

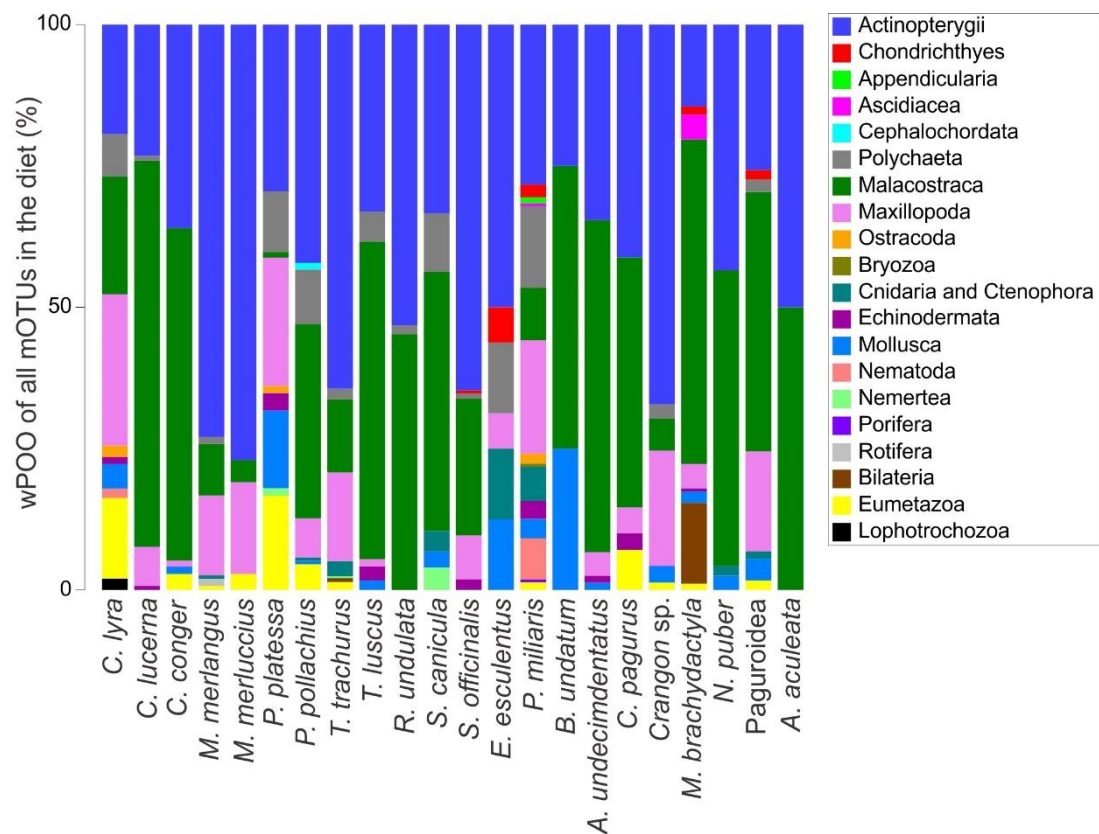
## Supplementary Figures and Tables for the manuscript

### ‘Gut content metabarcoding reveals potential importance of fisheries discards consumption in marine fauna.’

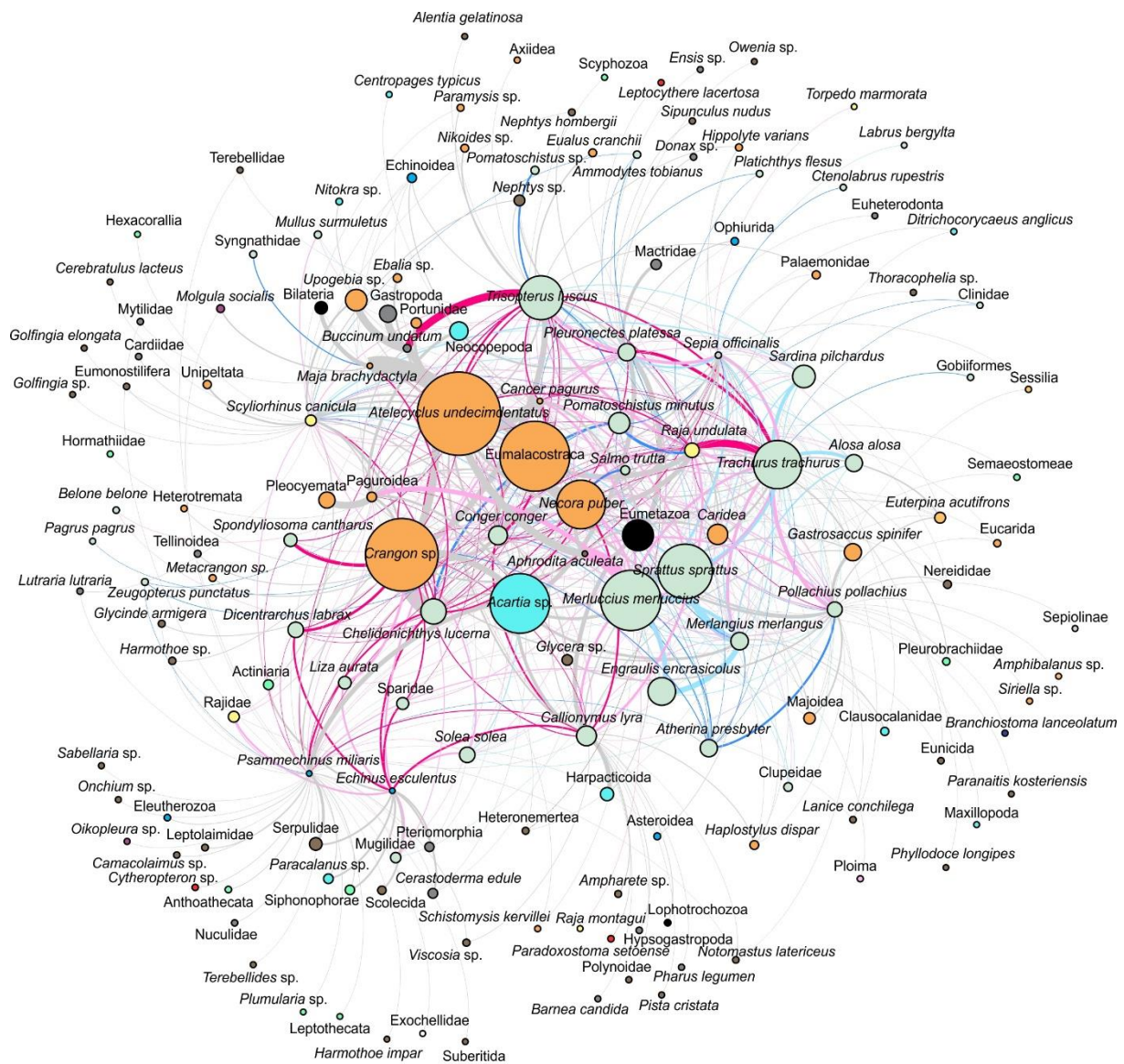
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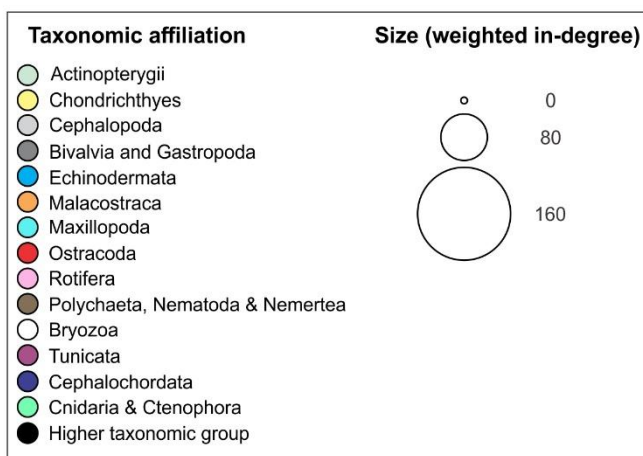
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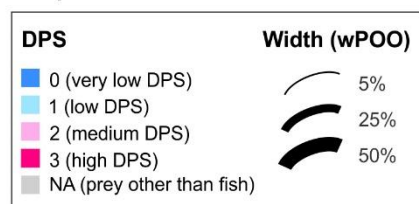
**Figure S1:** Weighted percentage of occurrence (wPOO [%]) of prey mOTUs identified via gut contents metabarcoding of the 22 consumers. mOTUs are regrouped according to their taxonomic affiliation (Class or higher taxonomic group).



**Node**



**Trophic link**



Published version of the article and its supplementary material are available at: <https://doi.org/10.1139/cjfas-2021-0267>.

**Figure S2:** Molecular network constructed from gut content metabarcoding data, depicting trophic interactions among the 22 consumers and their prey, and including all trophic links. Nodes are representing the consumers and prey mOTUs. Node color depicts taxonomic affiliation (by Class). Node size is proportional to its importance as a food source for other taxa (weighted in-degree centrality). Link width is proportional to wPOO of prey. Link color varies according to prey DPS.

**Table S1:** Observational data of discarding activity in the Bay of Bourgneuf. 'Discard frequency per haul per trip' is based on in situ observations on board of a commercial trawler during the sampling for stomach content data (i.e. 'Pers Obs', see details in Material and Methods): 0 = No discarding observed, <25% = Discarding observed in less than 25% of the hauls and with a maximum of 10 individuals per haul, >25% = Discarding observed in more than 25% of the hauls and with minimum 10 individuals per haul. 'Mean discard weight (g) per haul per trip' are the estimates of discarded quantities of each species based on data available from the OBSMER samplings of 2018 and 2019 in the area of study (see details in Material and Methods). 'Discard importance score': 0 = No discarding observed in any dataset, 1 = low discard importance (i.e. species was discarded and met the following criteria: (a) discarded in less than 25% of the hauls and with less than 10 individuals per haul according to our own observational data OR (b) discarded in mean quantities (weight) per haul lower than the mean of all discarded fish per haul according to OBSMER data) and 2 = high discard frequency (all discarded species above the aforementioned thresholds in at least one dataset. Cells are coloured according to the resulting discard importance score: 0 = white, 1 = light grey, 2 = dark grey. 'Reason for discarding': TQ = Individual below MCRS because of TAC & Quotas or because of quota limitation, NC = Noncommercial species or no market, TF = Too few individuals to be sold, non-target species, TS = Too small individuals, DG = Eviscerated on board and discarding of the guts. Note that these correspond to reasons for discarding at the time of study, which may not be acceptable anymore after the full implementation of the Landing Obligation.

Discard importance measure	Discard frequency per haul per trip	Mean discard weight (g) per haul per trip		Discard importance	Reason for discarding
Data Source	Pers Obs	OBSMER	OBSMER	score	
Year	2019	2018	2019		
<i>Ammodytes tobianus</i>	0	0	0	0	NC - TF
<i>Atherina presbyter</i>	0	0	0	0	NC
Clinidae	0	0	0	0	NC
<i>Ctenolabrus rupestris</i>	0	0	0	0	NC
Gobiiiformes	0	0	0	0	NC
<i>Platichthys flesus</i>	0	0	0	0	TF-TS
<i>Salmo trutta</i>	0	0	0	0	TF
Syngnathidae	0	0	0	0	NC
<i>Zeugopterus punctatus</i>	0	0	0	0	NC
<i>Alosa alosa</i>	< 25%	0	0	1	NC - TF
<i>Belone belone</i>	< 25%	0	0	1	TS
<i>Engraulis encrasicolus</i>	< 25%	0	0	1	TF
<i>Labrus bergylta</i>	< 25%	0	0	1	TS
<i>Liza aurata</i>	< 25%	0	0	1	TS
<i>Merlangius merlangus</i>	< 25%	262	0	1	TQ
<i>Merluccius merluccius</i>	< 25%	0	0	1	TQ
Mugilidae	< 25%	400	0	1	TS
<i>Mullus surmuletus</i>	< 25%	492	0	1	TS
<i>Pagrus pagrus</i>	< 25%	0	0	1	TS
<i>Pleuronectes platessa</i>	< 25%	115	0	1	TQ
<i>Pollachius pollachius</i>	< 25%	0	0	1	TQ
<i>Raja montagui</i>	< 25%	0	0	1	TQ
Rajidae	< 25%	0	0	1	TQ
<i>Sardina pilchardus</i>	< 25%	0	0	1	TF
<i>Scyliorhinus canicula</i>	< 25%	0	0	1	TS
<i>Solea solea</i>	< 25%	157	0	1	TQ
<i>Sprattus sprattus</i>	< 25%	0	0	1	TF
<i>Callionymus lyra</i>	< 25%	1223	5102	2	NC
<i>Chelidonichthys lucerna</i>	> 25%	219	1253	2	TS
<i>Conger conger</i>	> 25%	4413	0	2	TS - DG
<i>Dicentrarchus labrax</i>	> 25%	2212	0	2	TQ
<i>Raja undulata</i>	> 25%	2542	2858	2	TQ
Sparidae	< 25%	3702	0	2	TF - TS
<i>Spondylisoma cantharus</i>	> 25%	2254	114	2	TS
<i>Torpedo marmorata</i>	0	559	6386	2	NC
<i>Trachurus trachurus</i>	> 25%	2919	0	2	TQ
<i>Trisopterus luscus</i>	> 25%	5586	1032	2	TS -NC
Global mean weight of discarded fish per haul		1804	2791		

**Table S2:** Species constituting the local reference database for metabarcoding analysis, sequenced for the two markers: Tele02 (targetting the the 12S rDNA gene) and Euka02 (targetting the the 18S rDNA gene). Taxon = the sequenced taxon. TaxID = identification number of the sequenced taxon, which can be found in the curated NCBI Taxonomy Browser database.

<b>Taxon</b>	<b>TaxID</b>	<b>Tele02</b>	<b>Euka02</b>
<i>Scyliorhinus canicula</i>	7830	x	x
<i>Spondyliosoma cantharus</i>	50595	x	
<i>Solea solea</i>	90069	x	x
<i>Pollachius pollachius</i>	185739	x	x
<i>Trisopterus luscus</i>	27722	x	x
<i>Cancer pagurus</i>	6755		x
<i>Trachurus trachurus</i>	36212	x	x
<i>Sprattus sprattus</i>	196075	x	x
<i>Atelecyclus undecimdentatus</i>	483410		x
<i>Zeugopterus punctatus</i>	526623	x	x
<i>Belone belone</i>	129037	x	x
<i>Buccinum undatum</i>	37541		x
<i>Echinus esculentus</i>	7648		x
<i>Callionymus lyra</i>	34785	x	x
<i>Pleuronectes platessa</i>	8262	x	x
<i>Alosa alosa</i>	278164	x	x
<i>Chelidonichthys lucerna</i>	2576622	x	x
<i>Necora puber</i>	338210		x
<i>Scomber scombrus</i>	13677	x	x
<i>Dicentrarchus labrax</i>	13489	x	x
<i>Sardina pilchardus</i>	27697	x	x
<i>Mullus surmuletus</i>	87757	x	x
<i>Conger conger</i>	82655	x	
<i>Merlangius merlangus</i>	8058	x	x
<i>Merluccius merluccius</i>	8063	x	x
<i>Pagrus pagrus</i>	8173	x	x



**Table S3: Summary statistics of sequence read counts per consumer for each marker's dataset: (A) after the removal of self-hits and (B) of removed self-hits.**

(A) Total reads without self-hits	Euka02				Tele02			
	Mean	SD	min	max	Mean	SD	min	max
<i>Aphrodita aculeata</i>	4952	3253	2651	7252	263	NA	NA	NA
<i>Atelecyclus undecimdentatus</i>	27226	43275	10025	177002	18210	19933	2212	68747
<i>Buccinum undatum</i>	7800	NA	NA	NA	3696	NA	NA	NA
<i>Callionymus lyra</i>	47726	42202	13097	124848	3931	11138	0	33621
<i>Cancer pagurus</i>	66903	45689	3290	160235	50389	60817	1184	175126
<i>Chelidonichthys lucerna</i>	63134	41777	11322	183042	3510	8464	0	34129
<i>Conger conger</i>	46329	33966	1582	138724	3332	9976	0	44131
<i>Crangon</i> sp.	21540	14238	7083	43590	76170	74835	9068	268118
<i>Echinus esculentus</i>	494	NA	NA	NA	2477	NA	NA	NA
<i>Maja brachydactyla</i>	71128	52329	3914	162602	8488	12950	279	30717
<i>Merlangius merlangus</i>	55964	34129	2389	146744	43678	34606	541	111101
<i>Merluccius merluccius</i>	53548	41931	10018	158153	49702	40141	10	133492
<i>Necora puber</i>	16202	7449	6748	37469	21229	25006	178	73347
Paguroidae	36945	44513	2562	148627	39432	60476	1173	109153
<i>Pleuronectes platessa</i>	57891	30014	22121	121583	6851	18771	42	77684
<i>Pollachius pollachius</i>	47503	50031	6132	156152	20528	35837	0	134286
<i>Psammechinus miliaris</i>	33864	42711	1193	140814	31893	78601	171	273176
<i>Raja undulata</i>	58294	36750	4003	159391	23514	45075	0	170145
<i>Scyliorhinus canicula</i>	50108	32046	16721	148857	16282	28473	0	105049
<i>Sepia officinalis</i>	30154	25460	1810	88740	64408	52608	394	196395
<i>Trachurus trachurus</i>	32777	37763	1738	160953	48100	52727	956	194902
<i>Trisopterus luscus</i>	39828	25028	6856	90830	11668	15261	0	46148

(B) Removed self-hits reads	Euka02				Tele02			
	Mean	SD	min	max	Mean	SD	min	max
<i>Aphrodita aculeata</i>	51781	1076	51020	52542	NA	NA	NA	NA
<i>Atelecyclus undecimdentatus</i>	14207	14475	824	43719	NA	NA	NA	NA
<i>Buccinum undatum</i>	60992	NA	NA	NA	NA	NA	NA	NA
<i>Callionymus lyra</i>	27784	29620	597	83416	72194	96860	6184	315396
<i>Cancer pagurus</i>	7040	5485	0	15740	NA	NA	NA	NA
<i>Chelidonichthys lucerna</i>	20087	17855	0	64527	64072	51585	753	183496
<i>Conger conger</i>	7978	8109	581	25537	34476	31709	7233	147250
<i>Crangon</i> sp.	16829	11260	879	30728	NA	NA	NA	NA
<i>Echinus esculentus</i>	25496	NA	1	1	NA	NA	NA	NA
<i>Maja brachydactyla</i>	7969	8762	0	28579	NA	NA	NA	NA
<i>Merlangius merlangus</i>	15872	18636	566	66876	24884	38872	0	169646
<i>Merluccius merluccius</i>	21546	24761	646	66964	29400	20903	2079	75264
<i>Necora puber</i>	10683	14538	420	46435	NA	NA	NA	NA
Paguroidae	24360	22715	0	62895	NA	NA	NA	NA
<i>Pleuronectes platessa</i>	11599	13711	128	51006	69387	57010	213	171454
<i>Pollachius pollachius</i>	11741	11675	545	40991	27305	22768	100	86580
<i>Psammechinus miliaris</i>	19430	14942	0	46216	NA	NA	NA	NA
<i>Raja undulata</i>	13231	16675	0	64092	19436	15496	192	49939
<i>Scyliorhinus canicula</i>	21776	31215	0	115500	10054	18294	0	71541
<i>Sepia officinalis</i>	16459	27300	0	138564	NA	NA	NA	NA
<i>Trachurus trachurus</i>	13355	14108	142	48605	34973	30643	2132	108191
<i>Trisopterus luscus</i>	5855	6052	0	19256	24672	26502	0	90642

Note: For Tele02 in table B, non-fish consumers do not yield any self-hits as Tele02 was used to target fish DNA. Because summary statistics are calculated including samples in which only the consumer's own DNA was amplified with Tele02 (i.e. individuals that likely did not ingest other fish), they participate to high self-hits read counts in some species.

**Table S4:** Details on the calculation of the discard probability score (DPS). Discard importance score (a) = score estimated based on personal observations on board of a commercial ship during gut contents sampling and discarding data obtained via the OBSMER sampling campaigns of 2018 and 2019; 0 = No discarding, 1 = low discarding and 2 = high discarding. Trophic link score (b) = likelihood of natural predation vs. discard consumption based on prior knowledge of the consumer diet and that of its trophic guild; 0 = documented natural predation, 1 = no documented natural predation by the consumer or other species in its trophic guild. Discard Probability Score (DPS) = score classifying prey according to their probability of being ingested as fisheries discard based on the scores from (a) and (b); 0 = very low, 1 = low, 2 = medium, 3 = high probability of being ingested as fisheries discard. Diet and trophic guild references = databases and literature consulted to determine the likelihood of natural predation vs discard consumption for each consumer-prey interaction revealed by DNA metabarcoding of gut contents.

Consumer	Ingested fish MOTUs	Discard importance score (a)	Trophic link score (b)	Discard probability score (DPS)	Diet and trophic guild references
<i>Callionymus lyra</i>	<i>Conger conger</i>	2	1	3	Fricke, R. (1986) Callionymidae. Fishes of the North-eastern Atlantic and the Mediterranean (eds P.J.P. Whitehead, M.-L. Bauchot, J.-C. Hureau, J. Nielsen & E. Tortonese), pp. 1086–1093. UNESCO, Paris. ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-enclosed sea. <i>Limnology and Oceanography</i> , 62(5), 2213–2226. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Callionymus lyra</i>	<i>Merlangius merlangus</i>	1	1	2	
<i>Callionymus lyra</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Callionymus lyra</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Chelidonichthys lucerna</i>	<i>Conger conger</i>	2	1	3	Richards, W. J., V. P. Saksena. (1990). Triglidae. Check-list of the fishes of the eastern tropical Atlantic (CLOFETA) (eds J.C. Quero, J.-C. Hureau, C. Karrer, A. Post & L. Saldanha), pp. 680-684. JNICT/SEI/UNESCO, Paris, Vol 2. ; Serrano, A., Velasco, F., Olaso, I., & Sánchez, F. (2003). Macrobenthic crustaceans in the diet of demersal fish in the Bay of Biscay in relation to abundance in the environment. <i>Sarsia</i> , 88, 36–48. doi: 10.1080/00364820308469 ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-enclosed sea. <i>Limnology and Oceanography</i> , 62(5), 2213–2226. ; Thompson, Murray S. A., Pontalier, H.,
<i>Chelidonichthys lucerna</i>	<i>Belone belone</i>	1	1	2	
<i>Chelidonichthys lucerna</i>	<i>Trachurus trachurus</i>	2	1	3	
<i>Chelidonichthys lucerna</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Chelidonichthys lucerna</i>	<i>Sprattus sprattus</i>	1	0	1	
<i>Chelidonichthys lucerna</i>	<i>Trisopterus luscus</i>	2	1	3	
<i>Chelidonichthys lucerna</i>	<i>Merlangius merlangus</i>	1	1	2	



<i>Chelidonichthys lucerna</i>	<i>Merluccius merluccius</i>	1	1	2	M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Chelidonichthys lucerna</i>	<i>Pomatoschistus minutus</i>	0	0	0	
<i>Chelidonichthys lucerna</i>	<i>Dicentrarchus labrax</i>	2	1	3	
<i>Chelidonichthys lucerna</i>	<i>Pleuronectes platessa</i>	1	1	2	
<i>Chelidonichthys lucerna</i>	<i>Raja undulata</i>	2	1	3	
<i>Conger conger</i>	<i>Atherina presbyter</i>	0	0	0	Serrano, A., Velasco, F., Olaso, I., & Sánchez, F. (2003). Macrobenthic crustaceans in the diet of demersal fish in the Bay of Biscay in relation to abundance in the environment. <i>Sarsia</i> , 88, 36–48. doi: 10.1080/00364820308469 ; O’Sullivan, S., Moriarty, C., & Davenport, J. (2004). Analysis of the stomach contents of the European conger eel <i>Conger conger</i> in Irish waters. <i>Journal of the Marine Biological Association of the United Kingdom</i> , 84, 823–826. ; Sallami, B., Ben Salem, M., Reynaud, C., & Canapé, C. (2015). Diet of European conger eel <i>Conger conger</i> (Osteichthyes: Congridae) from the northeastern coast of Tunisia (central Mediterranean). <i>Cahiers de Biologie Marine</i> , 56, 253–262. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Conger conger</i>	<i>Trachurus trachurus</i>	2	0	2	
<i>Conger conger</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Conger conger</i>	<i>Sprattus sprattus</i>	1	0	1	
<i>Conger conger</i>	<i>Engraulis encrasicolus</i>	1	0	1	
<i>Conger conger</i>	Clupeidae	1	0	1	
<i>Conger conger</i>	<i>Trisopterus luscus</i>	2	0	2	
<i>Conger conger</i>	<i>Merluccius merluccius</i>	1	0	1	
<i>Conger conger</i>	<i>Pomatoschistus minutus</i>	0	0	0	
<i>Conger conger</i>	<i>Zeugopterus punctatus</i>	0	0	0	
<i>Conger conger</i>	<i>Solea solea</i>	1	0	1	
<i>Conger conger</i>	<i>Callionymus lyra</i>	1	0	1	
<i>Conger conger</i>	<i>Scyliorhinus canicula</i>	1	1	2	
<i>Conger conger</i>	<i>Raja undulata</i>	2	1	3	
<i>Merlangius merlangus</i>	<i>Atherina presbyter</i>	0	0	0	Patterson, K. R. (1985). The trophic ecology of whiting <i>Merlangius merlangus</i> in the Irish Sea and its significance to the Manx herring stock. <i>Journal Du Conseil / Conseil Permanent International Pour l’Exploration de La Mer</i> , 42, 152–161. ; Hislop, J. R. G., Robb, A. P., Bell, M. A., & Armstrong, D. W. (1991). The diet and food consumption of whiting <i>Merlangius merlangus</i> in the North Sea. <i>ICES Journal of Marine Science</i> , 48, 139–156.; Bergstad, O. A. (1991). Distribution and trophic ecology of some gadoid fish of the Norwegian deep. <i>Sarsia</i> , 75, 269–313. doi: 10.1080/00364827.1991.10413455 ; Fjøsne, K., & Gjøsæter, J. (1996). Dietary composition and the potential of food competition between 0-group cod ( <i>Gadus morhua</i> L.) and some other fish species in the littoral zone. <i>ICES Journal of Marine Science</i> , 53, 757–770. ; Pedersen, J. (1999). Diet comparison between pelagic and demersal whiting in the North Sea. <i>Journal of Fish Biology</i> , 55, 1096–1113. ; Thompson,
<i>Merlangius merlangus</i>	<i>Trachurus trachurus</i>	2	0	2	
<i>Merlangius merlangus</i>	<i>Alosa alosa</i>	1	1	2	
<i>Merlangius merlangus</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Merlangius merlangus</i>	<i>Sprattus sprattus</i>	1	0	1	
<i>Merlangius merlangus</i>	<i>Engraulis encrasicolus</i>	1	0	1	
<i>Merlangius merlangus</i>	Clupeidae	1	0	1	
<i>Merlangius merlangus</i>	<i>Trisopterus luscus</i>	2	0	2	
<i>Merlangius merlangus</i>	<i>Merluccius merluccius</i>	1	0	1	
<i>Merlangius merlangus</i>	<i>Pomatoschistus minutus</i>	0	0	0	

<i>Merlangius merlangus</i>	<i>Pleuronectes platessa</i>	1	0	1	Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Merluccius merluccius</i>	<i>Atherina presbyter</i>	0	0	0	Bergstad, O. A. (1991). Distribution and trophic ecology of some gadoid fish of the Norwegian deep. <i>Sarsia</i> , 75, 269–313. doi: 10.1080/00364827.1991.10413455 ; Guichet, R. (1995). The diet of European hake ( <i>Merluccius merluccius</i> ) in the northern part of the Bay of Biscay. <i>ICES Journal of Marine Science</i> , 52, 21–31. ; Du Buit, M. H. (1996). Diet of hake ( <i>Merluccius merluccius</i> ) in the Celtic Sea. <i>Fisheries Research</i> , 28, 381–394. ; Cabral, B. H. N., & Murta, A. G. (2002). The diet of blue whiting, hake, horse mackerel and mackerel off Portugal. <i>Journal of Applied Ichthyology</i> , 18, 14–23. ; Serrano, A., Velasco, F., Olasso, I., & Sánchez, F. (2003). Macrobenthic crustaceans in the diet of demersal fish in the Bay of Biscay in relation to abundance in the environment. <i>Sarsia</i> , 88, 36–48. doi: 10.1080/00364820308469 ; Mahe, K., Amara, R., Bryckaert, T., Kacher, M., & Brylinski, J. M. (2007). Ontogenetic and spatial variation in the diet of hake ( <i>Merluccius merluccius</i> ) in the Bay of Biscay and the Celtic Sea. <i>ICES Journal of Marine Science</i> , 64, 1204–1219. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Merluccius merluccius</i>	<i>Trachurus trachurus</i>	2	0	2	
<i>Merluccius merluccius</i>	<i>Alosa alosa</i>	1	0	1	
<i>Merluccius merluccius</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Merluccius merluccius</i>	<i>Sprattus sprattus</i>	1	0	1	
<i>Merluccius merluccius</i>	<i>Engraulis encrasicolus</i>	1	0	1	
<i>Merluccius merluccius</i>	Clupeidae	1	0	1	
<i>Merluccius merluccius</i>	<i>Trisopterus luscus</i>	2	0	2	
<i>Merluccius merluccius</i>	<i>Merlangius merlangus</i>	1	0	1	
<i>Pleuronectes platessa</i>	<i>Trachurus trachurus</i>	2	1	3	Rijnsdorp, A. D., & Vingerhoed, B. (2001). Feeding of plaice <i>Pleuronectes platessa</i> L. and sole <i>Solea solea</i> (L.) in relation to the effects of bottom trawling. <i>Journal of Sea Research</i> , 45, 219–229. ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-enclosed sea. <i>Limnology and Oceanography</i> , 62(5), 2213–2226. ; Heindler, F. M., Maes, G. E., Delerue-ricard, S., Vanden, A., Hostens, K., & Volckaert, F. A. M. (2019). Diet composition and gut microbiome of 0-group European plaice <i>Pleuronectes platessa</i> L. - Strong homogeneity and subtle spatial and temporal differences. <i>Journal of Sea Research</i> , 144, 67–77. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Pleuronectes platessa</i>	<i>Sardina pilchardus</i>	1	1	2	
<i>Pleuronectes platessa</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Pleuronectes platessa</i>	<i>Trisopterus luscus</i>	2	1	3	
<i>Pleuronectes platessa</i>	<i>Merlangius merlangus</i>	1	1	2	
<i>Pleuronectes platessa</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Pleuronectes platessa</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Pleuronectes platessa</i>	<i>Ammodytes tobianus</i>	0	1	0	
<i>Pollachius pollachius</i>	<i>Atherina presbyter</i>	0	1	0	Vea Salvanes, A. G. (1986). Preliminary report from a comparative study of the diet of four gadoid fishes in a fjord of western Norway. <i>International Council for the Exploration of the</i>
<i>Pollachius pollachius</i>	<i>Trachurus trachurus</i>	2	0	2	

<i>Pollachius pollachius</i>	<i>Alosa alosa</i>	1	1	2	Sea, 1–18. Bergstad, O. A. (1991). Distribution and trophic ecology of some gadoid fish of the Norwegian deep. <i>Sarsia</i> , 75, 269–313. doi: 10.1080/00364827.1991.10413455 ; Sarno, B., Glass, C. W., & Smith, G. W. (1994). Differences in diet and behaviour of sympatric saithe and pollack in a Scottish sea loch. <i>Journal of Fish Biology</i> , 45(Supplement A), 1–11. ; Gibson, R. N., & Robb, L. (1996). Piscine predation on juvenile fishes on a Scottish sandy beach. <i>Journal of Fish Biology</i> , 49, 120–138. ; * Høines, Å. S., & Bergstad, O. A. (1999). Resource sharing among cod, haddock, saithe and pollack on a herring spawning ground. <i>Journal of Fish Biology</i> , 55, 1233–1257. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.	
<i>Pollachius pollachius</i>	<i>Sardina pilchardus</i>	1	0	1		
<i>Pollachius pollachius</i>	<i>Sprattus sprattus</i>	1	0	1		
<i>Pollachius pollachius</i>	<i>Engraulis encrasicolus</i>	1	0	1		
<i>Pollachius pollachius</i>	<i>Trisopterus luscus</i>	2	0	2		
<i>Pollachius pollachius</i>	<i>Merlangius merlangus</i>	1	0	1		
<i>Pollachius pollachius</i>	<i>Merluccius merluccius</i>	1	1	2		
<i>Pollachius pollachius</i>	<i>Pomatoschistus minutus</i>	0	0	0		
<i>Pollachius pollachius</i>	<i>Solea solea</i>	1	1	2		
<i>Pollachius pollachius</i>	<i>Callionymus lyra</i>	1	0	1		
<i>Pollachius pollachius</i>	<i>Raja undulata</i>	2	1	3		
<i>Trachurus trachurus</i>	<i>Atherina presbyter</i>	0	0	0		Cabral, H. N., & Murta, A. G. (2002). The diet of blue whiting, hake, horse mackerel and mackerel off Portugal. <i>Journal of Applied Ichthyology</i> , 18, 14–23. ; Jardas, I., Šantić, M., & Pallaoro, A. (2004). Diet composition and feeding intensity of horse mackerel, <i>Trachurus trachurus</i> (Osteichthyes : Carangidae) in the eastern Adriatic. <i>Marine Biology</i> , 144, 1051–1056. doi: 10.1007/s00227-003-1281-7 ; Šantić, M., Jardas, I., & Pallaoro, A. (2005). Feeding habits of horse mackerel, <i>Trachurus trachurus</i> (Linnaeus, 1758), from the central Adriatic Sea. <i>Journal of Applied Ecology</i> , 21, 125–130. ; Garrido, S., & Murta, A. G. (2011). Interdecadal and spatial variations of diet composition in horse mackerel <i>Trachurus trachurus</i> . <i>Journal of Fish Biology</i> , 79, 2034–2042. doi: 10.1111/j.1095-8649.2011.03148.x ; Rumolo, P., Basilone, G., Fanelli, E., Barra, M., Calabrò, M., Genovese, S., ... Bonanno, A. (2017). Linking spatial distribution and feeding behavior of Atlantic horse mackerel ( <i>Trachurus trachurus</i> ) in the Strait of Sicily (Central Mediterranean sea). <i>Journal of Sea Research</i> , 121, 47–58. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Trachurus trachurus</i>	Clinidae	0	1	0		
<i>Trachurus trachurus</i>	<i>Alosa alosa</i>	1	1	2		
<i>Trachurus trachurus</i>	<i>Sardina pilchardus</i>	1	0	1		
<i>Trachurus trachurus</i>	<i>Sprattus sprattus</i>	1	0	1		
<i>Trachurus trachurus</i>	<i>Engraulis encrasicolus</i>	1	0	1		
<i>Trachurus trachurus</i>	Clupeidae	1	0	1		
<i>Trachurus trachurus</i>	<i>Trisopterus luscus</i>	2	0	2		
<i>Trachurus trachurus</i>	<i>Merlangius merlangus</i>	1	1	2		
<i>Trachurus trachurus</i>	<i>Merluccius merluccius</i>	1	1	2		
<i>Trachurus trachurus</i>	<i>Pomatoschistus minutus</i>	0	0	0		
<i>Trachurus trachurus</i>	Gobiiformes	0	0	0		
<i>Trachurus trachurus</i>	<i>Solea solea</i>	1	1	2		
<i>Trachurus trachurus</i>	<i>Callionymus lyra</i>	1	0	1		
<i>Trachurus trachurus</i>	<i>Mullus surmuletus</i>	1	1	2		
<i>Trisopterus luscus</i>	<i>Conger conger</i>	2	1	3	Van Den Broek, W. L. F. (1978). Dietary habits of fish populations in the Lower Medway	

<i>Trisopterus luscus</i>	<i>Atherina presbyter</i>	0	0	0	Estuary. Journal of Fish Biology, 13, 645–654. ; Vea Salvanes, A. G. (1986). Preliminary report from a comparative study of the diet of four gadoid fishes in a fjord of western Norway. International Council for the Exploration of the Sea, 1–18. ; Serrano, A., Velasco, F., Olaso, I., & Sánchez, F. (2003). Macrobenthic crustaceans in the diet of demersal fish in the Bay of Biscay in relation to abundance in the environment. Sarsia, 88, 36–48. doi: 10.1080/00364820308469 ; Reubens, J. T., Degraer, S., & Vincx, M. (2011). Aggregation and feeding behaviour of pouting ( <i>Trisopterus luscus</i> ) at wind turbines in the Belgian part of the North Sea. Fisheries Research, 108(1), 223–227. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. Journal of Applied Ecology 57, 1769–1781.
<i>Trisopterus luscus</i>	<i>Trachurus trachurus</i>	2	1	3	
<i>Trisopterus luscus</i>	<i>Alosa alosa</i>	1	1	2	
<i>Trisopterus luscus</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Trisopterus luscus</i>	<i>Sprattus sprattus</i>	1	0	1	
<i>Trisopterus luscus</i>	<i>Pomatoschistus minutus</i>	0	0	0	
<i>Trisopterus luscus</i>	<i>Pomatoschistus</i> sp.	0	0	0	
<i>Trisopterus luscus</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Trisopterus luscus</i>	<i>Scyliorhinus canicula</i>	1	1	2	
<i>Trisopterus luscus</i>	<i>Raja undulata</i>	2	1	3	
<i>Raja undulata</i>	<i>Conger conger</i>	2	1	3	Serrano, A., Velasco, F., Olaso, I., & Sánchez, F. (2003). Macrobenthic crustaceans in the diet of demersal fish in the Bay of Biscay in relation to abundance in the environment. Sarsia, 88, 36–48. doi: 10.1080/00364820308469 ; Moura, T., Figueiredo, I., Farias, I., Serra-pereira, B., Neves, A., de Fátima Borges, M., & Serrano Gordo, L. (2008). Ontogenetic dietary shift and feeding strategy of <i>Raja undulata</i> Lacepède , 1802 (Chondrichthyes : Rajidae) on the Portuguese continental shelf. Scientia Marina, 72, 311–318. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. Journal of Applied Ecology 57, 1769–1781.
<i>Raja undulata</i>	Clinidae	0	1	0	
<i>Raja undulata</i>	<i>Trachurus trachurus</i>	2	1	3	
<i>Raja undulata</i>	<i>Alosa alosa</i>	1	1	2	
<i>Raja undulata</i>	<i>Sardina pilchardus</i>	1	1	2	
<i>Raja undulata</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Raja undulata</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Raja undulata</i>	<i>Pomatoschistus minutus</i>	0	0	0	
<i>Raja undulata</i>	<i>Platichthys flesus</i>	0	1	0	
<i>Raja undulata</i>	<i>Solea solea</i>	1	1	2	
<i>Raja undulata</i>	<i>Callionymus lyra</i>	1	1	2	
<i>Raja undulata</i>	<i>Ammodytes tobianus</i>	0	1	0	
<i>Raja undulata</i>	<i>Scyliorhinus canicula</i>	1	1	2	
<i>Scyliorhinus canicula</i>	<i>Conger conger</i>	2	1	3	
<i>Scyliorhinus canicula</i>	<i>Atherina presbyter</i>	0	1	0	
<i>Scyliorhinus canicula</i>	<i>Trachurus trachurus</i>	2	0	2	
<i>Scyliorhinus canicula</i>	<i>Alosa alosa</i>	1	1	2	
<i>Scyliorhinus canicula</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Scyliorhinus canicula</i>	<i>Sprattus sprattus</i>	1	0	1	

<i>Scyliorhinus canicula</i>	<i>Engraulis encrasicolus</i>	1	0	1	of twenty-three species of chondrichthyes on a feeding ground area. <i>Hydrobiologia</i> , 842, 77–99. ; Martinho, F., Sá, C., Falcão, J., Cabral, H. N., & Pardal, M. Á. (2012). Comparative feeding ecology of two elasmobranch species, <i>Squalus blainville</i> and <i>Scyliorhinus canicula</i> , off the coast of Portugal. <i>Fishery Bulletin</i> , 110, 71–84. ; Mnasri, N., El Kamel, O., Boumaïza, M., Reynaud, C., & Capapé, C. (2012). Food and feeding habits of the small-spotted catshark, <i>Scyliorhinus canicula</i> (Chondrichthyes : Scyliorhinidae) from the northern coast of Tunisia (central Mediterranean). <i>Cahiers de Biologie Marine</i> , 53, 139–150. ; Kousteni, V., Karachle, P. K., & Megalofonou, P. (2017). Diet of the small-spotted catshark <i>Scyliorhinus canicula</i> in the Aegean Sea (eastern Mediterranean). <i>Marine Biology Research</i> , 13, 161–173. ; Wieczorek, A. M., Power, A. M., Browne, P., & Graham, C. T. (2011). Stable-isotope analysis reveals the importance of soft-bodied prey in the diet of lesser spotted dogfish <i>Scyliorhinus canicula</i> . <i>Journal of Fish Biology</i> , 93, 685–693. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Scyliorhinus canicula</i>	<i>Pollachius pollachius</i>	1	1	2	
<i>Scyliorhinus canicula</i>	<i>Trisopterus luscus</i>	2	0	2	
<i>Scyliorhinus canicula</i>	<i>Merluccius merluccius</i>	1	0	1	
<i>Scyliorhinus canicula</i>	<i>Pomatoschistus minutus</i>	0	1	0	
<i>Scyliorhinus canicula</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Scyliorhinus canicula</i>	<i>Pleuronectes platessa</i>	1	0	1	
<i>Scyliorhinus canicula</i>	<i>Solea solea</i>	1	0	1	
<i>Scyliorhinus canicula</i>	<i>Ammodytes tobianus</i>	0	1	0	
<i>Scyliorhinus canicula</i>	<i>Raja undulata</i>	2	0	2	
<i>Sepia officinalis</i>	<i>Conger conger</i>	2	1	3	Castro, B. G., & Guerra, Á. (1990). The diet of <i>Sepia officinalis</i> (Linnaeus, 1758) and <i>Sepia elegans</i> (D’Orbigny, 1835) (Cephalopoda, Sepioidea) from the Ría de Vigo (NW Spain). <i>Scientia Marina</i> , 54, 375–388. ; Pinczon du Sel, G., & Daguzan, J. (1997). A note on sex ratio, length and diet of a population of cuttlefish <i>Sepia officinalis</i> L. (Mollusca : Cephalopoda) sampled by three fishing methods. <i>Fisheries Research</i> , 32, 191–195. ; Pinczon du Sel, G., Blanc, A., & Daguzan, J. (2000). The diet of the cuttlefish <i>Sepia officinalis</i> L. (mollusca : cephalopoda) during its life cycle in the Northern Bay of Biscay (France). <i>Aquatic Sciences</i> , 61, 167–178. ; Alves, D. M., Cristo, M., Sendão, J., & Borges, T. C. (2006). Diet of the cuttlefish <i>Sepia officinalis</i> (Cephalopoda : Sepiidae) off the south coast of Portugal (eastern Algarve). <i>Journal of Marine Biological Association of the United Kingdom</i> , 86, 429–436. ; Neves, A., Sequeira, V., Vieira, A. R., Paiva, R., & Gordo, L. S. (2009). Feeding habits of the cuttlefish <i>Sepia officinalis</i> during its life cycle in the Sado estuary (Portugal). <i>Hydrobiologia</i> , 636, 479–488. ; Thompson, Murray S. A., Pontalier, H., M. A. Spence, J. K. Pinnegar, S. Greenstreet, M. Moriarty, P. Hélaouët, and C. P. Lynam. (2020). A feeding guild indicator to assess environmental change impacts on marine ecosystem structure and functioning. <i>Journal of Applied Ecology</i> 57, 1769–1781.
<i>Sepia officinalis</i>	<i>Atherina presbyter</i>	0	0	0	
<i>Sepia officinalis</i>	<i>Trachurus trachurus</i>	2	0	2	
<i>Sepia officinalis</i>	<i>Sardina pilchardus</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Sprattus sprattus</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Engraulis encrasicolus</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Pollachius pollachius</i>	1	1	2	
<i>Sepia officinalis</i>	<i>Trisopterus luscus</i>	2	0	2	
<i>Sepia officinalis</i>	<i>Merlangius merlangus</i>	1	1	2	
<i>Sepia officinalis</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Sepia officinalis</i>	<i>Pomatoschistus minutus</i>	0	0	0	
<i>Sepia officinalis</i>	<i>Ctenolabrus rupestris</i>	0	0	0	
<i>Sepia officinalis</i>	<i>Labrus bergylta</i>	1	0	1	
<i>Sepia officinalis</i>	Mugilidae	1	1	2	
<i>Sepia officinalis</i>	<i>Dicentrarchus labrax</i>	2	1	3	
<i>Sepia officinalis</i>	<i>Chelidonichthys lucerna</i>	2	1	3	



<i>Sepia officinalis</i>	<i>Pleuronectes platessa</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Solea solea</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Spondyliosoma cantharus</i>	2	0	2	
<i>Sepia officinalis</i>	Sparidae	1	0	1	
<i>Sepia officinalis</i>	<i>Callionymus lyra</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Mullus surmuletus</i>	1	0	1	
<i>Sepia officinalis</i>	<i>Raja undulata</i>	2	1	3	
<i>Sepia officinalis</i>	<i>Torpedo marmorata</i>	0	NA	0	
<i>Echinus esculentus</i>	<i>Liza aurata</i>	1	1	2	
<i>Echinus esculentus</i>	Mugilidae	1	1	2	
<i>Echinus esculentus</i>	<i>Dicentrarchus labrax</i>	2	1	3	Bonsdorff, E. & Vahl, O. (1982) Food preference of the sea urchins <i>Echinus acutus</i> and <i>E. esculentus</i> . Marine Behaviour and Physiology, 8,243-248. ; MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021] ; MARGERIE Bernard, PETIT DE VOIZE Patrice, JEGLOT Samuel in : DORIS, 08/11/2020 : <i>Echinus esculentus</i> Linnaeus, 1758, <a href="https://doris.ffessm.fr/ref/specie/491">https://doris.ffessm.fr/ref/specie/491</a>
<i>Echinus esculentus</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Echinus esculentus</i>	<i>Pleuronectes platessa</i>	1	1	2	
<i>Echinus esculentus</i>	<i>Solea solea</i>	1	1	2	
<i>Echinus esculentus</i>	Sparidae	1	1	2	
<i>Echinus esculentus</i>	<i>Callionymus lyra</i>	1	1	2	
<i>Echinus esculentus</i>	Rajidae	1	1	2	
<i>Psammechinus miliaris</i>	<i>Conger conger</i>	2	1	3	Picton, B. E. (1983). A field guide to the shallow-water echinoderms of the british isles, Immel Publishing, UK, p. 96. ; Gros, P. & Hamon, D. (1988). Typologie biosédimentaire de la baie de Saint-Brieuc (Manche ouest), et estimation de la biomasse des catégories trophiques macrozoobenthiques. Rapport IFREMER, DERO-88-27 EL, Euphorbe. 153 pp. ; MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021] ; Le Pape, O., L. Baulier, A. Cloarec, J. Martin, F. Le Loc'h and Y. Désaunay. (2007) Habitat suitability for juvenile common sole ( <i>Solea solea</i> , L.) in the Bay of Biscay (France): a quantitative description using indicators based on epibenthic fauna. Journal of Sea Research 57,126-136. ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-enclosed sea. Limnology and Oceanography, 62, 2213–2226. ; MARAN Vincent, ZIEMSKI Frédéric, JEGLOT Samuel in : DORIS, 07/11/2020 : <i>Psammechinus miliaris</i> (P.L.S. Müller, 1771), <a href="https://doris.ffessm.fr/ref/specie/133">https://doris.ffessm.fr/ref/specie/133</a>
<i>Psammechinus miliaris</i>	<i>Atherina presbyter</i>	0	1	0	
<i>Psammechinus miliaris</i>	<i>Trachurus trachurus</i>	2	1	3	
<i>Psammechinus miliaris</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Psammechinus miliaris</i>	<i>Engraulis encrasicolus</i>	1	1	2	
<i>Psammechinus miliaris</i>	<i>Trisopterus luscus</i>	2	1	3	
<i>Psammechinus miliaris</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Psammechinus miliaris</i>	Mugilidae	1	1	2	
<i>Psammechinus miliaris</i>	<i>Dicentrarchus labrax</i>	2	1	3	
<i>Psammechinus miliaris</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Psammechinus miliaris</i>	<i>Pleuronectes platessa</i>	1	1	2	
<i>Psammechinus miliaris</i>	<i>Zeugopterus punctatus</i>	0	1	0	



<i>Psammechinus miliaris</i>	<i>Solea solea</i>	1	1	2	
<i>Psammechinus miliaris</i>	<i>Spondyliosoma cantharus</i>	2	1	3	
<i>Psammechinus miliaris</i>	<i>Callionymus lyra</i>	1	1	2	
<i>Psammechinus miliaris</i>	<i>Scyliorhinus canicula</i>	1	1	2	
<i>Psammechinus miliaris</i>	<i>Raja undulata</i>	2	1	3	
<i>Psammechinus miliaris</i>	Rajidae	1	1	2	
<i>Buccinum undatum</i>	<i>Trisopterus luscus</i>	2	1	3	Gros, P. & Hamon, D. (1988). Typologie biosédimentaire de la baie de Saint-Brieuc (Manche ouest), et estimation de la biomasse des catégories trophiques macrozoobenthiques. Rapport IFREMER, DERO-88-27 EL, Euphorbe. 153 pp. ; MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021] ; Le Pape, O., L. Baulier, A. Cloarec, J. Martin, F. Le Loc'h and Y. Désaunay. (2007) Habitat suitability for juvenile common sole ( <i>Solea solea</i> , L.) in the Bay of Biscay (France): a quantitative description using indicators based on epibenthic fauna. Journal of Sea Research 57,126-136. ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-enclosed sea. Limnology and Oceanography, 62, 2213–2226. ; FEY Laurent, BURON Daniel, MÜLLER Yves in : DORIS, 21/01/2021 : <i>Buccinum undatum</i> Linnaeus, 1758, <a href="https://doris.ffessm.fr/ref/specie/860">https://doris.ffessm.fr/ref/specie/860</a>
<i>Atelecyclus undecimdentatus</i>	<i>Conger conger</i>	2	1	3	
<i>Atelecyclus undecimdentatus</i>	<i>Atherina presbyter</i>	0	1	0	
<i>Atelecyclus undecimdentatus</i>	<i>Trachurus trachurus</i>	2	1	3	
<i>Atelecyclus undecimdentatus</i>	<i>Sardina pilchardus</i>	1	1	2	Le Pape, O., L. Baulier, A. Cloarec, J. Martin, F. Le Loc'h and Y. Désaunay. (2007) Habitat suitability for juvenile common sole ( <i>Solea solea</i> , L.) in the Bay of Biscay (France): a quantitative description using indicators based on epibenthic fauna. Journal of Sea Research 57, 126-136. ; NOËL Pierre, ZIEMSKI Frédéric, ANDRÉ Frédéric in : DORIS, 30/01/2021 : <i>Atelecyclus undecimdentatus</i> (Herbst, 1783), <a href="https://doris.ffessm.fr/ref/specie/1187">https://doris.ffessm.fr/ref/specie/1187</a>
<i>Atelecyclus undecimdentatus</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Engraulis encrasicolus</i>	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Pollachius pollachius</i>	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Trisopterus luscus</i>	2	1	3	
<i>Atelecyclus undecimdentatus</i>	<i>Pomatoschistus minutus</i>	0	0	0	
<i>Atelecyclus undecimdentatus</i>	Mugilidae	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Dicentrarchus labrax</i>	2	1	3	

<i>Atelecyclus undecimdentatus</i>	<i>Pleuronectes platessa</i>	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Zeugopterus punctatus</i>	0	1	0	
<i>Atelecyclus undecimdentatus</i>	<i>Solea solea</i>	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Spondyliosoma cantharus</i>	2	1	3	
<i>Atelecyclus undecimdentatus</i>	Sparidae	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Scyliorhinus canicula</i>	1	1	2	
<i>Atelecyclus undecimdentatus</i>	<i>Raja undulata</i>	2	1	3	
<i>Cancer pagurus</i>	<i>Conger conger</i>	2	1	3	
<i>Cancer pagurus</i>	<i>Trachurus trachurus</i>	2	1	3	
<i>Cancer pagurus</i>	<i>Sardina pilchardus</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Engraulis encrasicolus</i>	1	1	2	
<i>Cancer pagurus</i>	Clupeidae	1	1	2	MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021]; Tonk, A. L., & Rozemeijer, M. J. C. (2019). Ecology of the brown crab ( <i>Cancer pagurus</i> ) and production potential for passive fisheries in Dutch offshore windfarms. p. 49. Yerseke: Wageningen Marine Research report C064/19A. ; ZIEMSKI Frédéric, BOUCHARD Jean-Marie, NOËL Pierre in : DORIS, 07/11/2020 : <i>Cancer pagurus</i> Linnaeus, 1758, <a href="https://doris.ffessm.fr/ref/specie/116">https://doris.ffessm.fr/ref/specie/116</a>
<i>Cancer pagurus</i>	<i>Pollachius pollachius</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Trisopterus luscus</i>	2	1	3	
<i>Cancer pagurus</i>	<i>Merlangius merlangus</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Dicentrarchus labrax</i>	2	1	3	
<i>Cancer pagurus</i>	<i>Chelidonichthys lucerna</i>	2	1	3	
<i>Cancer pagurus</i>	<i>Platichthys flesus</i>	0	1	0	
<i>Cancer pagurus</i>	<i>Scyliorhinus canicula</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Raja montagui</i>	1	1	2	
<i>Cancer pagurus</i>	<i>Raja undulata</i>	2	1	3	
<i>Crangon</i> sp.	<i>Conger conger</i>	2	1	3	Gros, P. & Hamon, D. (1988). Typologie biosédimentaire de la baie de Saint-Brieuc (Manche ouest), et estimation de la biomasse des catégories trophiques macrozoobenthiques. Rapport IFREMER, DERO-88-27 EL, Euphorbe. 153 pp. ; Ansell, A.D., Comely, C.A. & Robb, L. (1999) Distribution, movements and diet of macrocrustaceans on a Scottish sandy beach with particular reference to predation on juvenile fishes. Marine Ecology Progress Series. 176,115-130. ; Oh, C., Hartnoll, R. G., & Nash, R. D. M. (2001). Feeding ecology of the common shrimp <i>Crangon crangon</i> in Port Erin Bay, Isle of Man, Irish Sea. Marine Ecology
<i>Crangon</i> sp.	<i>Atherina presbyter</i>	0	1	0	
<i>Crangon</i> sp.	<i>Alosa alosa</i>	1	1	2	
<i>Crangon</i> sp.	<i>Sprattus sprattus</i>	1	1	2	
<i>Crangon</i> sp.	<i>Engraulis encrasicolus</i>	1	1	2	
<i>Crangon</i> sp.	<i>Pollachius pollachius</i>	1	1	2	
<i>Crangon</i> sp.	<i>Trisopterus luscus</i>	2	1	3	

<i>Crangon</i> sp.	<i>Merlangius merlangus</i>	1	1	2	Progress Series, 214, 211–223. ; MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021] ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-enclosed sea. <i>Limnology and Oceanography</i> , 62(5), 2213–2226. ; COROLLA Jean-Pierre, NOËL Pierre, ZIEMSKI Frédéric in : DORIS, 11/11/2020 : <i>Crangon crangon</i> (Linnaeus, 1758), <a href="https://doris.ffessm.fr/ref/specie/1747">https://doris.ffessm.fr/ref/specie/1747</a>
<i>Crangon</i> sp.	<i>Merluccius merluccius</i>	1	1	2	
<i>Crangon</i> sp.	<i>Liza aurata</i>	1	1	2	
<i>Crangon</i> sp.	Mugilidae	1	1	2	
<i>Crangon</i> sp.	<i>Dicentrarchus labrax</i>	2	1	3	
<i>Crangon</i> sp.	<i>Chelidichthys lucerna</i>	2	1	3	
<i>Crangon</i> sp.	<i>Solea solea</i>	1	1	2	
<i>Crangon</i> sp.	<i>Pagrus pagrus</i>	0	1	0	
<i>Crangon</i> sp.	<i>Spondyliosoma cantharus</i>	2	1	3	
<i>Crangon</i> sp.	Sparidae	1	1	2	
<i>Crangon</i> sp.	<i>Callionymus lyra</i>	1	1	2	
<i>Crangon</i> sp.	<i>Mullus surmuletus</i>	1	1	2	
<i>Maja brachydactyla</i>	<i>Conger conger</i>	2	1	3	
<i>Maja brachydactyla</i>	<i>Atherina presbyter</i>	0	1	0	
<i>Maja brachydactyla</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Maja brachydactyla</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Maja brachydactyla</i>	<i>Chelidichthys lucerna</i>	2	1	3	
<i>Maja brachydactyla</i>	<i>Pleuronectes platessa</i>	1	1	2	
<i>Maja brachydactyla</i>	Syngnathidae	0	1	0	
<i>Maja brachydactyla</i>	Rajidae	1	1	2	
<i>Necora puber</i>	<i>Conger conger</i>	2	1	3	Norman, C. P., & Jones, M. B. (1992). Influence of depth, season and moult stage on the diet of the velvet swimming crab <i>Necora puber</i> (Brachyura , Portunidae). <i>Estuarine, Coastal and Shelf Science</i> , 34, 71–83. ; Freire, J., & González-Gurriarán, E. (1995). Feeding ecology of the velvet swimming crab <i>Necora puber</i> in mussel raft areas of the Ría de Arousa (Galicia, NW Spain). <i>Marine Ecology Progress Series</i> , 119, 139–154. ; MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021] ; Giraldo, C., Ernande, B., Cresson, P., Kopp, D., Cachera, M., Travers-Trolet, M., & Lefebvre, S. (2017). Depth gradient on the resource use of a fish community from a semi-
<i>Necora puber</i>	<i>Sprattus sprattus</i>	1	1	2	
<i>Necora puber</i>	<i>Pollachius pollachius</i>	1	1	2	
<i>Necora puber</i>	<i>Trisopterus luscus</i>	2	0	2	
<i>Necora puber</i>	<i>Merlangius merlangus</i>	1	1	2	
<i>Necora puber</i>	<i>Merluccius merluccius</i>	1	1	2	
<i>Necora puber</i>	<i>Liza aurata</i>	1	1	2	
<i>Necora puber</i>	<i>Dicentrarchus labrax</i>	2	1	3	

<i>Necora puber</i>	<i>Chelidonichthys lucerna</i>	2	1	3	enclosed sea. <i>Limnology and Oceanography</i> , 62(5), 2213–2226.
<i>Necora puber</i>	<i>Pleuronectes platessa</i>	1	1	2	
<i>Necora puber</i>	<i>Solea solea</i>	1	1	2	
<i>Necora puber</i>	<i>Salmo trutta</i>	0	1	0	
<i>Necora puber</i>	<i>Spondyliosoma cantharus</i>	2	1	3	
<i>Necora puber</i>	<i>Callionymus lyra</i>	1	1	2	
<i>Necora puber</i>	<i>Scylliorhinus canicula</i>	1	1	2	
<i>Necora puber</i>	<i>Raja undulata</i>	2	1	3	
Paguroidea	<i>Conger conger</i>	2	1	3	
Paguroidea	<i>Belone belone</i>	1	1	2	
Paguroidea	<i>Sprattus sprattus</i>	1	1	2	
Paguroidea	<i>Pollachius pollachius</i>	1	1	2	
Paguroidea	<i>Trisopterus luscus</i>	2	1	3	
Paguroidea	<i>Merlangius merlangus</i>	1	1	2	
Paguroidea	<i>Merluccius merluccius</i>	1	1	2	
Paguroidea	<i>Callionymus lyra</i>	1	1	2	
Paguroidea	<i>Scylliorhinus canicula</i>	1	1	2	
Paguroidea	<i>Raja undulata</i>	2	1	3	
Paguroidea	Rajidae	1	1	2	
<i>Aphrodita aculeata</i>	<i>Pollachius pollachius</i>	1	1	2	Gros, P. & Hamon, D. (1988). Typologie biosédimentaire de la baie de Saint-Brieuc (Manche ouest), et estimation de la biomasse des catégories trophiques macrozoobenthiques. Rapport IFREMER, DERO-88-27 EL, Euphorbe. 153 pp. ; MarLIN. (2006) BIOTIC - Biological Traits Information Catalogue. URL <a href="http://www.marlin.ac.uk/biotic">www.marlin.ac.uk/biotic</a> [accessed 4 April 2021] ; Carlier, A., Riera, P., Amouroux, J.-M., Bodiou, J.-Y. & Grémare, A. (2007) Benthic trophic network in the Bay of Banyuls-sur-Mer (northwest Mediterranean, France): an assessment based on stable carbon and nitrogen isotopes analysis. <i>Estuarine, Coastal and Shelf Science</i> 72,1-15. ; ZIEMSKI Frédéric, MÜLLER Yves in : DORIS, 21/01/2021 : <i>Aphrodita aculeata</i> Linnaeus, 1758, <a href="https://doris.ffessm.fr/ref/specie/149">https://doris.ffessm.fr/ref/specie/149</a>
<i>Aphrodita aculeata</i>	<i>Merluccius merluccius</i>	1	1	2	

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**Table S5:** Weighted percentage of occurrence (wPOO [%]) and frequency of occurrence (FOO [%]) of fish mOTUs (Actinopterygii and Chondrichthyes) in the gut contents of the (a) fish and (b) invertebrates sampled taxa in the Bay of Biscay. Colour code corresponds to the discard probability score (DPS), following: dark blue: DPS = 0 (very low probability of being ingested as discard), light blue: DPS = 1 (low probability), light pink: DPS = 2 (medium probability), dark pink: DPS = 3 (high probability). Bold font indicates fish prey that are under TAC or quota and therefore subject to the European Landing Obligation.







**Table S6:** Centrality metrics (weighted in-degree centrality and eigenvector centrality) of each taxon in the constructed trophic network of a fishing ground in the Bay of Bourgneuf.

<b>Taxon</b>	<b>Category</b>	<b>Class</b>	<b>Weighted in-degree centrality</b>	<b>Eigenvector centrality</b>
<i>Callionymus lyra</i>	Sampled consumer	Actinopterygii	31.3	0.489
<i>Chelidonichthys lucerna</i>	Sampled consumer	Actinopterygii	43.4	0.447
<i>Conger conger</i>	Sampled consumer	Actinopterygii	29	0.632
<i>Merlangius merlangus</i>	Sampled consumer	Actinopterygii	26.1	0.602
<i>Merluccius merluccius</i>	Sampled consumer	Actinopterygii	120	0.785
<i>Pleuronectes platessa</i>	Sampled consumer	Actinopterygii	25.8	0.376
<i>Pollachius pollachius</i>	Sampled consumer	Actinopterygii	20.6	0.341
<i>Trachurus trachurus</i>	Sampled consumer	Actinopterygii	93.8	0.714
<i>Trisopterus luscus</i>	Sampled consumer	Actinopterygii	84.2	0.842
<i>Sepia officinalis</i>	Sampled consumer	Cephalopoda	1.7	0.105
<i>Raja undulata</i>	Sampled consumer	Chondrichthyes	19.6	0.560
<i>Scyliorhinus canicula</i>	Sampled consumer	Chondrichthyes	11.6	0.461
<i>Echinus esculentus</i>	Sampled consumer	Echinoida	0	0.000
<i>Psammechinus miliaris</i>	Sampled consumer	Echinoida	0	0.000
<i>Buccinum undatum</i>	Sampled consumer	Gastropoda	4.7	0.134
<i>Atelecyclus undecimdentatus</i>	Sampled consumer	Malacostraca	170.5	0.593
<i>Cancer pagurus</i>	Sampled consumer	Malacostraca	0	0.000
<i>Crangon</i> sp.	Sampled consumer	Malacostraca	147	0.503
<i>Maja brachydactyla</i>	Sampled consumer	Malacostraca	0	0.000
<i>Necora puber</i>	Sampled consumer	Malacostraca	94.6	0.815
Paguroidea	Sampled consumer	Malacostraca	8.9	0.267
<i>Aphrodita aculeata</i>	Sampled consumer	Polychaeta	0	0.000
<i>Atherina presbyter</i>	Prey mOTU	Actinopterygii	25.6	0.694
<i>Belone belone</i>	Prey mOTU	Actinopterygii	2	0.089
Clinidae	Prey mOTU	Actinopterygii	1.4	0.158
<i>Alosa alosa</i>	Prey mOTU	Actinopterygii	25.2	0.598
<i>Sardina pilchardus</i>	Prey mOTU	Actinopterygii	37.4	0.803
<i>Sprattus sprattus</i>	Prey mOTU	Actinopterygii	107.3	1.000
<i>Engraulis encrasicolus</i>	Prey mOTU	Actinopterygii	48.6	0.589
Clupeidae	Prey mOTU	Actinopterygii	6.9	0.340
<i>Pomatoschistus minutus</i>	Prey mOTU	Actinopterygii	34.1	0.658
<i>Pomatoschistus</i> sp.	Prey mOTU	Actinopterygii	5.6	0.105
Gobiiformes	Prey mOTU	Actinopterygii	1.9	0.089
<i>Ctenolabrus rupestris</i>	Prey mOTU	Actinopterygii	1.8	0.013
<i>Labrus bergylta</i>	Prey mOTU	Actinopterygii	0.5	0.013
<i>Liza aurata</i>	Prey mOTU	Actinopterygii	15.3	0.164
Mugilidae	Prey mOTU	Actinopterygii	10.5	0.150

<i>Dicentrarchus labrax</i>	Prey mOTU	Actinopterygii	21.8	0.307
<i>Platichthys flesus</i>	Prey mOTU	Actinopterygii	2.4	0.070
<i>Zeugopterus punctatus</i>	Prey mOTU	Actinopterygii	3	0.152
<i>Solea solea</i>	Prey mOTU	Actinopterygii	21.7	0.587
<i>Salmo trutta</i>	Prey mOTU	Actinopterygii	7.5	0.101
<i>Pagrus pagrus</i>	Prey mOTU	Actinopterygii	1.1	0.063
<i>Spondylisoma cantharus</i>	Prey mOTU	Actinopterygii	16.4	0.251
Sparidae	Prey mOTU	Actinopterygii	13.9	0.149
<i>Mullus surmuletus</i>	Prey mOTU	Actinopterygii	4	0.164
Syngnathidae	Prey mOTU	Actinopterygii	3.4	0.000
<i>Ammodytes tobianus</i>	Prey mOTU	Actinopterygii	4	0.174
<i>Raja montagui</i>	Prey mOTU	Chondrichthyes	0.8	0.000
Rajidae	Prey mOTU	Chondrichthyes	11.7	0.033
<i>Torpedo marmorata</i>	Prey mOTU	Chondrichthyes	0.5	0.013
<i>Oikopleura</i> sp.	Prey mOTU	Appendicularia	1	0.000
<i>Molgula socialis</i>	Prey mOTU	Ascidiacea	4.8	0.000
<i>Branchiostoma lanceolatum</i>	Prey mOTU	Cephalochordata	1.2	0.042
<i>Notomastus latericeus</i>	Prey mOTU	Polychaeta	2.1	0.103
Eunicida	Prey mOTU	Polychaeta	1.9	0.122
<i>Golfingia elongata</i>	Prey mOTU	Polychaeta	0.9	0.057
<i>Golfingia</i> sp.	Prey mOTU	Polychaeta	0.9	0.057
<i>Sipunculus nudus</i>	Prey mOTU	Polychaeta	1.8	0.047
Scolecida	Prey mOTU	Polychaeta	6.3	0.000
<i>Thoracophelia</i> sp.	Prey mOTU	Polychaeta	1.6	0.070
<i>Glycera</i> sp.	Prey mOTU	Polychaeta	10.8	0.100
<i>Glycinde armigera</i>	Prey mOTU	Polychaeta	3.2	0.063
<i>Nephtys hombergii</i>	Prey mOTU	Polychaeta	2.9	0.104
<i>Nephtys</i> sp.	Prey mOTU	Polychaeta	11.4	0.297
Nereididae	Prey mOTU	Polychaeta	5.4	0.260
<i>Paranaitis kosteriensis</i>	Prey mOTU	Polychaeta	0.6	0.042
<i>Phyllodoce longipes</i>	Prey mOTU	Polychaeta	0.4	0.089
<i>Alentia gelatinosa</i>	Prey mOTU	Polychaeta	0.8	0.105
<i>Harmothoe impar</i>	Prey mOTU	Polychaeta	0.4	0.000
<i>Harmothoe</i> sp.	Prey mOTU	Polychaeta	3.8	0.094
Polynoidae	Prey mOTU	Polychaeta	1.3	0.061
<i>Owenia</i> sp.	Prey mOTU	Polychaeta	0.6	0.047
<i>Sabellaria</i> sp.	Prey mOTU	Polychaeta	1	0.000
Serpulidae	Prey mOTU	Polychaeta	15.4	0.000
<i>Ampharete</i> sp.	Prey mOTU	Polychaeta	2.9	0.061
<i>Lanice conchilega</i>	Prey mOTU	Polychaeta	2.2	0.060
<i>Pista cristata</i>	Prey mOTU	Polychaeta	1.1	0.061
Terebellidae	Prey mOTU	Polychaeta	1.2	0.105
<i>Terebellides</i> sp.	Prey mOTU	Polychaeta	0.9	0.000

<i>Metacrangon</i> sp.	Prey mOTU	Malacostraca	3.1	0.063
<i>Eualus cranchii</i>	Prey mOTU	Malacostraca	5	0.193
<i>Hippolyte varians</i>	Prey mOTU	Malacostraca	3.5	0.236
<i>Ebalia</i> sp.	Prey mOTU	Malacostraca	6.3	0.074
Axiidea	Prey mOTU	Malacostraca	1.3	0.105
Caridea	Prey mOTU	Malacostraca	32.1	0.615
Heterotremata	Prey mOTU	Malacostraca	1.8	0.079
Majoidea	Prey mOTU	Malacostraca	12.9	0.407
Pleocyemata	Prey mOTU	Malacostraca	23.3	0.033
Palaemonidae	Prey mOTU	Malacostraca	5.7	0.192
Portunidae	Prey mOTU	Malacostraca	10.3	0.074
<i>Nikoides</i> sp.	Prey mOTU	Malacostraca	5	0.162
<i>Upogebia</i> sp.	Prey mOTU	Malacostraca	35	0.304
<i>Gastrosaccus spinifer</i>	Prey mOTU	Malacostraca	25.5	0.520
<i>Haplostylus dispar</i>	Prey mOTU	Malacostraca	7	0.248
<i>Paramysis</i> sp.	Prey mOTU	Malacostraca	3.3	0.162
<i>Schistomysis kervillei</i>	Prey mOTU	Malacostraca	1.3	0.160
<i>Siriella</i> sp.	Prey mOTU	Malacostraca	1	0.042
Eucarida	Prey mOTU	Malacostraca	3.5	0.168
Eumalacostraca	Prey mOTU	Malacostraca	141.3	0.779
Unipeltata	Prey mOTU	Malacostraca	5.8	0.057
<i>Acartia</i> sp.	Prey mOTU	Maxillopoda	118.6	0.754
<i>Centropages typicus</i>	Prey mOTU	Maxillopoda	1	0.000
Clausocalanidae	Prey mOTU	Maxillopoda	5.8	0.241
<i>Paracalanus</i> sp.	Prey mOTU	Maxillopoda	8.9	0.000
<i>Nitokra</i> sp.	Prey mOTU	Maxillopoda	2.5	0.074
<i>Euterpina acutifrons</i>	Prey mOTU	Maxillopoda	11.7	0.317
Harpacticoida	Prey mOTU	Maxillopoda	16.7	0.158
Maxillopoda	Prey mOTU	Maxillopoda	0.7	0.042
Neocopepoda	Prey mOTU	Maxillopoda	28	0.481
<i>Ditrichocorycaeus anglicus</i>	Prey mOTU	Maxillopoda	1.2	0.102
<i>Amphibalanus</i> sp.	Prey mOTU	Maxillopoda	0.7	0.042
Sessilia	Prey mOTU	Maxillopoda	1.4	0.055
<i>Cytheropteron</i> sp.	Prey mOTU	Ostracoda	1.8	0.000
<i>Leptocythere lacertosa</i>	Prey mOTU	Ostracoda	1.3	0.047
<i>Paradoxostoma setoense</i>	Prey mOTU	Ostracoda	2	0.061
Exochellidae	Prey mOTU	Gymnolaemata	0.5	0.000
Hormathiidae	Prey mOTU	Anthozoa	1.3	0.033
Actiniaria	Prey mOTU	Anthozoa	9.3	0.057
Hexacorallia	Prey mOTU	Anthozoa	0.7	0.057
Anthoathecata	Prey mOTU	Hydrozoa	2.2	0.000
Leptothecata	Prey mOTU	Hydrozoa	0.5	0.000
<i>Plumularia</i> sp.	Prey mOTU	Hydrozoa	0.4	0.000

Siphonophorae	Prey mOTU	Hydrozoa	8.5	0.000
Scyphozoa	Prey mOTU	Scyphozoa	0.9	0.101
Semaeostomeae	Prey mOTU	Scyphozoa	1.5	0.176
Pleurobrachiidae	Prey mOTU	Tentaculata	4.1	0.131
Asteroidea	Prey mOTU	Asteroidea	3	0.013
Echinoidea	Prey mOTU	Echinoidea	7.9	0.234
Eleutherozoa	Prey mOTU	Eleutherozoa	2.1	0.000
Ophiurida	Prey mOTU	Ophiuroidea	4.4	0.108
<i>Barnea candida</i>	Prey mOTU	Bivalvia	1.1	0.061
Mytilidae	Prey mOTU	Bivalvia	1.8	0.000
Euheterodonta	Prey mOTU	Bivalvia	0.9	0.101
Pteriomorphia	Prey mOTU	Bivalvia	8.1	0.074
Nuculidae	Prey mOTU	Bivalvia	1.6	0.000
Cardiidae	Prey mOTU	Bivalvia	1.8	0.057
<i>Cerastoderma edule</i>	Prey mOTU	Bivalvia	9	0.180
<i>Donax</i> sp.	Prey mOTU	Bivalvia	2.3	0.047
<i>Lutraria lutraria</i>	Prey mOTU	Bivalvia	1.9	0.057
Mactridae	Prey mOTU	Bivalvia	10.4	0.047
Tellinoidea	Prey mOTU	Bivalvia	2.9	0.063
<i>Ensis</i> sp.	Prey mOTU	Bivalvia	1	0.047
<i>Pharus legumen</i>	Prey mOTU	Bivalvia	1.3	0.061
Sepiolinae	Prey mOTU	Cephalopoda	0.6	0.042
Gastropoda	Prey mOTU	Gastropoda	25	0.017
Hypsogastropoda	Prey mOTU	Gastropoda	2	0.061
<i>Camacolaimus</i> sp.	Prey mOTU	Gastropoda	1.8	0.000
Leptolaimidae	Prey mOTU	Chromadorea	2.6	0.000
<i>Onchium</i> sp.	Prey mOTU	Chromadorea	1.1	0.000
<i>Viscosia</i> sp.	Prey mOTU	Enopla	3.5	0.061
<i>Cerebratulus lacteus</i>	Prey mOTU	Anopla	0.8	0.057
Heteronemertea	Prey mOTU	Anopla	3.1	0.193
Eumonostilifera	Prey mOTU	Enopla	1.8	0.057
Suberitida	Prey mOTU	Demospongiae	0.5	0.000
Ploima	Prey mOTU	Monogononta	1.3	0.075
Bilateria	Prey mOTU	Bilateria	15	0.089
Eumetazoa	Prey mOTU	Eumetazoa	55.6	0.586
Lophotrochozoa	Prey mOTU	Lophotrochozoa	2	0.061