DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



THERAPEUTIC OPTIONS FOR CTLA-4 INSUFFICIENCY

David Egg, Ina Caroline Rump, MD, Noriko Mitsuiki, MD, PhD, Jessica Rojas-Restrepo, Maria-Elena Maccari, MD,^{a,b} Charlotte Schwab, MD,^a Annemarie Gabrysch, MD,^a Klaus Warnatz, MD,^{a,c} Sigune Goldacker, MD, a,c Virginia Patiño, MD, Daniel Wolff, MD, Satoshi Okada, MD, PhD, Seiichi Hayakawa, MD, Yoshiaki Shikama, MD, Kenji Kanda, MD, Kohsuke Imai, MD, PhD, Manabu Sotomatsu, MD, Makoto Kuwashima, MD,^k Takahiro Kamiya, MD, PhD,^l Tomohiro Morio, MD, PhD,^m Kazuaki Matsumoto, MD,^m Takeshi Mori, MD, PhD,ⁿ Yuri Yoshimoto, MD,^o Ingunn Dybedal, MD, PhD,^p Maria Kanariou, MD, Zeynep Yesim Kucuk, MD, Hugo Chapdelaine, MD, Lenka Petruzelkova, MD, PhD, t Hanns-Martin Lorenz, MD, Wathleen E. Sullivan, MD, PhD, Jennifer Heimall, MD, Michel Moutschen, MD, PhD, Jiri Litzman, MD, Mike Recher, MD, Michael H. Albert, MD, Fabian Hauck, MD, PhD, aa Suranjith Seneviratne, MD, PhD, bb Jana Pachlopnik Schmid, MD, PhD, cc Antonios Kolios, MD, dd Gary Unglik, MBBS, ee Christian Klemann, MD, a,b Scott Snapper, MD, PhD, ff Lisa Giulino-Roth, MD, gg Michael Svaton, MD, hh Craig D. Platt, MD, PhD, Sophie Hambleton, FRCPCH, DPhil, Olaf Neth, MD, PhD, kk Geraldine Gosse, MSc, Steffen Reinsch, MD, Dirk Holzinger, MD, Yae-Jean Kim, MD, PhD, O Shahrzad Bakhtiar, MD,^{pp} Faranaz Atschekzei, MD, PhD,^{qq} Reinhold Schmidt, MD,^{qq} Georgios Sogkas, MD, PhD, qq Shanmuganathan Chandrakasan, MD, r William Rae, MD, ss,t Beata Derfalvi, MD, PhD, uu, Hanne Vibeke Marquart, MD, PhD, Ahmet Ozen, MD, Ayca Kiykim, MD, Elif Karakoc-Aydiner, MD, ww Pavlína Králíčková, MD, PhD, xx Godelieve de Bree, MD, PhD, yy Dimitra Kiritsi, MD, zz Markus G. Seidel, MD, aaa Robin Kobbe, MD, bbb Jennifer Dantzer, MD, MHS, ccc Laia Alsina, MD, PhD, ddd Thais Armangue, MD, PhD, eee Vassilios Lougaris, MD, fff Philipp Agyeman, MD, ggg Sofia Nyström, MD, PhD, hhh David Buchbinder, MD, iii Peter D. Arkwright, MD, PhD, iii and Bodo Grimbacher, MD^{a,c,bb,kkk,lll,mmm,nnn}

Freiburg, Regensburg, Heidelberg, Munich, Jena, Essen, Frankfurt am Main, Hannover, and Hamburg, Germany; Montevideo, Uruguay; Hiroshima, Yokohama, Shiga, Tokyo, Shibukawa, Kiryū, and Kobe, Japan; Oslo, Norway; Athens, Greece; Cincinnati, Ohio; Montreal, Quebec, and Halifax, Nova Scotia, Canada; Prague, Brno, and Hradec Kralove, Czech Republic; London, Newcastle upon Tyne, Southampton, and Manchester, United Kingdom; Zurich and Bern, Switzerland; Melbourne, Australia; Boston, Mass; New York, NY; Sevilla and Barcelona, Spain; Seoul, Korea; Atlanta, Ga; Copenhagen, Denmark; Istanbul, Turkey; Amsterdam, The Netherlands; Graz, Austria; Baltimore, Md; Brescia, Italy; Linköping, Sweden; and Orange, Calif

From ^a the Institute for Immunodeficiency, Center for Chronic Immunodeficiency, ^bthe Division of Pediatric Hematology and Oncology, Department of Pediatrics and Adolescent Medicine, ^cthe Department of Rheumatology and Clinical Immunology, ^{zz}the Department of Dermatology, and ^{mmm} the Centre for Integrative Biological Signaling Studies, Medical Center, Faculty of Medicine, Albert Ludwig University of Freiburg; ^dthe Immunology Team, American Insurance, Montevideo; ^ethe Department of Internal Medicine III, University Hospital Regensburg; ^fthe Department of Pediatrics, Hiroshima University Graduate School of Biomedical and Health Sciences; ^gthe Division of Infection, Immunology and Infection, Kanagawa Children's Medical Center, Yokohama, ^hthe Department of Pediatrics, Hikone

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



Municipal Hospital, Shiga; 'the Department of Community Pediatrics, Perinatal and Maternal Medicine, Tokyo Medical and Dental University; ithe Department of Hematology/Oncology, Gunma Children's Medical Center, Shibukawa; kthe Department of Pediatrics, Kiryu Kosei General Hospital, Kiryu; Ithe Department of Lifetime Clinical Immunology and mthe Department of Pediatrics and Developmental Biology, Tokyo Medical and Dental University, Tokyo; "the Department of Hematology and Oncology, Hyogo Prefectural Kobe Children's Hospital; othe Department of Pediatrics, Center Hospital of the National Center for Global Health and Medicine, Tokyo; pthe Department of Hematology, Oslo University Hospital; 4the Department of Immunology and Histocompatibility, Center for Primary Immunodeficiencies-Paediatric Immunology, Aghia Sophia Children's Hospital, Athens; 'the Division of Bone Marrow Transplantation and Immune Deficiency, Children's Hospital Medical Center, Cincinnati; sthe Division of Clinical Immunology, Montreal Clinical Research Institute; the Department of Paediatrics, Motol University Hospital, Second Medical Faculty in Prague, Charles University; "the Division of Rheumatology, Department of Internal Medicine V, University of Heidelberg; "The Children's Hospital of Philadelphia, Perelman School of Medicine, University of Pennsylvania; ™the Department of Infectious Diseases and General Internal Medicine, University Hospital of Liege; *the Department of Clinical Immunology and Allergology, Medical Faculty, Masaryk University, Brno; 'the Department of Clinical Immunology and Allergology, St. Anne's University Hospital, Brno; ^zthe Immunodeficiency Clinic, Medical Outpatient Unit and Immunodeficiency Lab, Department Biomedicine, University Hospital, Basel; at the Department of Pediatrics, Dr von Hauner Children's Hospital, University Hospital, Ludwig Maximilians Uni- versitat Munchen; bbthe Institute of Immunology and Transplantation, Royal Free Hospital, University College London; "the Division of Immunology, University Children's Hospital Zurich, and ddthe Department of Immunology, University Hospital Zurich, University of Zurich; eethe Department of Clinical Immunology and Allergy, Royal Melbourne Hospital; "the Division of Pediatric Gastroenterology, Hepatology, and Nutrition and "the Division of Immunology, Boston Children's Hospital and Department of Pediatrics, Harvard Medical School; 99the Department of Pediatrics, Division of Pediatric Hematology/Oncology, Weill Cornell Medicine, New York; hhChildhood Leukaemia Investigation Prague, Department of Paediatric Haematology and Oncology, Second Faculty of Medicine, Charles University and University Hospital Motol, Prague; "Ithe Great North Children's Hospital, Newcastle upon Tyne Hospitals NHS Foundation Trust, and Newcastle University Translational and Clinical Research Institute; kkthe Pediatric Infectious Diseases, Rheumatology and Immunology Unit, Hospital Virgen del Rocío/Instituto de Biomedicina de Sevilla, Sevilla, RECLIP; "the Montreal Clinical Research Institute, Universite de Montreal; mm the Jena University Hospital, Pediatric Gastroenterology; ""the Department of Pediatric Hematology-Oncology, University of Duisburg-Essen; oothe Division of Infectious Diseases and Immunodeficiency, Department of Pediatrics, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul; ppthe Division of Stem Cell Transplantation and Immunology, Department of Children and Adolescents, University Hospital Frankfurt, Goethe University Frankfurt, Frankfurt am Main; ⁹⁹the Department for Clinical Rheumatology and Immunology, Hannover Medical School; "the Division of Bone Marrow Transplant, Aflac Cancer and Blood Disorders Center, Children's Healthcare of Atlanta, Emory University School of Medicine, Atlanta; ssthe Department of Immunology, University Hospital Southampton NHSFT; ttthe Southampton National Institute for Health Research Clinical Research Facility, University Hospital Southampton

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



NHSFT; "the Division of Immunology, IWK Health Centre and Dalhousie University, Department of Pediatrics, Halifax; "the Department of Clinical Immunology, Rigshospitalet, University of Copenhagen; wwthe Marmara University School of Medicine, Division of Pediatric Allergy and Immunology, Istanbul; **the Institute of Clinical Immunology and Allergy, University Hospital Hradec Kra- love, Charles University, Faculty of Medicine in Hradec Kralove; "ythe Department of Internal Medicine, Division of Infectious Diseases, Amsterdam Institute for Infection and Immunity, Amsterdam University Medical Center, University of Amsterdam; and the Research Unit for Pediatric Hematology and Immunology, Division of Pediatric Hemato-Oncology, Department of Pediatric and Adolescent Medicine, Medical University Graz; bbb the Division of Infectious Diseases, First Department of Medicine, University Medical Center Hamburg-Eppendorf; ccthe Division of Pediatric Allergy and Immunology, and Rheumatology, Department of Pediatrics, John Hopkins University School of Medicine, Baltimore; dddthe Clinical Immunology and Primary Immunodeficiencies Unit, Pediatric Allergy and Clinical Immunology Department, Hospital Sant Joan de Deu and Institut de Recerca Sant Joan de Deu, Barcelona; Clinical Immunology Unit Hospital Sant Joan de Deu-Hospital Clinic Barcelona, Universitat de Barcelona; eeethe Neuroimmunology Program, Institut d'Investigacions Biomediques August Pi i Sunyer, Hospital Clinic, Universitat de Barcelona; ffthe Pediatric Neuroimmunology Unit, Neurology Department, Sant Joan de Deu Children's Hospital, University of Barcelona; ⁹⁹⁹the Pediatrics Clinic and Institute for Molecular Medicine A. Nocivelli, Department of Clinical and Experimental Sciences, University of Brescia, ASST-Spedali Civili of Brescia; hhhthe Department of Pediatrics, Inselspital, Bern University Hospital, University of Bern; "the Department of Clinical Immunology and Transfusion Medicine, and Department of Biomedical and Clinical Sciences, Linkoping University; ""the Division of Pediatric Hematology, Children's Hospital of Orange County, Orange; kkkthe Lydia Becker Institute of Immunology and Inflammation, University of Manchester, Royal Manchester Children's Hospital; "the German Center for Infection Research, Satellite Center Freiburg; nnn the RESISTCluster of Excellence 2155 to Hannover Medical School, Satellite Center Freiburg.

Supported by the Deutsche Forschungsgemeinschaft (grant SFB1160/2_B5), under Germany's Excellence Strategy (CIBSS-EXC-2189 [project identifier 390939984] and RESIST-EXC 2155 [project identifier 39087428] [to B.D.]); the E-Rare Program of the European Union, managed by the Deutsche Forschungsgemeinschaft (grant code GR1617/14-1/iPAD [to B.D.]); the Netzwerke Seltener Erkrankungen of the German Ministry of Education and Research (grant code GAIN 01GM1910A [to B.D.]); the Ministry of Health of the Czech Republic (grant code NV19-05-00332 [to M.S.]); the Torrons Vicens Foundation (grant code PFNR0144 [to L.A. and T.A.]); the Else Kroner-Fresenius-Stiftung (EKFS, 2017_A110 [to F.H.]), and the German Federal Ministry of Education and Research (BMBF, 01GM1910C [to F.H.]); and the Cancer Research Center Funding Program (CRC, C3Z) of the University of Zurich, Zurich, Switzerland (to J.P.S.).

Disclosure of potential conflict of interest: The authors declare that they have no relevant conflicts of interest.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



Corresponding author: Bodo Grimbacher, MD, Center for Chronic Immunodeficiency, Universitatsklinikum Freiburg, Breisacher Straße 115, 79106 Freiburg, Germany. E-mail: bodo.grimbacher@uniklinik-freiburg.de.

KEYWORDS: CTLA-4, common variable immunodeficiency, primary immunodeficiency, diagnosis, treatment, abatacept, sirolimus, LRBA, rituximab, HSCT

ABBREVIATIONS USED

AIHA: Autoimmune hemolytic anemia

CNS: Central nervous system

CSF: Cerebrospinal fluid

CVID: Common variable immunodeficiency

CTLA-4: Cytotoxic T lymphocyte-associated antigen-4

DEF6: Differentially expressed in FDCP6 homolog

GFP: Green fluorescent protein

GLILD: Granulomatous-lymphocytic interstitial lung disease

HSCT: Hematopoietic stem cell transplantation

ICOS: Inducible costimulator

IEI: Inborn error of immunity

IPEX: Immune dysregulation, polyendocrinopathy, enteropathy, X-linked

IgRT: Immunoglobulin replacement therapy

ITP: Immune thrombocytopenia

IVIG: Intravenous immunoglobulin

LRBA: Lipopolysaccharide-responsive beige-like anchor protein

MOG: Myelin oligodendrocyte glycoprotein

MRI: Magnetic resonance imaging

PRCA: Pure red cell aplasia

TPO-RA: Thrombopoietin receptor agonist

Treg: Regulatory T

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



BACKGROUND: Heterozygous germline mutations in cytotoxic T lymphocyte–associated antigen-4 (CTLA4) impair the immunomodulatory function of regulatory T cells. Affected individuals are prone to life-threatening autoimmune and lymphoproliferative complications. A number of therapeutic options are currently being used with variable effectiveness.

OBJECTIVE: Our aim was to characterize the responsiveness of patients with CTLA-4 insufficiency to specific therapies and provide recommendations for the diagnostic workup and therapy at an organ-specific level.

METHODS: Clinical features, laboratory findings, and response to treatment were reviewed retrospectively in an international cohort of 173 carriers of CTLA4 mutation. Patients were followed between 2014 and 2020 for a total of 2624 months from diagnosis. Clinical manifestations were grouped on the basis of organ-specific involvement. Medication use and response were recorded and evaluated.

RESULTS: Among the 173 CTLA4 mutation carriers, 123 (71%) had been treated for immune complications. Abatacept, rituximab, sirolimus, and corticosteroids ameliorated disease severity, especially in cases of cytopenias and lymphocytic organ infiltration of the gut, lungs, and central nervous system. Immunoglobulin replacement was effective in prevention of infection. Only 4 of 16 patients (25%) with cytopenia who underwent splenectomy had a sustained clinical response. Cure was achieved with stem cell transplantation in 13 of 18 patients (72%). As a result of the aforementioned methods, organspecific treatment pathways were developed. Conclusion: Systemic immunosuppressants and abatacept may provide partial control but require ongoing administration. Allogeneic hematopoietic stem cell transplantation offers a possible cure for patients with CTLA-4 insufficiency. (J Allergy Clin Immunol 2022;149:736-46.)

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



Heterozygous germline mutations in cytotoxic T lymphocyte- associated antigen-4 (CTLA4) cause an immune dysregulation syndrome in humans that is termed *CTLA-4 insufficiency.*^{1,2} The disease was first described in 2014, and the number of published cases has risen rapidly.¹⁻³ The diagnosis is made by gene sequencing and study of CTLA-4 function and expression.

Loss-of-function mutations in *CTLA4* result in immune activation by impairing the negative regulation of T-cell activation.² The regulatory role of CTLA-4 was first described in *CTLA4* knockout mice, which showed fatal autoimmunity caused by an expansion of activated T cells.⁴ Patients with heterozygous *CTLA4* mutations present a similar but less severe phenotype with incomplete penetrance.^{1,2}

The phenotype consists of hypogammaglobulinemia associated with recurrent respiratory infections, autoimmune features (cytopenias, (poly)endocrinopathy, and type-1 diabetes), and lymphoproliferation, which can be malignant. Lymphoid or granulomatous infiltrations can be found in the lung, brain, gut, spleen, liver, kidney, and skin. Additionally, *CTLA4* mutation carriers suffer from chronic inflammatory bowel and rheumatic diseases. Currently, therapy focuses on T- or B-cell-specific immunosuppression, including biologics, whereas stem cell transplantation may be curative.^{3,5} However, treatment regimens or symptom-specific recommendations are missing.

Here, we present the detailed outcomes of 123 symptomatic carriers of *CTLA4* mutation. On an organ-specific level, we review the effectiveness of the different therapies used. Our therapeutic regimens were driven by literature searches and our retrospective treatment evaluation, aiming for improved treatment options for our patients. We add new pathogenic mutations and demonstrate the need for functional studies of new gene variants in *CTLA4*.

METHODS

PATIENT COHORT

We collected data on a worldwide cohort of 173 *CTLA4* mutation carriers over a 4-year period (July 2014 to September 2017). The data capture reported by Schwab et al³ ended in September 2016. We continued data collection with a focus on treatment and its outcome. Hence, all 133 individuals in the work of Schwab et al³ and their allocated identifiers are identical to the first 133 patients of this report (see Table E1 in the Online Repository at www.jacionline.org).

The current cohort consists of 81 families. The individuals were between the ages of 1 and 73 years (88 females and 85 males, witha median age of onset of 13 years and median age of evaluation of 24 years [for details, see Table E1]). Of the entire cohort, 123 individuals (penetrance of71%) had symptoms related to CTLA-4 insufficiency requiring treatment.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



DATA COLLECTION

All contributing physicians were provided with a detailed questionnaire, usually by mail contact. We recorded the mutation carriers' phenotype and their clinical history, including therapy. Within 173 mutation carriers, 54 different mutations were reported. For all 54 mutations, we collected data on pathogenicity from the literature, our own data, and the ClinVar database (https://www.ncbi.nlm.nih.gov/clinvar/) (see Table E1).

For those mutation carriers who were registered before September 2016 (n = 133), we contacted the collaborating physicians for a second time and sent a specific follow-up form to obtain the most recent clinical data and treatment outcome, so that the patient's full clinical history was validated and all treatment relevant data and existing information were updated.

Three individuals were excluded from the analyses: 1 with a large chromosome 2 deletion (2q33.2-2q33.3; including the genes *CD28* and inducible costimulator *[ICOS]*), possibly confounding the phenotype; 1 with the missense variant p.N145S; and 1 with the missense variant p.T207A (located in exon 4 of *CTLA4*). For both missense variants, the results of functional testing (CTLA-4 surface expression and CTLA-4-mediated transendocytosis) were normal.

ETHICS

The study was carried out in accordance with the recommendations of the scientific committee at the University Medical Center of Freiburg for studies involving human subjects. All physicians confirmed that their patients had signed an informed consent under local ethics-approved protocols and in accordance with the Declaration of Helsinki. The study protocol was approved by the ethics committee of the University Medical Center of Freiburg (approval nos. 295/13_140782 and 60/18). No financial incentive was provided to the patients or the contributing physicians. Data were reported after having been pseudonymized, and physician-to-physician contact allowed communication of treatment results and advice.

TREATMENT OUTCOME

With regard to the 123 clinically symptomatic individuals, we focused on treatment with immunosuppressants, disease-modifying antirheumatic drugs, biologics, antibiotics, splenectomy, and hematopoietic stem cell transplantation (HSCT). For all drugs, we requested indication, dosage and interval, side effects, and treatment response based on the physician's judgment. A total of 50 unaffected mutation carriers were analyzed regarding their clinical phenotype.

We created groups according to the most affected organ systems (hematologic; hypogammaglobulinemia; and lymphoid, lung, gastrointestinal, neurologic, and skin involvement).

The patient's phenotype determined the organ group; patients affected by multiple organs were part of multiple organ groups independently of whether the manifestations occurred concurrently or consecutively. In fact, immunosuppressive drugs were often administered to alleviate

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



multisystemic manifestations and were therefore listed for multiple organ groups despite given only once.

GENERATION OF MANAGEMENT CHARTS

To generate organ-specific diagnosis and treatment flowcharts, we took the following steps, as suggested by the World Health Organization for the generation of consensus documents: (1) we defined 7 groups of the most important clinical manifestations of CTLA4-4 insufficiency; (2) we identified existing guidelines and systematic reviews in the literature for specific symptoms; (3) we generated a draft topic list of opportunities for treatment/ diagnosis improvements in CTLA4-4 insufficiency and created the first draft charts; (4) we performed an internal review (by D.E., B.G., and K.W.) of the draft management charts and revised them accordingly; (5) we called for an external review by all 73 coauthors in an initial contact (via e-mail); (6) 7 coauthors (D.E., B.G., K.W., D.B., P.A., L.A, and T.A.) volunteered to form a task force for the improvement of the management charts; (7) the task force discussed, changed, and revised the management charts; (8) we called for a second external review (via mail) by all 73 coauthors (1 coauthor could not agree on the management protocols and therefore asked to be removed from the list of authors); (9) we performed another internal review for consistency and obtained approval from all coauthors for the final management charts; and (10) the consented management charts were submitted to the peer review process of the *Journal of Allergy and Clinical Immunology*.

Taken together, the charts are based on a literature search, retrospective treatment evaluation, and real-life experience of contributing authors. This process is actually in conformity with the World Health Organization guidelines for development of expert opinion-guided management guidelines with evidence level 4. The charts therefore represent the opinion of an expert panel based on a nonrandomized case series; hence the use of evidence level 4 (of 5) with a recommendation grade of C.

CTLA-4 STAINING AND TRANSENDOCYTOSIS ASSAY

When blood samples from patients carrying variants of uncertain significance in CTLA-4 were available, intracellular CTLA-4 expression and CTLA-4-dependent transendocytosis were evaluated by flow cytometry, as described by Schubert et al.¹ Briefly, primary human CD4¹T cells from patients and controls were purified from PBMCs by negative selection following activation with CD3/CD28 beads (Invitrogen) for 16 hours in the presence of either Chinese hamster ovary cells stably expressing CD80-green fluorescent protein (GFP) or mouse embryonic fibroblasts stably expressing CD80-mScarlet. After 16 hours of incubation, cells were harvested and stained for extracellular markers with anti-CD4-PercPCy5.5 (Invitrogen) and anti- CD45RO-PECy7 (Invitrogen). Following fixation and permeabilization, cells were stained with anti-FOXP3-PE (Invitrogen) and ant.iCTLA-4BV421 (BD Bioscience), and later acquired on a FACS Canto II flow cytometer. Analyses of CTLA-4 expression and CD80-GFP/mScarlet-uptake were evaluated in Tregs gated as CD4+CD45RO+FOXP3+CTLA4+ by using FlowJo 7.6.5 software (TreeStar Inc, Ashland, Ore).

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



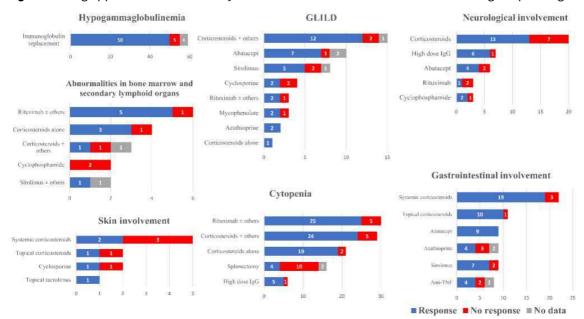


Figure 1. Drug applications in a clinically affected CTLA4 mutation carrier ordered in organ-specific groups.

STATISTICAL ANALYSIS

All statistics and figures were computed and created by using R (version 3.4.1), Microsoft Excel (2016), or FlowJo 7.6.5 (TreeStar Inc). The drug- related outcome was listed in 7 contingency tables to display how different treatments have produced different outcomes (classified as response, no response, or no data [see Table E2 in the Online Repository at www.jacionline.org]). The Fisher exact test was performed to test for independence as to whether treatment affected the outcome (alpha = 5%; 95% CI = 0.95). The percentage of responses was computed by dividing the number of positive effects by the number of total administrations.

RESULTS

PATIENT COHORT

We are reporting on 173 individuals bearing 54 distinct *CTLA4* mutations. Data on 139 patients have already been published.^{3,5-10} Hence, 34 individuals are described here for the first time. The clinical penetrance of the cohort was mutation independent and reached 71% (123 of 173) (see Fig E1 in the Online Repository at www.jacionline.org). The median age at diagnosis was 26 years. At data lock, 18 of 123 symptomatic mutation carriers were younger than 16 years. The clinical penetrance was not a function of age and even decreased to 52% in the cohort of mutation carriers aged 40 years and older, as 23 affected mutation carriers have died at a median age of 24 years. Patients were followed for a total of 47,340 months (median 344 months) since birth and 2,624 months (median 20 months) since molecular diagnosis.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



Treatment details were available for 117 (95%) of the 123 clinically symptomatic patients in our cohort. Most of these patients (111of 123 [90%]) had more than 1 organ system affected, reflecting the complex clinical phenotype of CTLA-4 insufficiency. The number of organ systems involved did not increase significantly at the follow-up visits compared with at the time at diagnosis. We constructed 7 groups with typical manifestations and analyzed their therapy outcomes (Fig 1). Liver involvement, including autoimmune hepatitis, fibrosis, and lymphocytic infiltrations, had been previously underestimated, as it occurred in 18 affected mutation carriers (15%). Regarding the treatment with abatacept (CTLA-4 Fc), we now are reporting on 29 patients (including 13 updated reports from Schwab et al³), as detailed in this article.

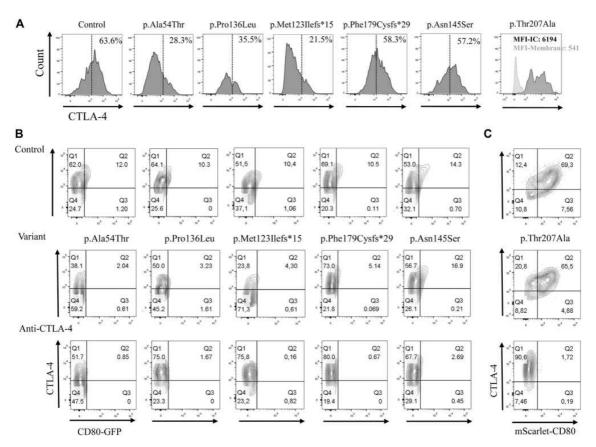
VARIANTS AND ASSAYS

Among the 34 individuals not previously reported in a publication, we documented 6 novel variants: p.L47P, p.A86fs, p.L133P, p.Q148*, p.Y150X, and p.S172W. We performed functional testing on 6 additional variants (all previously untested): p.A54T, p.M123Ifs*15, p.P136L, p.N145S, p.F179Cfs*29, and p.T207A. We measured CTLA-4 expression levels and CTLA-4-mediated transendocytosis (Fig 2). Regulatory T (Treg) cells with p.A54T, p.P136L, and p.M123Ifs*15 mutations had a reduced CTLA-4 expression, whereas the CTLA-4 expression levels in Treg cells with the p.F179Cfs*29, p.N145S, and p.T207A mutations were comparable to those in healthy controls (Fig 2, A). Although Treg cells with the variant p.F179Cfs*29 had normal CTLA-4 expression, the frequency of CD80 uptake in Treg cells was as reduced as in 3 additional deleterious variants (Fig 2, B). In contrast, the p.N145S and p.T207A variants showed a normal CD80 uptake and normal CTLA-4 expression (Fig 2, B and C) and were hence considered benign and not included in our cohort.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



Figure 2. CTLA-4 expression and relative CD80-GFP uptake in *CTLA4* mutation carriers and 1 wild-type control. **A,** Treg cellswith p.A54T, p.P136L, and p.M123Ifs*15showed reduced CTLA-4 expression. The expression levels of CTLA-4 in Treg cells of the patients with p.F179Cfs*29, p.N145S, and p.T207A mutationswere com- parableto those in the healthycontrols. **B,** Thefrequencyof CD80-GFP uptakewas reduced in Treg cells with p.A54T, p.P136L, p.M123Ifs*15, and p.F179Cfs*29 mutations. In contrast, p.N145S showed CD80-GFP uptake comparable to that in the healthy control. **C,** The frequency of mouse embryonic fibroblast (MEF)- CD80-Scarlet uptake was comparable to that in the healthy control. The data shown are representative of 2 experiments



TREATMENT

HSCT. Of our 173 patients, 18 underwent HSCT, including 12 who were initially reported by Schwab et al and whose condition has been updated in this article,³ which already included the 6 patients from Slatter et al.⁵ Of these 18 patients, 13 are alive, well, and no longer taking any medications, the only exception being 1 patient who is undergoing immunoglobulin replacement therapy (IgRT) (see Table E3 in the Online Repository at www. jacionline.org). The median time between the age of onset and the transplantation procedure was 6 years. The median followup after the transplantation procedure was 634 days. The indications for HSCT were refractory autoimmune cytopenias, lymphoid infiltration in nonlymphoid organs, severe enteropathy, or malignancy (in decreasing frequency). A total of 5 patients died: 2 of graft-versus-host disease, 1 of acute respiratory distress syndrome, 1 of multiple organ failure, and 1 of diabetic ketoacidosis unrelated to the HSCT.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



Hypogammaglobulinemia. Of the 123 affected patients, 94 (76%) had hypogammaglobulinemia, of whom 63 (76%) experienced recurrent respiratory infections. Of these 63 patients 59 received IgRT (0.4-0.6 g/kg every 3-4 weeks). In 50 (91%) of the 59 patients who received IgRT, susceptibility to respiratory infections clearly improved, as measured by the number of infections (see Table E2, A). Antibiotic prophylaxis was given in 48 (39%) of the 123 cases. The most used antibiotic for prophylaxis was trimethoprim-sulfamethoxazole (in 24 patients) followed by azithromycin (in 6 patients). Penicillin was used prophylactically in 3 patients after splenectomy. In 7 of the 59 patients (11%), side effects of the IgRT such as fever or rash were observed, but they were controlled either by premedication with systemic steroids or by switching to subcutaneous application. In addition to the 59 patients with primary hypogammaglobulinemia, 8 patients in our cohort received IgRT because of secondary loss of antibodies after therapy with rituximab.

Abnormalities in bone marrow and secondary lymphoid organs. Of the 123 affected patients, 90 (73%) developed noninfectious lymphoid disorders. The most common clinical manifestations were lymphoid expansion with splenomegaly (in 69 of 123 patients), lymph node enlargement (in 51 of 123), and hepatosplenomegaly (in 22 of 123). Lymphocytic infiltrations of organs were reported in 66 of 123 patients (54%). Lymphocytic infiltrations and lymphadenopathy with or without hepatomegaly and/or splenomegaly were treated only when associated with organ dysfunction, particularly with autoimmune cytopenias. Seven patients received corticosteroids (5-20 mg per day), with 4 patients (57%) responding favorably. Six patients received rituximab (2-4 X 375 mg/m²), with 5 positive responses (83%); 2 patients received sirolimus (2-4 mg per day); with a response in 1 of them; and 2 patients had cyclophosphamide (2 X 500 mg/m² and 2 X 200 mg/m², respectively) without response (see Table E2, B). In 1 patient lymphocytic infiltration in the bone marrow was treated with corticosteroids (10 mg per day) with transient improvement, which was more sustained after the corticosteroids were combined with sirolimus (2 mg per day). Five patients received abatacept because of lung and brain infiltrations and are discussed in the appropriate sections.

Cytopenias. In all, 77 patients (62%) were symptomatic; of these 77 patients, 45 developed immune thrombocytopenia (ITP [formerly known as idiopathic thrombocytopenic purpura]), 43 developed Evans syndrome, 35 developed autoimmune hemolytic anemia (AIHA),17 developed pancytopenia, 7 developed autoimmune neutropenia, and 3 developed pure red cell aplasia (PRCA). A total of 50 patients were treated with corticosteroids (mostly at doses of 0.25-1 mg/kg of prednisolone per day with slow wean), with 43 of them showing at least transient response (89%). A total of 30 patients received rituximab (1-4 X 375 mg/ m²), of whom 25 (83%) responded; 3 patients received an additional course of rituximab within 2 or 6 months on account of severe cytopenia and responded well; and 6 patients received immunoglobulin at immunomodulatory doses (1 g/kg for 1-2 days), with positive responses in 5 (83%). Of the cohort patients, 16 had a splenectomy (median age 20 years), with only 4 (25%) showing a sustained response (see Table E2, C). Abatacept was reported to be helpful in 1 patient with chronic ITP, in 1 patient with chronic AIHA, and in 1 patient with chronic PRCA (125 mg per week subcutaneously). Of the 77 symptomatic patients, 32 (42%) were receiving IgRT before the presentation of autoimmune cytopenia, indicating that IgRT does not protect against this complication in CTLA-4 insufficiency, although that is thought to be the case for patients with common variable immunodeficiency (CVID).¹¹

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



GLILD. Of the123 affected patients, 44 (36%) had symptomatic granulomatous-lymphocytic interstitial lung disease (GLILD) needing treatment (in 27 patients [61%], the disease was confirmed on biopsy). Symptoms included dyspnea, cough, and exercise intolerance. Bronchiectasis was present in 25 of 123 patients (20%). Therapeutic outcome was typically evaluated radiologically (by high-resolution computer tomography), physically (on the basis of lung function), and clinically by the physicians.

In all, 25 patients received IgRT (0.4-0.8 g/kg every 3-4 weeks), either for low IgG or specifically as part of the treatment for GLILD. A total of 16 patients received corticosteroids (mostly 5-20 mg per day, with 6 patients initially receiving 40-60 mg per day with prolonged tapering) with 13 positive responses (82%). Abatacept (sometimes with a loading dose of 500-1000 mg intravenously and then typically 125 mg per week subcutaneously) was used in 10 patients. Full resolution occurred in 5 patients, and a partial response occurred in an additional 2 patients (70%). Sirolimus (2-4 mg per day) was administered to 8 patients, with resolution of lesions in 5 of them (62%). Rituximab (3-4 X 375 mg/m²) was given to 3 patients for systemic lymphoproliferation and GLILD. The lung lesions receded but did not resolve in any of the 3 cases. Azathioprine (3 X 50 mg per day) was given to 2 patients, both of whom showed a reduction of interstitial lung disease. Mycophenolate (1000 mg per day) was given to 3 patients and 2 patients showed a reduction of the extent of GLILD (see Table E2, D).

Gastrointestinal involvement. Of 123 patients, 74 (60%) had gastrointestinal involvement, 72 with diarrhea. The underlying causes were autoimmune enteropathy (in 21 of 74 patients), inflammatory bowel disease (in 16 of 74), atrophic gastritis (in 14 of 74), and celiac disease (in 3 of 74).

In 33 patients, gastrointestinal involvement was treated with corticosteroids (22 systemic corticosteroids and 11 oral budeso- nide). Systemic corticosteroids alleviated gastrointestinal symptoms in 19 cases temporarily (86%). Dosages varied between 5 and 50 mg per day and were often tapered to 5 mg. Seven patients received single high-dose bolus injections (1000 mg of methylprednisolone) without observable benefit. Of the 11 patients who received oral budesonide (9 mg per day), 10 achieved a good clinical response. Nine patients received abatacept (125 mg per week subcutaneously with a loading dose of 500 mg intravenously in 3 of them). All 9 showed an initial clinical response, but in 2 cases the diarrhea relapsed and abatacept was discontinued, with a 78% response rate. Nine patients received azathioprine (50-150 mg per day), with 4 (44%) showing a positive response. Eight received anti-TNF-a biologics (eg, adalimumab, 20-80 mg per week, or infliximab, 5-10 mg/kg), with 4 positive responses (56%). A total of 9 patients had sirolimus (1-2 mg per day), leading to 7 positive responses (78%) (for details, see Table E2, E).

Neurologic involvement. Of 123 patients, 41 (33%) had neurologic involvement. Of the latter 41 patients, 27 (65%) developed inflammatory cerebral lesions; 15 of the lesions were analyzed by biopsy. A total of 7 patients (17%) were diagnosed with encephalitis.

For the treatment of inflammatory central nervous system (CNS) lesions, 20 patients received corticosteroids (5-50 mg per day with slow wean, with 6 patients receiving methylprednisolone boluses of 1-3 X 1000 mg); 13 of these patients showed a symptomatic clinical response (65%). However, use of second- line medication was necessary on account of unremitting lesions or

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



relapses after steroid tapering. Abatacept (a loading dose of 500-750 mg intravenously followed by 125 mg per week subcutaneously) was added to steroids in 6 patients, with 4 patients (66%) showing stabilization of lesions and clinical improvement. Recently, the case of 1 of these patients has been published.¹⁰ High-dose intravenous immunoglobulin (IVIG) (1 g/kg) was added to steroids in 7 patients, with 6 (86%) showing beneficial effects. Rituximab (4 X 375 mg/m²) was added to steroids in 3 cases, but only 1 patient showed a good response (this patient additionally received cyclophosphamide). Cyclophosphamide (2 X 200 mg/m²) plus corticosteroids was tried in 3 cases (2 patients received 5 mg per day and 1 received 40 mg per day). Of these 3 patients, 2 showed a good clinical response (both received 5 mg per day) (see Table E2, *F*).

Skin involvement. Skin involvement was present in 61 of 123 patients (50%). We observed 59 cases of recurrent eczema, with 25 of them diagnosed as atopic dermatitis by the treating dermatologist. Additionally, we observed psoriatic skin irritations in 15 patients, alopecia in 14, warts in 11, and vitiligo in 5.

Four patients received systemic corticosteroids (5-20 mg per day): in 1 case because of granulomatous skin infiltration, in 2 cases because of severe psoriasis, and in 1 case because of atopic dermatitis. Of the 16 patients with psoriasis, 3 received methotrexate (7.5-20 mg per week) with good effect and 2 patients received cyclosporine (2-5 mg/kg) for severe infiltration of skin and lung and for concomitant cytopenia. In 1 patient cyclosporine alleviated the skin lesions (see Table E2, G).

DISCUSSION

This study reports the experience of physicians treating a large cohort of symptomatic *CTLA4* mutation carriers. We followed 123 affected *CTLA4* mutation carriers for a total of 2624 patient months. In all, 90% of patients had multiorgan autoimmunity. Abatacept, rituximab, sirolimus, and topical and systemic corticosteroids ameliorated disease severity, especially in cases of lymphocytic organ infiltration. Splenectomy should be avoided, as the response is not sustained. Cure was achieved with stem cell transplantation in 13 of 18 patients (72%). In detail, we propose the following diagnostic workup and treatment protocols for patients with a defect in CTLA-4-mediated biology. Adjustments for pediatric cases must be considered at the sections indicated in treatment protocols.

FUNCTIONAL TESTING OF CTLA4 VARIANTS

We functionally tested 6 variants, of which only 4 had a reduced CD80 transendocytosis; hence, the impairing effect of the mutation on the biology of CTLA-4 was proved. This highlights the importance of functional testing in cases in which a variant of unknown significance is identified in *CTLA4* in patients with clinical symptoms compatible with CTLA-4 insufficiency, as the differential diagnosis of CTLA-4 insufficiency includes several other monogenic traits for inborn errors of immunity (IEIs), such as the following: lipopolysaccharide-responsive beige-like anchor protein (LRBA) deficiency¹²; immune dysregulation, polyendocrinopathy, enteropathy, X-linked¹³; autoimmune

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



lymphoproliferative syndrome¹⁴; differentially expressed in FDCP6 homolog (DEF6) deficiency¹⁵; and nuclear factor-κB insufficiency.¹⁶ Interestingly, of these 4 mutations with proven pathogenicity, only 3 had reduced CTLA-4 staining on the surface of Treg cells, indicating that for diagnosing CTLA-4 insufficiency the CTLA-4 surface stain is not as sensitive as CTLA-4-mediated transendocytosis.

HYPOGAMMAGLOBULINEMIA

In CTLA-4 insufficiency, decreased serum levels of immunoglobulins are manifested with recurrent infections, as seen in other IEIs.¹⁷ In CTLA-4 insufficiency, hypogammaglobulinemia is possibly caused by bone marrow infiltration of activated effector T cells, which prevents long-lived plasma cell survival.¹ Moreover, there seems to be an additional bone marrowindependent loss of memory B cells, which is probably caused by a disturbed function of secondary lymphoid organs. Patients with CTLA-4 insufficiency with hypogammaglobulinemia should follow the European consensus protocols for dosing the IgRT (ie, 0.4-0.8 g/kg every 3-4 weeks).¹⁸ In addition, IgRT should be initiated in cases of isolated deficiency of the antibody subclasses IgG1 and IgG2 when associated with infections at values less than 3.0 g/L for IgG1 and less than 2.0 g/L for IgG2 (for adults), respectively (see Management Chart 1 in the Online Repository at www.jacionline.org). In patients with enteropathy, a secondary loss of serum antibodies via the gut should be considered.

ABNORMALITIES IN BONE MARROW AND SECONDARY LYMPHOID ORGANS

Lymphocytic infiltrations in CTLA-4 insufficiency typically involve the spleen and lymph nodes, as well as the gut, lung, skin, liver, kidney, bone marrow, and CNS.^{1,3} Additionally, the mucosa-associated lymphoid tissue may proliferate under sustained immune activation and cause gastrointestinal symptoms.^{1,3} Benign lymphadenopathy is common in IEIs¹⁹ and in CTLA-4 insufficiency.³ There is also an increased risk of cancer, especially lymphomas, in patients with CTLA-4-insufficiency.⁹ Hepatomegaly, splenomegaly, or regional lymphadenopathy should be carefully monitored, and if malignancy is suspected, a biopsy should be performed.

Our analysis showed that treatment was given only in cases of symptomatic lymphoid infiltrations, especially in the gut, lung, and brain, whereas asymptomatic cases were monitored. We currently advise treating symptomatic lymphoid infiltrations in CTLA-4 insufficiency with corticosteroids, rituximab, or a combination of both. Although histologic evaluations often show a predominant T-cell infiltrate, the B-cell-targeted anti-CD20 treatment with rituximab was clinically very efficient in our analysis. Alternatively, in cases of involvement of organs such as the lung or brain, a mechanistic target of rapamycin inhibitor or abatacept may be considered (see Management Chart 2 in the Online Repository at www.jacionline.org).

CYTOPENIAS

Autoimmune cytopenias in patients with CTLA-4 insufficiency often have a severe, chronic, and relapsing course.³ This might be caused by autoantibody-mediated peripheral consumption or T-cell-mediated bone marrow output failure. The pathophysiologic cause of the latter is currently the

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



subject of intense research. Drug treatment leads to some response in more than 80% of patients. On the basis of our documented outcomes, we suggest a high dose of steroids plus high-dose IVIG as first-line treatment for ITP in CTLA-4 insufficiency, as in CVID.²⁰ In AIHA, combining corticosteroids with rituximab was more effective than steroids alone, as reported by Birgens et al for AIHA in general.²¹ Rituximab also showed efficiency and safety in ITP^{21,22} complicating CVID,^{20,23} and it should be used in refractory or relapsing cases of any autoimmune cytopenias in the setting of CTLA-4 insufficiency (see Management Chart 3 in the Online Repository at www.jacionline.org). Rituximab helps to eliminate CD80- and CD86-expressing B cells, which activate effector T cells in the setting of CTLA-4 insufficiency. Thrombopoietin receptor agonists (TPO-RAs) are further second-line therapeutic agents for ITP.²⁴ We documented 4 applications of TPO-RAs in our CTLA-4 cohort, but a satisfying stabilization of platelet count could be reached in only 1 case. Abatacept may be worthwhile for keeping patients with a history of cytopenia (including PRCA) in remission. Splenectomy may improve the platelet count and complete blood count and reduce steroid dependence; however, the patients with CTLA-4 insufficiency in our cohort showed no sustained response, as already published for patients with Evans syndrome²⁵ and CVID in general.²⁶ In individual severe cases of cytopenia that are not being controlled even under high doses of steroids, rituximab, or TPO-RAs, HSCT may become necessary at an early stage. HSCT has been described as an effective therapy escalation for patients with CTLA-4 insufficiency by Slatter et al⁵ and Schwab et al.³ A total of 16 patients with CTLA-4-associated cytopenia were treated by HSCT; of these 16 patients, 12 were cured and 4 died.

GLILD

For the diagnosis of GLILD, computed tomography imaging, a 6-minute walk test, lung function testing with diffusion capacity, documentation of the Medical Research Council Dyspnea Scale score, and the exclusion of an infection by bronchoalveolar lavage are recommended. Lung biopsy may be helpful. For treatment, we advise following the British consensus protocols for GLILD in CVID.²⁷ In summary, we recommend initiating treatment with corticosteroids when patients are symptomatic or/and have abnormal or deteriorating lung function, focusing especially on the diffusion capacity. We start treatment with 0.5 mg/kg per day, but the dose recommendations range up to 2 mg/kg per day.²⁷ After steroids are tapered to less than 7.5 mg per day within the first 2 months, a computed tomography scan, 6-minute walk test, and lung function testing (including of diffusion capacity) should be performed to document the success of the therapy. In cases of a nonsatisfactory response to steroids, we suggest adding abatacept, rituximab, mycophenolate, or sirolimus as a second- line agent (see Management Chart 4 in the Online Repository at www.jacionline.org). Recently, abatacept was shown to be safe and effective in 10 patients with CVID with GLILD.²⁸

However, as GLILD is usually seen in the context of multisystem involvement and all of our 44 patients with GLILD had additional nonpulmonary features, one may argue that (1) the first attempt to treat GLILD with corticosteroid monotherapy is superfluous and (2) abatacept should be preferred over mycophenolate or sirolimus despite the fact that sirolimus has been successful in 1 of our

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



cases.²⁹ Antibiotics may be used for both prophylaxis and treatment of established infections. Antibiotic treatment of acute exacerbations in patients with bronchiectasis is advised to reduce bacterial load and decrease systemic inflammatory mediators. At times, this may require intravenous or inhaled antibiotics.

GASTROINTESTINAL INVOLVEMENT

Therapy for enteropathy in affected carriers of the *CTLA4* mutation depends on the type and severity of gastrointestinal involvement. In cases of gluten sensitivity or lactose intolerance, the respective diets should be followed. In cases of inflammation without known antigen, we recommend a stepwise protocol starting with (1) topical corticosteroids in the form of budesonide (a combination of standard budesonide for the small bowels and retard capsules for the large bowels) or use of oral sulfasalazine; (2) systemic courses of corticosteroids up to 1 mg/kg; (3) abatacept, anti-TNF-a, or sirolimus; or (4) a combination of the aforementioned. A beneficial effect of abatacept on the gastrointestinal symptoms of patients with CTLA-4 insufficiency has been reported before,^{3'30'31} and is it therefore recommended as a first-line steroid-sparing agent in patients with CTLA-4 insufficiency (see Management Chart 5 in the Online Repository at www.jacionline.org).

Alternative strategies should include consideration of sirolimus or the anti-a4β7 integrin mAb vedolizumab.³² The latter was successful in treating the diarrhea in 2 patients. Interestingly, vedolizumab has also proved successful for gastrointestinal adverse effects in anti-CTLA-4 treatment.³³ On the basis of our data, we suggest sirolimus rather than anti-TNF-a as a second-line agent in cases of CTLA-4 insufficiency. In 9 of our patients the gastrointestinal inflammation was so recalcitrant that HSCT was performed.^{3,5} Of these 9 patients, 6 were cured and 3 died of HSCT-related causes.

NEUROLOGIC INVOLVEMENT

We defined CTLA-4-associated CNS syndrome as noninfectious neurologic affection caused by intracerebral lesions in *CTLA4* mutation carriers. Some symptoms were caused by secondary bleeding.³ We recommend performing a magnetic resonance imaging (MRI) of the brain even in asymptomatic mutation carriers and repeating the scanning every 5 years or more frequently in children and when clinically indicated. If lesions are identified, we advise analyzing cerebrospinal fluid (CSF) and searching for autoantibodies. In patients with neurologic symptoms, MRI, lumbar puncture, and serology are mandatory.

CTLA4 mutation carriers typically show extensive white matter lesions with contrast enhancement.¹⁻ Despite no autoantibodies having been identified in these patients, we recommend excluding anti-myelin oligodendrocyte glycoprotein (antiMOG) antibodies, as patients with anti-MOG antibodies show similar CNS lesions and hence represent the main differential diagnosis.³⁴ Testing of anti-MOG antibodies in serum is more sensitive than in the CSF.³⁴ However, we recommend storing a biobank sample (serum and CSF) for possible future novel autoantibody detection, especially as patients undergoing treatment with CTLA-4 inhibitors may develop different types of autoimmune

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



encephalitis associated with neuronal surface antibodies (NMDAR and GABAR).³⁵ For this reason, neuronal surface antibodies in CSF should be considered in patients with CTLA-4 insufficiency with prominent psychiatric symptoms or movement disorder.

In all patients with CTLA-4-associated CNS syndrome, optimization of baseline treatment with abatacept or sirolimus is recommended to control lymphocyte activation. Abatacept showed benefits in 4 of our patients, including in 1 published case patient with regredient lesions. ¹⁰ As recommended for other demyelinating CNS disorders, ³⁶ as first-line treatment we suggest high-dose corticosteroids (methylprednisolone pulses [30 mg/kg/ m²] at a maximum dose of 1 g for 3-5 days). If no rapid clinical response is observed, adding high dose IVIG (1-2 g/kg) appeared to be valuable. Repeated MRI will be necessary to evaluate treatment response at 3 and 6 months following treatment initiation. As a second-line strategy, rituximab or cyclophosphamide should be considered while continuing the first-line therapy, as recommended for autoimmune encephalitis³⁷ (see Management Chart 6 in the Online Repository at www.jacionline.org). The data are not currently sufficient to recommend 1 regimen over the other.

SKIN INVOLVEMENT

Skin lesions in individuals with CTLA-4 insufficiency are a possible entry port for infections and should be carefully treated or monitored. As skin involvement is often mild to moderate in patients with CTLA-4 insufficiency, skin-specific therapy in CTLA-4 insufficiency should be performed in the context of possible additional organ involvement. As first-line therapy, we suggest topical treatment with corticosteroids and as second-line topical tacrolimus, respectively pimecrolimus for skin lesions in the face. For psoriasis-like lesions we refer to the psoriasis treatment protocols. If systemic treatment is warranted, we suggest considering abatacept, sirolimus, or cyclosporine with or without systemic corticosteroids (see Management Chart 7 in the Online Repository at www.jacionline.org). In 1 of our patients without detectable T_H17 cell infiltration abatacept did not improve the skin condition.

STUDY LIMITATIONS

The retrospective collection of data represents a limitation in our work. It was especially difficult to discriminate retrospectively between an adverse effect of the treatment and the natural course of CTLA-4 insufficiency. Drugs may have been chosen before the diagnosis of CTLA-4 insufficiency was known and therefore influenced treatment decisions. In many cases, insufficient treatment responses have driven further attempts until a sufficient disease control has been established. Differing concomitant steroid regimens make comparison of treatment responses difficult. Although the length of treatment was recorded, the interaction between different drug regimens was not studied. Finally, treatment outcomes are hard to compare in a multicenter approach without defined protocols. Nevertheless, we observed that in the majority of our patients therapeutic success could be achieved. Often, multiple changes of immunosuppressants were needed owing to adverse effects or steroid dependence. Nonetheless, some treated patients were stable over the

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



years. In contrast, untreated or insufficiently treated patients developed further CTLA-4-related symptoms, suggesting a progressive natural course of the disease.

PERSONALIZING PRACTICE AND PHENOTYPE-GENOTYPE CORRELATIONS

Our analysis suggests that treatment-refractory CNS involvement, cytopenias, enteropathy, or lung involvement represent "red flags" and should trigger early search for a possible stem cell donor. As about 90% of patients with CTLA-4 insufficiency experience multiorgan disease, treating physicians need to combine our treatment recommendations. Other involvement, such as hypogammaglobulinemia, thyroid disease, diabetes, skin involvement or similar conditions, may actually be managed by a more conservative approach. For pediatric patients developing severe symptoms, HSCT should be considered early before manifestation of disease-related organ dysfunction or consequences of long-term immunosuppression. Treatment in pediatric patients should be age-adjusted and should always be planned in consultation with experienced pediatricians.

Regarding the variable penetrance in affected mutation carriers, the timing of treatment and treatment escalation are central questions. A completely unanswered question is, for example, whether one should preemptively treat oligosymptomatic patients (eg, with abatacept). We are not at this point yet, as we do not know whether, for example, the early replacement of CTLA-4-Fc at a sufficient dose may prevent disease onset and/or disease progression and/or worsening. Basically, in an individual with CTLA4 mutation, abatacept replaces the individual CTLA-4 that is not being produced sufficiently and/or not functioning. Especially in patients with gastrointestinal involvement or lymphoid infiltrations (in the lung and CNS), abatacept has been shown to elicit effective responses.^{3,31} Previously, abatacept was successfully used in patients with LRBA deficiency.³⁸ The same study described T follicular helper cells as a biomarker of disease activity and treatment response.³⁸ Future studies of abatacept in patients with CTLA-4 insufficiency should examine different doses and aim for a scoring system to measure efficacy similar to that used by Kiykim et al in patients with LRBA.³⁸ Until further data are available, blood levels of EBV and cytomegalovirus should be carefully monitored while abatacept is being used because loss of control over oncogenic viruses leading to severe virus-associated viremia or neoplasms in selected patients has been described.^{3,9} Interestingly, not all of our patients had satisfactory responses to abatacept. Three of our patients became even worse. However, nonresponsiveness may depend on the amount of abatacept given to the patient. Data on long-term application and safety studies for abatacept in this cohort are missing. To this end, the authors have initiated an investigator-driven trial, called ABACHAI (EudraCT no. 2019-000972-40; DRKS no. DRKS00017736).

So far, only HSCT is regarded as a long-term cure for CTLA-4 insufficiency, but because of its treatment-related morbidity and mortality, it requires careful individual consideration. As we understand the increasing burden and complications of CTLA-4 insufficiency, the specific immunomodulatory therapies may just be "firefighting," whereas what is needed is definitive curative HSCT or, in the longer-term, gene therapy to effectively cure our patients. Tesch et al reported on 17 of 24 patients with LRBA deficiency who had a successful outcome with HSCT.³⁹ Here,

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



we have reported on 18 patients with CTLA-4 insufficiency with HSCT, but to our knowledge, at least 6 additional patients with CTLA-4 have since undergone HSCT and a task force to publish an article on HSCT in these patients is in place. Tesch et al associated severe disease activity, lung involvement, age at transplantation, and delayed HSCT with poor outcome.³⁹ So far, our data have not shown significant differences in outcome by age at HSCT, transplantation year, and time between age of onset and time of transplantation.

In this article, we have published functional data on 4 pathogenic variants but also added 2 variants to the list of most likely benign *CTLA4* sequence variants (p.N145S and p.T207A). Interestingly, so far only patients with variants in exons 1 to 3 have been published. Here, we have reported on 1 carrier who has a variant in exon 4, which codes for the cytoplasmic tail of CTLA-4. The cytoplasmatic tail of CTLA-4 binds to LRBA, which prevents CTLA-4 from lysosomal degradation. Unlike patients with LRBA deficiency, who are generally more severely affected with a phenocopy of CTLA-4 insufficiency, the 11- year-old carrier of the variant in exon 4 had low IgG serum levels but does not currently require immunosuppressive therapy. Functional testing of the variant showed no impairment in CD80 uptake.

Despite the lack of exon 4 mutations, however, no genotypephenotype correlation has been observed. Hence, the type and location of the heterozygous mutation in *CTLA4* may not have a considerable impact on the clinical outcome.³

Nonetheless, the incomplete clinical penetrance of CTLA-4 insufficiency suggests that there are factors in addition to the *CTLA4* mutation that are influencing the phenotype. These may be genetic variants, epigenetic modifications, a difference in the microbiota, infectious triggers, or an unknown environmental trigger. It is also conceivable that in different families there are different disease modifiers that may require different treatment protocols. All of the aforementioned issues should be further studied in the future by collecting new cases worldwide to identify disease modifiers and their possible impact on treatment response. Further studies may also indicate which individuals may possibly become affected and may hence need advice on preventive treatment options. As described, treatment in cases of CTLA-4 insufficiency is challenging, and its optimization is an ongoing process. We are still collecting data on more *CTLA4* mutation carriers worldwide for further improvement of patient management and development of additional targeted therapies.

We thank all patients, family members, and physicians who participated in this study.

Clinical implications: The benefits and risks of symptomatic and curative therapies need to be considered when deciding on the best treatment for patients with CTLA-4 insufficiency.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



References

- Schubert D, Bode C, Kenefeck R, Hou TZ, Wing JB, Kennedy A, et al. Autosomal dominant immune dysregulation syndrome in humans with CTLA4 mutations. Nat Med 2014;20:1410-6.
- Kuehn HS, Ouyang W, Lo B, Deenick EK, Niemela JE, Avery DT, et al. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. Science 2014;345:1623-7.
- 3. Schwab C, Gabrysch A, Olbrich P, Patiño V, Warnatz K, Wolff D, et al. Phenotype, penetrance, and treatment of 133 cytotoxic T-lymphocyte antigen 4-insufficient subjects. J Allergy Clin Immunol 2018;142:1932-46.
- 4. Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. Immunity 1995;3:541-7.
- 5. Slatter MA, Engelhardt KR, Burroughs LM, Arkwright PD, Nademi Z, Skoda- Smith S, et al. Hematopoietic stem cell transplantation for CTLA4 deficiency. J Allergy Clin Immunol 2016;138:615-9.e1.
- Rae W, Ward D, Mattocks CJ, Gao Y, Pengelly RJ, Patel SV, et al. Autoimmunity/ inflammation in a monogenic primary immunodeficiency cohort. Clin Transl Immunol 2017;6:e155.
- Zeissig S, Petersen B-S, Tomczak M, Melum E, Huc-Claustre E, Dougan SK, et al. Early-onset Crohn's disease and autoimmunity associated with a variant in CTLA- 4. Gut 2015;64:1889-97.
- 8. Kucuk ZY, Charbonnier L-M, McMasters RL, Chatila T, Bleesing JJ. CTLA-4 hap- loinsufficiency in a patient with an autoimmune lymphoproliferative disorder. J Allergy Clin Immunol 2017;140:862-4.e4.
- Egg D, Schwab C, Gabrysch A, Arkwright PD, Cheesman E, Giulino-Roth L, et al. Increased risk for malignancies in 131 affected CTLA4 mutation carriers. Front Immunol 2018;9:2012.
- van Leeuwen EM, Cuadrado E, Gerrits AM, Witteveen E, Bree GJ de. Treatment of intracerebral lesions with abatacept in a CTLA4-haploinsufficient patient. J Clin Immunol 2018;38:464-7.
- Scheuerlein P, Pietsch L, Camacho-Ordonez N, Reiser V, Patel S, Burns SO, et al. Is it safe to switch from intravenous immunoglobulin to subcutaneous immunoglobulin in patients with common variable immunodeficiency and autoimmune thrombocytopenia? Front Immunol 2018;9:116.
- Lopez-Herrera G, Tampella G, Pan-Hammarström Q, Herholz P, Trujillo-Vargas CM, Phadwal K, et al. Deleterious mutations in LRBA are associated with a syndrome of immune deficiency and autoimmunity. Am J Hum Genet 2012;90: 986-1001.
- Jamee M, Zaki-Dizaji M, Lo B, Abolhassani H, Aghamahdi F, Mosavian M, et al. Clinical, immunological, and genetic features in patients with immune dysregulation, polyendocrinopathy, enteropathy, X-linked (IPEX) and IPEX-like syndrome. J Allergy Clin Immunol Pract 2020;8:2747-60.e7.
- Matson DR, Yang DT. Autoimmune lymphoproliferative syndrome. An overview. Arch Pathol Lab Med 2020;144:245-51.
- 15. Serwas NK, Hoeger B, Ardy RC, Stulz SV, Sui Z, Memaran N, et al. Human DEF6 deficiency underlies an immunodeficiency syndrome with systemic autoimmunity and aberrant CTLA-4 homeostasis. Nat Commun 2019;10:3106.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



- ^{16.} Klemann C, Camacho-Ordonez N, Yang L, Eskandarian Z, Rojas-Restrepo JL, Frede N, et al. Clinical and immunological phenotype of patients with primary immunodeficiency due to damaging mutations in NFKB2. Front Immunol 2019;10: 297.
- Krivan G, Jolles S, Granados EL, Paolantonacci P, Ouaja R, Cissé OA, Bernatowska E. New insights in the use of immunoglobulins for the management of immune deficiency (PID) patients. Am J Clin Exp Immunol 2017;76-83.
- Sewell WAC, Kerr J, Behr-Gross ME, Peter HH Kreuth Ig Working Group European consensus proposal for immunoglobulin therapies. Eur J Immunol 2014;44: 9-10.
- 19. Gangemi S, Allegra A, Musolino C. Lymphoproliferative disease and cancer among patients with common variable immunodeficiency. Leuk Res 2015;39: 389-96.
- Wang J, Cunningham-Rundles C. Treatment and outcome of autoimmune hematologic disease in common variable immunodeficiency (CVID). J Autoimmun 2005; 25:57-62.
- Birgens H, Frederiksen H, Hasselbalch HC, Rasmussen IH, Nielsen OJ, Kjeldsen L, et al. A phase III randomized trial comparing glucocorticoid monotherapy versus glucocorticoid and rituximab in patients with autoimmune haemolytic anaemia. Br J Haematol 2013;163:393-9.
- Arnold DM, Vrbensky JR, Karim N, Smith JW, Liu Y, Ivetic N, et al. The effect of rituximab on anti-platelet autoantibody levels in patients with immune thrombocytopenia. Br J Haematol 2017;178:302-7.
- 23. Gobert D, Bussel JB, Cunningham-Rundles C, Galicier L, Dechartres A, Berezne A, et al. Efficacy and safety of rituximab in common variable immunodeficiency- associated immune cytopenias. A retrospective multicentre study on 33 patients. Br J Haematol 2011;155:498-508.
- Neunert C, Lim W, Crowther M, Cohen A, Solberg L, Crowther MA. The American Society of Hematology 2011 evidence-based practice guideline for immune thrombocytopenia. Blood 2011;117:4190-207.
- Michel M, Chanet V, Dechartres A, Morin A-S, Piette J-C, Cirasino L, et al. The spectrum of Evans syndrome in adults. New insight into the disease based on the analysis of 68 cases. Blood 2009;114:3167-72.
- Wong GK, Goldacker S, Winterhalter C, Grimbacher B, Chapel H, Lucas M, et al. Outcomes of splenectomy in patients with common variable immunodeficiency (CVID). A survey of 45 patients. Clin Exp Immunol 2013;172: 63-72.
- Hurst JR, Verma N, Lowe D, Baxendale HE, Jolles S, Kelleher P, et al. British Lung Foundation/United Kingdom Primary Immunodeficiency Network consensus statement on the definition, diagnosis, and management of granulomatous-lymphocytic interstitial lung disease in common variable immunodeficiency disorders. J Allergy Clin Immunol Pract 2017;5:938-45.
- von Spee-Mayer C, Echternach C, Agarwal P, Gutenberger S, Soededjo V, Goldacker S, et al. Abatacept use is associated with steroid dose reduction and improvement in fatigue and CD4-dysregulation in CVID patients with interstitial lung disease. J Allergy Clin Immunol Pract 2021;9:760-70.
- 29. Deyà-Martínez A, Esteve-Solé A, Vélez-Tirado N, Celis V, Costa J, Cols M, et al. Sirolimus as an alternative treatment in patients with granulomatous-lymphocytic lung disease and humoral immunodeficiency with impaired regulatory T cells. Pe- diatr Allergy 2018;29:425-32.
- Gupta NK, Yilmaz O, Fisher M, Yajnik V. Abatacept. A new treatment option for refractory adult autoimmune enteropathy. J Clin Gastroenterol 2014;48:55-8.

DOI: 10.1016/J.Jaci.2021.04.039 Status: Postprint (Author's version)



- Lee S, Moon JS, Lee C-R, Kim H-E, Baek S-M, Hwang S, et al. Abatacept alleviates severe autoimmune symptoms in a patient carrying a de novo variant in CTLA-4. J Allergy Clin Immunol 2016;137:327-30.
- ^{32.} Walter JE, Farmer JR, Foldvari Z, Torgerson TR, Cooper MA. Mechanismbased strategies for the management of autoimmunity and immune dysregulation in primary immunodeficiencies. J Allergy Clin Immunol Pract 2016;4:1089-100.
- 33. Shivaji UN, Jeffery L, Gui X, Smith SCL, Ahmad OF, Akbar A, et al. Immune checkpoint inhibitor-associated gastrointestinal and hepatic adverse events and their management. Therap Adv Gastroenterol 2019;12:1756284819884196.
- Ramanathan S, Dale RC, Brilot F. Anti-MOG antibody. The history, clinical phenotype, and pathogenicity of a serum biomarker for demyelination. Autoimmun Rev 2016;15:307-24.
- Pan PC, Haggiagi A. Neurologic immune-related adverse events associated with immune checkpoint inhibition. Curr Oncol Rep 2019;21:108.
- Varley J, Taylor J, Irani SR. Autoantibody-mediated diseases of the CNS. Structure, dysfunction and therapy. Neuropharmacology 2018;132:71-82.
- Stingl C, Cardinale K, van Mater H. An update on the treatment of pediatric autoimmune encephalitis. Curr Treatm Opt Rheumatol 2018;4: 14-28.
- Kiykim A, Ogulur I, Dursun E, Charbonnier LM, Nain E, Cekic S, et al. Abatacept as a long-term targeted therapy for LRBA deficiency. J Allergy Clin Immunol Pract 2019;7:2790-800.e15.
- Tesch VK, Abolhassani H, Shadur B, Zobel J, Mareika Y, Sharapova S, et al. Longterm outcome of LRBA deficiency in 76 patients after various treatment modalities as evaluated by the immune deficiency and dysregulation activity (IDDA) score. J Allergy Clin Immunol 2020;145:1452-63