

1 **Distinct blood protein profiles associated with the risk of short-term and mid/long-term**
2 **clinical relapse in Crohn's disease patients stopping infliximab: when the remission state**
3 **hides different types of residual disease activity**

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35 **ABSTRACT**

36 **Objective**

37 Despite being in sustained and stable remission, Crohn's disease (CD) patients stopping anti-
38 TNF α show a high rate of relapse (~50% within 2 years). Characterising non-invasively the
39 biological profiles of those patients is needed to better guide the decision of anti-TNF α
40 withdrawal.

41 **Design**

42 Ninety-two immune-related proteins were measured by proximity extension assay in serum of
43 CD patients (n=102) in sustained steroid-free remission and stopping anti-TNF α (infliximab).
44 As previously shown, a stratification based on time to clinical relapse was used to characterise
45 the distinct biological profiles of relapsers (short-term relapsers: <6 months vs mid/long-term
46 relapsers: >6 months). Associations between protein levels and time to clinical relapse were
47 determined by univariable Cox model.

48 **Results**

49 The risk (HR: hazard ratio) of mid/long-term clinical relapse was specifically associated with a
50 high serum level of proteins mainly expressed in lymphocytes (LAG3, SH2B3, SIT1; HR: 2.2-
51 4.5; p<0.05), a low serum level of anti-inflammatory effectors (IL-10, HSD11B1; HR: 0.2-0.3;
52 p<0.05) and cellular junction proteins (CDSN, CNTNAP2, CXADR, ITGA11; HR: 0.4;
53 p<0.05). The risk of short-term clinical relapse was specifically associated with a high serum
54 level of pro-inflammatory effectors (IL-6, IL12RB1; HR: 3.5-3.6; p<0.05) and a low or high
55 serum level of proteins mainly expressed in antigen presenting cells (CLEC4A, CLEC4C,
56 CLEC7A, LAMP3; HR: 0.4-4.1; p<0.05).

57 **Conclusion**

58 We identified distinct blood protein profiles associated with the risk of short-term and mid/long-
59 term clinical relapse in CD patients stopping infliximab. These findings constitute an advance
60 for the development of non-invasive biomarkers guiding the decision of anti-TNF α withdrawal.

61 **What is already known on this topic?**

- 62 ▶ Crohn's disease (CD) patients achieving sustained remission and stopping anti-TNF α
63 show a high rate of relapse (~50% within 2 years).
64 ▶ Characterising non-invasively the biological profiles of relapsers is needed to better
65 guide the decision of anti-TNF α withdrawal.

66 **What this study adds**

- 67 ▶ Blood proteins involved in inflammatory processes, anti-inflammatory defences,
68 cellular junctions and immunoregulation of immune cells showed distinct associations
69 with the risk of short-term (<6 months) and mid/long-term (>6 months) clinical relapse
70 following anti-TNF α withdrawal in CD patients.
71 ▶ The biological profiles of short-term and mid/long-term relapsers can be considered as
72 different degrees of unstable biological remission characterised by different types of
73 residual disease activity.

74 **How this study might affect research, practice or policy**

- 75 ▶ These findings represent a biological basis to develop non-invasive biomarkers guiding
76 the decision of anti-TNF α withdrawal in CD patients.

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87 INTRODUCTION

88 In Crohn's disease (CD), anti-tumour necrosis factor α (TNF α) therapy is well-known to
89 induce stable remission in a subset of patients[1]. When this objective is achieved, anti-TNF α
90 withdrawal can be considered and this choice could offer substantial benefits in terms of safety
91 and cost. Indeed, chronic exposure to anti-TNF α has been linked to serious complications (eg,
92 lymphoma, infections, melanoma and non-melanoma skin cancers)[2–5] and this treatment is a
93 burden for the healthcare system[6]. These arguments need to be balanced by the fact that CD
94 patients stopping anti-TNF α present a high rate of relapse (~50% within 2 years according to a
95 meta-analysis)[7]. This risk shows a remarkable homogeneity across studies and arises despite
96 that, at time of anti-TNF α withdrawal, most patients (>80%) are in sustained remission
97 according to clinical, endoscopic and biomarker (CRP and fecal calprotectin) criteria
98 classically used[7]. This situation underlines that the current definition of remission is not well
99 adapted to guide the decision of anti-TNF α withdrawal. Such option could be better considered
100 by knowing the biological profiles of relapsers.

101 We previously reported that, in CD patients stopping anti-TNF α (infliximab), the risk of
102 short-term (<6 months) and mid/long-term clinical relapse (>6 months) were associated with
103 specific serum proteins reflecting distinct pathological process[8]. This result was a first
104 advance to identify the biological profiles of relapsers. The aim of the present study was to
105 improve this knowledge by further characterising the blood protein profiles of short-term and
106 mid/long-term relapsers following infliximab withdrawal.

107 In our above mentioned study, the relatively low analytical sensitivity (down to ~1 $\mu\text{g.mL}^{-1}$)
108 of the measurement method (crude serum analysed by selected reaction monitoring, SRM, a
109 mass spectrometry-based technology)[8,9] led us to measure mainly the abundant and liver-
110 produced proteins of the serum (54 out of the 72 measured proteins). In the present study, we
111 hypothesised that targeting low abundance serum proteins which do not mainly reflect a liver

112 response could reveal novel information complementary to our previous work. To test this
113 assumption, we took advantage of the proximity extension assay (PEA) technology (Olink)
114 which, by using the amplification system of polymerase chain reaction (PCR), reaches a
115 sensitivity allowing to measure low-abundant proteins of human blood (down to $\sim 1 \text{ pg.mL}^{-1}$)
116 ¹[10,11]. We selected the immune response panel (n=92 proteins) which targets some
117 processes related to the development and perpetuation of CD: inflammation, cytokine-mediated
118 signalling pathways, adaptive immune response, defence response to virus, lymphocyte
119 activation (Olink). This panel contains a low number of liver-produced proteins (3/92). As in
120 our previous work[8], proteins were measured in the baseline serum of patients included in the
121 study of infliximab discontinuation in Crohn's disease patients in stable Remission on
122 combined therapy with Immunosuppressors (STORI), a trial set up to identify risk factors for
123 clinical relapse in CD patients stopping infliximab[12].

124

125 **METHODS**

126 **Patients and samples**

127 STORI is a study designed by the Groupe d'Etude Thérapeutique des Affections Inflammatoires
128 du tube Digestif (GETAID) in which CD patients were prospectively recruited across 20 centres
129 in Belgium and France between March 2006 and December 2009[12]. The inclusion and
130 exclusion criteria have been detailed previously[12]. Briefly, included patients were under a
131 combined therapy (infliximab and antimetabolites for at least 1 year) and in corticosteroid-free
132 clinical remission (CD activity index: CDAI<150 for at least 6 months) when they stopped
133 infliximab (baseline). At baseline, sera were obtained after centrifugation of coagulated blood
134 and stored at $-80 \text{ }^{\circ}\text{C}$ until analysis. A clinical relapse was declared when patients presented a
135 CDAI higher than 250 or a CDAI between 150 and 250 with an increase of 70 point from
136 baseline over two consecutive weeks. The data presented in this study were obtained from 102

137 out of the initial 115 patients of the STORI cohort (Table 1). The study was approved by the
138 French Ethics Committee-Hôpital Saint-Louis (CPP 2005/14) and the AFSSAPS
139 (0809/ALV/EG05). The investigational review board at each of the participating centre
140 approved the protocol and, before screening, all patients gave their written informed consent.
141 Patients or the public were not involved in the design, or conduct, or reporting, or dissemination
142 plans of our research.

143

144 **Proximity extension assay**

145 The serum abundance of 92 proteins were measured by PEA (immune response panel, Olink,
146 Sweden). For each protein, the analytic performances of the assay (limit of detection, lower
147 limit of quantification, upper limit of quantification, hook effect, dynamic range, intra-assay
148 and inter-assay run precision) are available on the manufacturer's website
149 (<https://www.olink.com/>). The principle of PEA technology has been previously explained[11].
150 Briefly, this method relies on dual recognition of protein by a pair of antibodies labelled with
151 complementary single-stranded DNA oligonucleotides. These PEA probes become close when
152 antibody pair binds to its target, this allows their hybridization and subsequent elongation by a
153 DNA polymerase thus resulting in the synthesis of a double-stranded DNA with a unique
154 sequence for each targeted protein (barcode). This DNA template is then amplified and
155 quantified by quantitative PCR. Data are normalised with internal and external controls to
156 reduce technical variability, they are finally reported on a Log₂ scale with an arbitrary unit
157 called normalized protein expression (NPX)[11].

158

159 **Statistical analysis**

160 Among the 102 studied patients, 44 were declared relapsers (R) and 58 were declared non-
161 relapsers (NR). To study separately the short-term relapsers (<6 months) from the mid/long-

162 term relapsers (>6 months), the cohort was stratified according to a time to clinical relapse of 6
163 months as previously described[8]. In the short-term clinical relapse dataset, the follow-up of
164 non-relapsers was censored at 6 months and mid/long-term relapsers (n=29) were considered
165 as non-relapsers. In the mid/long-term clinical relapse dataset, non-relapsers presenting a
166 follow-up inferior to 6 months (n=13) and short-term relapsers (n=15) were excluded. Thus, the
167 short-term clinical relapse dataset and the mid/long-term clinical relapse datasets were
168 composed of 102 (15 R, 87 NR) and 74 (29 R, 45 NR) patients, respectively.

169 For each targeted protein, an optimal cut-off value was selected as the one maximizing the
170 sum of sensitivity and specificity (Youden's index) in classifying relapsers versus non-
171 relapsers. By using these cut-offs, the univariable Cox model was applied to determine the
172 association of each protein with the time to clinical relapse, this relation was depicted by the
173 hazard ratio (HR=hazard in patients above the cut-off/hazard in patients under the cut-off). The
174 proportional hazard assumption was verified by a statistical test (Schoenfeld's residuals) and
175 by a graphical criterion (crossing of survival curves). The non-respect of this assumption (HR
176 not constant over time) led to exclusion of some variables (mid/long-term clinical relapse
177 dataset: ITM2A, LAMP3; non-stratified dataset: TRIM21, IL-10, IRAK1, ITGA6, TRIM5,
178 PRDX5, IRF9, CDSN, DDX58).

179 The lifelines Python library and the ggplot2 R package were used to perform the Cox model
180 and to generate volcano plots, respectively[13,14]. The p-values inferior to 0.05 were
181 considered significant.

182

183 **Strategy of analysis to search biological patterns differentiating the short-term relapsers** 184 **from the mid/long-term relapsers**

185 Proteins only associated with the risk of short-term clinical relapse (n=25) were compared to
186 those only associated with the risk of mid/long-term clinical relapse (n=17) (online

187 supplemental table 1). This research was conducted by investigating heterogeneous information
188 related to the studied proteins: 1) functions, cellular origin and involvement in signalling
189 pathways which were determined with the help of literature and databases (Uniprot, Human
190 Protein Atlas, Kyoto Encyclopedia of Genes and Genomes, Reactome) (online supplemental
191 table 1); 2) association between the risk of clinical relapse and the serum level (high: HR>1;
192 low: HR<1; online supplemental table 1); 3) links with CD pathophysiology; 4) biological
193 convergences with our previous findings[8]. By integrating this prior knowledge, we found
194 relevant to focus our analysis on: pro-inflammatory and anti-inflammatory effectors, proteins
195 mainly expressed in antigen presenting cells (APCs) or lymphocytes, cellular junction proteins,
196 downstream signalling of cytokine receptors and pattern recognition receptors. The information
197 used to classify proteins in the abovementioned categories are provided in online supplemental
198 tables 2, 3, 4, 5.

199 The proteins not highlighted in the core manuscript and showing a significant association
200 with the risk of short-term and/or mid/long-term clinical relapse (n=19) were included in the
201 online supplemental table 1 and the online supplemental figure 1.

202

203 **RESULTS**

204 **Study population, experimental design and outcomes of patients after infliximab** 205 **withdrawal**

206 The clinical characteristics of the study population are presented in table 1. The Figure 1
207 presents the experimental design and outcomes of patients after infliximab withdrawal. After
208 stopping infliximab, 44 patients relapsed and 58 did not relapse with a median (IQR) time of
209 clinical relapse of 6.7 (4.2-11.2) months and a median (IQR) time of follow-up of 25.4 (7.9-
210 30.1) months, respectively. Among relapsers, 15 were considered short-term relapsers (< 6

211 months) and 29 mid/long-term relapsers (> 6 months) with a median (IQR) time of clinical
212 relapse of 3.6 (2.8-4.1) and 9.8 (6.7-12.5), respectively (figure 1).

213

214 **Risk of clinical relapse associated with pro-inflammatory and anti-inflammatory effectors**

215 Among the 6 pro-inflammatory or anti-inflammatory effectors of the PEA panel (online
216 supplemental table 2), 4 showed an association with the risk of clinical relapse: interleukin-6
217 (IL-6); interleukin-12 receptor subunit beta-1 (IL12RB1); corticosteroid 11-beta-
218 dehydrogenase isozyme 1 (HSD11B1), interleukin-10 (IL-10) (online supplemental table 1).
219 Remarkably, the risk of mid/long-term clinical relapse was associated with a low serum level
220 of anti-inflammatory effectors (IL-10 and HSD11B1) while the risk of short-term clinical
221 relapse was associated with high serum level of pro-inflammatory effectors (IL-6 and IL12RB1)
222 (figure 2A,B; online supplemental table 1).

223

224 **Risk of clinical relapse associated with proteins mainly expressed in antigen presenting 225 cells or lymphocytes**

226 Some proteins of the PEA panel are mainly expressed in lymphocytes or APCs where they exert
227 immunoregulatory functions since, most of them (17/18), are known to promote immunity
228 and/or tolerance (online supplemental table 3). Among the proteins mainly expressed in APCs,
229 2 were associated with the risk of mid/long-term clinical relapse (C-type lectin domain family
230 4 member G, CLEC4G; Allergin-1, MILR1; figure 3A,B) while 6 were associated with the risk
231 of short-term clinical relapse (C-type lectin domain family 4 member A, CLEC4A; C-type
232 lectin domain family 4 member C, CLEC4C; CLEC4G; C-type lectin domain family 7 member
233 A, CLEC7A; MILR1; lysosome-associated membrane glycoprotein 3, LAMP3; figure 3A,B).
234 Three of these proteins presented the strongest statistical associations with the risk of short-
235 term clinical relapse (CLEC4C; CLEC4G; LAMP3; figure 3A). Among the proteins mainly

236 expressed in lymphocytes, 1 was associated with the risk of short-term clinical relapse (Natural
237 killer cells antigen CD94, KLRD1) while 3 were associated with the risk of mid/long-term
238 clinical relapse (lymphocyte activation gene 3 protein, LAG3; SH2B adapter protein 3, SH2B3;
239 signaling threshold-regulating transmembrane adapter 1, SIT1) (figure 3A,B). Of note, SIT1
240 showed the strongest statistical association with the risk of mid/long-term clinical relapse
241 (figure 3B).

242

243 **Risk of clinical relapse associated with cellular junction proteins**

244 The PEA panel contains 6 cellular junction proteins which are either implicated in cell-cell
245 junctions (corneodesmosin: CDSN; contactin-associated protein-like 2: CNTNAP2;
246 coxsackievirus and adenovirus receptor: CXADR) or in cell-matrix junctions (integrin α -11:
247 ITGA11; integrin alpha-6: ITGA6; integrin beta-6: ITGB6) (online supplemental table 4). The
248 risk of mid-long-term clinical relapse was characterised by a low serum level of 4 cellular
249 junction proteins (CDSN; CNTNAP2; CXADR; ITGA11) (figure 4A,B). Finally, a low or a
250 high serum level of ITGA6 was associated with respectively the risk of short-term clinical
251 relapse or the risk of mid/long-term clinical relapse (figure 4A,B).

252

253 **Risk of clinical relapse associated with downstream signalling of cytokine receptors and** 254 **pattern recognition receptors**

255 In the PEA panel, some proteins are part of immune pathways such as nuclear factor- κ B (NF-
256 κ B) (n=10), interferon (n=3) and mitogen-activated protein kinases (n=1) (online supplemental
257 table 5). All those proteins belong to the downstream signalling of cytokine receptors and
258 pattern recognition receptors, and remarkably, most of them (10/12) exhibited an opposite
259 pattern (HR<1 vs HR>1) between the short-term clinical relapse dataset and mid/long-term
260 clinical relapse dataset (figure 5).

261

262 **DISCUSSION**

263 The present work supports that, in CD patients stopping infliximab, the risk of short-term
264 clinical relapse (<6 months) and mid/long-term clinical relapse (>6 months) are associated with
265 distinct pathophysiological processes. Previously, we came to the same conclusion by using
266 another technology (SRM) and by evaluating other biological phenomena[8]. These converging
267 results extend and consolidate the knowledge on the distinct biological profiles of short-term
268 relapsers, mid/long-term relapsers and, by contrast, non-relapsers. As a corollary, we now
269 propose to consider that non-relapsers experience a stable biological remission, while short-
270 term relapsers and mid/long-term relapsers present different degrees of unstable biological
271 remission characterised by different types of residual disease activity. This identification and
272 characterisation of CD patient subsets could constitute a basis to develop non-invasive
273 biomarkers guiding the decision of anti-TNF α withdrawal.

274 Our previous study revealed that monitoring inflammation as currently performed in clinics
275 can only have a limited utility when contemplating anti-TNF α withdrawal. Indeed,
276 inflammatory markers were essentially associated with the risk of short-term clinical relapse[8],
277 while the majority of relapsers present a time to relapse greater than 6 months after anti-TNF α
278 withdrawal (66% in the present work and >80% in another study[15]). Thus, characterising the
279 biological profile of mid/long-term relapsers is a research priority that will require to study
280 other targets than inflammatory markers. Our study constitutes a progress in this direction since
281 we highlighted that the risk of mid/long-term clinical relapse is associated with markers
282 involved in anti-inflammatory defences, cellular junctions, immunoregulation of APCs and
283 lymphocytes.

284 Our study also adds new elements in the understanding of clinical relapse after infliximab
285 withdrawal. According to our results, a reduced anti-inflammatory capacity could be implicated

286 in the pathophysiological mechanisms leading to clinical relapse. Indeed, we showed an
287 association between the risk of mid/long-term clinical relapse and a low circulating levels of
288 potent anti-inflammatory effectors (IL-10, the master anti-inflammatory cytokine; HSD11B1,
289 an enzyme converting inactive cortisone to active cortisol thus promoting the anti-inflammatory
290 effect of glucocorticoids[16,17]). This goes along the same line as our previous results showing
291 a similar association with the apolipoproteins A-I and A-II which are known to mediate the
292 anti-inflammatory properties of high density lipoprotein[8,18]. Although our study cannot
293 establish causal relationships, our results are concordant with the hypothesis that a reduction of
294 anti-inflammatory effectors could be a prelude to the inflammatory flare characterising the
295 relapse. This hypothesis needs to be tested in a dedicated work.

296 Despite its capital role in regulating the action of endogenous and synthetic
297 glucocorticoids[17], HSD11B1 has received little attention in the IBD literature so far. An
298 increase of HSD11B1 mRNA level has been reported in inflamed versus non-inflamed
299 intestinal biopsies of IBD and non-IBD patients[19]. In mice, similar results were reported in
300 dextran sulfate sodium (DSS)-induced colitis model[20]. Actually, up-regulation of HSD11B1
301 is a well-known homeostatic response to inflammatory stimuli which is observed in non-
302 immune and immune cells (monocytes, macrophages, leukocytes, neutrophils)[17]. In contrast,
303 it is intriguing and new to report in the present study that a future risk for patients (clinical
304 relapse) is associated with a low serum level of HSD11B1. Taken together, these observations
305 suggest to more deeply study the role of HSD11B1 in CD.

306 When compared to mucosa of healthy individual, the macroscopically normal mucosa of CD
307 patients showed abnormalities of the cellular junctions which were revealed through
308 microscopy techniques (confocal endomicroscopy, confocal and electron microscopy)[21–25]
309 and also supported by molecular-based evidence showing a decreased mRNA levels of α -
310 catenin (adherens junction) and tight junction protein ZO-1[26]. Remarkably, our results

311 showed that a perturbation of cellular junctions could be an early defect affecting the relapsing
312 patients. Indeed, we found that low serum levels of 4 cellular junction proteins were specifically
313 associated with the risk of mid/long-term clinical relapse. It now remains to elucidate whether
314 these results obtained in the blood reflect an alteration of the gut epithelium.

315 In a coherent manner, our previous and present studies well converge to support that acute
316 inflammation characterises the short-term relapsers. Indeed, we previously reported that a high
317 serum level of acute-phase reactants is specifically associated with the risk of short-term clinical
318 relapse[8] and, this outcome was herein associated with a high serum level of IL-6 which is a
319 canonical inducer of the acute-phase response[27].

320 Our study showed an association between the risk of short-term clinical relapse and a high
321 serum level of CLEC4C, a highly specific marker of plasmacytoid dendritic cells (pDCs) which
322 exerts a tolerogenic effect[28]. This result deserves to be discussed in the light of what is already
323 known on pDCs in CD. In mice, ablation of pDCs reduced DSS-induced colitis[29]. In CD
324 patients with acute flare ups, evidence supported that peripheral pDCs migrate to colonic
325 mucosa and mesenteric lymph nodes and this is accompanied by a modification of their
326 phenotype toward a pro-inflammatory profile[30,31]. According to our results on CLEC4C, we
327 can reasonably speculate that changes in peripheral pDCs could be detected, not only during,
328 but also before acute flare ups.

329 Prediction and causality are aims commonly confused in clinical studies, this situation leads
330 to inappropriate methodologies and erroneous conclusions[32]. In this context, it has been
331 recommended that researchers clearly explain and state their objective: causality vs
332 prediction[32]. Etiological research generally aims to determine the causal effect of an
333 independent variable X on a dependent variable Y (outcome). In this situation, adjustment for
334 confounders is typically performed to reduce the omitted-variables bias which can distort the
335 relation between X and Y. On the other hand, prediction research aims to forecast Y by knowing

336 a set of variables called predictors and this whatever their relations (causal or not) with Y and
337 with omitted variables. The present study does not pursue a causal aim, i.e., we do not pretend
338 that modulating a PEA marker (X) will influence the clinical relapse (Y). Our objective clearly
339 concerns the prediction of a clinical outcome. More precisely, we aimed to characterise the
340 biological states of patients (depicted by PEA markers) predicting their clinical relapse and this
341 whatever the effect of other variables (eg, age, sex, treatments, infliximab trough level, disease
342 extension, smoking). In other words, considering other variables than the PEA markers is not
343 relevant to achieve our objective. Besides, adjusting for confounders would have been a
344 methodological error since this approach is appropriate to deal with causality but not
345 prediction[32].

346 The limitations of our work include the fact that the definition of relapse was only clinical
347 (CDAI), i.e., not confirmed by objective markers (CRP or calprotectin). Thus, the classification
348 of patients as relapsers can be affected by the subjectivity of symptoms used to calculate the
349 CDAI.

350 We also acknowledge a limitation regarding the interpretation of results obtained on non-
351 secreted proteins (79/92 in the PEA dataset according to UniProt and Human Protein Atlas,
352 including some proteins highlighted in the discussion: HSD11B1, CLEC4C). Indeed, the role
353 of non-secreted proteins in the serum is largely unknown and their presence outside the cells
354 probably depends on complex combination of mechanisms regulating cells death, protein
355 cleavage, cellular population and gene expression. For those proteins, the main interest of our
356 study was to show their distinct patterns in the short-term relapsers and mid/long-term relapsers
357 rather than to provide clear pathophysiological insights. In contrast, our results obtained on IL-
358 6 and IL-10 (secreted proteins) can be better interpreted given that function of these cytokines
359 in the blood is well known and their roles in CD are largely documented in the literature.

360

361 Another limitation of our work is the absence of independent cohort to generalise the results,
362 this prevents to directly determine the clinical interest of individual markers or their
363 combinations in statistical models. However, we think that part of our results can be
364 generalisable in a much broader context than our study. This proposal is supported by the idea
365 that, whatever the cause of a relapse, this is a failure of both treatments and human homeostasis
366 which is associated with some invariable pathophysiological processes. For instance, this is the
367 case of acute inflammation which is generally associated with a relapse whatever the clinical
368 context. In the present study, a high level of IL-6 was specifically associated with the risk of
369 short-term clinical relapse after infliximab withdrawal. Similar finding was reported in inactive
370 CD patients who were not treated with biologics and, for some of them, receiving no
371 treatments[33]. Hence, a high circulating level of IL-6 can announce a short-term clinical
372 relapse and this result is generalisable far beyond the STORI cohort and the specific context of
373 infliximab withdrawal. A similar reasoning can be applied with the association between a high
374 serum level of acute-phase reactants and the risk of short-term clinical relapse[8]. Even not
375 independently validated, the generalisability of this finding cannot be really contested given its
376 coherence with the CD pathophysiology. To our point of view, the main message of our study
377 (distinct biological profiles between short-term relapsers and mid/long-term relapsers) could be
378 also generalisable given that any disease flare is preceded by a succession of biological states.
379 In our study, these biological changes have been objectified and allow to better understand why,
380 after stopping the infliximab, the clinical relapse occurred in a wide period of time (2-26
381 months). Reasonably, we think that similar observations could be found in heterogeneous
382 contexts and that, time to relapse or more generally time to event, is a useful parameter to reveal
383 biological profiles.

384 Our method of analysis was built to anchor the results in a biological reality and, to our
385 opinion, this is an essential condition for external validity. Indeed, our findings rely on a

386 validated biological hypothesis (time to clinical relapse is related to the pathophysiological state
387 of patients) and a simple univariable statistics approach where variables were grouped based on
388 their biological rather than mathematical relations. On the other hand, data-driven approaches
389 performing complex mathematical relations between variables are prone to overfitting and the
390 biological meaning of their outputs is generally unknown or difficult to understand (black box
391 effect). These criticisms notably concern machine learning tools[34,35].

392 In summary, we identified distinct biological profiles between short-term relapsers and
393 mid/long-term relapsers following anti-TNF α withdrawal in CD patients. This knowledge is
394 essential to develop biomarkers guiding the therapeutic strategy.

395

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404

405 **Contributors**

406 M-AM, YB, DL, J-FC, MA and EL designed the study. The GETAID provided the samples
407 and the clinical information. VAH-T performed the statistical analysis. NP analysed and
408 interpreted the data with the help of M-AM, VAH-T, TM and EL. NP wrote the initial draft of
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413

414 **Competing interests**

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416

417 **References**

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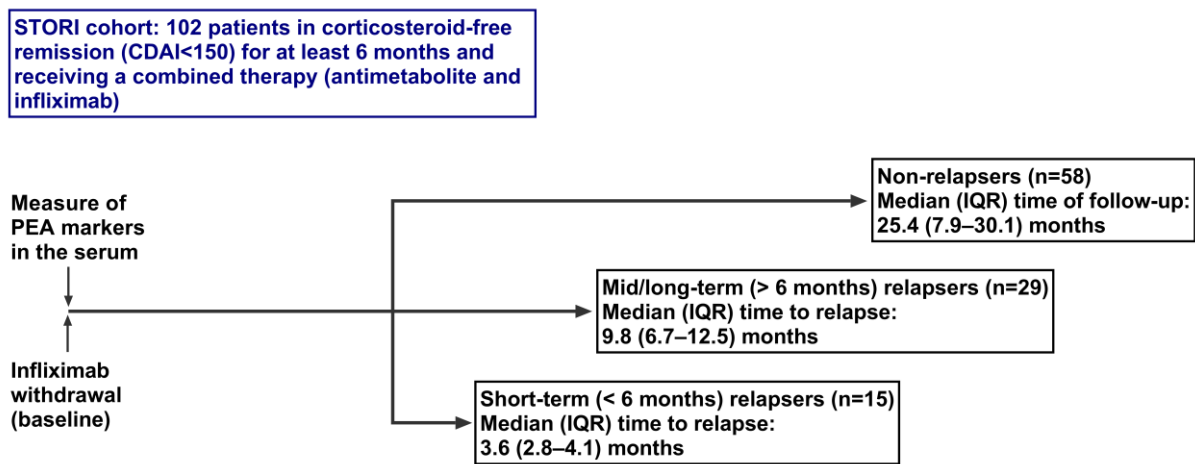
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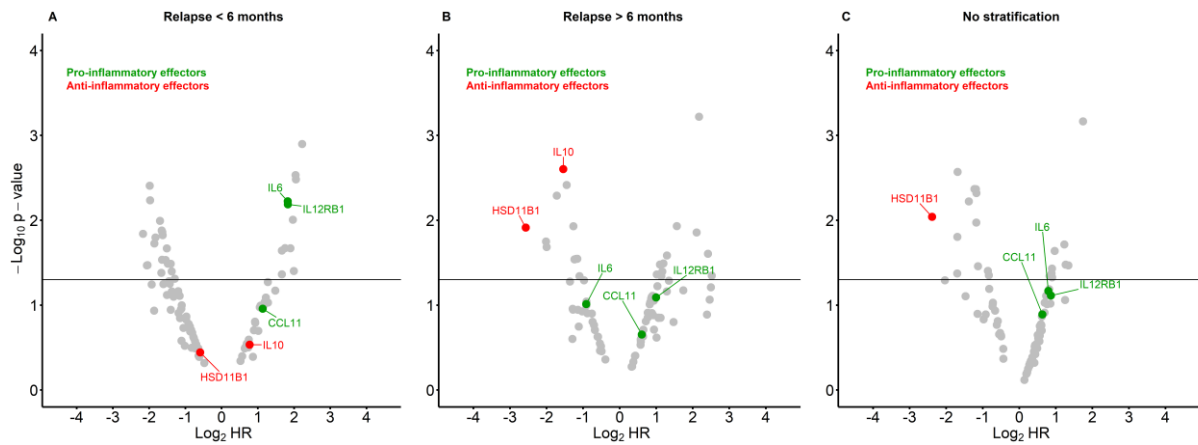
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525 **Figure 1. Experimental design and outcomes of patients after infliximab withdrawal**

526 IQR, interquartile range; PEA, proximity extension assay; STORI, diSconTinuation in CrOhn’s
 527 disease patients in stable Remission on combined therapy with Immunosuppressors.

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531 **Figure 2. Risk of clinical relapse associated with pro-inflammatory and anti-inflammatory**

532 **effectors.** The risk of clinical relapse (HR: hazard ratio) associated with each protein was

533 determined by univariable Cox model and represented by plotting its degree of significance (-

534 Log_{10} p-value) against its effect size (Log_2 HR) in the short-term clinical relapse (<6 months)

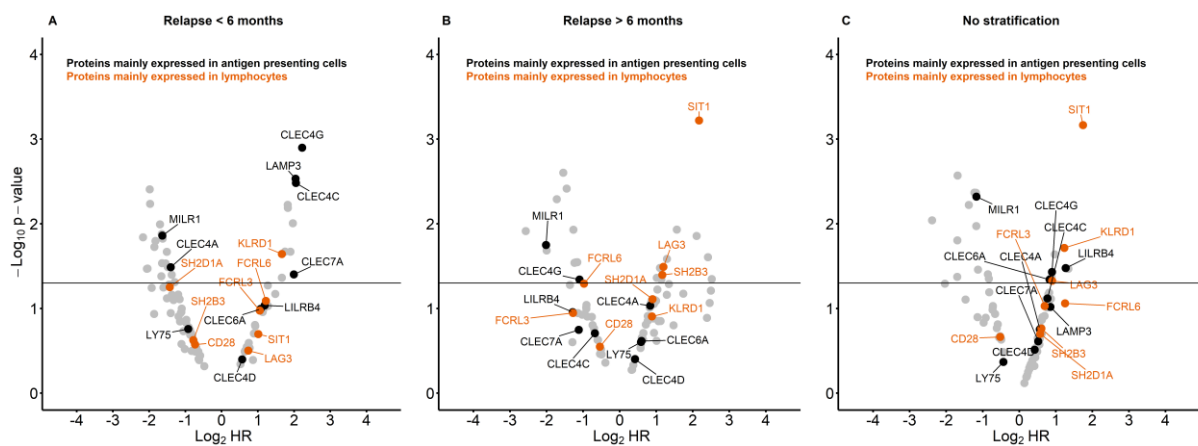
535 dataset (A) mid/long-term clinical relapse (>6 months) dataset (B) and non-stratified dataset

536 (C). The significance threshold (p-value=0.05) is represented by the horizontal lines. CCL11,

537 eotaxin; HSD11B1, corticosteroid 11-beta-dehydrogenase isozyme 1; IL6, interleukin-6; IL10,

538 interleukin-10; IL12RB1, interleukin-12 receptor subunit beta-1.

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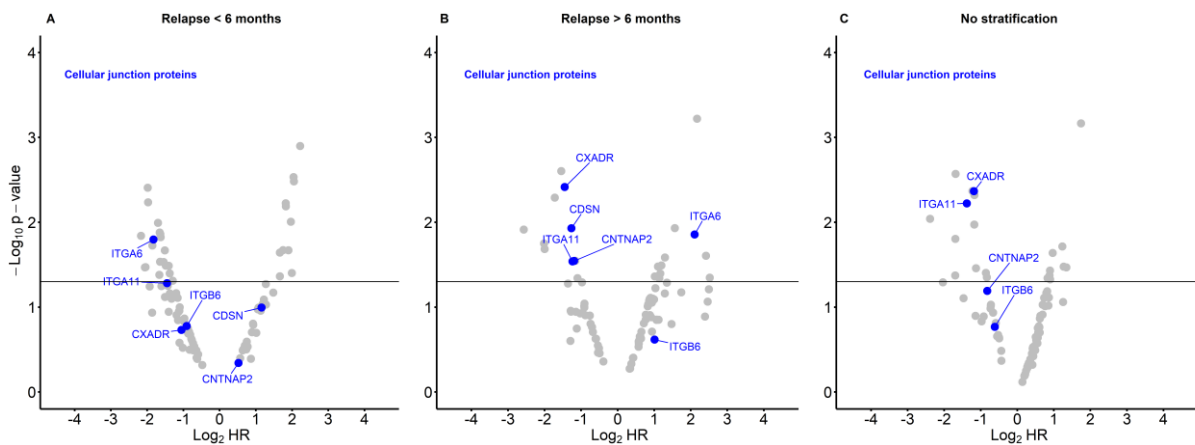
541 **Figure 3. Risk of clinical relapse associated with proteins mainly expressed in antigen**

542 **presenting cells or lymphocytes** The risk of clinical relapse (HR: hazard ratio) associated with

543 each protein was determined by univariable Cox model and represented by plotting its degree

544 of significance ($-\text{Log}_{10}$ p-value) against its effect size (Log_2 HR) in the short-term clinical
545 relapse (<6 months) dataset (A) mid/long-term clinical relapse (>6 months) dataset (B) and
546 non-stratified dataset (C). The significance threshold (p-value=0.05) is represented by the
547 horizontal lines. CD28, T-cell-specific surface glycoprotein CD28; CLEC4A, C-type lectin
548 domain family 4 member A; CLEC4C, C-type lectin domain family 4 member C; CLEC4D, C-
549 type lectin domain family 4 member D; CLEC4G, C-type lectin domain family 4 member G;
550 CLEC6A, C-type lectin domain family 6 member A; CLEC7A, C-type lectin domain family 7
551 member A; FCRL3, Fc receptor-like protein 3; FCRL6, Fc receptor-like protein 6; KLRD1,
552 natural killer cells antigen CD94; LAG3, lymphocyte activation gene 3 protein; LAMP3,
553 Lysosome-associated membrane glycoprotein 3; LILRB4, leukocyte immunoglobulin-like
554 receptor subfamily B member 4; LY75, lymphocyte antigen 75; MILR1, allergin-1; SIT1,
555 signaling threshold-regulating transmembrane adapter 1; SH2B3, SH2B adapter protein 3;
556 SH2D1A, SH2 domain-containing protein 1A.

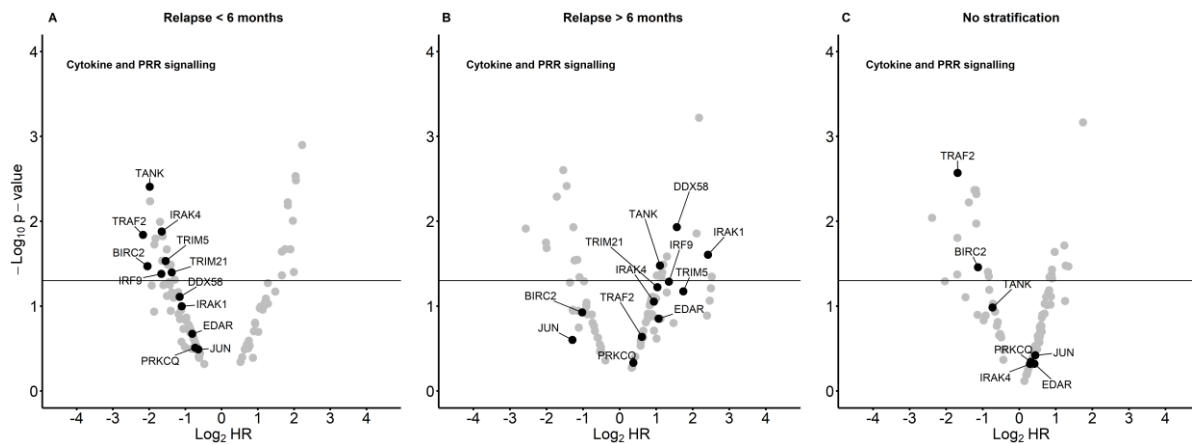
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559 **Figure 4. Risk of clinical relapse associated with cellular junction proteins.** The risk of
560 clinical relapse (HR: hazard ratio) associated with each protein was determined by univariable
561 Cox model and represented by plotting its degree of significance ($-\text{Log}_{10}$ p-value) against its
562 effect size (Log_2 HR) in the short-term clinical relapse (<6 months) dataset (A) mid/long-term
563 clinical relapse (>6 months) dataset (B) and non-stratified dataset (C). The significance

564 threshold (p-value=0.05) is represented by the horizontal lines. CDSN, corneodesmosin;
 565 CNTNAP2, contactin-associated protein-like 2; CXADR, coxsackievirus and adenovirus
 566 receptor; ITGA6, integrin alpha-6; ITGA11, integrin alpha-11; ITGB6, integrin beta-6.
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568
 569 **Figure 5. Risk of clinical relapse associated with downstream signalling of cytokine**
 570 **receptors and pattern recognition receptors.** The risk of clinical relapse (HR: hazard ratio)
 571 associated with each protein was determined by univariable Cox model and represented by
 572 plotting its degree of significance ($-\text{Log}_{10}$ p-value) against its effect size (Log_2 HR) in the short-
 573 term clinical relapse (<6 months) dataset (A) mid/long-term clinical relapse (>6 months) dataset
 574 (B) and non-stratified dataset (C). The significance threshold (p-value=0.05) is represented by
 575 the horizontal lines. BIRC2, baculoviral IAP repeat-containing protein 2; DDX58, antiviral
 576 innate immune response receptor RIG-I; EDAR, tumor necrosis factor receptor superfamily
 577 member EDAR; IRAK1, interleukin-1 receptor-associated kinase 1; IRAK4, interleukin-1
 578 receptor-associated kinase 4; IRF9, interferon regulatory factor 9; JUN, transcription factor AP-
 579 1; PRKCQ, protein kinase C theta type; TANK, TRAF family member-associated NF-kappa-B
 580 activator; TRAF2, TNF receptor-associated factor 2; TRIM5, tripartite motif-containing protein
 581 5; TRIM21, E3 ubiquitin-protein ligase TRIM21.

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Table 1. Patients' characteristics (n=102)

Male, n (%)	46 (45)
Age, median years (IQR)	31 (25-39)
Disease duration, median years (IQR)	8 (4-12)
Active smoker, n (%)	38 (37)
CDAI (IQR)	36.2 (16.1-59.5)
Disease site	
Ileal, n (%)	13 (13)
Colonic, n (%)	32 (31)
Ileocolonic, n (%)	56 (55)
Upper gastrointestinal tract, n (%)	8 (8)
Perianal lesions, n (%)	36 (35)
Treatment history	
Methotrexate, n (%)	17 (17)
Azathioprine/mercaptopurine, n (%)	85 (83)
Duration of antimetabolite treatment, median years (IQR)	2.6 (1.7-4.5)
Duration of infliximab treatment, median years (IQR)	2.2 (1.6-3.1)
Previous surgical resection, n (%)	22 (22)
Endoscopy	
CDEIS (IQR)	0.6 (0.0-2.8)
CDEIS=0, n (%)	38 (37)
Remaining ulcers, n (%)	32 (31)
Biologic variables	
Haemoglobin level, g/L, median (IQR)	136 (127-144)
Haematocrit, %, median (IQR)	40 (37-43)
Leukocyte count, 10 ⁹ /L, median (IQR)	6.0 (4.9-7.3)
Platelet count, 10 ⁹ /L, median (IQR)	265 (225-313)
hsCRP level, mg/L, median (IQR)	1.9 (0.8-4.2)
Infliximab trough level, mg/L, median (IQR)	3.7 (1.9-7.9)
Faecal calprotectin, µg/g, n=77, median (IQR)	49.4 (29.6-200.8)

CDAI: Crohn's disease activity index; CDEIS: Crohn's disease endoscopic index of severity; hsCRP: high-sensitivity CRP; IQR: interquartile range

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Merlin	NF2	P35240	Ubiquitous	Cytoskeleton, hippo pathway	Not shown	No association	0.3(0.1-1.4)	0.116	1.3(0.6-2.6)	0.528	1.1(0.6-2.1)	0.762
Nuclear factor of activated T-cells, cytoplasmic 3	NFATC3	Q12968	Ubiquitous	Immune response	Not shown	No association	0.5(0.2-1.2)	0.136	0.6(0.3-1.2)	0.157	0.6(0.3-1.1)	0.094
Neurotrophin-4	NTF4	P34130	Widely expressed	Growth factor	Not shown	No association	0.6(0.2-1.4)	0.244	0.7(0.4-1.4)	0.305	0.7(0.4-1.2)	0.228
Protein-arginine deiminase type-2	PADI2	Q9Y2J8	Widely expressed	Pleiotropic effects	Not shown	No association	0.5(0.2-1.2)	0.113	1.8(0.9-3.6)	0.084	1.5(0.8-2.9)	0.199
Phosphoinositide 3-kinase adapter protein 1	PIK3AP1	Q6ZUJ8	Widely expressed	Adaptive immunity	Not shown	No association	0.4(0.2-1.1)	0.069	1.7(0.9-3.4)	0.123	1.2(0.7-2.1)	0.562
Plexin-A4	PLXNA4	Q9HCM2	Widely expressed	Pleiotropic effects	Not shown	No association	0.5(0.2-1.4)	0.204	0.4(0.2-1)	0.053	1.5(0.8-2.7)	0.169
Neurabin-2	PPP1R9B	Q96SB3	Widely expressed	Pleiotropic effects	Sup Figure 1	Short-term relapse	0.3(0.1-0.8)	0.015	5.5(0.8-38.9)	0.086	0.6(0.3-1)	0.045
Peroxisiredoxin-1	PRDX1	Q06830	Ubiquitous	Antioxidant defence	Sup Figure 1	Short and mid/long-term relapse	0.4(0.1-0.9)	0.021	2.5(1.1-5.4)	0.026	0.6(0.3-1)	0.039
Thioredoxin-dependent peroxide reductase, mitocho	PRDX3	P30048	Ubiquitous	Antioxidant defence	Not shown	No association	1.9(0.8-4.7)	0.161	1.9(0.7-5.1)	0.194	1.4(0.7-2.8)	0.322
Peroxisiredoxin-5, mitochondrial	PRDX5	P30044	Ubiquitous	Antioxidant defence	Sup Figure 1	Short-term relapse	0.4(0.2-1)	0.049	2(0.9-4.1)	0.080	1.2(0.7-2.1)	0.617
Protein kinase C theta type	PRKCQ	Q04759	Widely expressed	NF-κB pathway	Figure 5	No association	0.6(0.2-1.6)	0.309	1.3(0.6-2.6)	0.465	1.2(0.7-2.2)	0.453
PC4 and SFRS1-interacting protein	PSIP1	O75475	Ubiquitous	Not well defined	Sup Figure 1	Short-term relapse	0.4(0.1-0.9)	0.032	1.8(0.9-3.6)	0.094	1.2(0.6-2.1)	0.615
Parathyroid hormone/parathyroid hormone-related p	PTH1R	Q03431	Widely expressed	Pleiotropic effects	Not shown	No association	0.7(0.3-1.8)	0.481	1.7(0.8-3.5)	0.140	1.6(0.9-2.7)	0.127
SH2B adapter protein 3	SH2B3	Q9UQQ2	Lymphocytes	Tolerance, adaptor protein	Figure 3	Mid/long-term relapse	0.6(0.2-1.4)	0.236	2.2(1-4.9)	0.040	1.5(0.8-2.8)	0.173
SH2 domain-containing protein 1A	SH2D1A	O60880	Lymphocytes	Immunity, adaptor protein	Figure 3	No association	0.4(0.1-1)	0.056	1.9(0.9-3.7)	0.078	1.5(0.8-2.7)	0.199
Signaling threshold-regulating transmembrane adapt	SIT1	Q9Y3P8	Lymphocytes	Tolerance, transmembrane adaptor	Figure 3	Mid/long-term relapse	2(0.7-5.9)	0.201	4.5(1.9-10.7)	0.001	3.4(1.7-6.7)	0.001
Protein sprouty homolog 2	SPRY2	O43597	Widely expressed	Inhibitor of the fibroblast growth factor signaling	Sup Figure 1	Short-term relapse	3.2(1-9.7)	0.043	0.7(0.3-1.5)	0.350	1.6(0.9-3)	0.123
SRSF protein kinase 2	SRPK2	P78362	Ubiquitous	Pleiotropic effects	Not shown	No association	0.5(0.2-1.3)	0.179	1.9(0.9-3.8)	0.085	1.3(0.8-2.4)	0.297
Stanniocalcin-1	STC1	P52823	Widely expressed	Pleiotropic effects	Sup Figure 1	Short-term relapse	3.7(1.2-11.4)	0.021	2.4(0.9-6.4)	0.069	1.5(0.8-2.8)	0.229
TRAF family member-associated NF-kappa-B activa	TANK	Q92844	Ubiquitous	NF-κB pathway	Figure 5	Short and mid/long-term relapse	0.3(0.1-0.6)	0.004	2.2(1.1-4.4)	0.033	0.6(0.3-1.1)	0.104
Tryptase alpha/beta-1	TPSAB1	Q15661	Mast cells	Protease	Not shown	No association	0.5(0.2-1.2)	0.110	0.8(0.4-1.5)	0.438	0.7(0.4-1.2)	0.213
TNF receptor-associated factor 2	TRAF2	Q12933	Ubiquitous	NF-κB pathway	Figure 5	Short-term relapse	0.2(0.1-0.7)	0.014	1.5(0.8-3)	0.230	0.3(0.1-0.7)	0.003
Triggering receptor expressed on myeloid cells 1	TREM1	Q9NP99	Widely expressed	Pro-inflammatory	Not shown	No association	1.9(0.7-4.7)	0.197	1.6(0.8-3.3)	0.155	1.5(0.8-2.7)	0.168
E3 ubiquitin-protein ligase TRIM21	TRIM21	P19474	Ubiquitous	NF-κB and interferon pathways	Figure 5	Short-term relapse	0.4(0.2-1)	0.040	2(1-4.3)	0.060	1.1(0.6-2)	0.665
Tripartite motif-containing protein 5	TRIM5	Q9C035	Ubiquitous	NF-κB and interferon pathways	Figure 5	Short-term relapse	0.3(0.1-0.9)	0.029	3.3(0.9-12.2)	0.067	1.1(0.5-2.4)	0.716
Zinc finger and BTB domain-containing protein 16	ZBTB16	Q05516	Widely expressed	Pleiotropic effects	Sup Figure 1	Short-term relapse	0.2(0.1-0.9)	0.034	2.1(0.8-5.6)	0.141	1.5(0.7-2.9)	0.266

Supplementary table 2. Pro-inflammatory and anti-inflammatory effectors

Protein names	Gene names	Uniprot accession number	Main cellular expression	Inflammatory balance
Interleukin-6	IL6	P05231	Ubiquitous	Pro-inflammatory ¹
Eotaxin	CCL11	P51671	Ubiquitous	Pro-inflammatory ²
Interleukin-12 receptor subunit beta-1	IL12RB1	P42701	Ubiquitous	Pro-inflammatory ³
Interleukin-10	IL10	P22301	Ubiquitous	Anti-inflammatory ⁴
Corticosteroid 11-beta-dehydrogenase isozyme 1	HSD11B1	P28845	Ubiquitous	Anti-inflammatory ⁵

Supplementary table 3. Proteins mainly expressed in antigen presenting cells or lymphocytes

Protein names	Gene names	Uniprot accession number	Main cellular expression	Immune function	Protein function and ligands
T-cell-specific surface glycoprotein CD28	CD28	P10747	T cells ⁶	Immunity ⁷	Receptor for CD80 and CD86 ⁸
Natural killer cells antigen CD94	KLRD1	Q13241	NK cells ⁹	Immunity and tolerance ⁹	Receptor for MHC I antigens ⁹
Lymphocyte activation gene 3 protein	LAG3	P18627	Lymphocytes ¹⁰	Tolerance ⁷	Receptor for MHC II antigens ⁸
Signaling threshold-regulating transmembrane adapter 1	SIT1	Q9Y3P8	Lymphocytes ¹¹	Tolerance ¹²	Transmembrane adaptor ¹²
SH2B adapter protein 3	SH2B3	Q9UQQ2	Lymphocytes ¹³	Tolerance ¹³	Adaptor protein ¹³
SH2 domain-containing protein 1A	SH2D1A	O60880	Lymphocytes ¹⁴	Immunity ¹⁴	Adaptor protein ¹⁴
Fc receptor-like protein 6	FCRL6	Q6DN72	Lymphocytes ¹⁵	Tolerance ¹⁶	Receptor for MHC II antigen ¹⁵
Fc receptor-like protein 3	FCRL3	Q96P31	Lymphocytes ¹⁷	Immunity and tolerance ¹⁷	Receptor for IgA ¹⁷
C-type lectin domain family 4 member C	CLEC4C	Q8WTT0	pDCs ¹⁸	Tolerance ¹⁹	PRRs, ligands not well known ²⁰
Leukocyte immunoglobulin-like receptor subfamily B member 4	LILRB4	Q8NHJ6	pDCs ²¹	Tolerance ²²	Receptor for BST2 ²¹
Lysosome-associated membrane glycoprotein 3	LAMP3	Q9UQV4	DCs ²³	Unknown	MHC II response ²³
C-type lectin domain family 4 member G	CLEC4G	Q6UXB4	LSECs, APCs ²⁰	Tolerance ⁷	PRRs, virus recognition ²⁰
C-type lectin domain family 4 member A	CLEC4A	Q9UMR7	APCs ²⁴	Tolerance ²⁰	PRRs, ligands not well known ²⁰
C-type lectin domain family 7 member A	CLEC7A	Q9BXN2	APCs ²⁴	Immunity and tolerance ^{18,20}	PRRs, fungi and DAMPs recognition ²⁰
C-type lectin domain family 6 member A	CLEC6A	Q6EIG7	APCs ²⁰	Immunity ²⁰	PRRs, fungi recognition ²⁵
C-type lectin domain family 4 member D	CLEC4D	Q8WXI8	APCs ²⁶	Immunity ¹⁸	PRRs, ligands not well known ²⁰
Allergin-1	MILR1	Q7Z6M3	APCs ²⁷	Tolerance ²⁷	Ig-like receptor with unknown ligand ²⁷
Lymphocyte antigen 75	LY75	O60449	APCs ²⁸	Immunity and tolerance ²⁹	PRRs, bacteria and DAMPs recognition ³⁰

APCs: antigen presenting cells; BST2: bone marrow stromal cell antigen 2; CD80: cluster of differentiation 80; DAMP: damage associated molecular pattern; DCs: dendritic cells; LSECs: liver sinusoidal endothelial cells; MHC I; major histocompatibility complex class I; MHC II: major histocompatibility complex class II; pDCs: plasmacytoid dendritic cells; PRRs: pattern recognition receptors.

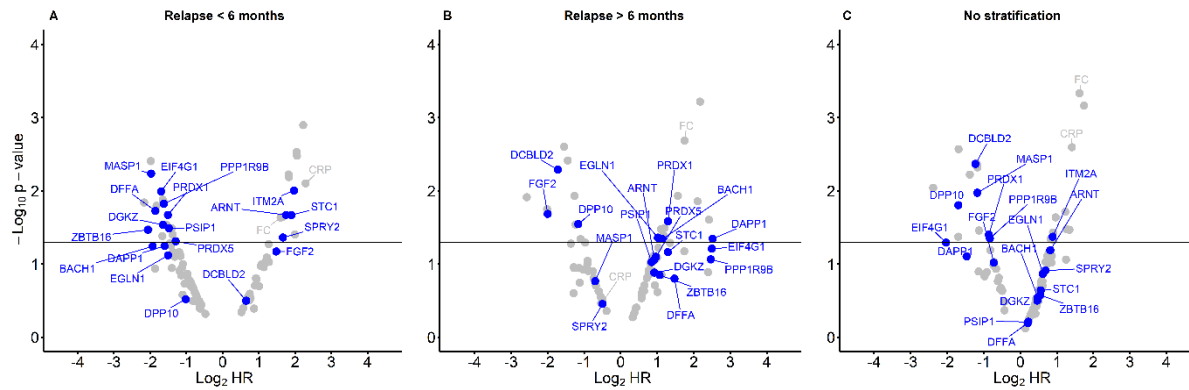
Supplementary table 4. Cellular junction proteins

Protein names	Gene names	Uniprot accession number	Main cellular expression	Functions
Corneodesmosin	CDSN	Q15517	Keratinocytes ³¹	Cell-cell junction (corneodesmosomes) ³¹
Contactin-associated protein-like 2	CNTNAP2	Q9UHC6	Nervous system cells ³²	Cell-cell junction (synapse) ³²
Coxsackievirus and adenovirus receptor	CXADR	P78310	Ubiquitous	Cell-cell junction (tight junction) ³³
Integrin alpha-6	ITGA6	P23229	Ubiquitous	Cell-matrix junction ³⁴
Integrin alpha-11	ITGA11	Q9UKX5	Ubiquitous	Cell-matrix junction ³⁴
Integrin beta-6	ITGB6	P18564	Ubiquitous	Cell-matrix junction ³⁴

Supplementary table 5. Downstream signalling of cytokine receptors and pattern recognition receptors

Protein names	Gene names	Uniprot accession number	Main cellular expression	Pathways
Antiviral innate immune response receptor RIG-I	DDX58	O95786	Ubiquitous	NF- κ B ^a
Interleukin-1 receptor-associated kinase 1	IRAK1	P51617	Ubiquitous	NF- κ B ^a
Protein kinase C theta type	PRKCQ	Q04759	Ubiquitous	NF- κ B ^a
TNF receptor-associated factor 2	TRAF2	Q12933	Ubiquitous	NF- κ B ^a
Baculoviral IAP repeat-containing protein 2	BIRC2	Q13490	Ubiquitous	NF- κ B ^a
TRAF family member-associated NF-kappa-B activator	TANK	Q92844	Ubiquitous	NF- κ B
Interleukin-1 receptor-associated kinase 4	IRAK4	Q9NWZ3	Ubiquitous	NF- κ B ^a
Tumor necrosis factor receptor superfamily member EDAR	EDAR	Q9UNE0	Ubiquitous	NF- κ B ^a
E3 ubiquitin-protein ligase TRIM21	TRIM21	P19474	Ubiquitous	Interferon ^b ; NF- κ B ³⁵
Tripartite motif-containing protein 5	TRIM5	Q9C035	Ubiquitous	Interferon ^b ; NF- κ B ³⁶
Interferon regulatory factor 9	IRF9	Q00978	Ubiquitous	Interferon ^b
Transcription factor AP-1	JUN	P05412	Ubiquitous	MAPKs ^c

MAPKs: mitogen-activated protein kinases; NF- κ B: nuclear factor- κ B. ^aKEGG pathway hsa04064 (NF-kappa B signaling pathway); ^bReactome pathway R-HSA-877300 (interferon gamma signalling); ^cKEGG pathway hsa04010 (MAPK signaling pathway).



Supplementary Figure 1. Proteins not highlighted in the core manuscript and showing a significant association with the risk of short-term and/or mid/long-term relapse.

The risk of relapse (HR: hazard ratio) associated with each protein was determined by univariable Cox model and represented by plotting its degree of significance ($-\text{Log}_{10}$ p-value) against its effect size (Log_2 HR) in the short-term relapse (<6 months) dataset (A) mid/long-term relapse (>6 months) dataset (B) and non-stratified dataset (C). The significance threshold ($p\text{-value}=0.05$) is represented by the horizontal lines. The protein names, their main cellular expression and function are presented in online supplemental table 1. ARNT, Aryl hydrocarbon receptor nuclear translocator; BACH1, Transcription regulator protein BACH1; DAPP1, Dual adapter for phosphotyrosine and 3-phosphotyrosine and 3-phosphoinositide; DCBLD2, Discoidin, CUB and LCCL domain-containing protein 2; DFFA, DNA fragmentation factor subunit alpha; DGKZ, Diacylglycerol kinase zeta; DPP10, Inactive dipeptidyl peptidase 10; EGLN1, Egl nine homolog 1; EIF4G1, Eukaryotic translation initiation factor 4 gamma 1; FGF2, Fibroblast growth factor 2; ITM2A, Integral membrane protein 2A; MASP1, Mannan-binding lectin serine protease 1;

PPP1R9B, Neurabin-2; PRDX1, Peroxiredoxin-1; PRDX5, Peroxiredoxin-5, mitochondrial; PSIP1, PC4 and SFRS1-interacting protein; SPRY2, Protein sprouty homolog 2; STC1, Stanniocalcin-1; ZBTB16, Zinc finger and BTB domain-containing protein 16.

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