

Starting datasets: FASTQ files from 3 samples

Species	Potato	Apple	Grapevine
Sequencing technology	Illumina		
Sequencing depth	23M	13M	12M
Plant	Potato	Apple	Grapevine rootstock Kober 125 aa, Vitis berlandieri x Vitis riparia
Viruses or viroids detected (one column per virus)	PNV	New Zealand virus, AOSV, grapevine yellow exocortis, grapevine yellow mosaic virus	Grapevine rootstock associated virus 1, Grapevine virus A, Grapevine virus B, Grapevine viroids stem girdling, grapevine yellow mosaic virus, Grapevine yellow mosaic virus 1, Grapevine yellow mosaic virus 2, Grapevine yellow exocortis, Grapevine yellow exocortis 2
Percentage of reads for each virus/viroid (%)	~23%	~5%	0.12%, 0.6%, 1.2%, 0.4%, 0.3%, 0.5%, 0.14%
Confirmations techniques for each virus/viroid	RT-PCR & Sanger sequencing	RT-PCR & Sanger sequencing, RT-PCR & Sanger sequencing	ELISA, RT-PCR, ELISA, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR, RT-PCR

Difficulties:

1. New unknown virus
2. Complex mix of 9 viruses/viroids

Methodology for evaluation of bioinformatic pipelines

- > sRNA datasets (length between 21 and 24 nt)
- > Worst case scenario by rarefaction at 3 sequencing depths:
 - ❖ 50,000 reads – 3 files
 - ❖ 250,000 reads – 4 files (2 grapevine replicates)
 - ❖ 2,500,000 reads – 3 files
- > 10 fastq files available on a server in double blind
- > 21 participants

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Difficulties:

1. New unknown virus
2. Complex mix of 9 viruses/viroids
3. Very low abundance virus/viroid

Methodology for evaluation of bioinformatic pipelines

Participants free to apply their own bioinformatics strategy to identify the viruses in the 10 datasets (de novo assembly + annotation)

Interpret the data in a diagnostics setting

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Difficulties:

1. New unknown virus
2. Complex mix of 9 viruses/viroids
3. Very low abundance virus/viroid
4. Close virus species with poor genome knowledge

Results: analytical sensitivity

- 70% sensitivity overall
- Sensitivity from 35 to 100%
- Sensitivity increase with sequencing depth

labID	Sensitivity			Average
	50,000	250,000	2,500,000	
A	100%	53%	90%	51%
B	30%	35%	80%	46%
C	60%	71%	80%	70%
D	50%	82%	100%	78%
E	30%	82%	80%	68%
F	80%	88%	100%	89%
G	20%	53%	100%	57%
H	30%	65%	70%	57%
J	70%	94%	100%	89%
K	40%	71%	90%	68%
M	50%	94%	90%	82%
N	30%	82%	90%	70%
O	20%	41%	40%	35%
P	20%	59%	70%	51%
R	100%	100%	100%	100%
S	50%	100%	100%	86%
T	90%	100%	100%	97%
V	60%	88%	80%	78%
W1	40%	82%	90%	73%
W2	60%	82%	90%	78%
X	30%	71%	80%	62%
AVERAGE	46%	75%	86%	70%

Results: analytical sensitivity

- 70 % sensitivity overall
- Sensitivity from 35 to 100%
- Sensitivity increase with sequencing depth
- Seven strategies have 100% sensitivity at 2.5 M
- Participant R: 100% sensitivity (T close)
- > Both have an additional read mapping step

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- Sensitivity improved by lower k-mer lengths (13-15), decreased by higher length (17-21)
- > impact of k-mer length
- Sensitivity reduced by setting minimal contig length too high, a value <60nt should be preferred
- > Shorter minimal contig length preferred

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Results: analytical sensitivity per virus

- Correlation between percentage of viral reads in sample and detectability of a given agent
- Other parameters have an influence

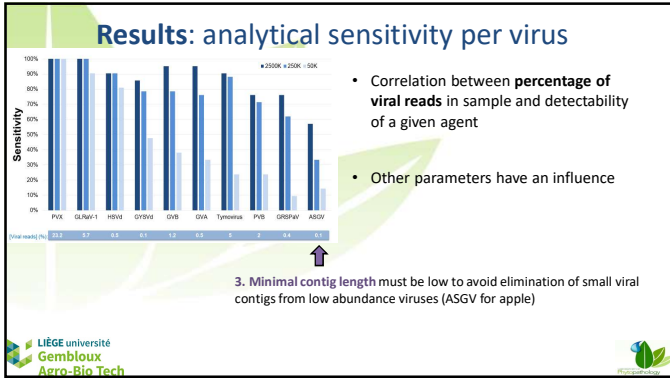
Results: analytical sensitivity

- Sensitivity improved by contig extension after de novo analysis
- > Contig extension to pass minimal contig length threshold

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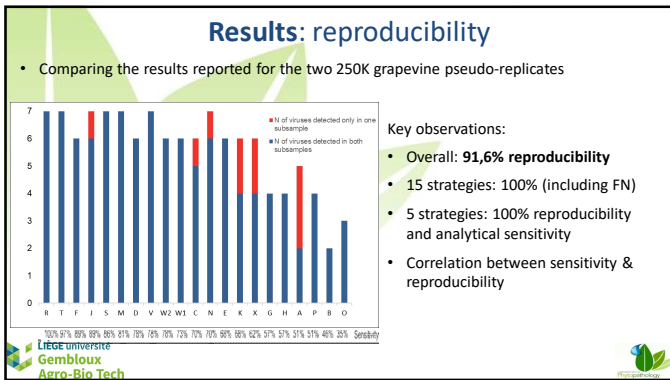
Results: analytical sensitivity per virus

- Correlation between percentage of viral reads in sample and detectability of a given agent
- Other parameters have an influence



Conclusions: bioinformatics matters !

- Some rather trivial things to avoid
 - BlastN only
 - Stringent Blast cut-off values
 - MegaBlast (high homology threshold)



Conclusions: bioinformatics matters !

- Some rather trivial things to avoid
 - BlastN only
 - High Blast cut-off values
 - MegaBlast (high homology threshold)
- Some less obvious lessons
 - Minimal contig length must be very low (<60nt) for optimal sensitivity with low abundance viruses
 - For Velvet, broad range of k-mer values including higher ones improve sensitivity
 - Additional contigs extension and combination of mapping and de novo assembly might improve detection

Conclusions: bioinformatics matters !

- A huge diversity in pipelines and pipeline performance
- Identification of trivial errors to avoid and other lessons
- **siRNA indexing data annotation is not such a straightforward business**

For more details: see our publication in Phytopathology



Phytopathology • 2019 • 109-488-497 • <https://doi.org/10.1094/PHYTO-02-19-0067-R>

Virology e-Xtra*

Virus Detection by High-Throughput Sequencing of Small RNAs: Large-Scale Performance Testing of Sequence Analysis Strategies

Sebastien Massart,¹ Michela Chiumenti, Kris De Jonghe, Rachel Glover, Annelies Haegeman, Igor Koloniuk, Petr Kominek, Jan Kreuze, Denis Kutnjak, Leonidas Lotos, François Maclot, Varvara Maliogka, Hans J. Maree, Thibaut Olivier, Antonio Olmos, Mikhail M. Pwoggin, Jean-Sébastien Reynard, Ana B. Ruiz-García, Dana Safarova, Pierre H. H. Schneeburger, Noa Sela, Silvia Turco, Eva J. Vainis, Eva Varallyay, Eric Verdin, Marcel Westenberg, Yves Brostaux, and Thierry Candresse

One step further: applying HTS in plant pest diagnostics

- ✓ Climbing the diagnostic cliff of HTS technologies ?
- ✓ Writing of international guidelines for the use of HTS technologies for plant pest diagnostics
- ✓ 54 scientists & >30 countries
- ✓ For viruses, bacteria, fungi, nematodes and insects
- ✓ 70 pages soon available as scientific paper

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Thanks for the invitation

Thank you for your attention


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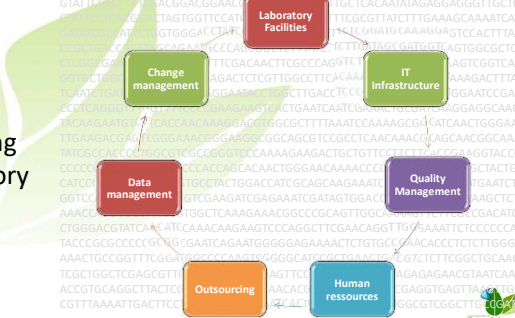


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One step further: applying HTS in plant pest diagnostics




1. Preparing the laboratory

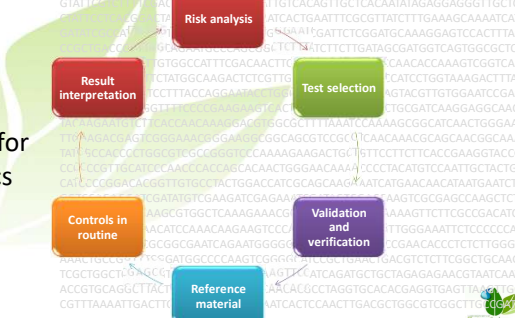


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One step further: applying HTS in plant pest diagnostics



2. HTS test for diagnostics



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