Biophysical Features and Local Early Conformational Propensities in Intrinsically **Disordered Regions of Rhabdoviral Glycoproteins**

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Across the Diverse Rhabdoviral Gps

Through meticulous examination of the Gps from the most well-known genera of Rhabdoviridae, a high level of conservation in biophysical and structural properties is apparent among genera. Furthermore, various gradients of secondary structures, phase separation determinants, amnio acid composition and properties display strong intra-genus impact. In contrast, Rabies viruses show weak sequence-based biophysical variations within a wide range of hosts, suggesting importantly this group adopts "biophysical hotspot" allowing efficient multi-host functioning.

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Rhabdoviral Glycoprotein (Gp)

Most harmful rhabdoviruses cause disease that is invariably lethal to humans, animals and plants. Throughout viral infection, protein refolding is a complex process critical to both receptor recognition and membrane-interacting fusion domains mediated by transmembrane rhabdoviral Gp. Little is known about the early stages of contextsensitive structural transitions of the rhabdoviral Gp. To solve this issue, we combined state-of-the-art machine learning-driven biophysical and intrinsic disorder-based metapreditcors.









Local Conformational Propensities in Intrinsically Disordered Regions (IDRs)

Gp are essential for virion attachment and penetration of host cells prior to transmission. Here, we investigate the host-specific biophysical IDRs-determinants encoded in the primary amino acid sequence of rhabdoviral Gps, which are predicted to modulate early conformational events. This can be related to host-specific biophysical features of the local backbone flexibility and secondary structure propensities mostly close to or in the C-terminal IDRs.



Determinant



High Conservation and Specificity in Biophysical Determinants

Importantly, the essential plant rhabdoviral Gp is still PFAM-unassigned and poorly characterized. While showing striking differences between IDRs of N- and C-termini and the core, our results give direct insights into the biophysical folding signals located in these variable regions and are in congruence with independent epidemiological observations.

On a quantitative scale, these termini-residues contain biophysical features and sequence segregation compositional biases, such as hydrophobicity, charges and aromaticity, involved mostly in binding interactions and display evolutionary covariation, reflecting a general tendency toward host-specificity.

Hence, an in-depth comparative structural study of the conserved core protein and the C-terminal IDRs patterns would most likely allow the robust identification and assignment of a new PFAM domain candidate. This analysis highlights these regions' functional significance and potential roles in the viral life cycle, host interactions, and viral adaptation across different hosts.

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Linking Biophysical Features to the **Biological Context**

Using supervised uniform manifold approximation and projection (UMAP) together with robust gradient boosting regressors (XGBoost) helped unravel the most relevant features.

Therefore, characterizing the impact of secondary structures, sidechains and conformational properties in IDRs could help elucidate vectorspecific transmission differences.

Conclusion and Perspectives

In conclusion, the accurate and statistically significant connection of structural features with both IDRs conformational propensities and context-sensitive folding data suggests their critical role in local biophysics with lasting effects on subsequent conformational changes during virus-host interactions and disease-related pathogenicity outcomes.

Our results also highlight the importance of a comprehensive understanding of the combined effect of the conserved structural patterns in the core protein domain and the biophysical features of the genus-specific Cterminal IDRs among rhabdoviral Gps, such as, for instance, in novel plant-virus-vectors associations.

| Entry | Genus | Species | Prediction |
|------------|-------------------|--|------------|
| A0A2D2PYL4 | Cytorhabdovirus | Cabbage cytorhabdovirus 1 | Aphid |
| A0A482PGH7 | Cytorhabdovirus | Raspberry vein chlorosis virus | Mite(?) |
| A0A482PGU2 | Cytorhabdovirus | Raspberry vein chlorosis virus | Aphid |
| A0A6F9EYP3 | Cytorhabdovirus | Trichosanthes associated rhabdovirus 1 | Aphid |
| A0A7G3W8J0 | Cytorhabdovirus | Medicago cytorhabdovirus A | Aphid |
| A0A7T7FQX1 | Cytorhabdovirus | Kenyan potato cytorhabdovirus | Aphid |
| A0A7T7FQY6 | Cytorhabdovirus | Kenyan potato cytorhabdovirus | Aphid |
| ΑΟΑ7Τ7ЈΡΚΟ | Cytorhabdovirus | Kenyan potato cytorhabdovirus | Aphid |
| A0A8E6YK00 | Cytorhabdovirus | Actinidia cytorhabdovirus JS27 | Aphid |
| A0A8F4M7Q7 | Cytorhabdovirus | Passionfruit-associated rhabdovirus YN | Whitefly |
| A0A1D8FVH5 | Nucleorhabdovirus | Physostegia chlorotic mottle virus | Leafhopper |
| A0A1Y0JW81 | Nucleorhabdovirus | Physostegia chlorotic mottle virus | Leafhopper |
| A0A4P2UVB8 | Nucleorhabdovirus | Green Sichuan pepper nucleorhabdovirus | Aphid |
| A0A4Y6GLZ0 | Nucleorhabdovirus | Physostegia chlorotic mottle virus | Leafhopper |



