

Characterization of putative acetate transporters in *Chlamydomonas reinhardtii*

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Introduction

The unicellular green alga *C. reinhardtii* can grow heterotrophically by consuming acetate in the dark and mixotrophically by using both carbon sources in the light. Despite significant knowledge gained on acetate metabolism, the genes coding for acetate transporter/permease are still unknown in this alga. However, recent analyses^{1,2} have shown five functionally uncharacterized members of the **GPR1/FUN34/yaaH (GFY)**, a protein family which includes genes involved in carboxylic organic acid uptake/sensing already described in bacteria, yeasts and filamentous fungi. Thus, the five genes identified in *C. reinhardtii* as GFY1-5 encode for putative acetate transporter proteins given that they are structured in 6 hydrophobic transmembrane helices. They are characterized by a close gene structure (Fig. 1) and very high similarity in their coding sequence (CDS) except at the N-terminus amino acid sequences (Fig. 2). Insertional mutants for the genes GFY1, 2 and 3 are available, and artificial microRNA (amiRNA) technique will be used to generate knock-down mutant for GFY4, 5 and all the 5 genes. Mutants will be used to investigate about the role of this putative acetate transporters by placing them in different culture conditions. Plus, protein localization experiments will already give some clues about a putative peroxisomal localization (Fig. 3). If confirmed, as far as we know this work could represent the first attempt to describe acetate transporters in this microalga.

Fig. 1 – Gene organisation into the genome

artificial microRNA silencing

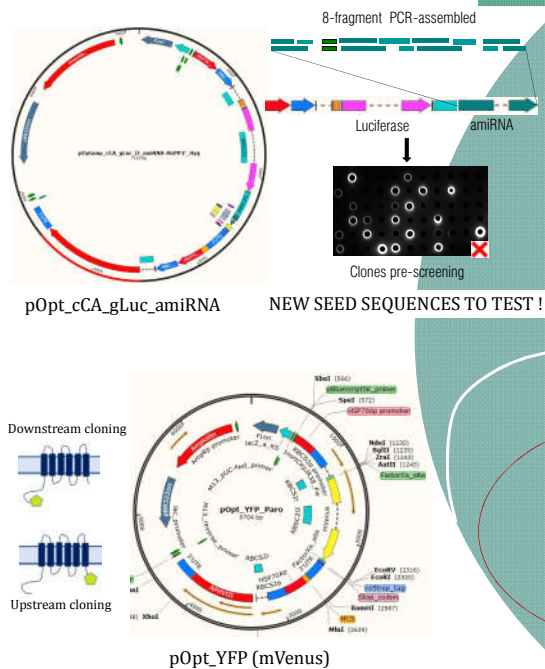
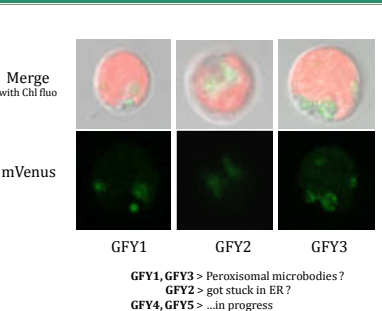


Fig. 3 Protein localization



Material & Methods

Chloroplast

Pyrenoid

Cytosol

Peroxisomal microbodies

Mitochondria

Conditions tested...

Photosynthesis/Respiration reactivation after anaerobiosis

Starch/Lipids accumulation

Dissection of respiratory pathways contribution

[acetate] vs pH
Fluoro-acetate as toxic analogue

Cells no./size distribution
Growth rate/dry weight
Acetate consumption rate

Results

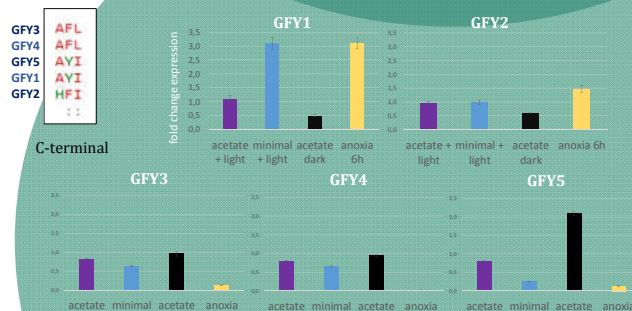


Fig. 3 – Expression level qRT-PCR: 2^{-ΔΔCt} normalization (CBLP/RPL13/BTUB)

Gene expression

The divergent N-terminus sequences and the distinct expression pattern in different cultivation conditions tested, point to a different situation. In particular, our qRT-PCR analyses showed that **GFY1 and GFY2 transcripts were more abundant in anaerobiosis** while **GFY3, GFY4, and GFY5 were mainly expressed during acetate assimilation**. In support of these findings, associated co-expressed genes also exhibited similar expression patterns, typical of each condition,

Conclusion

At first glance, GPR1/FUN34/yaaH genes found in *C. reinhardtii* seem to derive from gene redundancy. However, the N-terminal divergent amino acid composition and the distinct expression under the different culture conditions tested point to a different situation. Indeed, our preliminary data suggest two differentiated expression patterns, one co-expressed with fermentation pathway (GFY1, GFY2) and the second that match with acetate assimilation metabolism (GFY3, GFY4, GFY5). This putative acetate transporters will show eventually a different subcellular localization, from the cellular membrane (GFY2) to the inner membrane of peroxisomes (GFY1, 3, 4 and 5).