Investigations on Both Chemical Composition and Insecticidal Activities of Essential Oils of *Vepris heterophylla* (Rutaceae) from Two Localities of Northern Cameroon Towards *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae)

¹T.S.L. Ngamo, ²W.F. Noudjou, ¹M.B. Ngassoum, ¹P.M. Mapongmestsem,
¹A.B. Aminatou Boubakary, ²F. Malaisse, ²E. Haubruge, ²G. Lognay and ³T. Hance
¹University of Ngaoundéré Bp 454, Cameroon
²Faculty of Agronomy and Agricultural Sciences, Passage Des Déportés
2, B-5030 Gembloux, Belgium
³Laboratory of Ecology and Biogeography, Research Center on the Biodiversty,
4-5 Place Croix du Sud, 1348, Louvain-la-Neuve, Belgium

Abstract: Leaves of *Vepris heterophylla* (Rutaceae) are used traditionally as medicine and in crop protection for the reduction of post harvest losses due to insect pests. The present research aims to investigate the chemical composition, the toxicity and the repellent activities of the essential oil leaves of *V. heterophylla* collected in 2 localities (Mokolo and Meri). The research pointed out that, the 2 essential oils differ in their chemical composition. Sabinene and E-β-ocimene are the most common important compounds. Sabinene is particular to the essential oil from Meri (19.1%). E-caryophyllene, ą-humulene, elemol and germacrene, are the comon lowest compounds. E-carryophyllene (3.1%) and safrole (3%) are active compounds more present in the essential oil from Mokolo. Evaluation of the insecticidal properties of these 2 oils pointed out that the volume of essential oil killing half of the insect population tested (VL50) is not the same for the 2 oils. That from Mokolo is 49.44 μL from Meri is the highest, 61.2 μL. This useful plant is also cited as endangered, it is therefore an emergency to search strategy of exploitation that preserves this important component the local biodiversity.

Key words: Vepris heterophylla, tribolium castaneum, essential oil, Cameroon, toxicity, repellence

INTRODUCTION

In tropical countries, plants have been used and are still used to prevent post harvest losses. Most of these botanicals are used as insecticides or repellent agent (Boeke, 2002). However, the liberalisation of the insecticide market increases the availability of synthetic products for untrained farmers. One of the most important consequences of this is the loss of traditional knowledge on the use of botanicals in crop protection (Szafranski, 1991).

Important damages occur in untreated crops and the farmers are obliged to sell them at low price if possible. Stored products are treated many times during the conservation till the consumption or the selling. Many pests of stored products are Coleopterans and the most damaging species are from *Sitophilus* and *Tribolium* genus (Dal Bello *et al.*, 2001) for cereals. In developing countries as in Cameroon, many synthetic insecticides are used to prevent post harvest losses (Ngamo, 2004). In

most of the researches, they seem not to be used properly, consumers are therefore exposed to intoxication by their residues.

Moreover, synthetic insecticides are subject to dynamic trends. Target insects are able to develop resistance against single insect pesticide (Schmutterer and Ascher, 1986; Boeke, 2002). This can also enhance the development of non target pests, allowing a secondary pest in the past to be an important one with high tolerance towards popular pesticides. *T. castaneum* occurs in all infested grain during storage and is one of the most resistant pest to treatment.

The use of botanicals in crop protection may be a solution to prevent the environment and the consumer from the effects of the synthetic pesticides (Tapondjou et al., 2002; Regault-Roger and Hamraoui, 1995). One of the most used plants to protect grains in northern Cameroon is Vepris heterophylla (Engl.) Letouzey. This plant is not regularly distributed on the area, it occurs in patches from place to place. Its habitat

is at the sides of mountains, where it is not as present in sub Saharan Africa as observed in the past time (Geerling, 1982), this plant is signaled today in the UICN redlist as endangered plant with the reference EN A1c, B1+2c.

Botanicals produce secondary compounds which are concentrated in their essential oils. Their composition depends on many factors like the phenology of the plant and the ecological conditions. These factors influence the quantity and the quality of the essential oil and could explain the difference observed in activities of essential oils from different localities.

The present research combines investigations on the chemical composition of essential oils from 2 localities in northern Cameroon against *T castaneum*, in order to come to a user friendly method of protection of crops from the attacks of resistant pest.

MATERIALS AND METHODS

Production of essential olis

Sampling sites: Meri and Mokolo are located in the dry savana, where only 900 to 400 mm of rain are observed during the year. The dry season can be in seven months period. The mean temperature for the year is $28\pm7^{\circ}$ C. In each of the towns Meri, Mokolo, a patch of V. heterophylla was located and the leaves cut. Meri and Mokolo are in the Far North province, in the dry savana. The dry season is observed only during five months. In the Far North, peasant are mostly farmers cultivating sorghum, maize cowpea and others staple crops. V. heterophylla is known as a very important post harvest botanical insecticide.

Extraction of essential oils: The leaves of the plants of patch were collected, taken to the laboratory and dried without sun light at ambient temperature during 4 days. The essential oils were extracted by hydro distillation during 4 h using a Clevenger type apparatus. The oil obtained were collected and kept in brown bottles at 4 °C until use for bio assays or for analysis.

Chemical analysis of the essential oils GC/MS and GC/FID: The GC/MS was done with HP-5MS column (5% phenyl methyl siloxane) with 30m length and 250 µm in diameter. The carrier gas was helium, the temperature program applied was from 40 to 230°C with a speed of 5°C min⁻¹ with a stay at 280°C during 5 min. The pressure of the carrier gas was 49.9 KPa and the flux at 74.1 mL min⁻¹. The ion-source-eating at 230°C, the ion scan range was 50-350 amu. Mass spectra correlations for the identification of the compounds were done using Wiley 275 L, (Joulain and König, 1998; Adams Robert, 2001). After this qualitative analysis, the quantitative analysis was made using the GC/FID procedure in similar conditions.

Bio assays: A strain of *T. castaneum* associated with infected grains collected from farmer's granaries and reared in an incubator at 30 °C since 1999 was used for the tests. For the assays, only one-month aged adults were used.

Toxicity: Insecticidal activity of any oil was accessed through a contact and inhalation test. For each essential oil, precise volumes ranking from 0.5 to 3.2 mL were tested. The oil was diluted in 10 mL of acetone. For each trial, 0.5 mL of the preparation was deposited on a 9 cm diameter filter disk (Whatman n°1) put in a petri disk of 80 mL, 5 min after application, evaporation of the acetone was completed. Once acetone was evaporated, 20 T. castaneum were introduced in the petri disk, each one covered and sealed with parafilm. For each preparation, 5 replications were made. The control was the same volume of acetone without addition of essential oil. The mortality of insects was recorded 24 h after the treatment. Dose-response curves were analysed by the log-probit method. The mortalities observed were compared to that of control with an ANOVA test.

Repellence: The repellent activity of the essential oils was tested by using a linear olfactometer made of 30 cm glass tube having 2 cm diameter with a hole at its middle. At each end a small container was place with 2 g of flour of *Sorghum bicolour*. One contained a piece of filter paper with essential oil dilute in acetone and the control in the other hand had only acetone.

The hole in the middle was used to introduce insects there after it was covered with gauze. The choice of insects was observed for a period of one hour. Only the insect within the flour in either end were considered to have made a choice. Insect used was one month old. A group of 10 insects was used for each trial five replications were made.

The attractiveness via the Percentage of Repellence (PR) was evaluated according to a formula from Talukder and Howse (1993, 1995); Liu and Ho (1999).

 $PR = 2 \times (C-50)$ (where C = Percentage of insects choosing the untreated end or the control). If <math>PR > 0 the oil is repellent and if PR < 0 the oil is attractive.

RESULTS AND DISCUSSION

Yield and chemical composition of the essential oils:

The yield of essential oil obtained by hydro distillation was not the same for all the dried leaves of *V. heterophylla* from the 2 regions sampled. The production of essential oil is important with *V. heterophylla* leaves from Mokolo (1.12%) (Table 1) Those of Meri produced 0.65% of essential oil.

Table 1: Production of essential oil in relationship with the origin of the

| Sampling site | Amount of samples | Yield | SD |
|---------------|-------------------|--------|------|
| Meri | 4 | 0.65 b | 0.19 |
| Mokolo | 3 | 1.12 a | 0.61 |

In the same column, the values followed by the same letter do not differed significantly (p = 0.001)

These values indicate the low content of the 3 strains of *V. heterophylla* in essential oils. Even with this low yields observed, the range of the production of the essential oil is not the same for all these plants.

According to Fields *et al.* (2001), ecological factors like: water content in the soil, the quality and quantity of nutriments available, the luminosity, the temperature and relative humidity can influence the metabolism of the synthesis of essential oils. For Fields *et al.* (2001), Geographical factors is a significant parameter with respect to the essential production of oil and its composition (Azevedo *et al.*, 2002).

The samples studied contain merely equal composition in monoterpenes and sesquiterpenes. That from Mokolo contains 53.42% monoterpenes, the most important compounds being elemol (14.41%) and E-Bocimene (13.92%). That of Meri contains 52.90% of sesquiterpenes. Elemol (23.99%) and sabinene (19.11%) being most important compounds.

The essential oil of V. heterophylla from northern Cameroon contains mainly: Sabinene, E-ß ocimene, E-caryophyllene, α-humulene, germacrene and E-nerolidol (Table 2). The proportion of these compounds varies from a sample to another. Several chemotytes of Hyptis suaveolens and showed that differences among sesquiterpenes (Azevedo et al., 2002) this come from the fact that the plants were from different latitude and altitude. Other differences were explained by the growing conditions of the plants as light quality and intensity, availability in water and nutrients (Schoonhoven et al., 1998). These conditions may act on the accumulation of the secondary metabolites as it may be the case between plants from Meri and Mokolo. They seem to form 2 ecotypes of the plant in the soudano sahelian area of the Far North Cameroon.

Toxicity of the essential oils: The mortality of adult T. castaneum due to exposition to essential oil of V. heterophylla from 2 localities of northern Cameroon is shown the Table 3. The relationship between the volume of each essential oil and the insect mortality associated showed that, the volume of oil killing half of the insect population tested (VL50) for essential oil from Mokolo is $49.44~\mu L$ and that from Meri is the highest $61.2~\mu L$. The required volume of essential oil to kill all T. castaneum also changed in relationship with the origin of plants. A

Table 2: Chemical composition of essential oils of Vepris heterophylla from northern Cameroon

| northern Cameroon | | | % in essential oil of each locality | |
|---------------------------------------|--------------|---------------|-------------------------------------|--|
| Compounds | IK | Mokolo | Meri | |
| A-thujene | 930 | 0.4 | 0.4 | |
| A-pinene | 938 | 0.1 | 0.1 | |
| Sabinene | 977 | 14.0 | 19.1 | |
| β-pinene | 982 | 0.3 | 0.3 | |
| Myrcene | 992 | 4.2 | 2.8 | |
| A-phellandrene | 1009 | 0.1 | tr | |
| A-terpinene | 1021 | 0.8 | 0.5 | |
| Cymene (p/o) | 1028 | 0.5 | 0.2 | |
| Limonene | 1033 | 4.3 | 4.1 | |
| Z-β-ocimene | 1039 | 0.7 | 0.5 | |
| E-β-ocimene | 1049 | 14.0 | 9.2 | |
| γ-terpinene | 1063 | 1.3 | 0.9 | |
| Terpinolene | 1093 | 1.6 | 1.3 | |
| Allo-ocimene | 1131 | 0.3 | 0.3 | |
| Pregeijerene | 1288 | 0.2 | 0.2 | |
| Hydrocarbonated monoterpe | 1072 | 42.8 0.11 | 40.1 0.3 | |
| Cis-sabinene hydrate Linalool | 1100 | 0.11 | 0.5 | |
| Cis-p-menth-2-en-1-ol | 1100 | 0.9 | 0.0 | |
| Trans-p-menth-2-en-1-ol | 1145 | tr | tr | |
| Terpinen-4-ol | 1143 | 3.3 | 2.1 | |
| A-terpineol | 1196 | 1.7 | 1.2 | |
| Methyl salicylate | 1201 | 0.2 | 1.2 | |
| Safrole | 1296 | 3.0 | 1.7 | |
| Neryl acetate | 1364 | tr | tr | |
| Geranyl acetate | 1395 | tr | u u | |
| Methyl eugenol | 1406 | 0.3 | | |
| Methyl-n-methyl anthranilate | 1.00 | 1407 | tr | |
| Oxygenated monoterpenes | 9.7 | 6.2 | | |
| Monoterpenes NI | | tr | 0.5 | |
| Bicycloelemene | 1348 | tr | tr | |
| A-cubebene | 1360 | tr | tr | |
| A-ylangene | 1383 | 0.4 | 0.1 | |
| A-copaene | 1389 | 0.3 | 0.1 | |
| β-bourbonene | 1400 | tr | tr | |
| β-elemene | 1402 | 0.8 | 0.6 | |
| E-caryophyllene | 1438 | 3.1 | 1.7 | |
| β-copalene | 1445 | 0.3 | 0.3 | |
| Alloaromadendrene | 1451 | | 0.1 | |
| γ-elemene | 1457 | tr | | |
| A-humulene | 1473 | 1.9 | 1.2 | |
| Acora-3(10).14-diene | 1479 | 0.1 | tr | |
| γ-muurolene | 1490 | 0.2 | tr | |
| Germacrene D | 1499 | 3.4 | 2.3 | |
| Ledene | 1504 | 0.1 | tr | |
| γ-amorphene | 1512 | 0.4 | 0.4 | |
| Δ-amorphene | 1517 | 0.1 | tr | |
| γ-cadinene | 1521 | 0.1 | tr | |
| ∆-cadinene | 1536 | 0.6 | 0.5 | |
| β-cadinene | 1540 | 0.2 | 0.4 | |
| Germacrene D-4-ol | 1593 | 10.2 | tr | |
| Hydrocarbonated sesquiterp Cubebol | | 12.3 | 8.1 | |
| Elemol | 1531 1562 | 0.16 14.37 | 0.25 23.99 | |
| Caryophillene oxide | 1604 | | | |
| Guaiol Caryopninene oxide | 1613 | 0.05 12.85 | 0.12 13.47 | |
| Humulene epoxide II | 1630 | 0.11 | 0.21 | |
| 10-epi-y-eudesmol | 1641 | 0.11 | 0.21 | |
| y-eudesmol | 1650 | 1.26 | 0.61 | |
| a-muurolol | 1658 | 0.08 | 0.01 | |
| β-eudesmol | 1671 | 0.73 | 0.08 | |
| A-eudesmol + valerianol | 1673 | 1.78 | 1.63 | |
| Bulnesol | 1684 | 1.78 | 1.59 | |
| Oxygenated sesquiterpenes | 33.20 | 42.72 | 1.39 | |
| Sesquiterpenes NI | 55.20 | 1.95 | 2.06 | |
| Transporter 141 | 1 | 1.70 | 2.00 | |

KI = Kovalt index; NI = non identified

Table 3: Insecticidal effect of the essential oils of *Vepris heterophylla* from 2 localities of the northern Cameroon on *Tribolium castaneum*

| | Mortality (%) of adult T.castaneum | | |
|-----------------------------------|------------------------------------|-----------------------|--|
| Volume of the essential oil (μL) | Mokolo | Meri | |
| 24 | 1±0 | 3.8±0.4 | |
| 32 | 7.6±1.7 | 3±0 | |
| 40 | 22.4±6.11 | 8.6±2.5 | |
| 48 | 49±6.96 | 7.8±3.2 | |
| 56 | 54.6±4.04 | 14.8±5.81 | |
| 64 | 52.4±8.11 | 30.8±9.47 | |
| 72 | 86±6.4 | 41.21±5.7 | |
| 80 | 93±3.2 | 42.8±13.2 | |
| 88 | 100±0 | 50.6±17.04 | |
| 96 | 100±0104 | 67±13 | |
| 112 | 100±0 | 80±8.54 | |
| 120 | 100±0 | 82.6±9.6 | |
| VL50 | 49.44 uL⁵ | 61.21 µL ^b | |

In the same line the values followed by the same letter does not differed significantly (p = 0.001)

Table 4: Repellence of essential oil in relationship with the origin of the plant material toward *Tribolium castaneum*

| Origin of the oil | PR (%) | Observation |
|-------------------|--------|-------------|
| Meri | - 32 | Attractive |
| Mokolo | - 32 | Attractive |

maximum of 88 μ L could kill 100% of the insect tested with oil from leaves collected at Mokolo. But with that of Meri, to kill all these insects, more than 120 μ L of essential oil is required.

In general, results obtained by contact and inhalation test confirm the toxicity of V. heterophylla towards T. castaneum. The 2 oils tested contain toxic compound like limolene which is highly toxic to T. castaneum (Prates et al., 1998). A variation of efficiency between chemotypes of V. heterophylla would be due to difference in the amount of compound present such as sabinene which is most present amongst the plants collected at Meri (19.1%) than in that collected at Mokolo (14.0%). Sabinene is toxic to insects (Park et al., 2003). Other differences are observed with compounds such as α -pinene, myrcene, α -terpinene toxic to insects especially Callosobruchus maculatus (Park et al., 2003) occur most in the oil from Mokolo which is the most toxic to the pest.

Repellent properties of the essential oils: The essential oil of *V. heterophylla* expressed attractiveness or repellence depending on the origin of the leaves used for the extraction. Those from Meri and Mokolo are repellent to *T. castaneum*. The percentage of the attractive is 32% (Table 4). The essential oil from Ngaoundéré is repellent to *T. castaneum* at 20%.

The essential oils are both attractive and toxic to *T. castaneum*. This observation is the same as that made with *Hyptis suaveolens* which is very attractive to *Callosobruchus maculatus* (Boeke, 2002).

Leaves of *V. heterophylla* are used in traditional medicine against rheumatism, malaria and also as anthelminthic, parasiticide or purgative. They are

promising for their use in protection of grains during storage from the attack of post harvest insect pests. This useful plant is also endangered, it is therefore an emergency to search strategy of exploitation that preserve this important component the local biodiversity.

ACKNOWLEDGEMENTS

The authors gratefully acknowledge the Belgian CUD (Coopyration Universitaire au Développement) for a financial support through the convention Storeprotect PIC 2003.

REFRENCES

Adams Robert, P., 2001. Identification of essential oil components by gas chromatography/quadrupole mass spectroscopy. Allured Publishing Corporation, Carol Stream II.

Azevedo, N.R., I.F.P. Campos, H.D. Ferreira, T.A. Portes, J.C. Seraphin, J. Realino de Paula, S.C. Santos and P.H. Ferri, 2002. Essential oil chemotypes in *Hyptis* suaveolens from brazilian cerrado. Biochem. Sys. Ecol., 30: 205-216.

Boeke, S.J., 2002. Traditional African plant products to protect stored cowpeas against insect damage; the battle against the beetle. Ph D thesis, Wageningen Univ., pp. 151.

Dal Belo, G., S. Pandin, L. Lopez and M. Fabrizio, 2001. Laboratory evaluation of chemical biological control of rice weevil (*Sitophilus oryzae* L.) in stored grains. J. Stored Prod. Res., 37: 77-84.

Fields, P.G., Y.S. Xie and X. Hou, 2001. Repellent effect of pea (*Pisum sativum*) fractions against stored product insects. J. Stored Prod. Res., pp. 359-370.

Geerling, C., 1982. Guide de Terrain des Ligneux Sahyliens et Soudano Guinyens. H. Veennan and Zonen BV (Eds.) Wageningen, NL.

Joulain, D. and W.A. König, 1998. The Atlas of Spectral Data of Sesquiterpene Hydrocarbons. Hamburg, EB-Verl., Germany.

Liu, Z.L. and S.H. et Ho, 1999. Bioactivity of essential oil extracted from *Evodia rutaecarpa* Hook f. et Thomas against the grain storage insects, Sitophilus zeamais Motsch and *Tribolium castaneum* (Herbst).

Ngamo, T.L.S., 2004. A la recherche d'une alternative aux Polluants Organiques Persistants utilisys pour la protection des végétaux. Bulletin d'Informations Phytosanitaires. N° 43 Avril-Juin, 2004. pp. 23.

Park, I.K., S.G. Lee, D.W. Choi, J.D. Park and Y.J. Ahn, 2003. Insecticidal activities of constituents identified in the essential oil from leaves of *Chamaecyparis* obtusa against *Callosobruchus chinensis* (L.) and Sitophilus oryzae (L.). J. Stored Prod. Res., 39: 375-384.

- Prates, H.T., J.P. Santos, J.M. Waquil, J.D. Fabris, A. Oliveira and J.E. Forster, Embrapa, 1998. Insecticidal activity of monoterpenes against *Rhyzoperhta domonica* (F.) and *Tribolium castaneum* (Herbst). J. Stored Prod. Res., 34: 243-249.
- Regnault-Roger, C. and A. Hamaraoui, 1995. Fumigant toxic activity and reproductive inhibitory induced by monoterpenes on *Acathoscelides obtectus* (Say) (Coleoptera) bruchid of kidney bean (*Phaseolus vulgaris* L.). J. Stored Prod. Res., 31: 291-299.
- Schmutterer, H. and K.R.S. Ascher, 1986. Natural pesticides from the neem tree (*Azadirachta indica* A Juss) and other tropical plants. Proc 3rd Neem Conference Nairobi, pp. 517-523.
- Schoonhoven, L.M., T. Jermy and J.J.A. Van Loon, 1998. Insect-plant biology: from physiology to evolution. London, Chapman and Hall, pp. 490.

- Szafranski, F., 1991. Activité biologique des extraits de quelques plantes des environs de Kinsngani (Zaïre). Belgian J. Bot., 124: 60-70.
- Talukder, F.A. and P.E. Howse, 1993. Deterrent and insecticidal effects of extracts pithraj, Aphanamixis polystachya (Meliaceae), against Tribolium castaneum in storage. J. Chemi. Ecol., 19: 2463-2471.
- Talukder, F.A. and P.E. Howse, 1995. Evaluation of *Aphanamixis polystachya* as a source of repellents, antifeedants, toxicants and protectants in storage against *Tribolium castaneum* (Herbst). J. Stored Prod. Res., 34: 55-61.
- Taponjou, L.A., C. Adler, H. Bouda and D.A. Fontem, 2002. Efficacy of powder and essential oif from *Chenopodium ambrosioides* leaves as post-harvest grain protectants against six-stored product beetles. J. Stored Prod. Res., 38: 395-402.