho, J.; Francini-Filho, R.; et al. South-Western Atlantic Reef Fishes: Zoogeographical Patterns and Ecological Drivers Reveal a Secondary Biodiversity Centre in the Atlantic Ocean. *Divers. Distrib.* **2018**, *24*, 951–965. [[CrossRef](#)]
40. Steinke, D.; Connell, A.D.; Hebert, P.D.N. Linking Adults and Immatures of South African Marine Fishes. *Genome* **2016**, *59*, 959–967. [[CrossRef](#)]
41. Reid, K.; Crochelet, E.; Bloomer, P.; Hoareau, T.B. Investigating the Origin of Vagrant Dusky Groupers, *Epinephelus Marginatus* (Lowe, 1834), in Coastal Waters of Réunion Island. *Mol. Phylogenet. Evol.* **2016**, *103*, 98–103. [[CrossRef](#)]
42. Weigt, L.A.; Baldwin, C.C.; Driskell, A.; Smith, D.G.; Ormos, A.; Reyier, E.A. Using DNA Barcoding to Assess Caribbean Reef Fish Biodiversity: Expanding Taxonomic and Geographic Coverage. *PLoS ONE* **2012**, *7*, e41059. [[CrossRef](#)]
43. Healey, A.J.E.; McKeown, N.J.; Taylor, A.L.; Provan, J.; Sauer, W.; Gouws, G.; Shaw, P.W. Cryptic Species and Parallel Genetic Structuring in Lethrinid Fish: Implications for Conservation and Management in the Southwest Indian Ocean. *Ecol. Evol.* **2018**, *8*, 2182–2195. [[CrossRef](#)]
44. Ponton, D.; Carassou, L.; Raillard, S.; Borsa, P. Geometric Morphometrics as a Tool for Identifying Emperor Fish (Lethrinidae) Larvae and Juveniles. *J. Fish Biol.* **2013**, *83*, 14–27. [[CrossRef](#)]
45. Wibowo, A.; Wahlberg, N.; Vasemägi, A. DNA Barcoding of Fish Larvae Reveals Uncharacterised Biodiversity in Tropical Peat Swamps of New Guinea, Indonesia. *Mar. Freshw. Res.* **2017**, *68*, 1079. [[CrossRef](#)]
46. Durand, J.-D.; Pham, M.H.; Tran, T.T.V.; Hoang, D.H.; Van Vo, Q. Sorting the Wheat from the Chaff: A Review of BINs Associated with Groupers of Vietnam and the Implications for Species Identification from DNA Barcoding. *Mar. Biodivers.* **2020**, *50*, 38. [[CrossRef](#)]
47. Collins, R.A.; Cruickshank, R.H. The Seven Deadly Sins of DNA Barcoding. *Mol. Ecol. Resour.* **2013**, *13*, 969–975. [[CrossRef](#)]
48. Semmens, B.; Buhle, E.; Salomon, A.; Pattengill-Semmens, C. A Hotspot of Non-Native Marine Fishes: Evidence for the Aquarium Trade as an Invasion Pathway. *Mar. Ecol. Prog. Ser.* **2004**, *266*, 239–244. [[CrossRef](#)]
49. Blaxter, M. Imagining Sisyphus Happy: DNA Barcoding and the Unnamed Majority. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2016**, *371*, 20150329. [[CrossRef](#)]