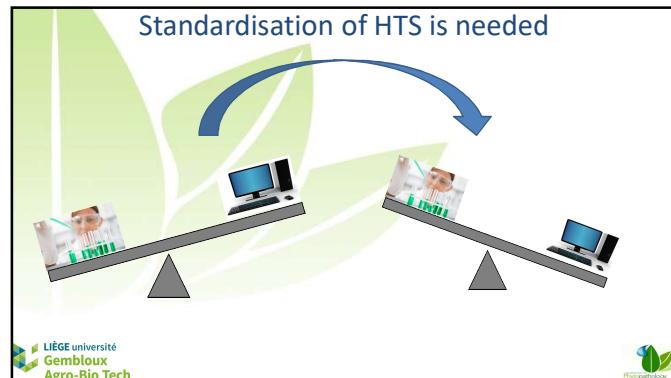


Untangling bioinformatics bias for the diagnostic of plant viruses by HTS:

Lessons from an international performance testing of sequence analysis strategies



Objective: evaluation of bio-informatic strategies

Evaluate and compare the performance of bioinformatic pipelines for the analysis of high-throughput siRNA sequencing data

The image is a collage of two distinct scenes. The upper scene is a photograph of a crowded event, likely a scientific conference or party, with many people raising their hands. The background is a large, circular screen displaying a complex, abstract pattern of blue dots arranged in concentric rings, resembling a microscopic view or a data visualization. The lower scene is a stylized illustration of a desert landscape. It features several prominent rock formations, including mesas and buttes, silhouetted against a clear blue sky. A winding, light-colored path or riverbed cuts through the foreground. The ground is depicted in shades of orange and brown, representing sand and rock.

Starting datasets: FASTQ files from 3 samples

Species	Potato	Apple	Grapevine
Sequencing technology	Illumina		
Sequencing depth	23M	13M	12M
Plant	Potato	Apple	Grapevine rootstock Kober 125 sa, Vitis berlandieri x Vitis riparia
Viruses or viroids detected (one column per virus)	Rf-PCR	New Viraloids AGV (2 variants)	Grapevine rootstock Kober 125 sa, Vitis berlandieri x Vitis riparia
Percentage of reads for each virus/viroid (*)	~23%	~5%	0.12%
Conformation(s) technique(s) for each virus/viroid	Rf-PCR &anger sequencing RT-PCR sequencing RT-PCR sequencing	Rf-PCR &anger sequencing RT-PCR sequencing RT-PCR sequencing	Rf-PCR RT-PCR RT-PCR RT-PCR RT-PCR RT-PCR RT-PCR RT-PCR RT-PCR RT-PCR

Difficulties:

1. New unknown virus
2. Complex mix of 9 viruses/viroids

Methodology for evaluation of bioinformatic pipelines

➤ sRNA datasets (length between 21 and 24 nt)

➤ Worst case scenario by rarefaction at 3 sequencing depths:

- ❖ 50,000 reads – 3 files
- ❖ 250,000 reads – 4 files (2 grapevine replicates)
- ❖ 2,500,000 reads – 3 files

➤ 10 fastq files available on a server in double blind

➤ 21 participants

Starting datasets: FASTQ files from 3 samples

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Difficulties:

1. New unknown virus
2. Complex mix of 9 viruses/viroids
3. Very low abundance virus/viroid

Methodology for evaluation of bioinformatic pipelines

Participants free to apply their own bioinformatics strategy to identify the viruses in the 10 datasets (de novo assembly + annotation)

Interpret the data in a diagnostics setting

Starting datasets: FASTQ files from 3 samples

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Difficulties:

1. New unknown virus
2. Complex mix of 9 viruses/viroids
3. Very low abundance virus/viroid
4. Close virus species with poor genome knowledge

Results: analytical sensitivity

LabID	Sensitivity			Average
	50,000	250,000	2,500,000	
A	10%	53%	90%	51%
B	30%	35%	80%	46%
C	60%	71%	80%	70%
D	50%	62%	100%	76%
E	30%	82%	80%	68%
F	80%	88%	100%	89%
G	20%	53%	100%	57%
H	30%	65%	70%	57%
J	70%	94%	100%	89%
K	40%	71%	90%	68%
M	50%	94%	90%	81%
N	30%	82%	90%	70%
P	20%	41%	40%	35%
R	100%	100%	100%	100%
S	50%	100%	100%	86%
T	90%	100%	100%	97%
V	60%	88%	80%	78%
W1	40%	82%	90%	73%
W2	60%	82%	90%	78%
X	30%	71%	80%	62%
AVERAGE	46%	75%	86%	70%

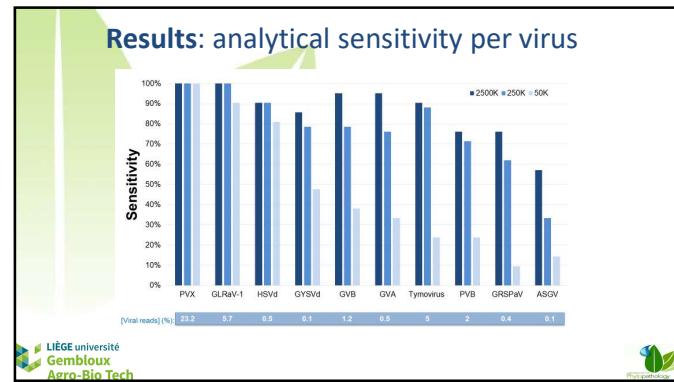
Results: analytical sensitivity

- 70 % sensitivity overall
- Sensitivity from 35 to 100%
- Sensitivity increase with sequencing depth
- Seven strategies have 100% sensitivity at 2.5 M
- Participant R: 100% sensitivity (T close)

-> Both have an additional read mapping step

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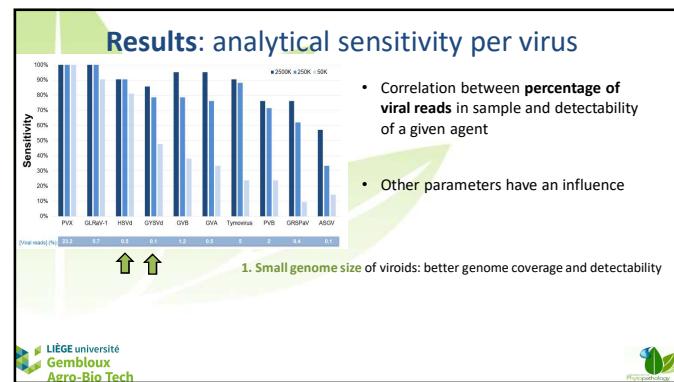


Results: analytical sensitivity

- Sensitivity improved by lower k-mer lengths (13-15), decreased by higher length (17-21)
- > impact of k-mer lenght
- Sensitivity reduced by setting minimal contig length too high, a value <60nt should be preferred
- > Shorter minimal contig lenght prefered

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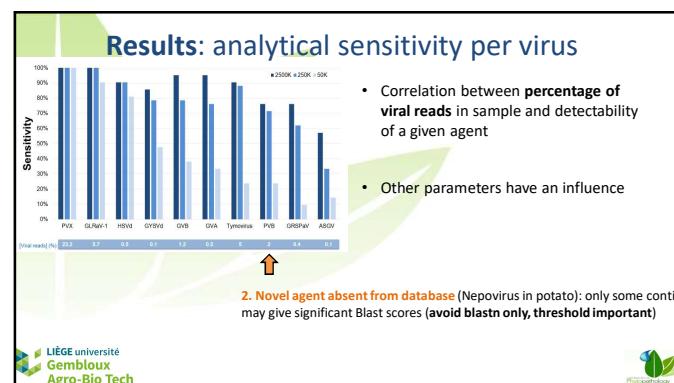


Results: analytical sensitivity

- Sensitivity improved by contig extension after de novo analysis
- > Contig extension to pass minimal contig lenght threshold

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Results: analytical sensitivity per virus

Virus	Total reads (%)	Sensitivity (%)
PVX	23.2	~95%
GLRV-Y-1	5.7	~90%
HSV	0.5	~85%
GYVW	0.1	~80%
GVB	1.0	~75%
GVA	0.5	~70%
Turnip	5	~65%
PVb	2	~60%
GRSPV	3.4	~55%
ASGV	6.1	~45%

- Correlation between percentage of viral reads in sample and detectability of a given agent
- Other parameters have an influence

3. Minimal contig length must be low to avoid elimination of small viral contigs from low abundance viruses (ASGV for apple)

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Conclusions: bioinformatics matters !

- ✓ Some rather trivial things to avoid
 - ✓ BlastN only
 - ✓ Stringent Blast cut-off values
 - ✓ MegaBlast (high homology threshold)

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Results: reproducibility

- Comparing the results reported for the two 250K grapevine pseudo-replicates

Strategy	N of viruses detected only in one subsample	N of viruses detected in both subsamples
R	7	0
T	6	0
F	5	0
J	7	0
S	6	0
M	5	0
D	6	0
V	7	0
W	6	0
I	5	0
C	7	0
N	6	0
E	5	0
K	7	0
X	6	0
G	4	0
H	5	0
A	3	0
P	4	0
B	2	0
O	0	0

Key observations:

- Overall: 91.6% reproducibility
- 15 strategies: 100% (including FN)
- 5 strategies: 100% reproducibility and analytical sensitivity
- Correlation between sensitivity & reproducibility

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Conclusions: bioinformatics matters !

- ✓ Some rather trivial things to avoid
 - ✓ BlastN only
 - ✓ High Blast cut-off values
 - ✓ MegaBlast (high homology threshold)
- ✓ Some less obvious lessons
 - ✓ Minimal contig length must be very low (<60nt) for optimal sensitivity with low abundance viruses
 - ✓ For Velvet, broad range of k-mer values including higher ones improve sensitivity
 - ✓ Additional contigs extension and combination of mapping and de novo assembly might improve detection

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Conclusions: bioinformatics matters !

- ✓ A huge diversity in pipelines and pipeline performance
- ✓ Identification of trivial errors to avoid and other lessons

➤ siRNA indexing data annotation is not such a straightforward business

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For more details: see our publication in Phytopathology

Phytopathology • 2019 • 109:488-497 • <https://doi.org/10.1093/PHYTO/02-18-0967-R>

Virology e-Xtra*

Virus Detection by High-Throughput Sequencing of Small RNAs: Large-Scale Performance Testing of Sequence Analysis Strategies

Sebastien Massart, Mélanie Chambon, Kris De Jonghe, Rachel Glover, Amélie Hagenman, Igor Kohanski, Pier Koninkx, Jan Krausz, Denis Kunatik, Léonidas Lotis, François Maes, Virginie Mattinga, Hans J. Moree, Thibaut Olivier, Antonio Olmos, Mikhail M. Pooggin, Jean-Sébastien Reynard, Ana B. Ruiz-Garcia, Dana Safanova, Pierre H. H. Schneegescher, Noa Sela, Silvia Turco, Eva J. Vainio, Eva Varallyay, Eric Verlin, Marcel Westenberg, Yves Brustinx, and Thierry Caudresse

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One step further: applying HTS in plant pest diagnostics

Valitest
www.valitest.eu

- ✓ Climbing the diagnostic cliff of HTS technologies ?
- ✓ Writing of international guidelines for the use of HTS technologies for plant pest diagnostics
- ✓ 54 scientists & >30 countries
- ✓ For viruses, bacteria, fungi, nematodes and insects
- ✓ 70 pages soon available as scientific paper

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Thanks for the invitation

Thanks for the invitation

Thank you for your attention ✓

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The diagram illustrates the integration of HTS into plant pest diagnostics across various laboratory facilities. It features a central red box labeled "HTS" connected by arrows to six surrounding green boxes representing different management and support functions:

- Laboratory Facilities**
- IT Infrastructure**
- Quality Management**
- Human resources**
- Outsourcing**
- Data management**
- Change management**

Each green box contains a smaller diagram showing a green leaf shape with a blue circular area representing the integration of HTS into specific laboratory processes.

```

graph TD
    A[One step further: applying HTS in plant pest diagnostics] --> B[Risk analysis]
    B --> C[Result interpretation]
    C --> D[Test selection]
    D --> E[Controls in routine]
    E --> F[Validation and verification]
    F --> G[Reference material]
    
```

The diagram illustrates the sequential steps in developing an HTS test for plant pest diagnostics:

- Risk analysis**
- Result interpretation**
- Test selection**
- Controls in routine**
- Validation and verification**
- Reference material**

Valitest
www.valitest.eu

2. HTS test for diagnostics