









The genetic history of Mayotte and Madagascar cattle breeds mirrors the complex pattern of human exchanges in Western Indian Ocean

Jessica Magnier ^{1,2} Tom Druet ³ Michel Naves ⁴ Mélissa Ouvrard⁵ Solène Raoul⁵ Jérôme Janelle ^{1,6}
 Katayoun Moazami-Goudarzi ⁷ Matthieu Lesnoff^{1,2} Emmanuel Tillard ^{1,6} Mathieu Gautier ⁸ Laurence Flori ^{9,*}

¹SELMET, University of Montpellier, CIRAD, INRAE, L'Institut Agro, Montpellier 34398, France,

²CIRAD, UMR SELMET, Montpellier 34398, France,

³Unit of Animal Genomics, GIGA-R, Faculty of Veterinary Medicine, University of Liège, Liège 4000, Belgium,

⁴URZ, INRAE, Guadeloupe 97170, France,

⁵COOPADEM, Mayotte 97670, France,

⁶CIRAD, UMR SELMET, Saint-Pierre 97410, France,

⁷University Paris-Saclay, INRAE, AgroParisTech, GABI, Jouy-en-Josas 78350, France,

⁸CBGP, INRAE, CIRAD, IRD, L'Institut Agro, University of Montpellier, Montpellier sur Lez 34988, France,

⁹SELMET, INRAE, CIRAD, L'Institut Agro, University of Montpellier, Montpellier 34398, France.

*Corresponding author: SELMET, INRAE, CIRAD, L'Institut Agro, University of Montpellier, Montpellier 34398, France. Email: laurence.flori@inrae.fr

Abstract

Despite their central economic and cultural role, the origin of cattle populations living in Indian Ocean islands still remains poorly documented. Here, we unravel the demographic and adaptive histories of the extant Zebus from the Mayotte and Madagascar islands using high-density SNP genotyping data. We found that these populations are very closely related and both display a predominant indicine ancestry. They diverged in the 16th century at the arrival of European people who transformed the trade network in the area. Their common ancestral cattle population originates from an admixture between an admixed African zebu population and an Indian zebu that occurred around the 12th century at the time of the earliest contacts between human African populations of the Swahili corridor and Austronesian people from Southeast Asia in Comoros and Madagascar. A steep increase in the estimated population sizes from the beginning of the 16th to the 17th century coincides with the expansion of the cattle trade. By carrying out genome scans for recent selection in the two cattle populations from Mayotte and Madagascar, we identified sets of candidate genes involved in biological functions (cancer, skin structure, and UV-protection, nervous system and behavior, organ development, metabolism, and immune response) broadly representative of the physiological adaptation to tropical conditions. Overall, the origin of the cattle populations from Western Indian Ocean islands mirrors the complex history of human migrations and trade in this area.

Keywords: cattle; genomics; demography; adaptation; Indian Ocean

Introduction

The Indian Ocean has played a prominent role in the human-mediated migration of cattle populations between East-Africa, Middle-East, and South-West Asia. However, the origin and genetic diversity of cattle populations living in Indian Ocean islands remain poorly investigated and relatively unclear. Their understanding may provide insights into the recent history of human populations in this area that has long displayed important levels of interaction, trade, migration, and domestic species translocation over time (Boivin *et al.* 2013; Beaujard 2019a, 2019b).

Within the large Indian Ocean area, the Comoro Islands (including Ngazidja, Ndzuwani, Mwali, and Mayotte) and Madagascar occupy a key position in the maritime trade routes that has linked the East-African coast, Middle East, and Asia over the past two millennia. Interestingly, cattle may have been

introduced early in this area and have subsequently represented an important domestic species. In both Mayotte and Madagascar where human have permanently settled from the 6th century CE, the most ancient archaeological and skeletal evidence of cattle presence traces back to the 9th –10th centuries CE but the quantities of identified cattle bones only increased from 14th to 15th centuries CE (Boivin *et al.* 2013; Pauly 2013). The first Portuguese eyewitnesses reported the presence of cattle on the Western Indian Ocean islands from the 16th century like almost all 17th and 18th century visitors in Comoros (Cheke 2010).

Nowadays, among the Comoros, the cattle population of Mayotte is the best-characterized thanks to a recent effort that lead to the official recognition of a local humped breed named “Zebu of Mayotte” (MAY) by the French government for conservation purpose (France 2011; Ouvrard *et al.* 2018). Overall this breed,

Received: December 13, 2021. **Accepted:** January 11, 2022

© The Author(s) 2022. Published by Oxford University Press on behalf of Genetics Society of America.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

traditionally and extensively raised in small herds of a handful of heads, represents about 70% of the 20,000 identified individuals identified in the island. Other individuals consist of recently imported individuals from European taurine (EUT) breeds (i.e. Montbeliarde, Jersey, French Brown Swiss, Gasconne) and MAYx EUT admixed individuals. In contrast, nine million of cattle heads are identified in Madagascar (Ministère de l'Agriculture 2007), including about 85% of Zebu of Madagascar (ZMA) raised in large herds, imported taurine breeds (e.g. Holstein, Norwegian Red), several synthetic admixed breeds with ZMA (such as Renitelo, Manjan'i Boina, and Rana) and Baria, a small wild and free-roaming population (Porter 2007, www.fao.org). Both MAY and ZMA populations are considered to be well adapted to their respective islands conditions. In particular, they are generally regarded as more resistant to heat stress and tick-borne diseases than imported European breeds, such as African zebu and N'Dama breeds (Mattioli et al. 1995; Bock et al. 1999; Hansen 2004; Glass et al. 2005).

If the MAY zebu population has not been genetically characterized yet, previous genetic characterization of ZMA populations based on the analysis of a few tens of microsatellite markers or medium-density SNP genotyping data suggested a hybrid composition between African taurine (AFT) and indicine (ZEB) ancestries, the latter being predominant (Zafindrajaona and Lauvergne 1993; Hanotte 2002; Gautier et al. 2009). However, these analyses remained mostly descriptive and only provided limited insights into the origins of these populations. In particular, from the history of human migration routes, cattle may have been introduced possibly repeatedly with subsequent exchanges between the 8th and the 13th centuries in the Comoro and Madagascar islands from several places including East Africa (as a result of movements of Bantu populations) or Indonesia (with Austronesian navigator populations) (Beaujard 2005, 2007, 2011, 2015; Fuller and Boivin 2009; Fuller et al. 2011).

The purpose of our study was to clarify the origin of the local Mayotte and Madagascar cattle breeds to better understand the demography of Western Indian Ocean cattle breeds based on their refined genetic characterization. To that end, we genotyped 32 MAY and 24 ZMA individuals on the bovineHD high-density SNP genotyping assay (comprising >770,000 SNPs) together with 113 individuals belonging to the Somba (SOM, $n=44$) and the Lagune (LAG, $n=44$) West-African taurine breeds and to the Fulani West-African Zebu (ZFU, $n=25$). These newly generated data were combined to publicly available BovineHD genotyping data for 363 individuals belonging to eight other breeds representative of EUT (Angus, Holstein, Jersey, and Limousine), another AFT (N'Dama), Indian Zebus (Gir and Nelore), and East African Zebus (Bahbahani et al. 2017). We first provided a refined analysis of the structuring of genetic diversity among the combined data sets and carried out a detailed inference of the demographic history of the MAY and ZMA populations with respect to the other extant populations by constructing admixture graphs (Patterson et al. 2012; Lipson 2020; Gautier et al. 2021) and estimating the timing of some admixture events using linkage-disequilibrium (LD) information (Loh et al. 2013). We further estimated their recent changes in effective population sizes using the recently developed method GONE (Santiago et al. 2020) and characterized the levels of genomic inbreeding in MAY and ZMA (Druet and Gautier 2017; Bertrand et al. 2019; Druet and Gautier 2021). We finally investigated the patterns of genetic adaptation of MAY and ZMA cattle breeds which recently diverged and live in slightly different tropical island conditions by searching for footprints of positive selection. The identified signals were subjected to a

detailed functional analysis to identify putative physiological pathways and their possible underlying selective pressures (see e.g. Flori et al. 2019).

Materials and methods

Animal sampling and genotyping

For MAY individuals (Zebus from Mayotte), we selected a group of 32 presumably nonrelated individuals (based on the newly created French National Registration Database) representative of the phenotypic diversity of the local population. They each originated from 32 different farms located in 17 townships spread over the Grande and Petite terres of the Mayotte Island (Supplementary Fig. 1, Supplementary Table 1). Blood samples were collected during year 2016 from the tail vein of the individuals using 10 ml EDTA vacutainer tubes, strictly following the recommendations of the directive 2010/63/EU for animal care. Genomic DNA was further extracted using the Wizard Genomic DNA purification kit (Promega, France) and stored at -20°C . MAY DNA samples were genotyped (Supplementary Table 1) on the Illumina BovineHD genotyping beadchip at Labogena platform (Jouy-en-Josas) using standard procedures (www.illumina.com) together with 24 Zebus of Madagascar (ZMA) and 113 West-African cattle DNA samples (i.e. 44 Lagune, 44 Somba, and 25 Zebu Fulani), sampled in the 1990s and previously genetically characterized on the Illumina BovineSNP50 beadchip (Gautier et al. 2009). The genotyping data were then added to the WIDDE database and combined with other publicly available ones, using WIDDE utilities (Sempéré et al. 2015).

To obtain the position of the 777,962 SNPs included in the BovineHD genotyping assay onto the latest ARS-UCD1.2 (aka bosTau9) bovine genome assembly (Rosen et al. 2020), we first retrieved the sequences of each SNP (length ranging from 61 to 121 bp with only 227 sequences <121 bp long) from the Illumina manifest file `bovinehd_15013478_b.csv`. These sequences were further realigned onto the ARS-UCD1.2 assembly using `pblat` (Wang and Kong 2019) run with options `-out=ps1x -minIdentity=98 -threads=4` and the resulting alignment file was parsed with a custom awk script. Unambiguous positions (i.e. with a single alignment hit of the underlying sequence) could then be obtained for 721,583 SNPs (92.8%). Among the 53,750 remaining SNPs, the number of alignment hits ranged from 2 to 1,790 and followed a L-shaped distribution with a median and mean values of 3 and 20.2, respectively. As a matter of expedience, we thus chose to discard from further analysis the 15,533 SNPs with more than 9 alignment hits (i.e. the third quartile of the aforementioned distribution) and to assign to the 40,846 other SNPs (with 2–9 hits) the position given by the alignment with the highest score. We indeed noticed that for these latter SNPs, multiple alignments mostly consisted of a single high scoring hit (score > 100 for 99% of them), the other alternative hits being generated by partial alignment of only a few tens bases of the SNP sequence.

The complete genotyping dataset consisted of 532 animals including 363 individuals from eight other bovine populations (Bahbahani et al. 2017, www.illumina.com) representative of the bovine worldwide genetic diversity (Table 1). The minimal individual genotyping call rate was set to 90% and the minimal SNP genotyping call rate to 90% overall populations and 75% within each population (i.e. SNPs genotyped for less than 75% of the animals from at least one population were discarded). SNPs with a $\text{MAF} < 0.01$ or departing from Hardy-Weinberg equilibrium expectation (exact test P -value $< 10^{-3}$ in at least one breed) were

Table 1. Sample description and origin of the Illumina BovineHD chip genotyping data (* before filtering).

Code	Population name	Country	Sampling year	Nb	Reference
ANG	Angus	Scotland	n.a.	42	Illumina (Sempéré et al. 2015)
EAZ	East African Shorthorn Zebu	Kenya	n.a.	92	Bahbahani et al. (2017)
GIR	Gir	India	n.a.	27	Illumina (Sempéré et al. 2015)
HOL	Holstein	Netherlands	n.a.	60	Illumina (Sempéré et al. 2015)
JER	Jersey	Jersey	n.a.	38	Illumina (Sempéré et al. 2015)
LAG	Lagune	Benin	1996	44	This study
LMS	Limousine	France (mainland)	n.a.	50	Illumina (Sempéré et al. 2015)
MAY	Zebu from Mayotte	France (Mayotte)	2016	30 (32)*	This study
NDA	N'Dama	Guinea	n.a.	23	Illumina (Sempéré et al. 2015)
NEL	Nelore	India	n.a.	31	Illumina (Sempéré et al. 2015)
SOM	Somba	Togo	1996	44	This study
ZFU	Zebu Fulani	Benin	1996	24 (25)*	This study
ZMA	Zebu from Madagascar	Madagascar	1991	23 (24)*	This study

also discarded. A total of 680,338 SNPs distributed throughout the 29 autosomes of the *bosTau9* bovine genome assembly passed finally all our filtering criteria.

Inference of the population demographic history

Characterization of the structuring of genetic diversity

Principal component analysis (PCA) based on individual SNP genotyping data was performed with *smartpca* (Patterson et al. 2006) and visualized with the R package *ggplot2* (Wickham 2016). Unsupervised genotype-based hierarchical clustering of the individual animal samples was carried out using the maximum-likelihood method implemented in ADMIXTURE 1.06 (Alexander et al. 2009). Results were visualized with custom functions in R environment (www.r-project.org). Finally, the overall and pairwise-population F_{ST} were computed with version 2.0 of the R package *poolfstat* (Gautier et al. 2021) using the *computeFST* ran with default settings (i.e. *method=Anova*) and option *nsnp.per.bjack.block = 5000* to estimate standard errors (and 95% CI as ± 1.96 s.e.) with *block-jackknife*.

f-statistics-based demographic inference

f-statistics based demographic inference (Patterson et al. 2012) were carried out with the new functionalities of the version 2.0 of the R package *poolfstat* (Gautier et al. 2021). We used the *compute.fstats* function to estimate the different *f*-statistics (including F_3 and F_4 for all the population triplets and quadruplets respectively) and within-population heterozygosities. As for F_{ST} , standard-errors of the estimated statistics (and their corresponding Z-scores for f_3 and f_4) were estimated using *block-jackknife* defining blocks of 5,000 consecutive SNPs (i.e. option *nsnp.per.bjack.block = 5,000*). In addition, to mitigate SNP ascertainment bias by favoring SNPs of most remote ancestry (i.e. discard SNPs of exclusive European ancestry), we only kept for *f*-statistics based demographic inference the 497,949 polymorphic SNPs that were polymorphic ($MAF > 0.05$) in both ZEB (GIR and NEL combined) and in AFT (NDA, SOM, and LAG combined) populations. Following Patterson et al. (2012) (see also Lipson 2020), we then carried out formal tests of population admixture using the estimated f_3 statistics. A negative Z-score ($Z < -1.65$ at the 95% significance threshold) associated to an f_3 for a given population triplet A; B, C showing that the target population A is admixed between two source populations each related to B and C. To further provide insights into the origins of MAY and ZMA, we build an admixture graph with *poolfstat* utilities (Gautier et al. 2021). Briefly, we first build a scaffold tree of presumably unadmixed populations (as suggested by both exploratory analyses and f_3 and f_4 based tests) consisting of two AFT breeds (NDA and LAG), two

ZEB breeds (GIR and NEL), and one EUT breed (HOL) with the *rooted.njtree.builder* function. We further relied on the *graph.builder* function (ran with default options) to jointly include EAZ, MAY, and ZMA that showed clear evidence for admixture on the graph considering all the six possible orders of inclusion. The fit of the best fitting graph (displaying a BIC more than 8 units lower than all the other graphs explored in the graph building process) was further validated with the *compare.fitted.fstats* function that allows to compare to which extent the estimated *f*-statistics depart from their predicted values based on the fitted admixture graph parameters via a Z-score (Patterson et al. 2012; Lipson 2020; Gautier et al. 2021).

Estimation of the timing of admixture events

We estimated the timing of admixture events (in generations) with the program *mALDER* (Pickrell et al. 2014) that implements a modified version of ALDER method originally described by Loh et al. (2013). This approach relies on the modeling of the exponential decay of admixture-induced LD in a target admixed population (here based on two-reference weighted LD curves, using a LD measure weighted by allele frequencies in two source population proxies) as a function of genetic distance. Genetic distances between pairs of SNPs were derived from physical distances assuming a cM to Mb ratio of 1 (Kadri et al. 2016). As commonly done in studies focusing on the population genetics of local cattle breeds, we here assumed a 6-year generation time to convert the timing from generations to years (e.g. Gautier et al. 2007; Flori et al. 2014; Mbole-Kariuki et al. 2014; Bahbahani et al. 2015, 2017). Such a long-term (i.e. across several generations) average generation time is in agreement with estimation by Keightley and Eyre-Walker (2000) for auroch and is also consistent with early estimation by Mahadevan (1955) (cited in Mbole-Kariuki et al. 2014) in the Red Sindhi East-African breed. We also obtained a very similar estimate of 5.6 years (± 0.3) by analyzing genotyping data publicly available for the synthetic Santa-Gertrudis breed (Matukumalli et al. 2009; Sempéré et al. 2015) which was established in 1910 in South-Texas by crossing taurine Shorthorns and Brahman zebus (Felius 2016). More precisely, to obtain this estimate of long-term generation time, we simply divided the presumed number of years separating birth of the genotyped samples from breed formation (ca. 95 years) by the timing of admixture estimated by ALDER (equal to 17.0 ± 0.99).

Inference of the recent population size histories

Historical effective population sizes (N_e) were inferred for the MAY and ZMA breeds with the program *GONE* that implements an approach recently developed by Santiago et al. (2020) to fit the

observed spectrum of LD of pairs of loci over a wide range of recombination rates (which we derived from physical map distances assuming a cM to Mb ratio equal to 1, see above). In practice, we adopted a block-jackknife approach to estimate confidence intervals for the inferred N_e trajectories by first identifying 55 nonoverlapping blocks of 10,000 consecutive SNPs (block size ranging from 32.8 to 40.2 Mb) out of the 680,338 genotyped ones. We then analyzed 55 different data sets of 670,338 SNPs that were each formed by removing from the original data sets one block of 10,000 SNPs. The resulting 55 inferred N_e trajectories were then summarized by computing a mean trajectory and a 95% confidence envelope defined by the 2.5 and 97.5 percentiles from the 55 estimated N_e at each time point.

Age-based partitioning of individual inbreeding in MAY and ZMA breeds

The characterization of individual levels of inbreeding at a global and local scale in MAY and ZMA breeds was performed with the model-based approach implemented in the RZooRoH package (Druet and Gautier 2017, 2021; Bertrand et al. 2019). This method models each individual genomes as a mosaic of Homozygous-by-Descend (HBD) and non-HBD segments using a multiple HBD-classes Hidden Markov Model (Druet and Gautier 2017). Each HBD class is specified by a rate R_k related to the expected length (equal to $\frac{1}{R_k}$ Morgans) of the associated HBD segments and that is approximately equal to twice the number of generations to the common ancestor that transmitted the DNA segment. Given the density of the HD chip, we considered a model with 11 HBD classes (with $R_k = 2^k$ for $k = 1, \dots, 11$) and one non-HBD class (with rate equal to R_{11}) allowing to capture the contribution to the overall individual inbreeding levels from each age-based classes of ancestors (living up to 1,024 generations in the past).

Whole-genome scan for footprints of selection

Computation of *iHS* and *Rsb* statistics

The genome-wide scan for footprints of positive selection within and between MAY and ZMA breeds was performed using extended haplotype homozygosity (EHH)-based tests. To that end we first jointly phased the genotyping data for MAY, ZMA, EAZ, GIR, and NEL individuals with version 1.4 of the fastPHASE software (Scheet and Stephens 2006) for each chromosome in turn using population label information and options `-Ku40 -k110 -ki10`. Genetic distances between consecutive SNPs were obtained from physical distances assuming a cM to Mb ratio equal to 1 (see above). Based on haplotype information, we further computed the *iHS* (Voight et al. 2006) and *Rsb* (Tang et al. 2007) statistics using the version 3.1.2 of the R package *rehh* (Gautier et al. 2017; Klassmann and Gautier 2020). These statistics are designed to detect regions with high level of haplotype homozygosity over an unexpected long distance (relative to neutral expectations), either within population (*iHS*) or across populations (*Rsb*). Note that for the computation of *iHS* within MAY and ZMA populations, we chose to not polarize the SNP alleles (i.e. `scan_hh` function was run with option `polarized=FALSE`) as discussed in section 7.6 of the online *rehh* package vignette (<https://cran.r-project.org/web/packages/rehh/vignettes/rehh.html>). The different SNP *iHS* and *Rsb* statistics were further transformed into $p_{iHS} = -\log_{10}(1 - 2|\Phi(iHS) - 0.5|)$ and $p_{Rsb} = -\log_{10}(\Phi(Rsb))$ where $\Phi(x)$ represents the Gaussian cumulative function. Assuming *iHS* and *Rsb* scores are normally distributed under neutrality, p_{iHS} and p_{Rsb} might thus be a 2-sided $-\log_{10}(P - \text{value})$ associated with the neutral hypothesis.

Identification of candidate genes

We used the approach previously described in Flori et al. (2019) to identify candidate genes under positive selection from *iHS* and *Rsb* estimates. Briefly, all the genotyped SNPs were annotated using as a gene set reference a list of 14,562 RefSeqGenes anchored on the Btau9 bovine genome assembly (`refGene.txt.gz`, 2019-06-07; <https://hgdownload.soe.ucsc.edu/goldenPath/bosTau9/database/>). A SNP was then considered as representative of one of these RefSeq genes if localized within its boundary positions extended by 15 kb upstream and downstream to account for persistence of LD (e.g. Gautier et al. 2007). Among the 680,338 SNP of HD-dataset participating to the analysis, 293,527 SNPs mapped to 13,696 different RefSeq Genes, corresponding to 13,536 gene symbols. On average, each SNP mapped within 1.2 RefSeq genes (from 1 to 34, median = 1) and each RefSeq gene was represented by 21 SNPs (from 1 to 641, median = 13). For a given statistic, genes with at least two representative SNPs with $-\log_{10}(P - \text{value}) > 4$ were considered as candidate genes.

Functional annotation of candidate genes under selection

Following the approach outlined in Flori et al. (2019), candidate genes were functionally annotated and submitted to gene network analysis using Ingenuity Pathway Analysis software (IPA, QIAGEN Inc., <https://www.qiagenbioinformatics.com/products/ingenuity-pathway-analysis>). Among the 13,535 genes symbols taken into account in the analysis, 12,324 were mapped in the Ingenuity Pathway Knowledge Base (IPKB) and were considered as the reference data set. Among the candidate genes identified for MAY ($N = 27$) and ZMA ($N = 47$) breeds, 24 and 45 were respectively mapped to IPKB. The top significant functions and diseases ($P - \text{value} < 0.05$) were obtained by comparing functions associated with the candidate genes under selection against functions associated with all genes in the reference set, using the right-tailed Fisher exact test. In the network analysis, a score S was computed for each network that contained at most 140 molecules (including candidate genes under selection) based on a right-tailed Fisher exact test for the over-representation of candidate genes under selection ($S = -\log_{10}(P - \text{value})$). A network was considered as significant when $S > 3$. The top significant functions and diseases associated with significant networks were also reported.

Results

Genetic relationships of Mayotte and Madagascar cattle breeds with other world cattle breeds

We first carried out an individual-based PCA of the SNP genotyping dataset consisting of 680,338 autosomal SNPs genotyped on individuals from MAY ($n = 32$) and ZMA ($n = 23$) populations together with 473 individuals belonging to 11 cattle breeds representative of the bovine genetic diversity (Table 1). The first factorial plan of the PCA is represented in Fig. 1a. The first two components accounting for 21.09% and 7.85% of the total variation, respectively. In agreement with previous studies (e.g. Gautier et al. 2010), the first two PCs revealed a clear structuring of individual genetic diversity according to their population of origins and highlighted a triangle-like 2-dimensional global structure of the cattle populations with apexes corresponding to three main groups: (1) EUT represented by ANG, HOL, JER, and LMS; (2) AFT represented by LAG, NDA, and SOM; and (3) zebu (ZEB) represented by NEL and GIR. Similarly, as previously observed with data from medium-density SNP genotyping data (on the same sample), ZMA and West-African hybrids (ZFU and EAZ) were

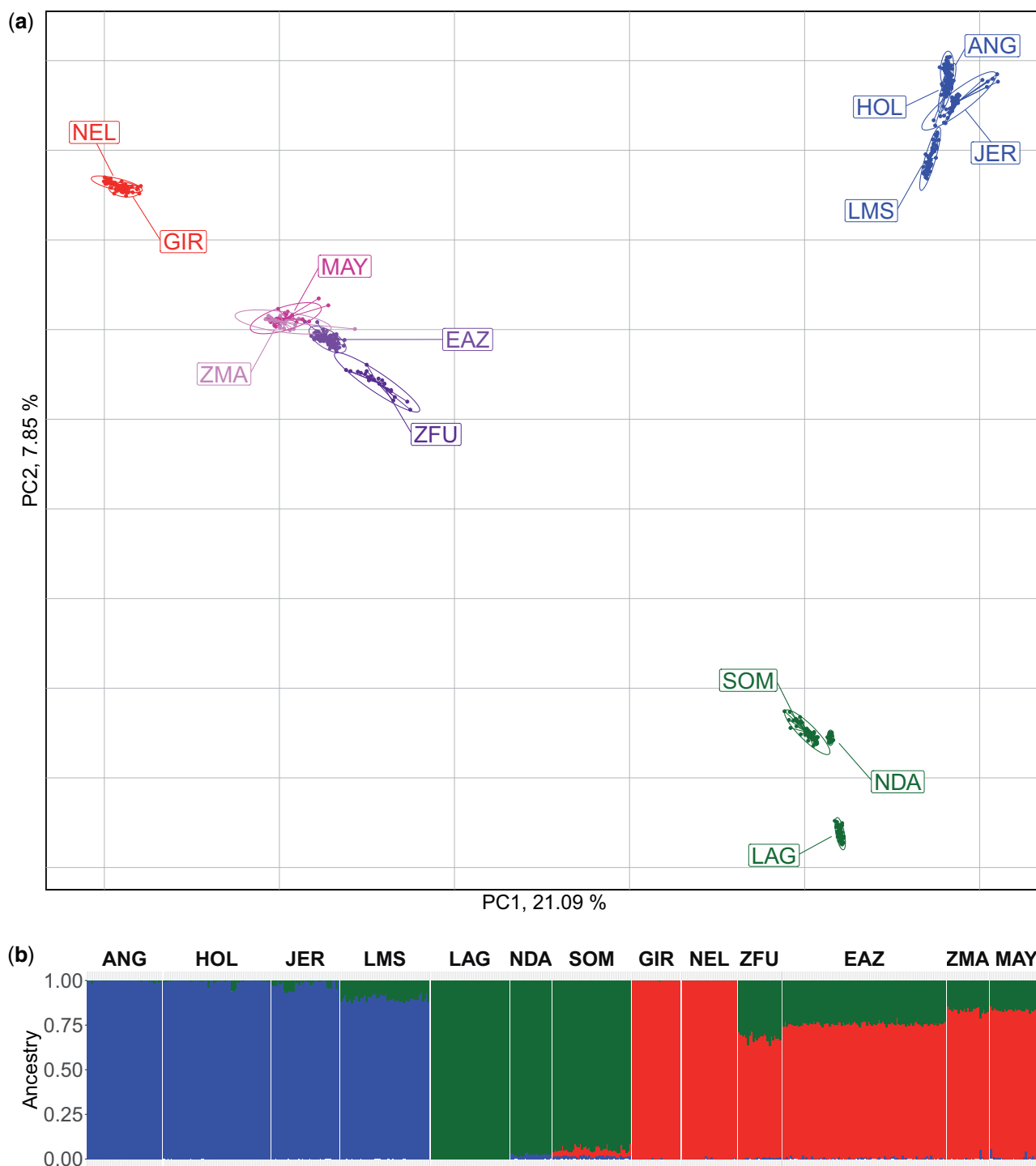


Fig. 1. Results of the PCA and unsupervised hierarchical clustering including HD genotyping data (528 individuals from 13 populations genotyped for 680,338 SNPs). a) PCA results. The individuals are plotted on the first two principal components according to their coordinates. Ellipses characterize the dispersion of each population around its center of gravity. MAY and ZMA individuals are plotted in dark-pink and pink and EAZ and ZFU individuals in purple and dark purple, respectively. EUT, AFT, and ZEB individuals are plotted in blue, green, and red, respectively. b) Unsupervised hierarchical clustering results with $K = 3$ predefined clusters. For each individual, the proportions of each cluster (y-axis) which were interpreted as representative of EUT, AFT, and ZEB ancestries are plotted in blue, green, and red, respectively.

located at intermediate positions on the AFT/ZEB segment, the ZMA individuals being closer to the ZEB apex (Gautier et al. 2010). Finally, the newly characterized MAY individuals appeared mostly confounded with ZMA individuals which is consistent with their close geographical proximity. Unsupervised hierarchical clustering analysis of the individuals using $K = 3$ predefined

clusters provided a complementary view in close agreement with PCA results while allowing to provide first estimates of ancestry proportions (Fig. 1b). Roughly interpreting the blue, red, and green cluster in Fig. 1b as representative of EUT, ZEB, and AFT ancestries respectively suggested that ZMA and MAY individuals had similarly high levels of ZEB ancestry (0.83 ± 0.02 and

0.82 ± 0.01 on average, respectively), a weaker level of AFT ancestry (0.16 ± 0.01), and almost no detectable EUT ancestry except for three individuals (2 MAY and 1 ZMA) with more than 5% of EUT ancestry, which were discarded from further analyses.

The different analyses performed at an individual scale showed that the partitioning of cattle into distinct populations (and breeds) is relevant to study their genetic history at the population level (e.g. Gautier et al. 2010). Accordingly, the overall F_{ST} among the 13 populations was found equal to 0.271 (95% CI, [0.262;0.279]). Population pairwise F_{ST} ranged from 2.40% (±0.01%) for the ZMA/MAY pair to 51.1% (±0.06%) for the LAG/NEL pair (Supplementary Fig. 2) confirming the close relatedness of MAY and ZMA populations. Among the 11 other populations, EAZ was found the most closely related to both MAY and ZMA ($F_{ST}^{may,eaz} = 7.69\%$ and $F_{ST}^{zma,eaz} = 8.07\%$) followed by ZFU and the two ZEB populations (GIR and NEL).

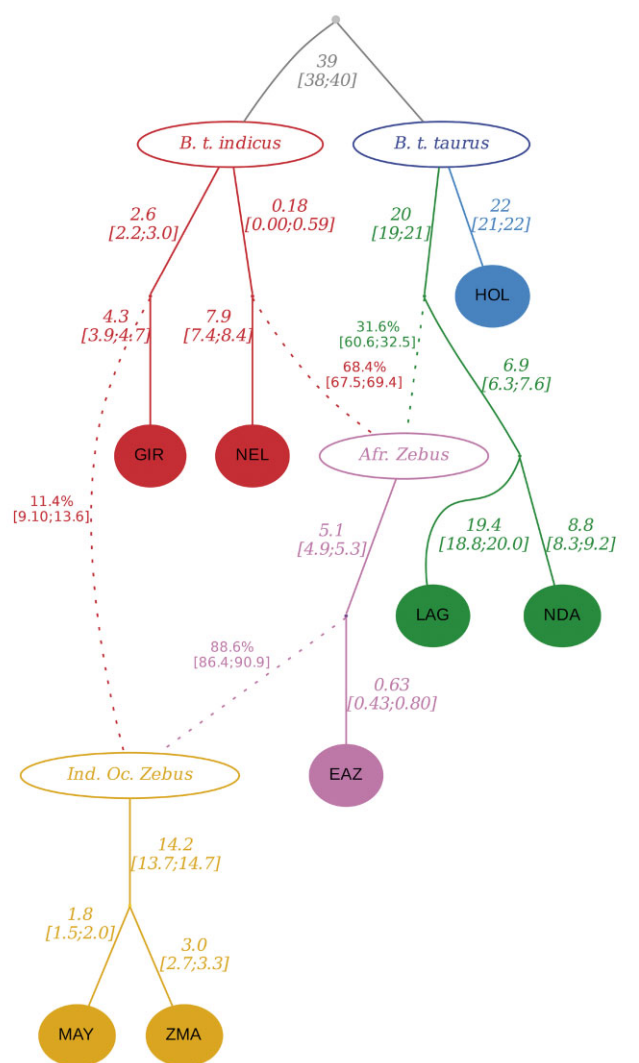
Inferring the history of Mayotte and Madagascar cattle populations

f_3 -based tests show clear evidence for admixture in the history of both MAY and ZMA

Out of the 66 possible f_3 -based tests when considering either MAY or ZMA as a target population, six were significantly negative (Z-score < -2.33 at the 99% significance threshold) providing strong evidence for admixture events in their history. They all involved the same pair of source proxies that consisted of one AFT population (LAG, NDA, or SOM) and one ZEB population (NEL or GIR), the most significant signal being observed with the LAG/NEL pair for ZMA [f_3 (ZMA; LAG, NEL) associated Z-score equal to -3.92] and the NDA/NEL pair for MAY [f_3 (MAY; NDA, NEL) associated Z-score equal to -6.38]. Among the 11 other populations, EAZ, ZFU, and SOM also showed clear evidence for admixture with 14, 14, and 6 f_3 -based tests with a significantly negative Z-score (at the 99% significance threshold), respectively. For each of these three target populations, the lowest Z-score was always associated with the LAG/NEL pair of source proxies [f_3 (EAZ; LAG, NEL), f_3 (ZFU; LAG, NEL), and f_3 (SOM; LAG, NEL) associated Z-scores being equal to -36.5, -33.6, and -9.13 respectively].

Admixture graph construction

To infer the history of MAY and ZMA ancestry, we constructed an admixture graph relating these two populations with two out of the three AFT breeds (LAG and NDA, discarding SOM as it displayed strong signal of presumably recent admixture with ZEB ancestry, see above), the two ZEB breeds (GIR and NEL), one (HOL) out of four EUT breeds and the EAZ African hybrid. We did not seek to include ZFU as it was more remotely related to East-African populations with a recent origin not directly informative for the MAY and ZMA history (Flori et al. 2014). The inferred best fitting admixture graph is represented in Fig. 2. It shows that MAY and ZMA breeds both derived from an admixed population we referred to Indian Ocean Zebu in Fig. 2, although it should be noticed that we have no clear evidence about the actual geographic origin of this ancestral population. This Indian Ocean Zebu had an admixed origin with a predominant ancestry (88.6%) from an admixed Zebu, likely of African origin since the most closely related to extant EAZ, and the remaining ancestry (11.4%) originating from a presumably Indian Zebu related to extant GIR. The former African Zebu was itself admixed with a Zebu related to extant NEL and a Taurine related to AFT ancestries (in 68.4% vs 31.6% respective proportions). Note that these inferred origins lead to overall ZEB (resp. AFT) related ancestries for MAY and ZMA similar to the ones obtained in Fig. 1.



Worst fitted f -statistic: f_4 (MAY,ZMA;HOL,LAG) [Z-score=-1.04]

Fig. 2. Inferred admixture graph connecting cattle breeds from Mayotte and Madagascar (MAY and ZMA, in yellow) with two Indian indicine breeds (GIR and NEL in red), one African zebu breed (EAZ in purple), two AFT breeds (LAG and NDA in green) and one European taurine breed (HOL in blue). Admixture events are shown by dotted arrows. Estimates of branch lengths ($\times 10^3$ in drift units of $\frac{2N_e}{\mu}$) and admixture rates are indicated next to the corresponding edges. The Z-score of the worst fitted f -statistics f_4 (MAY, ZMA; HOL, LAG) is equal to -1.04.

Timing of admixture events

We estimated the timing of the two aforementioned admixture event t_4^{afz} (leading to African Zebus, closely related to EAZ) and t_4^{ioz} (leading to Indian Ocean Zebu, the direct ancestor of MAY and ZMA) with the program mALDER Pickrell et al. (2014). Based on the inferred admixture graph (Fig. 2), we estimated t_4^{afz} by using EAZ as a population target and a pair of source population proxies consisting of one AFT (either NDA or LAG) and one ZEB (either GIR or NEL). In agreement with admixture graph fitting, the highest amplitude (i.e. y-intercept) estimate of the fitted weighted LD curve was obtained with NDA and NEL as source proxies which suggests that these populations are the closest (among the sampled ones) to the actual source population (Loh et al. 2013). The corresponding estimated timing for the admixture events leading to African Zebu was found equal to $t_4^{afz} = 174 \pm 19$ generations (i.e. 1,044 ± 114 years). This admixture events thus traces back to the 10th century CE (ca. year 950 ± 150). We similarly estimated

t_{α}^{ioz} by considering either MAY or ZMA as a population target and a pair of source population proxies consisting of EAZ and one ZEB (either GIR or NEL). For both target populations, the amplitude was the highest (although only slightly) with GIR as the ZEB represent. The estimated timing for the admixture events leading to Indian Ocean Zebus was found equal to $t_{\alpha}^{ioz} = 153 \pm 6$ and $t_{\alpha}^{ioz} = 152 \pm 9$ generations with MAY and ZMA as a target population respectively. The (slightly) lower latter estimate being consistent with the difference in sampling time (ca. 4 generations) between MAY and ZMA samples. This second admixture events thus traces back to the early 12th century CE (ca. year 1,100 \pm 40).

Recent population size history of MAY and ZMA

Figure 3 plots the recent effective population size histories of MAY and ZMA as inferred with GONE (Santiago et al. 2020). The two trajectories were found similar before the beginning of the 16th century (i.e. ca. 80 generations before MAY sampling time) when the two populations likely started to diverge. From that time onward, the two populations experienced a rapid growth in a few generations toward a mostly constant size (approximately twice higher for ZMA and MAY) followed by a new moderate increase from the beginning of the 20th century. As expected from the large differences in census cattle population sizes between Mayotte and Madagascar islands, the current estimated N_e was about twice lower in MAY compared to ZMA. More precisely, the harmonic mean of the (mean) historical N_e from generation 80 (76 for ZMA) onward was found equal to 2,160 and 1,045 for ZMA and MAY, respectively. Surprisingly, a close inspection of the inferred admixed graph in Fig. 2 seems to contradict these large differences in N_e since the estimated (leave) branch lengths was about twice higher for ZMA compared to MAY. This thus rather suggested increased drift in ZMA since these estimates (on a

diffusion timescale) might be interpreted under a pure-drift model of divergence as inversely proportional to N_e (i.e. equal to $\frac{t}{2N_e}$ where t is the divergence time in generation). Population-specific F_{ST} estimates for MAY and ZMA essentially confirmed this trend (1.59% and 3.15% respectively, Supplementary Fig. 2). Conversely, when considering only the 518,315 SNPs with a MAF >1% in MAY and ZMA (computed over MAY and ZMA combined individuals), the estimated heterozygosity was slightly higher in MAY (33.2%) than in ZMA (32.7%). As detailed in Supplementary Fig. 3, these apparent discrepancies may actually be explained by some residual asymmetric gene flow between MAY and ZMA (or from a common gene pool related to some other continental population, e.g. from the East-African coast) after these two populations split. More specifically, very similar level of differentiation and heterozygosities among MAY and ZMA could be obtained with data simulated with msprime (Kelleher et al. 2016) under a simplified scenario assuming MAY and ZMA split 80 generations ago and maintained a constant N_e (equal to the 1 estimated above) with MAY receiving twice as many migrants as ZMA from a common gene pool ($m_{\rightarrow MAY} = 4 \times 10^{-3}$ vs. $m_{\rightarrow ZMA} = 2 \times 10^{-3}$) (Supplementary Fig. 3).

MAY and ZMA individuals levels of inbreeding

In order to characterize individual levels of recent inbreeding at both a global (genome-wide) and local (locus-specific) scale, we estimated the individual inbreeding coefficients in MAY and ZMA populations by applying the method implemented in RZooRoH software (Druet and Gautier 2017; Bertrand et al. 2019). The distribution of the genome-wide coefficients of individual inbreeding estimated with the R package RZooRoH are detailed in Fig. 4a for the MAY and ZMA populations together with their age-based partitioning to assess the contribution of various class of ancestors

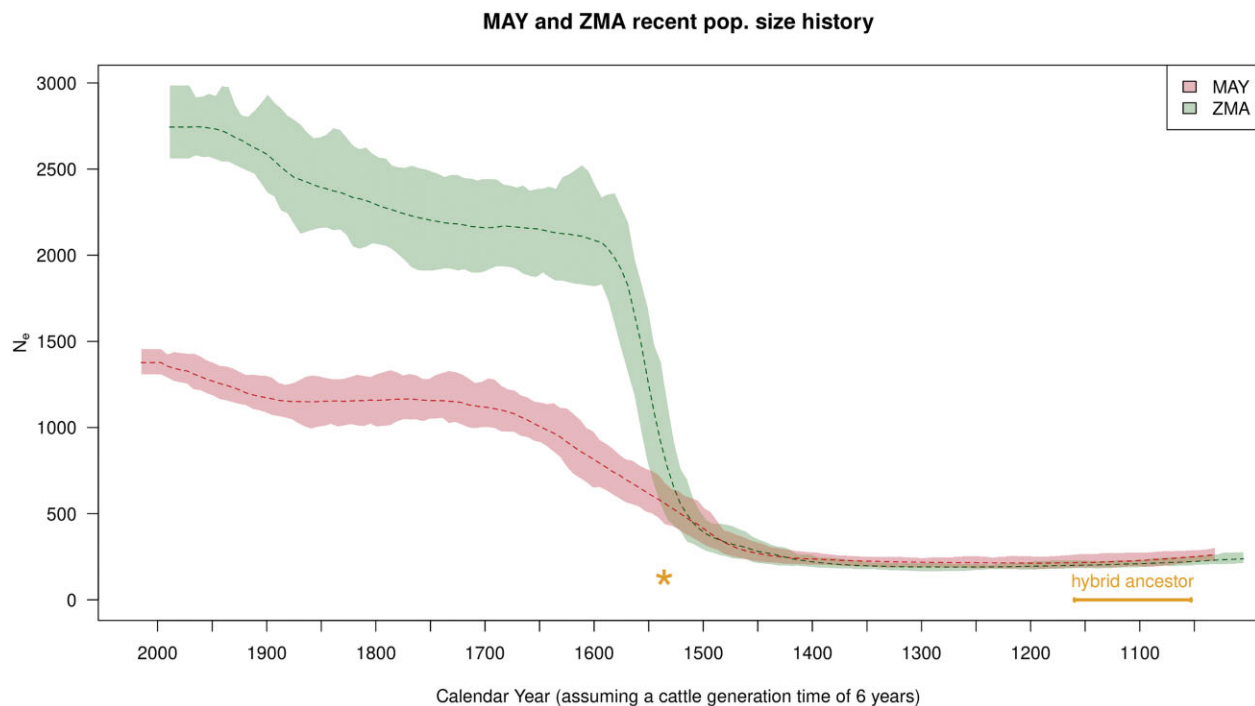


Fig. 3. Population size history (N_e) of MAY and ZMA populations estimated with GONE (Santiago et al. 2020). The average N_e trajectories (dashed line) and 95% confidence envelope estimated from block-jackknife samples are plotted in green for ZMA and red for MAY. The time scale was transformed into calendar assuming a 6-year generation time for cattle and accounting for the difference in sampling time between ZMA (1990) and MAY (2016). The estimated timing of the admixture event (t_{α}^{ioz}) that led to the common hybrid ancestor (named Indian Ocean Zebus in the main text) of MAY and ZMA is given in orange. The orange asterisk gives the likely splitting time (beginning of the 16th century, i.e., $t_s \approx 80$ generations before MAY sampling time) between MAY and ZMA roughly estimated from the separation of the two trajectories.

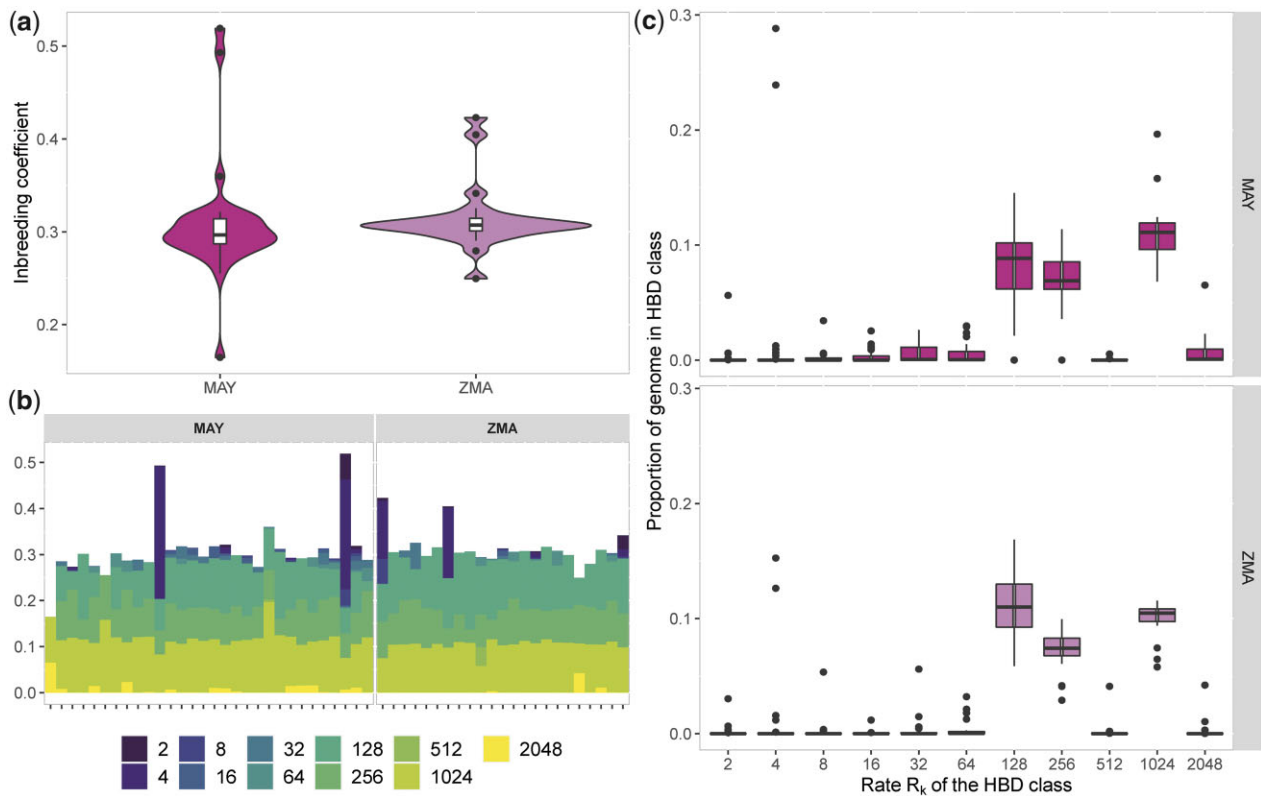


Fig. 4. Characterization of individual inbreeding levels in Mayotte and Madagascar breeds. a) Violin plots representing the distribution of inbreeding coefficients for the MAY ($N = 30$) and ZMA ($N = 23$) breeds, colored in pink and purple, respectively. b) Partitioning of individual genomes in different HBD classes. Each bar represents an individual and its total height the overall level of inbreeding. The height of the different stacks, which appear in different colors, represents the proportion of the genome associated with each HBD class, defined by their rate R_k . c) Average proportion of individual genomes associated with different HBD classes for MAY (pink) and ZMA (purple) populations. Individual proportions of the genome associated with a specific HBD class are obtained by averaging the corresponding HBD-classes probabilities over all marker positions.

(Fig. 4, b and c, Supplementary Fig. 4). Overall, the average individual inbreeding levels were found similar and equal to 0.308 ± 0.06 and 0.3136 ± 0.03 in MAY and ZMA respectively, MAY and ZMA individuals all displaying at least 16% and 25% of their genome in autozygous (HBD) segments. However, in agreement with the inferred trajectories of historical N_e (Fig. 3), most HBD segments were concentrated in ancestor age classes associated with R_k equal to 128 and 256 (i.e. approximately 64 and 128 generations before sampling thus spanning the split time period between MAY and ZMA). Hence, most of the estimated inbreeding level originated from the foundation of the two populations. Consistently, only a few individuals displayed a high proportion of HBD segments associated with the most recent common ancestors, two generations ago. More precisely, two MAY individuals showed more than 49% of their genome in HBD classes and two ZMA individuals showed more than 40% of their genome classified as HBD.

Identification of selection footprints in the MAY and ZMA genomes

The genome of ZMA and MAY individuals was scanned for footprints of selection with the package *rehh* 3.1.2 (Gautier et al. 2017), following the approach described in Flori et al. (2019) to identify directly candidate genes. According to our criteria (see Materials and Methods), the computation of the *iHS* statistic for each SNP over the whole MAY and ZMA genomes detected two (i.e. GRIK2 and RBM47) and 12 (i.e. AASS, ACTR6, ATP10D, COL15A1, DPP6,

DPYSL5, EXOC4, ITGA7, MAPRE3, SLC17A8, TNFSF11, and ZCCHC7) genes under selection located on two (i.e. BTA6 and 9) and six (i.e. BTA4, 5, 6, 8, 11, and 12) chromosomes, respectively (see details in Supplementary Table 2; Supplementary Figs. 5 and 6), with no overlap between these two lists of genes. By computing different *Rsb* statistics comparing the local extend of haplotype homozygosities between MAY and ZMA, and between these two breeds and each of their different main ancestries (i.e. AFZ and ZEB), 3, 16, 12, 18, 11 and 11 candidate genes were identified with the *Rsb*_{MAY/ZMA}, *Rsb*_{MAY/EAZ}, *Rsb*_{MAY/ZEB}, *Rsb*_{ZMA/MAY}, *Rsb*_{ZMA/EAZ}, and *Rsb*_{ZMA/ZEB} tests, respectively (Supplementary Table 2, Supplementary Figs. 5 and 6). Most of the candidate genes identified by the *Rsb* involving MAY or ZMA populations were different (74% and 85%, respectively). However, seven candidate genes (i.e. GRIK2, HMGSC2, LPCAT2, MIR2285BM, PHGDH, TRIM10, and TRIM15) were identified under selection in both breeds by some of these tests. Specifically, GRIK2, located on BTA9 was found under selection by five tests (*iHS*_{MAY}, *Rsb*_{MAY/EAZ}, *Rsb*_{MAY/ZEB} and *Rsb*_{ZMA/EAZ} and *Rsb*_{ZMA/ZEB} tests), HMGSC2 and PHGDH located on BTA3 and LPCAT2 located on BTA18 by *Rsb*_{MAY/ZEB} and *Rsb*_{ZMA/ZEB} tests, MIR2285BM located on BTA16 by *Rsb*_{MAY/ZEB} and *Rsb*_{ZMA/EAZ} tests and TRIM10 and 15 located on BTA23 by *Rsb*_{MAY/EAZ} and *Rsb*_{ZMA/EAZ} tests. *Rsb*_{MAY/ZMA} test detected three (i.e. LRRK2, GSTCD, and GPC5) and 18 genes (i.e. ATP2C1, SCN2A, STPG1, KCND2, ANKS1B, NR1H4, MIR2434, ANO4, SPIC, KCTD8, TG, LRRC6, CACNA1E, KCNH1, SLC4A7, ZSCAN31, PGBD1, and ZSCAN26) under selection in MAY and in

ZMA, respectively. Considering each breed separately, four candidate genes (i.e. GBE1, GPC5, SLC6A2, and ZMAT4) were detected in MAY population and four (i.e. CPS1, KCND2, KCTD8, and KCNH1) in ZMA population by two *Rsb* tests.

Overall, a total of 27 and 47 candidate genes under selection were detected in at least one of the above tests for MAY and ZMA, respectively. To obtain a global view of the main gene functions under selection, these 2 lists of candidate genes were annotated using IPA software. The main functional categories, in which these genes are involved, are listed in [Supplementary Table 3](#) (see [Supplementary Tables 4](#) and [5](#) for an exhaustive list of their functional annotation). For MAY, the top 5 significant functions belonging to the 3 IPA main categories (i.e. Diseases and Disorders, Molecular and Cellular Functions and Physiological System Development and Function) were related to inflammation (“Inflammatory Disease”), cancer and cell death (“Cancer” and “Cell Death and Survival”), gastrointestinal and hepatic diseases (“Hepatic Disease,” “Gastrointestinal disease”), nervous system (“Nervous System Development and Function”), organ and tissue development and morphology (“Organismal Injury and Abnormalities,” “Cell Morphology,” “Cellular development,” “Embryonic Development,” “Organ development,” “Organ morphology,” and “Organismal development”) and inflammation (“Inflammatory Disease”). For ZMA, the top 5 significant functions were related to cancer, dermatology (“Dermatological Diseases and Conditions”), organ and embryonic development (“Organismal Injury and Abnormalities,” “Organismal Development,” “Embryonic Development,” “Organ Development”), metabolism and gastrointestinal disease (“Molecular Transport,” “Lipid Metabolism,” “Small Molecule Biochemistry,” “Gastrointestinal Disease”), respiratory (“Respiratory System Development and Function”) and nervous (“Nervous System Development and Function”) systems ([Supplementary Table 3](#)). Interestingly, many functional categories or subcategories (nine among the top five significant functions) were found in both populations such as functions related to cancer, nervous system, organ development, and gastrointestinal disease.

Moreover, candidate genes identified in each breed participated to one significant network ([Supplementary Table 6](#)) which included 85% and 93% of candidates genes for MAY (score = 54) and ZMA (score = 108) breeds, respectively ([Fig. 5, a and b](#)). Ten molecules are in common between both networks (i.e. AR, GRIK2, HMGCS2, LPCAT2, MAP3K10, MAP3K11, PHGDH, TRIM10, TRIM15, and kynurenic acid) among which six were detected as candidate genes under selection (i.e. GRIK2, HMGCS2, LPCAT2, PHGDH, TRIM10, and TRIM15). Candidate genes were located in periphery or sub-periphery of both networks and no main node were detected under selection, except TNFSR11, associated with a significant *iHS* for ZMA breed. The functional annotation of each network gave a more complete picture of the functions involved in adaptation of MAY and ZMA breeds ([Supplementary Tables 7–9](#)). Molecules integrated in the MAY network play a role in carbohydrate metabolism, development, nervous and respiratory systems function, dermatology, inflammation, and behavior. The ZMA network is mainly involved in cancer, in organismal abnormalities, cell signaling growth and proliferation, reproductive system, dermatology, gastrointestinal disease, endocrine and nervous systems, and inflammation.

Discussion

The genetic characterization of Zebus from Mayotte and Madagascar (MAY and ZMA), two geographically closely related

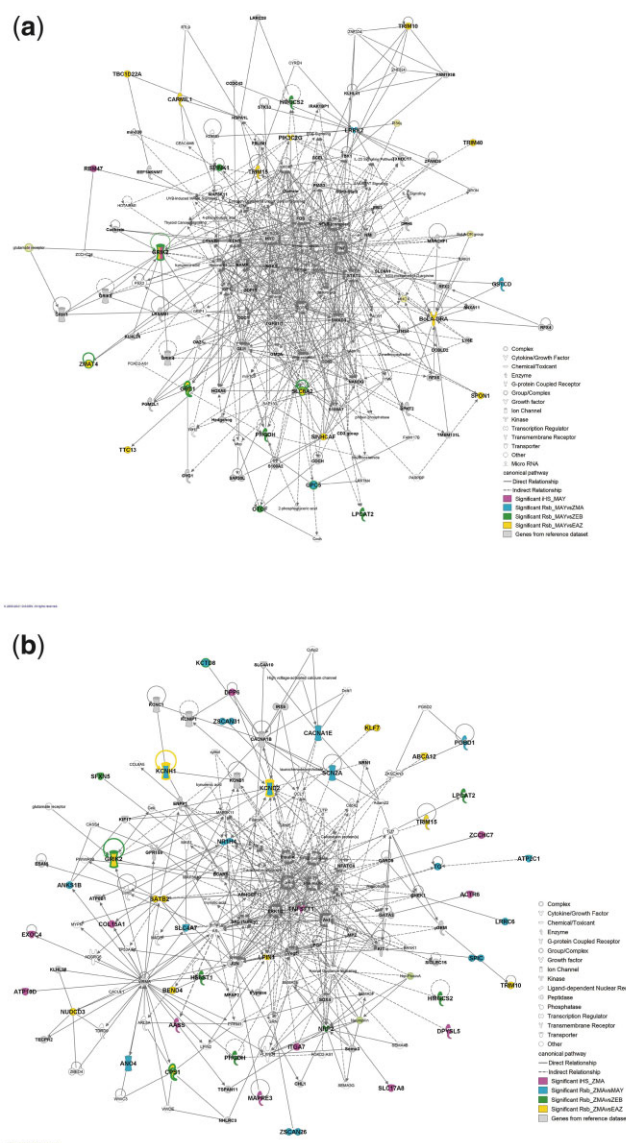


Fig. 5. Gene networks including candidate genes under selection detected in at least one *iHS* or *Rsb* test in the genome of breeds from Mayotte (a) and Madagascar (b)

island cattle populations from the Western Indian Ocean, showed that they share a similar recent and rather complex demographic history. In particular, we detected in these two populations a weak African taurine ancestry and a predominant indicine ancestry. The indicine ancestry in MAY and ZMA was even higher than in the East-African zebu breed, the African continental breed with the highest estimated indicine ancestry proportion in our dataset. This pattern had already been reported for ZMA in some previous studies ([Hanotte 2002](#); [Gautier et al. 2009](#)) but still remained difficult to explain from an historical point of view. Our demographic inference, in the form of admixture graph construction, demonstrated that the higher indicine ancestry in the MAY and ZMA populations, as compared to other African continental zebus, could actually be explained by a second pulse of Indian Zebu introgression, limited to the island populations, into an already admixed African taurine x Zebu population of likely East-African origin that traces back to the 12th century. These results are in agreement with the recent findings in human population dynamics in the area ([Brucato et al. 2017, 2018](#)) based

on genetic data analysis, which report that Comorian and Malagasy communities resulted from admixture between Swahili people from East-Africa and individuals from islands of Southeast Asian (Austronesian) dating from 8th to 12th centuries (Brucato et al. 2018). Migrations of Austronesian populations were promoted by the significant development of seaborne trade, from the Arabian Peninsula and Indonesia to East Africa and Madagascar during this period (Brucato et al. 2016, 2018; Bahbahani et al. 2017; Choudhury et al. 2018). These excellent navigators should have transported indicine cattle (and probably other livestock species) toward Western Indian Ocean that must have been crossed with AFZ individuals brought from African East coast by Swahili people.

Estimation of past effective population sizes for both MAY and ZMA populations showed similar trend and allowed clarifying the timing of their recent split to the 16th century which coincides with the arrival of Europeans in the area and the modification of the trade network (Newitt 1983; McPherson 1984; Beaujard 2005). Some residual cattle migration may have still remained after this split with either a differential contribution of an unknown continental population to MAY and ZMA or some weak and asymmetric gene flow between them. Further discriminating between these two scenarios would require additional samples from the area that may be difficult to collect (e.g. ancient DNA). Looking forward in time, after the split of MAY and ZMA, we estimated a steep increase in their ancestral populations sizes starting from the beginning of the 16th century to the 17th century, which was more pronounced for ZMA probably due to the far larger size of Madagascar. This observed cattle population expansion may be related to the increase of the livestock production by islands communities, in particular to meet the growing demand of European ships in fresh meat (Newitt 1983). Indeed, as the Portuguese retained control of the East African coast, the other Europeans (Dutch, English, and French), who began to enter Indian Ocean, needed revictualling ports and bases that Madagascar and the Comoros islands could offer. Cattle then represented one of the main trading resources that contributed to the economic development in this area during the early 17th century. From the middle of the 17th century, the population sizes of both MAY and ZMA tended to stabilize suggesting a maintenance of cattle production until the beginning of the 20th century, when both population sizes started to increase again but at a more moderate rate. This period actually coincides with French colonization of both islands that possibly lead to a new development in the use of local bovine resources. As expected, the characterization and partitioning of inbreeding levels into age-related HBD classes for MAY and ZMA individuals was found consistent with the inferred recent history of their corresponding population N_e . More specifically, in both populations, the contribution to individual inbreeding levels of HBD classes of most recent origin (with rates $R_k < 64$, i.e. tracing back to ancestors living less than 32 generations ago) remained limited, except for a few individuals that may result from sporadic unintentional consanguineous matings. This is thus quite encouraging from a breed conservation perspective. Conversely, the highest contributing HBD classes pointed to ancestors living between 64 and 128 generations (i.e. HBD classes with rates $R_k = 128$ and $R_k = 256$). Assuming a 6-year generation time (see above), this corresponds to the middle of the 13th century and the middle of the 17th century. Accordingly, very similar profile of the most remote HBD classes contribution (with $R_k \geq 128$) are observed for both MAY and ZMA individuals since these pertain to their common ancestral population. It should also be noticed, that considering a smaller (i.e. 5

years) or larger (i.e. 7 years) generation time did not fundamentally challenge our interpretation of the demographic history of MAY and ZMA with respect to either the effective population size histories or timing of admixture events (Supplementary Fig. 7).

From the 12th century (i.e. ca. 150 generations ago) onward, cattle living in Comoros and Madagascar islands have likely experienced various environmental and artificial selection pressures. To identify their footprints in the MAY and ZMA breeds, we relied on EHH-based tests considering different contrasts that our detailed inference of the demographic history helped interpreting. First, it should be noticed that only a few candidate genes were found in common in the two populations when comparing the various population-specific signals. These include (1) two genes (TRIM10 and TRIM15) with significant $Rsb_{MAY/EAZ}$ and $Rsb_{ZMA/EAZ}$ signals; and (2) five genes (HMGSC2, PHGDH, GRIK2, MIR2285BM, and LPCAT2) with significant $Rsb_{MAY/ZEB}$ and $Rsb_{ZMA/ZEB}$ signals. Note that among the latter, GRIK2 also displayed a significant *iHS* signal in the MAY breed. These different common footprints may be considered of older origin in response to environmental (e.g. tropical climate) and human-driven constraints imposed to the common ancestral population of MAY and ZMA breeds (i.e. predating their divergence). Among these genes, TRIM10 and TRIM15 are involved in innate immune response and were already detected under selection in Muturu cattle in Africa (Tijani et al. 2019). In addition, missense mutations (with respect to the Taurine assembly) within TRIM10 were found fixed in Nelore suggesting a putative role in tropical adaptation (Júnior et al. 2020). HMGSC2, PHGDH, and LPCAT2 are related to lipid or serine metabolism (Hegardt 1999; Vilà-Brau et al. 2011; Morimoto et al. 2014; Reid et al. 2018), the first two being reported as associated with some fatty acid in intramuscular fat of Nelore cattle (Cesar et al. 2014). GRIK2 that participates to the metabolism of glutamate, an important brain neurotransmitter (Purves et al. 2001), is involved in tropical cattle behavior. For instance, deficiency of this gene was found associated with a significant reduction in anxiety, fear memory and the flight time temperament phenotype in tropical cattle with varying amount of indicine ancestry (Ko et al. 2005; Porto-Neto et al. 2014).

Conversely, significant signals with $Rsb_{MAY/ZMA}$ based tests only, provided insights into footprints of selection of recent origin postdating the divergence of the two populations. These signals might be related to local differences between the tropical climate of Mayotte and Madagascar. In the small Mayotte island, the climate is homogeneous with a hot, humid and rainy season during the northeastern monsoon and a cooler dry season. In contrast, Madagascar island whose surface area is 15 times greater than that of Mayotte, presents a wider variety of climates being humid tropical on the East coast, dry tropical on the West coast and temperate on the Highlands. In addition, the differential evolution of husbandry practices in the two islands after the divergence of the two populations may also have contributed to leave different selection footprints. For instance, in Mayotte, Zebus have been reared in the recent past in traditional extensive systems with small herds (a few heads) with regular close contacts with breeders. Besides, in Madagascar, the sampled individuals originated from an area where breeding practices are based on larger herds (from four to few hundred heads) often dispersed on large pasture territory (Zafindrajaona 1991). A higher number of ZMA driven than MAY driven signals ($n = 18$ vs $n = 3$) were found with this test, as expected from the twice highest N_e of ZMA over the period which may result in improved detection power. Among the three genes that display significant signals in the MAY breed (LRRK2, GSTCD, and GPC5), GSTCD is involved in lung

function and was identified as CNVR-harbored gene associated with adaptation to hypoxia in yak population (Wang et al. 2019). GPC5 was associated with feed efficiency in cattle (Serão et al. 2013; Buitenhuis et al. 2014). The 18 genes with significant $Rsb_{MAY/ZMA}$ signals driven by ZMA breed included ANO4, which is located in a region under selection in New World Creole cattle and in Korean Brindle Hanwoo cattle (Gautier and Naves 2011; Kim et al. 2018). This gene belongs to the anoctamin protein family, whose members are expressed in sweat glands and are involved in thermal sweating, one of the mechanisms of heat tolerance (Jian et al. 2014; Cui and Schlessinger 2015). Within this gene list, KCTD8, encoding a potassium channel, was found associated with the Temperature Humidity Index in Mediterranean cattle breeds and also identified as a heat stress responsive gene in chicken (Sun et al. 2015; Flori et al. 2019). It is also associated with milk fat percentage in buffaloes (De Camargo et al. 2015) and located within selection signatures detected in dairy cattle breeds (Hayes et al. 2008; Flori et al. 2009). Potassium channels are also related to the production of prolactin, a key hormone for mammary development, milk production, hair development, thermoregulation and water balance during heat stress (Underwood and Suttle 1999; Silanikove et al. 2000; Czarniecki et al. 2003; Littlejohn et al. 2014). KCNH1 represents another remarkable gene since it is associated with a syndrome characterized by dysmorphism and hypertrichosis in human (Kortüm et al. 2015) and was associated with several climatic variables in local Mediterranean cattle breeds (Flori et al. 2019). Finally, the thyroglobulin gene (TG), encodes a protein required for thyroid hormone synthesis and iodine storage and contains variants associated with carcass and meat quality traits in beef cattle (Hou et al. 2011).

The timing of the footprints underlying the other 18 and 26 candidate genes found specific to MAY ($iHS_{MAY}, Rsb_{MAY/EAZ}, Rsb_{MAY/ZEB}$) or to ZMA ($iHS_{ZMA}, Rsb_{ZMA/EAZ}, Rsb_{ZMA/ZEB}$) breeds is more difficult to interpret. For instance, these footprints may either result from recent breed-specific selective constraints or older selective constraints further relaxed in one breed. In both cases, the fact that these genes were not identified with $Rsb_{MAY/ZMA}$ could be due to the overall limited power of the different tests (see above). Yet, some noticeable genes showed up in the list of these breed-specific candidate genes. For instance, among the genes specifically detected in MAY, SPINK1 promotes the progression of various types of cancer and is involved in abnormal morphology of thin skin (Lin 2021). PIK3C2G, under selection in Normande and Holstein breeds (Flori et al. 2009), is involved in the regulation of insulin-mediated activation of the PI3K/Akt pathway in the liver that controls metabolism (Braccini et al. 2015), PI3K and Akt corresponding to central nodes of the MAY network (Fig. 5a). TRIM40, is involved in innate immune response and was already detected under selection in Muturu cattle in Africa (Tijjani et al. 2019) and BOLA-DRA encodes the alpha subunit of the class II Major Histocompatibility Complex, expressed at the Antigen-Presenting Cells surface, which presents peptides from foreign origin to the immune system. Among the genes specific to ZMA breed, DDP6 was associated with feed efficiency (Serão et al. 2013; Buitenhuis et al. 2014; de Camargo et al. 2015), COL15A1 with iris hypopigmentation in Holstein Friesian cattle (Hollmann et al. 2017) with a possible involvement in UV-protection and NUDCD3, with milk yield and components in Swedish cattle breeds (Ghoreishifar et al. 2020), this latter being under positive selection in dairy cattle breeds and across Holstein, Angus, Charolais, Brahman, and N'Dama breeds (Flori et al. 2009; Xu et al. 2015).

Overall, the functional annotation of all the candidate genes identified with at least one test and of their inferred underlying

networks provided some insights into the main functions involved in adaptation of cattle to Western Indian Ocean tropical conditions and breeding practices. Some genes (e.g. SPINK1 in MAY and ANO4, KCTD8, COL15A1, and KCNH1 in ZMA) are related to cancer and skin properties and may have played a role in adaptation to Mayotte and Madagascar climates by increasing tolerance to thermal stress, humidity, and UV exposition. Respiratory function in which some candidate genes are involved (e.g. GSTCD in MAY) may also be related to tropical environment adaptation, breeds from warm climates presenting lower respiration rates under heat stress to maintain their body temperature. Several genes carrying footprints of selection are involved in nervous system and cattle behavior (e.g. GRIK2 in MAY and ZMA) which indicates also a physiological adaptation to the livestock system and the proximity with breeders in both islands. Development of different organs or physiological structures, gastrointestinal and hepatic system function, metabolism, and hormone production are also among the main functions targeted by selection in the two breeds (e.g. HMGCS2, PHGDH, MIR2285BM, and LPCAT2, detected in MAY and ZMA breeds, GPC5 detected in MAY breed and NUDCD3, TG, and DDP6 detected in ZMA breed). If these signatures of selection could be related to artificial selection on conformation and production traits, they could also be linked to the adaptation to tropical climate (e.g. through thermoregulation) and to available food and water resources. At last, several candidate genes involved in inflammatory and immune response (e.g. TRIM10, TRIM15, TRIM40, and BOLA-DRA) play a role in adaptation of Mayotte and Madagascar breeds suggesting that tropical pathogens responsible for e.g. as East Coast Fever, Rift Valley Fever, or blackleg documented outbreaks, may have had a deep impact on the adaptive genetic diversity of MAY and ZMA breeds (De Deken et al. 2007; Cêtre-Sossah et al. 2012; Dommergues et al. 2015; Porter et al. 2016).

This study displays for the first time a genetic characterization of the local cattle breed of Mayotte and presents a deep analysis of demographic and adaptive histories of cattle breeds from Mayotte and Madagascar islands. The genetic proximity between these two populations reflects their closely tied demographic history until the 16th century before the population divergence. Their demographic history mirrors the complex pattern of human migrations and trade in Western Indian Ocean islands. Their adaptive history has been probably overall conditioned by the same selective pressures and also by some differences between climates and breeding practices in the two islands. This study highlights the great originality of Zebu from Madagascar and Mayotte compared to the cattle populations from Africa mainland and their relevance in the context of global changes, which promotes specific measures of conservation especially for Mayotte breed.

Data availability

SNP genotyping data are available in the WIDDE database (<http://widde.toulouse.inra.fr/widde/>) and in the portal Data INRAE (<https://data.inrae.fr/>; <https://doi.org/10.15454/IQL6GE>).

Supplemental material is available at G3 online.

Acknowledgments

The authors would like to thank Sitty Bahyat Chamassi et Abdou Achiraffi (Laboratoire Vétérinaire d'Analyses Départemental de Mayotte, Kaweni, Mayotte), Catherine Cêtre-Sossah (UMR ASTRE,

CIRAD, Sainte-Clotilde, La Réunion) and Anlidine Mkadara, Hidachi Attoumani, Oussoufi Saindou, Kamardine Moussa and Adjibou Grimaldine (Chambre de l'Agriculture, de la Pêche et de l'Aquaculture de Mayotte, Coconi, Mayotte) for their technical assistance and all the breeders of the Mayotte cattle breed who participated to this study. Tom Druet is Senior Research Associate from the F.R.S.-FNRS.

Funding

This work was supported by the EU (FORWARD RITA DEFANIMAL project, 2015-2020) and INRAE, France (SELTRYP project, AIP-Bioressources, 2010 and PERSAFRICA project, Animal Genetics Division, 2013).

Conflicts of interest

None declared.

Literature cited

- Alexander DH, Novembre J, Lange K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.* 2009;19(9):1655–1664.
- Bahbahani H, Clifford H, Wragg D, Mbole-Kariuki MN, Tassell CV, Sonstegard T, Woolhouse M, Hanotte O. Signatures of positive selection in East African Shorthorn Zebu: a genome-wide single nucleotide polymorphism analysis. *Sci Rep.* 2015;5(1):11729.
- Bahbahani H, Tijjani A, Mukasa C, Wragg D, Almathen F, Nash O, Akpa GN, Mbole-Kariuki M, Malla S, Woolhouse M, et al. Signatures of selection for environmental adaptation and Zebu x Taurine hybrid fitness in East African Shorthorn Zebu. *Front Genet.* 2017;8:68.
- Beaujard P. The Indian Ocean in Eurasian and African world-systems before the sixteenth century. *J World Hist.* 2005;16(4):411.
- Beaujard P. East Africa, the Comoros Islands and Madagascar before the sixteenth century: on a neglected part of the world system. *Azania Archaeol Res Africa.* 2007;42(1):15–35.
- Beaujard P. The first migrants to Madagascar and their introduction of plants: linguistic and ethnological evidence. *Azania Archaeol Res Africa.* 2011;46(2):169–189.
- Beaujard P. East Africa and oceanic exchange networks between the first and the fifteenth centuries. *Afriques.* 2015;(06):WOS:000420334300001.
- Beaujard P. *The Worlds of the Indian Ocean: a Global History: Volume 1: from the Fourth Millennium BCE to the Sixth Century CE*, Cambridge: Cambridge University Press; 2019a.
- Beaujard P. *The Worlds of the Indian Ocean: A Global History: Volume 2: from the Seventh Century to the Fifteenth Century CE*, New York (NY): Cambridge University Press; 2019b.
- Bertrand AR, Kadri NK, Flori L, Gautier M, Druet T. Rzooroh: an R package to characterize individual genomic autozygosity and identify homozygous-by-descent segments. *Methods Ecol Evol.* 2019;10(6):860–866.
- Bock R, Kingston T, De Vos A. Effect of breed of cattle on transmission rate and innate resistance to infection with *Babesia bovis* and *Babesia bigemina* transmitted by *Boophilus microplus*. *Aust Vet J.* 1999;77(7):461–464.
- Boivin N, Crowther A, Helm R, Fuller DQ. East Africa and Madagascar in the Indian Ocean world. *J World Prehist.* 2013;26(3):213–281.
- Braccini L, Ciralo E, Campa CC, Perino A, Longo DL, Tibolla G, Pregnotato M, Cao Y, Tassone B, Damilano F, et al. PI3K-C2Y is a Rab5 effector selectively controlling endosomal Akt2 activation downstream of insulin signalling. *Nat Commun.* 2015;6(1):1–15.
- Brucato N, Fernandes V, Mazières S, Kusuma P, Cox MP, Ng'ang'a JW, Omar M, Simeone-Senelle M-C, Frassati C, Alshamali F, et al. The comoros show the earliest Austronesian gene flow into the Swahili corridor. *Am J Hum Genet.* 2018;102(1):58–68.
- Brucato N, Kusuma P, Beaujard P, Sudoyo H, Cox MP, Ricaut FX. Genomic admixture tracks pulses of economic activity over 2,000 years in the Indian Ocean trading network. *Sci Rep.* 2017;7(1):2919.
- Brucato N, Kusuma P, Cox MP, Pierron D, Purnomo GA, Adelaar A, Kivisild T, Letellier T, Sudoyo H, Ricaut FX. Malagasy genetic ancestry comes from an historical Malay trading post in Southeast Borneo. *Mol Biol Evol.* 2016;33(9):2396–2400.
- Buitenhuis B, Janss LL, Poulsen NA, Larsen LB, Larsen MK, Sørensen P. Genome-wide association and biological pathway analysis for milk-fat composition in Danish Holstein and Danish Jersey cattle. *BMC Genomics.* 2014;15(1):1112.
- Cesar AS, Regitano LC, Mourão GB, Tullio RR, Lanna DP, Nassu RT, Mudado MA, Oliveira PS, do Nascimento ML, Chaves AS, et al. Genome-wide association study for intramuscular fat deposition and composition in Nelore cattle. *BMC Genet.* 2014;15:39.
- Cheke A. The timing of arrival of humans and their commensal animals on Western Indian Ocean oceanic islands. *Phelsuma.* 2010;18:38–69.
- Choudhury A, Aron S, Sengupta D, Hazelhurst S, Ramsay M. African genetic diversity provides novel insights into evolutionary history and local adaptations. *Hum Mol Genet.* 2018;27(R2):R209–R218.
- Côte-Sossah C, Pédarrieu A, Guis H, Defernex C, Bouloy M, Favre J, Girard S, Cardinale E, Albina E. Prevalence of rift valley fever among ruminants, Mayotte. *Emerg Infect Dis.* 2012;18(6):972–975.
- Cui CY, Schlessinger D. Eccrine sweat gland development and sweat secretion. *Exp Dermatol.* 2015;24(9):644–650.
- Czamecki A, Dufy-Barbe L, Huet S, Odessa MF, Bresson-Bepoldin L. Potassium channel expression level is dependent on the proliferation state in the GH3 pituitary cell line. *Am J Physiol Cell Physiol.* 2003;284(4):C1054–C1064.
- De Camargo G, Aspilcueta-Borquis RR, Fortes M, Porto-Neto R, Cardoso DF, Santos D, Lehnert S, Reverter A, Moore S, Tonhati H. Prospecting major genes in dairy buffaloes. *BMC Genomics.* 2015;16(1):14.
- De Camargo GMF, Porto-Neto LR, Kelly MJ, Bunch RJ, McWilliam SM, Tonhati H, Lehnert SA, Fortes MR, Moore SS. Non-synonymous mutations mapped to chromosome X associated with andrological and growth traits in beef cattle. *BMC Genomics.* 2015;16(1):10.
- De Deken R, Martin V, Saido A, Madder M, Brandt J, Geysen D. An outbreak of East Coast Fever on the Comoros: a consequence of the import of immunised cattle from Tanzania? *Vet Parasitol.* 2007;143(3–4):245–253.
- ommergues L, Pannequin M, Cavalerie L, Cardinale E. *Etude épidémiologique sur le charbon symptomatique à Mayotte (2014)*. Technical report. Coopadem - GDS Mayotte; 2015.
- Druet T, Gautier M. A model-based approach to characterize individual inbreeding at both global and local genomic scales. *Mol Ecol.* 2017;26(20):5820–5841.
- Druet T, Gautier M. An improved hidden Markov model for the characterization of homozygous-by-descent segments in individual genomes. *bioRxiv.* 2021. <https://doi.org/10.1101/2021.05.25.445246>.
- Felius M. On the breeds of cattle: their history, classification and conservation [Ph.D. thesis]. University Utrecht, Utrecht; 2016.

- Flori L, Fritz S, Jaffrézic F, Boussaha M, Gut I, Heath S, Foulley JL, Gautier M. The genome response to artificial selection: a case study in dairy cattle. *PLoS One*. 2009;4(8):e6595.
- Flori L, Moazami-Goudarzi K, Alary V, Araba A, Boujenane I, Boushaba N, Casabianca F, Casu S, Ciampolini R, D'Acier AC, et al. A genomic map of climate adaptation in Mediterranean cattle breeds. *Mol Ecol*. 2019;28(5):1009–1029.
- Flori L, Thevenon S, Dayo GK, Senou M, Sylla S, Berthier D, Moazami-Goudarzi K, Gautier M. Adaptive admixture in the West African bovine hybrid zone: insight from the Borgou population. *Mol Ecol*. 2014;23(13):3241–3257. WOS:000338014900009.
- France M. *Synthese illustrée du recensement agricole 2010*. Technical report. Ministère de l'Agriculture, de l'Alimentation, de la Pêche, de la Ruralité et de l'Aménagement du Territoire. Mayotte. 2011.
- Fuller D, Boivin N, Hoogervorst T, Allaby R. Across the Indian Ocean: the prehistoric movement of plants and animals. *Antiquity*. 2011; 85(328):544–558.
- Fuller DQ, Boivin N. Crops, cattle and commensals across the Indian Ocean. *Etudes Ocean Indien*. 2009; 42/43:13–46.
- Gautier M, Faraut T, Moazami-Goudarzi K, Navratil V, Foglio M, Grohs C, Boland A, Garnier JG, Boichard D, Lathrop GM, et al. Genetic and haplotypic structure in 14 European and African cattle breeds. *Genetics*. 2007;177(2):1059–1070.
- Gautier M, Flori L, Riebler A, Jaffrézic F, Laloé D, Gut I, Moazami-Goudarzi K, Foulley JL. A whole genome Bayesian scan for adaptive genetic divergence in West African cattle. *BMC Genomics*. 2009;10:550.
- Gautier M, Klassmann A, Vitalis R. rehh 2.0: a reimplementation of the R package rehh to detect positive selection from haplotype structure. *Mol Ecol Resour*. 2017;17(1):78–90.
- Gautier M, Laloé D, Moazami-Goudarzi K. Insights into the genetic history of French cattle from dense SNP data on 47 worldwide breeds. *PLoS One*. 2010;5(9):e13038.
- Gautier M, Naves M. Footprints of selection in the ancestral admixture of a New World Creole cattle breed. *Mol Ecol*. 2011;20(15): 3128–3143.
- Gautier M, Vitalis R, Flori L, Estoup A. f-statistics estimation and admixture graph construction with Pool-Seq or allele count data using the R package poolfst. *Mol Ecol Resour*. 2021 Nov 27. doi: 10.1111/1755-0998.13557.
- Ghoreishifar SM, Eriksson S, Johansson AM, Khansefid M, Moghaddaszadeh-Ahrabi S, Parna N, Davoudi P, Javanmard A. Signatures of selection reveal candidate genes involved in economic traits and cold acclimation in five Swedish cattle breeds. *Genet Select Evol*. 2020;52(1):15.
- Glass EJ, Preston PM, Springbett A, Craigmile S, Kirvar E, Wilkie G, Brown CD. *Bos taurus* and *Bos indicus* (Sahiwal) calves respond differently to infection with *Theileria annulata* and produce markedly different levels of acute phase proteins. *Int J Parasitol*. 2005;35(3): 337–347.
- Hanotte O, Bradley DG, Ochieng JW, Verjee Y, Hill EW, Rege JEO. African pastoralism: genetic imprints of origins and migrations. *Science*. 2002;296(5566):336–339.
- Hansen P. Physiological and cellular adaptations of zebu cattle to thermal stress. *Anim Reprod Sci*. 2004;82–83:349–360.
- Hayes BJ, Lien S, Nilsen H, Olsen HG, Berg P, Maceachern S, Potter S, Meuwissen THE. The origin of selection signatures on bovine chromosome 6. *Anim Genet*. 2008;39(2):105–111.
- Hegardt FG. Mitochondrial 3-hydroxy-3-methylglutaryl-coa synthase: a control enzyme in ketogenesis. *Biochem J*. 1999;338(3):569–582.
- Hollmann AK, Bleyer M, Tipold A, Neßler JN, Wemheuer WE, Schütz E, Brenig B. A genome-wide association study reveals a locus for bilateral iridal hypopigmentation in Holstein Friesian cattle. *BMC Genet*. 2017;18(1):1–8.
- Hou GY, Yuan ZR, Zhou HL, Zhang LP, Li JY, Gao X, Wang DJ, Gao HJ, Xu SZ. Association of thyroglobulin gene variants with carcass and meat quality traits in beef cattle. *Mol Biol Rep*. 2011;38(7): 4705–4708.
- Jian W, Duangjinda M, Vajrabukka C, Katawatin S. Differences of skin morphology in *Bos indicus*, *Bos taurus*, and their crossbreds. *Int J Biometeorol*. 2014;58(6):1087–1094.
- Júnior GAF, de Oliveira HN, Carvalheiro R, Cardoso DF, Fonseca LFS, Ventura RV, de Albuquerque LG. Whole-genome sequencing provides new insights into genetic mechanisms of tropical adaptation in Nellore (*Bos primigenius indicus*). *Sci Rep*. 2020;10(1):7.
- Kadri NK, Harland C, Faux P, Cambisano N, Karim L, Coppieters W, Fritz S, Mullaart E, Baurain D, Boichard D, et al. Coding and non-coding variants in *HFM1*, *MLH3*, *MSH4*, *MSH5*, *RNF212*, and *RNF212B* affect recombination rate in cattle. *Genome Res*. 2016; 26(10):1323–1332.
- Keightley PD, Eyre-Walker A. Deleterious mutations and the evolution of sex. *Science*. 2000;290(5490):331–333.
- Kelleher J, Etheridge AM, McVean G. Efficient coalescent simulation and genealogical analysis for large sample sizes. *PLoS Comput Biol*. 2016;12(5):e1004842.
- Kim S, Cheong HS, Shin HD, Lee SS, Roh HJ, Jeon DY, Cho CY. Genetic diversity and divergence among Korean cattle breeds assessed using a BovineHD single-nucleotide polymorphism chip. *Asian-Australas J Anim Sci*. 2018;31(11):1691–1699.
- Klassmann A, Gautier M. Detecting selection using extended haplotype homozygosity (EHH)-based statistics in unphased or unpolarized data. *Authorea*. *PLoS ONE*. 2022;17(1):e0262024. <https://doi.org/10.1371/journal.pone.0262024>.
- Ko S, Zhao MG, Toyoda H, Qiu CS, Zhuo M. Altered behavioral responses to noxious stimuli and fear in Glutamate Receptor 5 (GluR5)- or GluR6-deficient mice. *J Neurosci*. 2005;25(4):977–984.
- Kortüm F, Caputo V, Bauer CK, Stella L, Cioffi A, Alawi M, Bocchinfuso G, Flex E, Paolacci S, Dentici ML, et al. Mutations in *KCNH1* and *ATP6V1B2* cause Zimmermann-Laband syndrome. *Nat Genet*. 2015;47(6):661–667.
- Lin TC. Functional roles of spink1 in cancers. *Int J Mol Sci*. 2021;22(8): 3814.
- Lipson M. Applying f-statistics and admixture graphs: theory and examples. *Mol Ecol Resour*. 2020;20(6):1658–1667.
- Littlejohn MD, Henty KM, Tiplady K, Johnson T, Harland C, Lopdell T, Sherlock RG, Li W, Lukefahr SD, Shanks BC, et al. Functionally reciprocal mutations of the prolactin signalling pathway define hairy and slick cattle. *Nat Commun*. 2014;5(1):8.
- Loh PR, Lipson M, Patterson N, Moorjani P, Pickrell JK, Reich D, Berger B. Inferring admixture histories of human populations using linkage disequilibrium. *Genetics*. 2013;193(4):1233–1254.
- Mahadevan P. Population and production characteristics of red Sindhi cattle in Ceylon. *J Dairy Sci*. 1955;38(11):1231–1241.
- Mattioli R, Bah M, Kora S, Cassama M, Clifford D. Susceptibility to different tick genera in Gambian N'Dama and Gobra zebu cattle exposed to naturally occurring tick infestations. *Trop Anim Health Prod*. 1995;27(2):95–105.
- Matukumalli LK, Lawley CT, Schnabel RD, Taylor JF, Allan MF, Heaton MP, O'Connell J, Moore SS, Smith TPL, Sonstegard TS, et al. Development and characterization of a high density SNP genotyping assay for cattle. *PLoS One*. 2009;4(4):e5350.
- Mbole-Kariuki MN, Sonstegard T, Orth A, Thumbi S, de C, Bronsvort B, Kiara H, Toye P, Conradie I, Jennings A, Coetzer K, et al. Genome-wide analysis reveals the ancient and recent admixture

- history of east African shorthorn zebu from western Kenya. *Heredity*. 2014;113(4):297–305.
- McPherson K. Cultural exchange in the Indian Ocean region. *Westerly*. 1984;29(4):5–16.
- Ministère de l'Agriculture dledpdM. Recensement de l'agriculture (RA) Campagne agricole 2004–2005: Tome 1 Généralités, méthodologies et principaux résultats. Technical report. Ministère de l'Agriculture, de l'élevage et de la pêche de Madagascar. 2007.
- Morimoto R, Shindou H, Tarui M, Shimizu T. Rapid production of platelet-activating factor is induced by protein kinase α -mediated phosphorylation of lysophosphatidylcholine acyltransferase 2 protein. *J Biol Chem*. 2014;289(22):15566–15576.
- Newitt M. The Comoro islands in Indian ocean trade before the 19th century les comores et le commerce dans l'océan indien avant le xixe siècle0. *CEA*. 1983;23(89):139–165.
- Ouvrard M, Magnier J, Raoul S, Anlidine M, Attoumani H, Oussoufi A, Kamardine M, Janelle J, Naves M, Flori L, et al. *Caractérisation phénotypique et génétique: le cas du zébu mahorais*. 2018. Actes de séminaire de clôture de la phase 1 des projets du RITA Mayotte (2015-2017), Bandrélé, Mayotte; 9–16.
- Patterson N, Price AL, Reich D. Population structure and eigenanalysis. *PLoS Genet*. 2006;2(12):e190.
- Patterson NJ, Moorjani P, Luo Y, Mallick S, Rohland N, Zhan Y, Genschoreck T, Webster T, Reich D. Ancient admixture in human history. *Genetics*. 2012;192(3):1065–1093.
- Pauly M. Acoua-Agnala M'kiri, Mayotte (976), Archéologie d'une localité médiévale (XIe-XVe siècles), entre Afrique et Madagascar. *Nyame Akuma, Bulletin of the Society of Africanist Archaeologists*; 2013;80:73–90.
- Pickrell JK, Patterson N, Loh PR, Lipson M, Berger B, Stoneking M, Pakendorf B, Reich D. Ancient west Eurasian ancestry in southern and eastern Africa. *Proc Natl Acad Sci USA*. 2014;111(7):2632–2637.
- Porter V. *Cattle: A Handbook to the Breeds of the World*. UK: Crowood Press; 2007.
- Porter V, Alderson L, Hall SJG, Sponenberg DP. *Mason's World Encyclopedia of Livestock Breeds and Breeding, 2 Volume Pack*. Wallingford, UK: CABI; 2016.
- Porto-Neto LR, Reverter A, Prayaga KC, Chan EK, Johnston DJ, Hawken RJ, Fordyce G, Garcia JF, Sonstegard TS, Bolormaa S, et al. The genetic architecture of climatic adaptation of tropical cattle. *PLoS One*. 2014;9(11):e113284.
- Purves D, Augustine GJ, Fitzpatrick D, Katz LC, Lamantia AS, McNamara JO, Williams SM. *Neuroscience*. 2nd ed. Sunderland, USA: Sinauer Associates; 2001.
- Reid MA, Allen AE, Liu S, Liberti MV, Liu P, Liu X, Dai Z, Gao X, Wang Q, Liu Y, et al. Serine synthesis through PHGDH coordinates nucleotide levels by maintaining central carbon metabolism. *Nat Commun*. 2018;9(1):11.
- Rosen BD, Bickhart DM, Schnabel RD, Koren S, Elsik CG, Tseng E, Rowan TN, Low WY, Zimin A, Couldrey C, et al. De novo assembly of the cattle reference genome with single-molecule sequencing. *GigaScience*. 2020;9(3):giaa021.
- Santiago E, Novo I, Pardiñas AF, Saura M, Wang J, Caballero A. Recent demographic history inferred by high-resolution analysis of linkage disequilibrium. *Mol Biol Evol*. 2020;37(12):3642–3653.
- Scheet P, Stephens M. A fast and flexible statistical model for large-scale population genotype data: applications to inferring missing genotypes and haplotypic phase. *Am J Hum Genet*. 2006;78(4):629–644.
- Sempéré G, Moazami-Goudarzi K, Eggen A, Laloë D, Gautier M, Flori L. WIDDE: a Web-Interfaced next generation database for genetic diversity exploration, with a first application in cattle. *BMC Genomics*. 2015;16(1):940.
- Serão NV, González-Peña D, Beever JE, Faulkner DB, Southey BR, Rodriguez-Zas SL. Single nucleotide polymorphisms and haplotypes associated with feed efficiency in beef cattle. *BMC Genetics*. 2013;14:1–20.
- Silanikove N, Shamay A, Shinder D, Moran A. Stress down regulates milk yield in cows by plasmin induced B-casein product that blocks K^+ channels on the apical membranes. *Life Sci*. 2000;67(18):2201–2212.
- Sun L, Lamont SJ, Cooksey AM, McCarthy F, Tudor CO, Vijay-Shanker K, DeRita RM, Rothschild M, Ashwell C, Persia ME, et al. Transcriptome response to heat stress in a chicken hepatocellular carcinoma cell line. *Cell Stress Chaperones*. 2015;20(6):939–950.
- Tang K, Thornton KR, Stoneking M. A new approach for using genome scans to detect recent positive selection in the human genome. *PLoS Biol*. 2007;5(7):e171.
- Tijjani A, Utsunomiya YT, Ezekwe AG, Nashiru O, Hanotte O. Genome sequence analysis reveals selection signatures in endangered trypanotolerant west African muturu cattle. *Front Genet*. 2019;10:442.
- Underwood E, Suttle N. *The Mineral Nutrition of Livestock*. 3rd edition. Wallingford, UK: CABI; 1999.
- Vilà-Brau A, De Sousa-Coelho AL, Mayordomo C, Haro D, Marrero PF. Human hmgcs2 regulates mitochondrial fatty acid oxidation and fgf21 expression in hepg2 cell line. *J Biol Chem*. 2011;286(23):20423–20430.
- Voight BF, Kudaravalli S, Wen X, Pritchard JK. A map of recent positive selection in the human genome. *PLoS Biol*. 2006;4(3):e72.
- Wang H, Chai Z, Hu D, Ji Q, Xin J, Zhang C, Zhong J. A global analysis of CNVs in diverse yak populations using whole-genome resequencing. *BMC Genomics*. 2019;20(1):12.
- Wang M, Kong L. pblat: a multithread blat algorithm speeding up aligning sequences to genomes. *BMC Bioinformatics*. 2019;20(1):28.
- Wickham H. *ggplot2: Elegant Graphics for Data Analysis*. New York (NY): Springer-Verlag; 2016.
- Xu L, Bickhart DM, Cole JB, Schroeder SG, Song J, Tassell CPV, Sonstegard TS, Liu GE. Genomic signatures reveal new evidences for selection of important traits in domestic cattle. *Mol Biol Evol*. 2015;32(3):711–725.
- Zafindrajaona PS. *Profils genetiques du zébu malgache* [thesis]. Université Paris XI; 1991.
- Zafindrajaona PS, Lauvergne J. Comparaison de populations de zébu malgache à l'aide des distances génétiques. *Genet Sel Evol*. 1993;25(4):373–395.

Communicating editor: M. Jakobsson