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Diffusive emissions of methane and nitrous oxide from a

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Lakes

Reservoirs

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Abstract

cascade of tropical hydropower reservoirs in Kenya

The present study investigated diffusive emissions of methane (CH_{4}) and nitrous oxide (N₂O) to the atmosphere from three relatively small (3–120 km^2) reservoirs (Masinga, Kamburu and Gitaru) on the Tana River (Kenya). Sampling was conducted biweekly in 2011, 2012 and 2013, at sampling sites upstream and downstream of these reservoirs while five sampling campaigns were carried out in 2011, 2012 and 2013 for different sites within each of the reservoirs. The dissolved CH_4 (range: 19-2101 nmol/L) and N₂O (range: 6.2-11.5 nmol/L) concentrations in the surface waters were generally very low in the three reservoirs, compared with other reservoirs globally. The lower diffusive emissions of CH_4 (20-216 μ mol/m² day⁻¹) and N_2O (1.0–1.6 μ mol/m² day⁻¹) from these reservoirs, compared with other tropical reservoirs, are probably related to their age (30-40 years), and lower vegetation biomass (savannah) originally present and submerged during their commissioning. The reservoirs with longer water residence times were characterized by higher diffusive CH₄ fluxes (216 ± 666 μ mol/m² day⁻¹) and slightly lower N₂O fluxes (1.0 ± 1.5 μ mol/ m^2 day⁻¹). The relative contribution of turbine fluxes of CH₄ and N₂O, compared to diffusive fluxes, was also highly variable among the three dams, being lower in Masinga Reservoir and higher in Gitaru Reservoir.

KEYWORDS

African (tropical) reservoirs, greenhouse gases, Methane emissions, nitrous oxide emissions

1 | INTRODUCTION

The damming of rivers has been integral to human population growth, economic development and technological innovation through the provision of a reliable water supply, security from flooding, irrigation water for agriculture and generation of electricity (Deemer et al., 2016; Snoussi et al., 2007). About 16.7 million reservoirs have been constructed, with a combined storage capacity of \sim 8,070 km³ and resultant increase in freshwater surface area by more than 305,000 km², and an associated retention and degradation of carbon (C) (Maavara, Lauerwald, Regnier, & Cappellen, 2016). The increased C retention and processing results in the reservoirs producing elevated levels of greenhouse gases (GHGs), including carbon dioxide

 (CO_2) , methane (CH_4) and nitrous oxide $(N_2O; Barros et al., 2011;$ Deemer et al., 2016; Maeck et al., 2013; St. Louis, 2000; Yang et al., 2014). CH_4 and N_2O are powerful GHGs with global warming potential (GWPs), being 32 and 310 times worse than CO₂ over a 100-year time horizon (IPCC, 2013). GHG emissions from reservoirs result from degradation of submerged biomass, allochtonous inputs and autochtonous production (Kemenes, Forsberg, & Melack, 2007, 2011; Teodoru et al., 2012), and are influenced by the age of a reservoir (Abril et al., 2005G; Barros et al., 2011).

Reservoirs (manmade lakes) are currently considered significant contributors of atmospheric GHGs (Barros et al., 2011; Bastviken, Tranvik, Downing, Crill, & Enrich-Prast, 2011; St. Louis et al., 2000). Raymond et al. (2013), for example, estimated inland waters emit carbon dioxide (CO $_2$) at a rate of 2.1 Pg C/year, with emissions from hydroelectric reservoirs representing around 2% of this flux (49 Tg C/year), whereas Barros et al. (2011) estimated hydroelectric reservoirs emit about 48 and 3 Tg C/year as CO₂ and methane (CH₄), respectively, corresponding to about 4% of global carbon (C) emissions from inland waters. Deemer et al. (2016) revised upward the global reservoir GHGs emissions to 13.4 Tg CH_4 and 0.03 Tg N₂O per year. The latter CH_4 fluxes were higher than the previous estimates since they consider fluxes from temperate reservoirs (Beaulieu et al., 2014; Maeck et al., 2013) and sub-tropical reservoirs (Grinham, Dunbabin, Gale, & Udv. 2011; Sturm, Yuan, Gibbes, Werner, & Grinham, 2014) not included in previous global estimates, but whose emissions are as significant as those from tropical systems. Nevertheless, only a handful of studies globally have analysed the contribution of nitrous oxide (N₂O) to total GHG emissions from reservoirs (Deemer et al., 2016; Descloux, Chanudet, Serça, & Guérin, 2017; Guérin, Abril, Tremblay, & Delmas, 2008; Tremblay, Varfalvy, Roehm, & Garneau, 2005), despite N₂O having a higher global warming potential (GWP) compared with CH₄. Further, there are sparse data from the tropical areas, although tropical reservoirs exhibit high run-offs and associated high organic carbon loads resulting from irregular and heavy precipitation (Tundisi, Matsumura-Tundisi, & Calijuri, 1993), as well as nearly constant and elevated temperatures that causes thermal stratification and deoxygenation of bottom waters (Barros et al., 2011; Tundisi & Tundisi, 2012). These conditions are known to be favourable conditions for enhanced CH₄ and N₂O production and emissions (Demarty & Bastien, 2011; Fearnside, 1995; Galy-Lacaux, Delmas, Kouadio, Richard, & Gosse, 1999; Guérin et al., 2008).

Reservoirs

GHG emissions from reservoirs have led to controversy on the contribution of reservoirs to global warming, despite justifications by various authorities that hydroelectric power generation is a green energy (i.e. produces lower GHG emissions compared to fossil fuel) (Deemer et al., 2016; Hoffert et al., 1998; Victor, 1998). According to IPCC guidelines, tropical countries relying heavily on hydroelectricity are at a risk of having their national GHG emission inventories increased by as much as 7% (IPCC2006). Thus, it is a requirement that these countries work towards reducing uncertainties regarding the role of reservoirs in climate change and global warming by including them in their National GHGs Inventory Programme. Hydropower is an important renewable energy in Kenya, accounting for about 50% of installed capacity, with more reservoirs vet to be commissioned. Thus, there is a need to better understand the role of reservoirs in GHG emissions in Kenya. To this end, the present study investigated the role of cascading reservoirs in the production and emission of CH_4 and N_2O from three large reservoirs (Masinga; Kamburu; Gitaru) in the Tana River network in Kenya.

2 | MATERIALS AND METHODS

2.1 | Study sites

The present study was carried out in the Tana River basin, Kenya (Figure 1). The Tana River has a length of ~1,000 km, originating in the Aberdare ranges and Mount Kenya of central Kenya and running through the arid and semi-arid lands in the eastern part of the country to enter the Indian Ocean through a fan-shaped delta. The Tana River basin experiences a bimodal precipitation pattern, with dominant rainy seasons occurring during April–June and October-December. The basin experiences variable rainfall patterns, decreasing from the headwaters (~1,800 mm/year), upper highlands (~2,200 mm/year), mid-altitude catchment (900–2,000 mm/year) to the lower semi-arid Tana catchment (450–900 mm/year; Brown & Schneider, 1998).



FIGURE 1 Sampling sites in the three cascading reservoirs of Tana River, Kenya

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Five cascading reservoirs were constructed along the Tana River between 1968 and 1981. The Masinga, Kamburu and Gitaru reservoirs were selected for the present study because of their different sizes, depth profiles and residence times (Table 1).

The samples river sections were upstream of Masinga Reservoir (R1), between the three dams (below Masinga Reservoir (R2) and below Kamburu Reservoir (R3)), and below Gitaru Reservoir (R4). Three sampling sites also were selected within Masinga Reservoir (along the water flow path: Masinga-1, -2 and -3) because of its larger size, while two sites were selected within Kamburu Reservoir (Kamburu-1 and -2) and Gitaru Reservoir (Gitaru-1 and -2; Figure 1). The sampling sites on the upper reaches of the reservoir were located near the reservoir inlet (characterized by high water flow rates), while the lower reach sites were located towards the reservoir outlets (close to the turbines and characterized by lower water flow rates), whereas the mid-reach sites were located roughly halfway across the reservoir length. Other small rivers flowing into the reservoirs (e.g., Sabasaba, Thika, Chania and Thiba rivers) were not sampled in the present study because of logistical constraints.

2.2 | Field measurements and sample collection

Samples were collected biweekly at the inflows and outflows from the three study reservoirs during five sampling campaigns (2011– 2013) at sites within the reservoirs (Figure 2). No spillover discharges from the reservoirs occurred during the five campaigns. Water samples from the reservoirs were collected using a vertical Niskin bottle (4 L), whereas the sampling sites upstream and downstream of reservoirs and the sites between the reservoirs were sampled with a plastic bucket (10 L).

Temperature, electric conductivity, pH and dissolved oxygen (DO) concentrations were measured in situ using a hand-held multiparameter metre (YSI Pro Plus). Water samples for determining dissolved ammonium ($NH_4^{+}-N$) and nitrate + nitrite {($NO_3^{-} + NO_2^{-})-N$ } concentrations were collected in acid pre-washed polyethylene bottles following double filtration on pre-combusted 47 mm GF/F filters and through 0.2-µm syringe filters. Nutrient samples (50 ml) were preserved by adding 50 µl of a saturated mercuric chloride (HgCl₂) solution. Methane (CH₄) and nitrous oxide (N_2O) samples were collected with 50-mL serum bottles, poisoned with 100 µl of a saturated solution of HgCl₂ and capped with a butyl-rubber stopper and aluminium cap. To avoid gas exchange with the atmosphere, the CH₄ and N_2O samples were collected with a rubber tube from the bottom of the Niskin bottles. In those cases where that Niskin bottles could

not be used (e.g., sampling sites between the study reservoirs), water samples were drawn from a 0.5m depth below the river surface directly into sampling bottles.

2.3 | Sample preparation and analysis

CH₄ and N₂O concentrations were determined via a headspace equilibration technique (20 ml N₂ headspace in 50-ml serum bottles) and measured with a gas chromatograph (SRI 8610C) with flame ionization detection, and electron capture detection calibrated with CH₄:CO₂:N₂O:N₂ mixtures (Air Liquide Belgium) of 1, 10 and 30 ppm CH₄ and of 0.2, 2.0 and 6.0 ppm N₂O (Borges et al., 2015). The methods described by Parsons, Maita, and Lally (1984) and APHA (1998) were used to analyse the dissolved ammonium (NH₄⁺-N), nitrate + nitrite {(NO₃⁻ + NO₂⁻)-N concentrations in the water samples. Dissolved NH₄⁺-N concentrations were determined using the indophenol method and measured at 630 nm after at least 6 hr' incubation. Dissolved (NO₃⁻+NO₂⁻)-N concentrations were determined using the cadmium reduction method, measured colorimetrically at 543 nm.

Diffusive fluxes of CH_4 and N_2O at the air-water interface were calculated using gas exchange velocities calculated from the empirical relationships with wind speed as proposed by Cole and Caraco (1998), using wind speed obtained from the National Centers for Environmental Prediction (NCEP) gridded daily product (grid point: $-0.95237^{\circ}N$, 37.50000°E). The emissions were further converted to CO_2 equivalents (1 mass unit of trace GHG to the emission caused by 1 mass unit of CO_2 over a 100 years) to obtain the relative contribution of the various GHG emissions to global warming (Deemer et al., 2016).

The turbine flux was calculated as the product of the turbine discharge and the difference between the observed CH_4 (or N_2O) concentration at the water withdrawal depth and the concentrations at equilibrium with the atmosphere. This flux estimate corresponds to the maximum potential emission since it assumes full degassing and equilibration with the atmosphere during the transit through the turbines. The computations were made for March 2011, July 2011, January 2012, May 2012 and February 2013, when vertical profiles were obtained for the reservoir intakes.

3 | RESULTS AND DISCUSSION

There were seasonal variations and different patterns in regard to the biogeochemical variables sampled in the river and study reservoirs (Table 2). The surface water temperature was higher

TABLE 1Main characteristics of Masinga, Kamburu and Gitaru reservoirs on the Tana River (modified from Brown, Schneider, & Harper,1996; water residence time calculated using discharge data for 2011 and 2012)

Reservoir	Altitude (m)	Year commissioned	Capacity (×10 ⁶ m ³)	Surface area (km²)	Maximum depth at commissioning (m)	Installed ca- pacity (MW)	Residence time (days)
Masinga	1,050	1981	1560	120	50	40	149
Kamburu	1,010	1974	123	15	56	96	16
Gitaru	924	1978	20	3.1	30	225	3

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FIGURE 2 Time series of discharges at different sampling sites (arrows showing sampling dates: 17/3/2011-21/3/2011; 15/7/2011-18/7/2011; 19/1/2012-22/12/2012; 20/5/2012-22/5/2012 and 8/2/2013-10/2/2013)

(*F* = 14.02; *p* < 0.01) in the reservoirs (range: 21.1–28.0°C), compared with the most upstream river station (R1; range: 20.6–25.7°C). The DO concentrations were similar (*F* = 0.091; *p* > 0.05) for both the reservoir surface (5.27–9.35 mg/L) and R1 (4.4–10.0 mg/L) sampling sites. The CH₄ concentrations in the surface water of the reservoirs

were generally low (compared with other tropical reservoirs), averaging 63, 55 and 28 nmol/L in Masinga, Kamburu and Gitaru reservoirs, respectively, indicating a decreasing pattern along the cascade of reservoirs, as well as along the size gradient, with the largest reservoirs having the longest water residence times, and

TABLE 2 Mean environmental variables at sampling sites (average ± standard deviation) in upstream and downstream rivers (R1, R2, R3 and R4) and within Masinga (Masinga-1, Masinga-2 and Masinga-3), Kamburu (Kamburu-1 and Kamburu-2) and Gitaru (Gitaru-1 and Gitaru-2) reservoirs

Sampling site	Temperature (°C)	Dissolved oxygen (DO) (mg/L)	Electrical conduc- tivity (µS/cm)	рН	Ammonia (NH₄⁺) (µmol/L)	Nitrate (NO ₃)⁻ (μmol/L)
R1	22.2 ± 1.0	7.49 ± 1.00	111 ± 13	7.30 ± 0.09	1.31 ± 0.57	3.90 ± 1.94
Masinga-1	25.5 ± 1.2	7.28 ± 0.43	125 ± 4	8.03 ± 0.11	1.33 ± 0.57	3.4 ± 1.6
Masinga-2	26.4 ± 0.4	6.82 ± 0.43	136 ± 10	8.19 ± 0.08	1.04 ± 0.48	3.64 ± 2.04
Masinga-3	25.2 ± 0.5	7.05 ± 0.63	129 ± 11	7.96 ± 0.07	0.80 ± 0.28	4.78 ± 2.55
R2	23.2 ± 0.5	3.46 ± 0.91	116 ± 11	7.00 ± 0.27	1.95 ± 0.62	4.59 ± 1.77
Kamburu-1	25.3 ± 1.0	7.48 ± 0.74	122 ± 8	8.15 ± 0.14	0.92 ± 0.39	3.81 ± 2.03
Kamburu-2	25.5 ± 0.6	6.96 ± 0.48	129 ± 10	8.03 ± 0.14	1.27 ± 0.45	3.99 ± 1.55
R3	24.0 ± 0.4	6.48 ± 0.40	124 ± 16	7.25 ± 0.36	1.31 ± 0.57	4.02 ± 2.13
Gitaru-1	23.8 ± 0.6	6.91 ± 0.24	126 ± 13	7.43 ± 0.16	0.97 ± 0.14	5.23 ± 2.38
Gitaru-2	25.5 ± 0.6	7.44 ± 0.42	126 ± 9	7.79 ± 0.19	1.36 ± 0.37	5.33 ± 1.99
R4	24.8 ± 0.3	6.57 ± 0.40	127 ± 11	7.64 ± 0.21	1.24 ± 0.39	5.09 ± 2.00

FIGURE 3 Seasonal evolution of methane (CH_4) and nitrous oxide (N_2O) concentrations in surface waters of Masinga, Kamburu and Gitaru reservoirs from December 2010 to July 2013 (inlet sites include Masinga-1, Kamburu-1 and Gitaru-1; intake sites include Masinga-3, Kamburu-2 and Gitaru 2)



slightly higher surface water concentrations. The overall amplitude of the CH₄ concentration temporal variations was also relatively modest, with minimum-maximum ranges of 8-454 nmol/L in Masinga, 7-425 nmol/L in Kamburu and 3-134 nmol/L in Gitaru. The overall amplitude of the CH_4 concentration temporal variations also followed the pattern of size as average concentration. Masinga and Gitaru reservoirs only exhibited small differences between the values at the inlet (upper reach sites) sites and the intake sites (lower reach sites), whereas a marked difference was observed for Kamburu Reservoir, with higher CH_4 concentrations in the upper reaches than in the lower reaches, possibly attributable to Thiba River inputs. Though not statistically significant, there were differences between the average CH₄ concentration in the river entering the reservoir and the reservoir itself, with systematically higher CH₄ average concentration values in the rivers, as well a higher temporal variability. The Sagana River (R1) exhibited an average CH₄ concentration of 272 nmol/L (range: 13-1,905 nmol/L), compared with 63 nmol/L in Masinga Reservoir. The river at the outlet of Masinga Reservoir exhibited an average CH₄ concentration of 839 nmol/L (range: 15-14,202 nmol/L), compared with 55 nmol/L in Kamburu Reservoir, while the Kamburu Reservoir outlet river exhibited an average CH₄ concentration of 38 nmol/L (range: 5-322 nmol/L) versus 33 nmol/L in Gitaru Reservoir.

The N₂O concentrations were significantly higher (F = 7.581; p < 0.01) and variable in the river upstream of the reservoirs (range: 6.9–23.0 nmol/L), compared with the sampling sites within the reservoirs (range: 6.2–11.9 nmol/L) and the rivers downstream of the reservoirs (Figures 3, 4 and 5). The concentrations remained relatively similar, however, within the three reservoir



FIGURE 4 Seasonal evolution of methane (CH_4) and nitrous oxide (N_2O) concentrations in surface waters of Sagana River (R1), and downstream rivers of Masinga (R2), Kamburu (R3) and Gitaru (R4) reservoirs from December 2010 to July 2013

sites, being 6.2–10.7 nmol/L; 6.9–8.9 nmol/L and 6.9–11.9 nmol/L for Masinga, Kamburu and Gitaru reservoirs, respectively (Figure 3).



FIGURE 5 Schematic diagram summarizing medians or range of medians of methane (CH₄; nM), nitrous oxide (N₂O;nM) and RT (water retention time, days), depth (D, m), surface area (SA; km²) and capacity (×10⁶ m³) in Masinga (MAS), Kamrubu (KAM) and Gitaru (GIT) reservoirs and riverine sites (R1, R2, R3 and R4).



FIGURE 6 Depth profiles of methane (CH₄) and nitrous oxide (N₂O) in reservoir lower reach sites (Masinga-3, Kamburu-2 and Gitaru-2) from December 2010 to July 2013

The CH₄ concentrations were generally higher in the rivers downstream of the reservoirs, whereas the N₂O was generally higher at the withdrawal depth than in the river downstream of the dams (Figure 6). For logistic reasons, the rivers downstream of the reservoirs were sampled several kilometres downstream, rather than immediately below the dams, which might have resulted in instream CH₄ production, and degassing of N₂O to the atmosphere.

The diffusive CH₄ flux in the three Tana reservoirs ranged between 20 and 216 μ mol/m² day⁻¹ (Table 3) with Masinga Reservoir (the largest reservoir with the longest water residence time) characterized by the highest CH₄ concentration and diffusive CH₄ fluxes. However, these fluxes fall within the lower end of the range of CH₄ flux intensities reported so far for tropical and sub-tropical reservoirs (15–100, 313 μ mol/m² day⁻¹), and well below the average of ~9,000 μ mol/m² day⁻¹, as reported by Yang et al. (2014).

The diffusive N₂O flux in the three Tana reservoirs ranged between 1.0 and 1.6 μ mol/m² day⁻¹ (Table 3), corresponding to the lowest N₂O flux intensities reported so far for tropical and sub-tropical reservoirs. Guérin et al. (2008) reported N₂O diffusive fluxes ranging between 3 and 103 μ mol/m² d⁻¹ for six South American tropical reservoirs, averaging 65 μ mol/m² day⁻¹. Diffusive N₂O fluxes were reported by Liu et al. (2011) and Zhu et al. (2013) for three sub-tropical Chinese reservoirs (Wujiangdu, 15 μ mol/m² day⁻¹; Hongjiadu, 10 μ mol/m² day⁻¹; Three Gorges Reservoir, 8 μ mol/m² day⁻¹), by Deshmukh (2013) for the sub-tropical Nam Theun 2 Reservoir (~15 μ mol/m² day⁻¹) and by Deemer et al., 2016 for the global mean estimate of reservoir N₂O fluxes (500 μ mol/m² day⁻¹).

The lower CH_4 and N_2O diffusive fluxes from the three Tana dams can be attributed to at least three reasons. First, the vegetation surrounding the reservoirs that was submerged during damming was open deciduous forest and grassland. Thus, the submerged biomass was less than that of tropical evergreen forests typical of South American reservoirs, for which higher CH_4 and N_2O were reported (Guerin & Abril, 2007; Guérin et al., 2008). Second, the three Tana dams are Lakes 🚫 Reservoirs

characterized by O_2 concentrations that decrease with depth, although strictly anoxic conditions were not encountered. This contrasts with the strongly meromictic reservoirs that exhibit anoxic bottom waters, which is typical with South American reservoirs (Guerin, & Abril, 2007; Guérin et al., 2008). Third, the measurements reported in the present study were carried out 30–40 years after the construction of the dams, typically when the CH₄ fluxes are low due to large degradation of the original submerged biomass (Abril et al., 2005G; Barros et al., 2011).

The total diffusive CH₄ flux (integrated over their respective surface areas) was 28,052 mol/day, with a contribution roughly equivalent to a relative surface area of 92% from Masinga Reservoir, 7% for Kamburu Reservoir and <1% for Gitaru Reservoir (Table 3). The CH_{4} emission by turbines was of the same order of magnitude among the three dams (range: 234-840 mol/day), although its relative importance with regard to the integrated diffusive emission was highly variable, being higher in the smaller reservoirs. The diffusive CH₄ emission in Masinga Reservoir was 32 times higher than the turbine flux while, in contrast, the turbine flux was six times higher than the diffusive flux in Gitaru Reservoir. Up to 70% of the total reservoir CH₄ emissions were attributed to turbine degassing (Abril et al., 2005G; Fearnside, 2002; Kemenes, Forsberg, & Melack, 2007). The contribution of turbine CH_4 degassing in the release of CH_4 from the three studied Kenyan reservoirs was lower, given the bottom waters in the reservoirs were not anoxic. The reported flux ranges, however, were much lower, compared with those reported for Lake Kariba, the only other African reservoir intensively studied for CH₄ fluxes. Diffusive fluxes in Lake Kariba ranged between 5 and 5,000 μ mol/m² day⁻¹, with a dominance of fluxes from ebullition ranging between 5 and 50,000 µmol/m² day⁻¹ (DelSontro, Beaulieu, & Downing, 2018).

The relative importance of the turbine flux, compared with the diffusive flux, was higher for N_2O than for CH_4 . In Masinga Reservoir, the diffusive N_2O flux was only ~4 higher than the N_2O turbine flux, while the turbine flux was higher than the diffusive flux by a factor of ~2 and ~5 in Kamburu and Gitaru reservoirs, respectively (Figure 7).

Reservoir Gitaru Masinga Kamburu Dissolved CH₄ concentration (nmol/L) 323 ± 1,015 192 ± 314 30 ± 25 Diffusive CH₄ flux (μ mol m⁻² day⁻¹) 216 ± 666 138 ± 226 20 ± 19 Diffusive CH_4 flux (mg- CO_2 eq m⁻² day⁻¹) 118 ± 362 75 ± 123 11 ± 10 Diffusive integrated CH₄ flux (mol/day) 25,926 ± 79,907 2,066 ± 3,397 60 ± 56 Turbine integrated CH₄ flux (mol/day) 804 ± 1,395 234 ± 199 350 ± 197 Dissolved N₂O concentration (nmol/L) 7.5 ± 2.2 7.4 ± 1.1 8.6 ± 1.9 Diffusive N₂O flux (μ mol m⁻² day⁻¹) 1.0 ± 1.5 1.1 ± 0.7 1.6 ± 1.3 Diffusive N₂O flux (mg-CO₂ eq m^{-2} day⁻¹) 13 ± 20 15 ± 9 22 ± 18 Diffusive integrated N₂O flux (mol/day) 117 ± 182 17 ± 10 5 ± 4 Turbine integrated N₂O flux (mol/day) 30 ± 35 28 ± 28 25 ± 20

Notes: Based on data collected from March 2011 to July 2013; concentrations and diffusive fluxes correspond to average intake and inlet data; CO_2 equivalent fluxes were computed using global warming potential values at 100 year time frame of 34 and 298 for CH_4 and N_2O , respectively (IPCC 2013); diffusive flux spatially integrated with average surface areas reported in Table 1.

TABLE 3 Average \pm standard deviation of dissolved methane (CH₄) and nitrous oxide (N₂O) concentrations, diffusive fluxes and turbine fluxes in Masinga, Kamburu and Gitaru reservoirs


FIGURE 7 Comparison of methane (CH_4) and nitrous oxide (N_2O) concentrations at withdrawal depth within reservoir and corresponding downstream river for Masinga, Kamburu and Gitaru reservoirs from March 2011 to July 2013 (CH_4 at withdrawal depth in the reservoir was lower than in downstream river 54% of the sampling occasions; solid line is 1:1 line)

Turbine fluxes are particularly important emission pathway for reservoirs, particularly when the spilled water is high in GHGs (Guérin & Abril, 2007) and when the water is withdrawn from the hypolimnion (Kemenes et al., 2007).

4 | CONCLUSIONS

Very low CH₄ and N₂O fluxes in the Masinga, Kamburu and Gitaru reservoirs were reported, compared with other tropical reservoirs, because of their age (30-40 years) and possibly because of the lower vegetation biomass initially submerged (mainly open deciduous forest and grassland), compared with other tropical reservoirs, such as in South America, which typically contain submerged tropical evergreen forest. The reservoirs with longer water residence times were characterized by higher diffusive CH_4 fluxes and slightly lower N_2O fluxes. The N₂O diffusive fluxes expressed in CO₂ equivalents were nine times lower than the CH₄ diffusive fluxes expressed in CO₂ equivalents in the reservoir with the longer residence time, but two times higher in the reservoir with the shortest water residence time. The relative contribution of turbine fluxes of CH₄ and N₂O, compared with diffusive fluxes, was also highly variable among the three dams, being smaller in Masinga Reservoir and larger in Gitaru Reservoir. Whereas the present study highlights the role of reservoirs in CH₄

and N₂O production, future research should involve a more detailed quantitative assessment of GHG emissions from the Tana River reservoirs, including dedicated field surveys combining CH_4 , N₂O and direct pCO₂ measurements (Abril et al., 2015) at high spatio-temporal resolution, coupled with chamber measurements to determine the water-air exchange fluxes of these GHGs. Future research should also attempt to investigate the emission pathways, especially ebullition, diffusion and turbine fluxes of CH₄ (sediment-water and sediment-air during low water level), and their variability in the littoral and pelagic zones.

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SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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