

A simple model of the electrostatic environment around the catalytic center of the ribosome and its significance for the elongation cycle kinetics

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INTRODUCTION

Ribosome Catalyzes Peptide Bond Formation

- The mechanisms of peptide bond formation at the **peptidyl transferase center (PTC)** have been studied for decades [1]
- There is a great **variation in peptide bond formation rate** during mRNA translation
- **The kinetics and the factors** that **impact** on the peptide bond formation rates **are not fully understood**

Key determinants of Peptide Bond Formation Rate

- **Nature of the amino acid** at A-site and **C-terminal amino acid** of the nascent chain **at the P-site**
- **Charged amino acids** interaction with **electric field at the catalytic center** of the ribosome
- Electrostatic interaction of the nascent chain with the **ribosome exit tunnel**
- **Proline** at the A site and/or the P-site

Approaches of this study

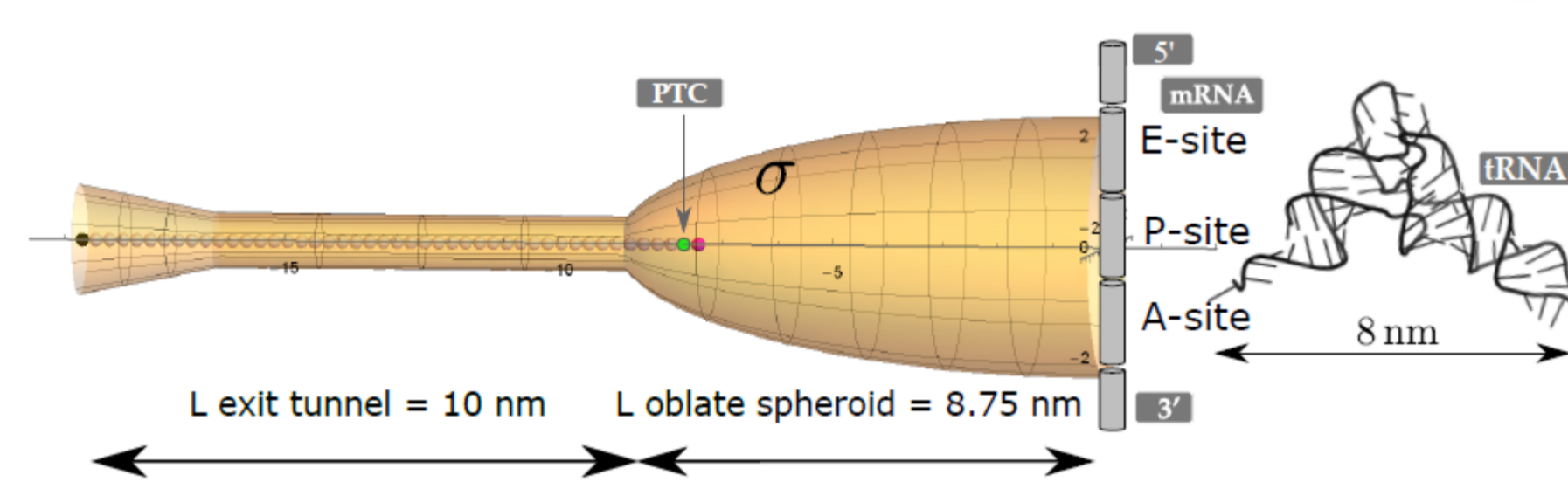
- **3D-Map** the charged groups that are close to the PTC from X-Ray crystallography
- **Relate** the **electrostatic potential profile** around the catalytic center of the ribosome to X-Ray solved structures
- Incorporate **mechano-chemistry** into the **transition state energy barrier** of the catalytic reaction and the kinetics calculations

MATERIALS & METHODS

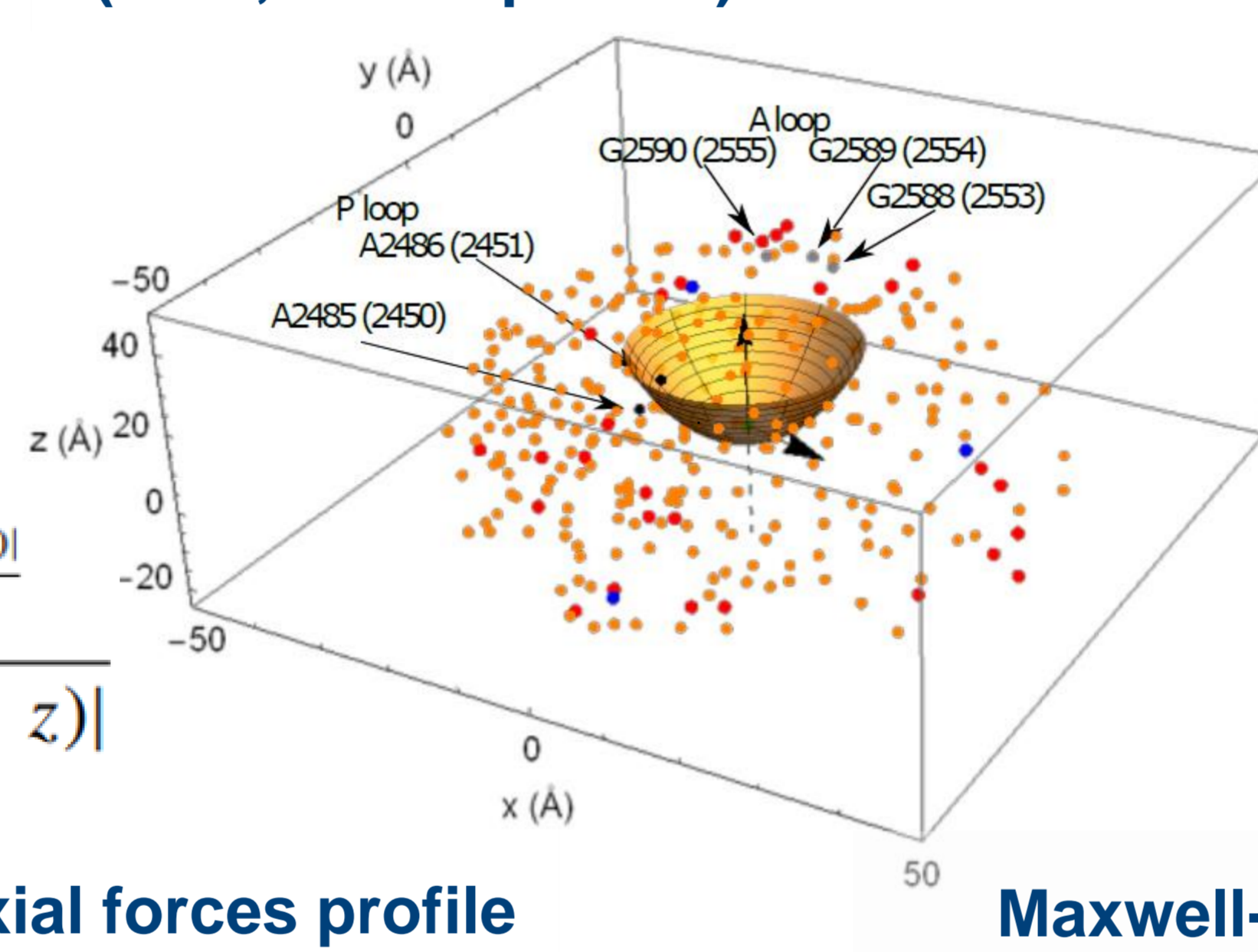
Ribosome large subunit X-Ray crystallography data mining and queueing time theory of the elongation cycle

- ✓ Our approach is to 3D map the charged groups (phosphate moieties of 23S/28S rRNA and charged amino acids from ribosomal proteins)
- ✓ Empirical parameters for dielectric response and screening lengths are estimated [2, 3]
- ✓ The electrostatic potential around the PTC cavity is calculated by the Yukawa-Debye-Hückel theory from the charged sources positions
- ✓ Comparison with previous experimental results of *Wohlgemuth et al.* [4] of elongation rate with the use of puromycin as A-site acceptor
- ✓ The elongation cycle queueing time is analyzed as the convolution product of 3 queueing times of the 3 sequential steps of elongation.

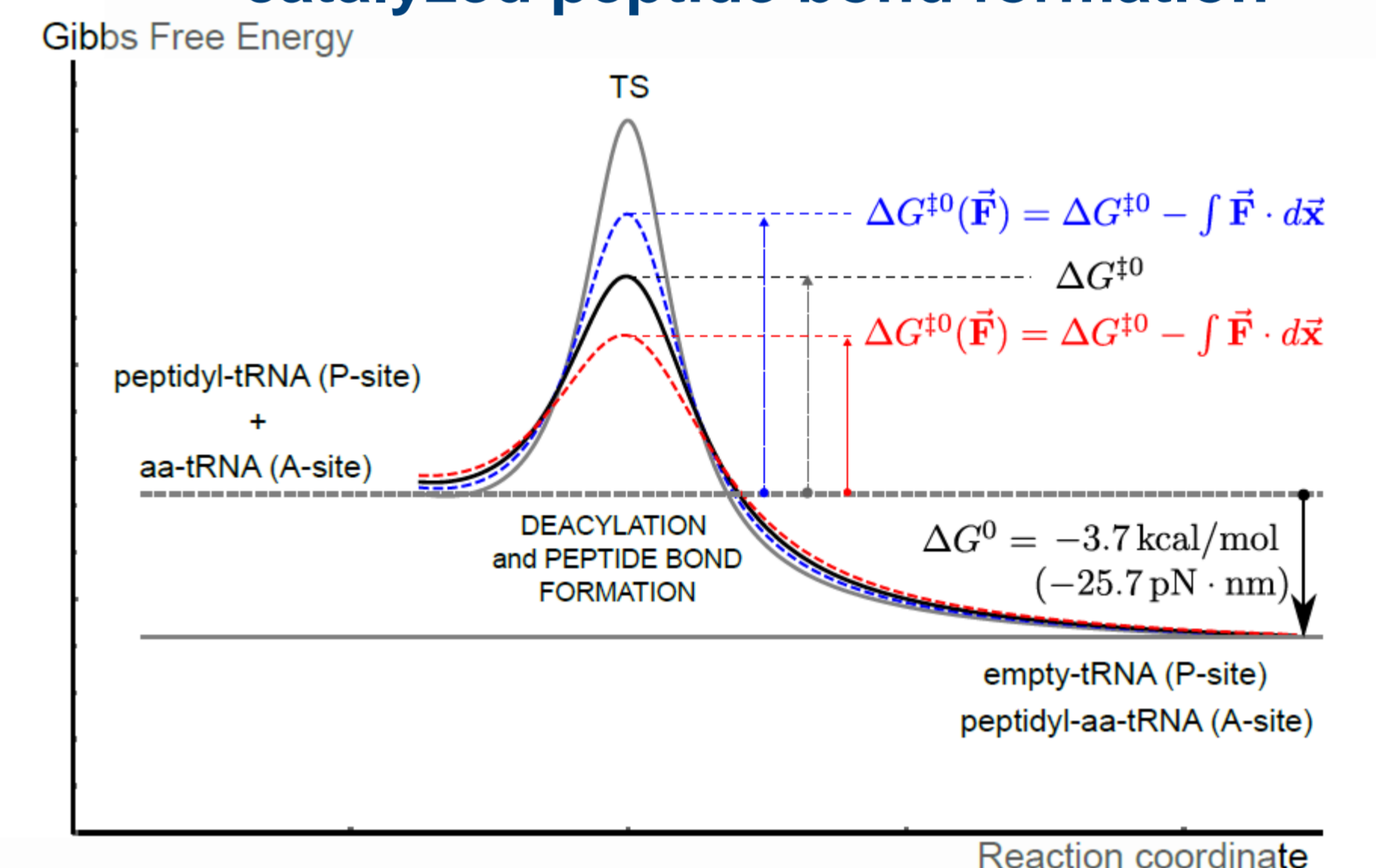
$$\Phi_{Yuk}(\vec{r}) = \iint_S \frac{\sigma^*(\vec{r}') da}{4\pi \epsilon_0} \cdot \frac{e^{-\frac{|\vec{r}-\vec{r}'|}{\xi}}}{|\vec{r}-\vec{r}'|}$$



Mapping the phosphate moieties (orange spheres) and amino acids (red+, blue- spheres) around the PTC

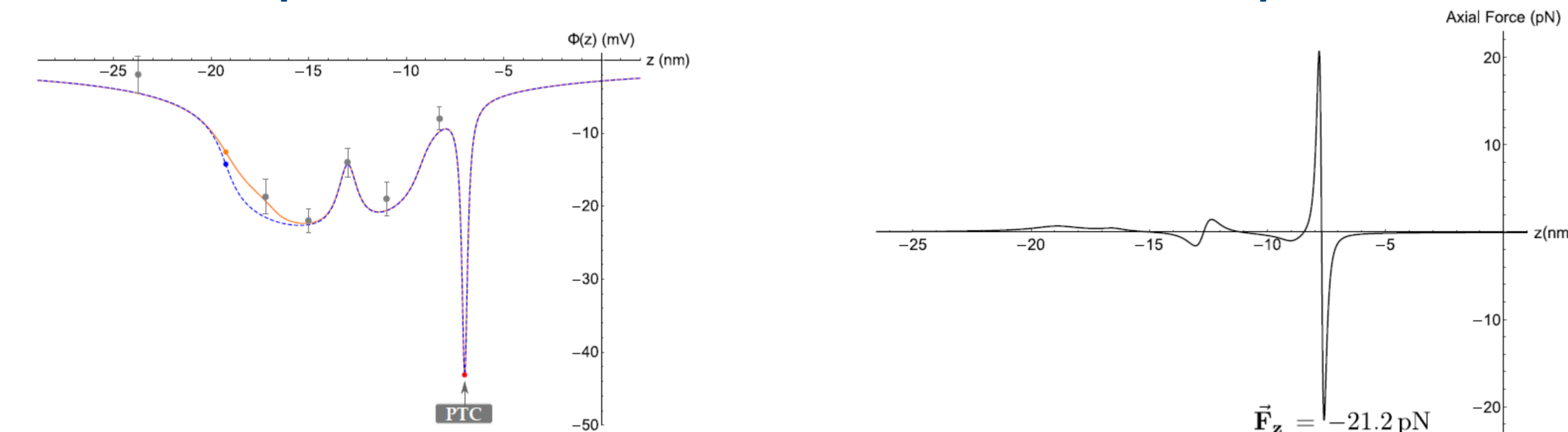


Mechano-chemistry modulates the transition state energy barrier of the catalyzed peptide bond formation



$$\Phi(z) = \sum_{k \in \text{regions}} \sum_{i \in \text{charged sources}} \frac{q_{i,k}}{4\pi \epsilon_0 \epsilon_r(k)} \frac{e^{-\frac{|r'_i - (0,0,z)|}{\xi_k}}}{|r'_i - (0,0,z)|}$$

Electrostatic potential model around the PTC and axial forces profile

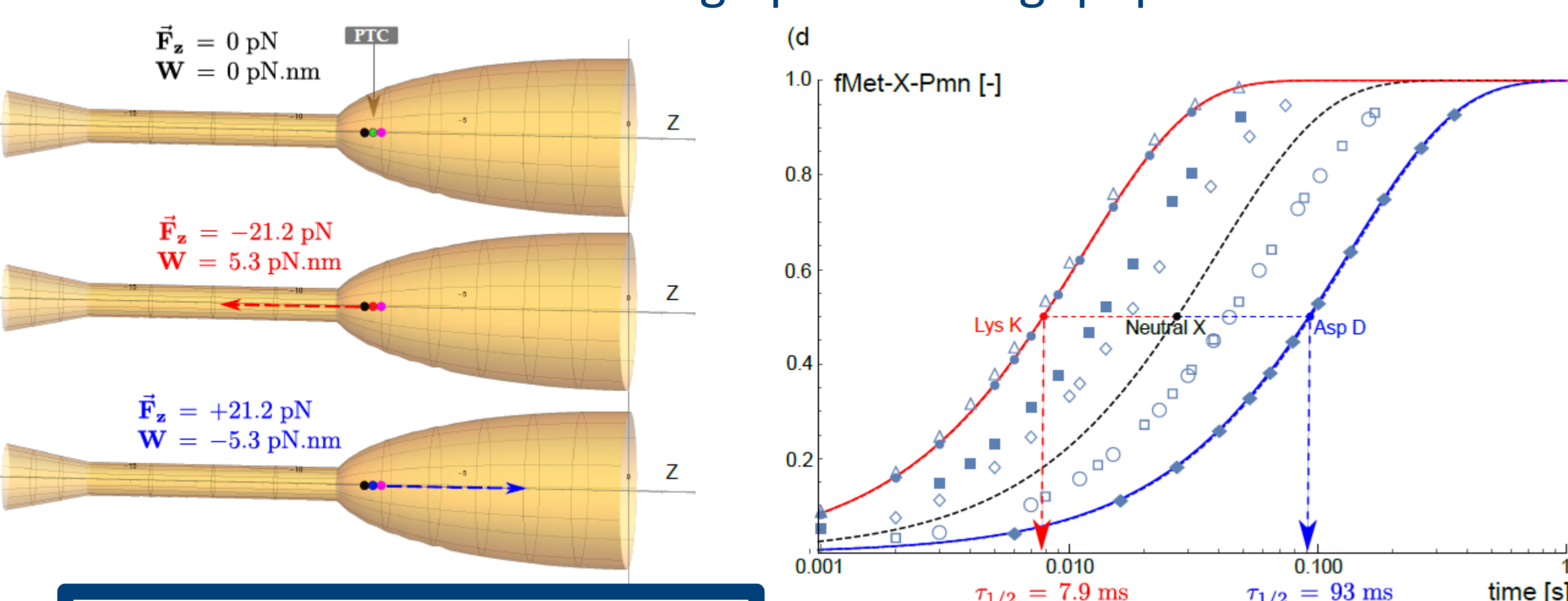


Maxwell-Boltzmann factor incorporating mechanical effects into the catalyzed reaction rate constant

$$k(\vec{F}) = k(0) \cdot e^{-\frac{\Delta G^{0\dagger}}{k_B \cdot T}} \cdot e^{+\frac{\int \vec{F} \cdot dx^\dagger}{k_B \cdot T}}$$

RESULTS AND PERSPECTIVES

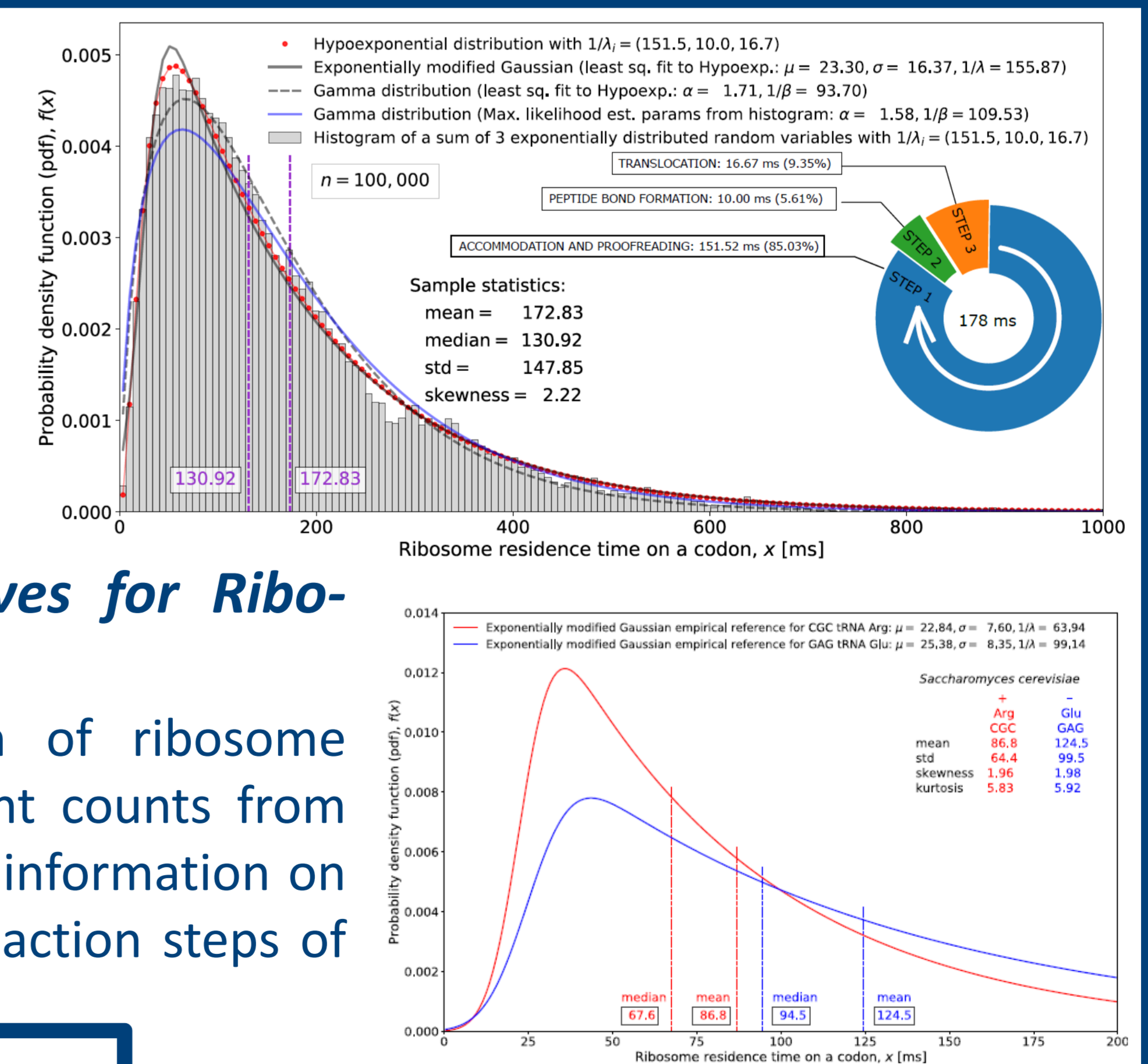
- ✓ The calculated electric field order of magnitude at the catalytic center of the ribosome (PTC) is 143 MV/cm, similar to the one observed in typical protein enzyme-substrate configuration [3, 5]
- ✓ The axial force around the PTC towards the exit tunnel is 21.2 pN for a positive unit test charge [3].
- ✓ The fold change in the peptide bond formation rate between positively and negatively charged amino acid as C-terminal residue at the P-site is consistent with the Maxwell-Boltzmann factor accounting for the mechanical forces acting upon the oligopeptide backbone at the P-site.



Biophysics and biochemistry

Computational Biology Perspectives for RiboSeq analysis

We suggest that the deconvolution of ribosome residence time or normalized footprint counts from **RiboSeq** data [6] can provide valuable information on the kinetics of the three sequential reaction steps of the elongation cycle.



REFERENCES

- [1] Rodnina et al., 2006, *Quat. Rev. of Biophys.*, 39(3) [2] Joiret et al., 2022, *Phys. Rev. E*, 105(1) [3] Joiret et al., 2022, *bioRxiv* <https://doi.org/10.1101/2022.03.07.483325> [4] Wohlgemuth et al., 2008, *J. Biol. Chem.*, 283 [5] Fried & Boxer., 2017, *An. Rev. of Biochem.*, 86 [6] Dana & Tuller., 2014, *Nucleic Acids Res.*, 42(9171).

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