
Endogenous Erythropoietin in the Anemia of Chronic Disorders

YVES BEGUIN

National Fund for Scientific Research,
Division of Hematology,
Department of Medicine; and Laboratory of Cell
and Gene Therapy, CHU Sart-Tilman, Center for
Cellular and Molecular Therapy,
University of Liege, Liege, Belgium

INTRODUCTION

Anemia of chronic disease (ACD) is defined as the anemia associated with infection, inflammation, cancer, or trauma that has the characteristic picture of hypoferrremia, hyperferritinemia, decreased transferrin concentration, and increased iron stores (1). The pathogenesis of ACD involves the combination of a shortened erythrocyte survival in circulation with failure of the bone marrow to increase red cell production

in compensation (2-7). Inappropriate red cell production is itself related to a combination of factors, including impaired availability of storage iron, inadequate erythropoietin (Epo) response to anemia, and overproduction of cytokines, which are capable of inhibiting erythropoiesis (2-5). These cytokines are involved in the retention of iron in the reticuloendothelial system, gastrointestinal tract, and hepatocytes. They may interfere with Epo production by the kidney, and may exert direct inhibitory effects on erythroid precursors (3,4,8-12). Indeed, their effect is much wider, involving the whole hematopoietic system (13).

Cancer is one of the leading causes of ACD. However, the anemia observed in cancer patients may have multiple mechanisms (2,14,15). Hemodilution may artificially dilute the red cell mass. Bleeding, autoimmune or microangiopathic hemolysis, hypersplenism, and hemophagocytosis may all reduce the red cell life span. Nutritional deficiencies, including iron, folate, vitamin B12, and global malnutrition, may impair red cell production. The bone marrow may be involved by metastases, necrosis, myelodysplasia, and autoimmune red cell aplasia. These various causes, not including the "anemia of chronic disorders," have been reviewed in detail elsewhere (16). Surprisingly, there are no reports on the relative proportion of cancer patients in general or of patients with any form of cancer, in particular, that present the typical features of ACD. In other words, the true incidence of ACD in cancer patients is completely unknown. Hence, the relevance of the biologic features of ACD to the overall erythropoietic activity of cancer patients remains elusive.

Furthermore, chemotherapy and radiotherapy have a major impact on the incidence and severity of anemia in cancer patients. Compared to untreated cancer alone, chemotherapy may double the incidence of anemia (17). The incidence and severity of anemia largely depends on the form of cancer as well as the type and dose intensity of chemotherapy administered to patients (18). This is also true in children where the incidence of chemotherapy-induced anemia may even be greater because of the nature of the cancer being

production is itself impaired availability of (Epo) response to which are capable of which are involved endothelial system, may interfere with direct inhibitory). Indeed, their role hematopoietic

of ACD. However, they may have multiple causes, including artificially dilute blood or microangiopathic hemophagocytosis, renal deficiencies, iron deficiency, malnutrition, bone marrow may be aplastic, and autophagy, not including which have been reviewed in literature. There are no reports on the pathogenesis in general or of the pathogenesis of ACD, that present itself as follows, the true incidence is completely unknown. The pathogenesis of ACD to the patients remains

Chemotherapy have a variety of anemia in which cancer alone, anemia (17). The pathogenesis ends on the form of chemotherapy, the true pathogenesis in children with anemia may be the cancer being

treated (many leukemias) and of the relative intensity of therapies applied (19). Various models have mostly identified older age, lower baseline Hb, and rapid drop of Hb after the first cycle as additional factors that are predictive of transfusion requirements in patients receiving chemotherapy (20-23). Chemotherapy may directly affect erythropoiesis in the bone marrow and also impact on endogenous Epo production.

In this review, we will examine the evidence for defective Epo production in patients with ACD. We conducted a wide literature survey on the topic and critically analyzed the papers identified in this search. Solid experimental data indicate that several cytokines interfere with Epo production. However, it is unclear how these data can be directly applied in vivo. Many clinical papers reporting serum Epo levels in various disorders associated with ACD, in particular in cancer patients, have methodological problems. Two major such problems can be identified. The first problem relates to the heterogeneity of the patients studied in terms of disease and stage of the disease, as well as the simultaneous inclusion of patients at diagnosis, during treatment and after completion of therapy. The second problem involves the interpretation of serum Epo levels in individual patients or in groups of subjects, with lack of appropriate controls and inadequate interpretation of Epo data.

Therefore, we will first present the experimental data on the effect of various cytokines on Epo production. Second, we will comment on appropriate methods allowing interpretation of serum Epo levels in patients. We will then review the evidence for defective Epo production in patients with ACD, focusing in particular on HIV (as a model of chronic infection), rheumatoid arthritis (as a model of chronic inflammatory disorder), and cancer. In the case of cancer, we will attempt to examine various diseases independently whenever possible, and we will try to delineate the respective roles of cancer itself and of chemotherapy. Finally, we will illustrate how baseline serum Epo levels can help predict response to recombinant human erythropoietin (rHuEpo) therapy.

EFFECTS OF CYTOKINES ON ERYTHROPOIETIN PRODUCTION (Table 1)

Peripheral blood mononuclear cells from patients with chronic renal failure released soluble factors that suppressed Epo production by HepG2 cells, but these factors did not appear to be TNF- α or IL-1 (24). Neopterin also induces a suppression of hypoxia-induced Epo synthesis in HepG2 cells in a dose-dependent manner (25). It has been reported that IL-1 α , IL-1 β , TNF- α , IFN- γ , and TGF- β inhibited, whereas IL-6 stimulated, cobalt-induced or hypoxia-induced Epo production at the mRNA level by the hepatoma cell line Hep3B (26,27). The inhibitory effect of IFN- γ was found to be additive to that of IL-1 and even synergistic with that of TNF- α , and was capable of preventing any response to IL-6 (27). The same inhibition of Epo gene expression and protein production was observed with the HepG2 line for IL-1 and TNF but not for TGF- β , IFN- γ , or IL-6 (28-30). Contrary to phorbol-ester-induced inhibition of Epo production, inhibition by IL-1 β or TNF- α was independent of protein kinase C (31). Inhibition of hepatic Epo production by TNF- α appears to be mediated by the 55 kDa (TNF-RI) rather than the 75 kDa (TNF-RII) receptor (32). IL-1, TNF- α , and IL-6 also blocked hypoxia-induced Epo formation by the isolated rat kidney (29).

Cytokine-induced inhibition of Epo production by HepG2 cells is not mediated by impairment of hypoxia-induced factor-1 (HIF-1) whose activity is rather enhanced by IL-1 β

Table 1 Effect of Various Cytokines on Epo Production by the Hepatoma Cell Lines Hep 3B and G2, by the Isolated Rat Kidney and in Vivo

	Hep 3B	Hep G2	Rat kidney	In vivo
IL-1	↓	↓	↓	↓
TNF- α	↓	↓	↓	↓
TGF- β	↓	=		= ↓
IL-6	↑	= ↓	↓	↑
IFN- γ	↓	=		↑

or TNF- α , while VEGF expression remains unaffected (33). Several cytokines stimulate inducible nitric oxide (NO) synthase gene expression in several tissues. It is therefore not surprising that NO donors dose dependently reduced Epo production in the HepG2 cell line, either by directly influencing the cellular redox state or by increasing reactive oxygen species in the cell (34). Indeed, reactive oxygen species, including H₂O₂, have been shown to suppress the in vitro synthesis of Epo (35). H₂O₂, whose production is reduced in hypoxic conditions, has been proposed as a potential signaling molecule between the oxygen sensor and the transcriptional machinery (35). Desferrioxamine and cobalt chloride antagonize the inhibition of Epo production by reactive oxygen species, by reducing the action of H₂O₂, and by interfering with its production and/or scavenging, respectively (36). Similarly, the antioxidant vitamins A, E, and C significantly increased Epo production by the hypoxic isolated rat kidney (37). While vitamin A also dose dependently increased Epo synthesis in Epo-producing hepatoma cell cultures, vitamins E and C had no such effects (37). In another experiment in which Epo synthesis by HepG2 cells was reduced by monocyte-conditioned medium as well as IL-1 β , TNF- α , and IL-6, dexamethasone decreased cytokine secretion by monocytes but did not affect Epo production on its own (38).

Injection of bacterial lipopolysaccharide (LPS) or IL-1 β to normoxic or hypoxic rats resulted in increased TNF- α mRNA and reduced Epo mRNA in the kidney, as well as decreased serum Epo levels (39). In vivo administration of TGF- β was associated with depressed serum Epo levels in one study (40) but not in another (41). Administration of IL-6 to cancer patients resulted in elevated serum Epo levels that paralleled the development of anemia (42). Treatment of patients with chronic active hepatitis B with interferon- α resulted in a transient increase in plasma Epo levels (43). The exogenous administration of rHuEpo to mice treated with IL-1 was able to correct the suppression of CFU-E as well as of other erythroid parameters (44-46). Erythropoietin could also reverse the anemia of mice treated with single injections of TNF (47) but not always when mice were continuously exposed

ients with suppressed
rs did not
uces a sup-
2 cells in a
that IL-1 α ,
as IL-6 sti-
production
3B (26,27).
tive to that
, and was
The same
production
NF but not
o phorbol-
tion by IL-
31). Inhibi-
ars to be
he 75 kDa
so blocked
idney (29).
by HepG2
ia-induced
d by IL-1 β

in by the
at Kidney

In vivo
↓
= ↓
↑

to TNF (47-49). Exogenous Epo was nevertheless capable of preventing the anemia induced by TGF- β (40).

INTERPRETATION OF SERUM Epo LEVELS

What Is a Normal Epo Value?

Erythropoietin production is regulated through a feedback system between the bone marrow and the kidney, which depends on a renal oxygen sensor (50,51). The capacity of the kidney to respond acutely to hypoxia by increasing Epo production may be modulated by prior sensitization. Post-transfusion polycythemic mice exposed to hypoxia (52) or cobalt chloride (53) did not show the increased rate of Epo production observed in normal animals (52). Mice made polycythemic by exposure to intermittent hypoxia showed an apparent sensitization of Epo-producing cells to hypoxic stimuli, explaining their greater Epo response to acute hypoxia, dexamethasone, testosterone, or isoproterenol, compared to hypertransfused mice (54-56). This was true for renal but not for extrarenal Epo production (57).

Serum Epo levels may vary considerably (51,58). Levels are usually between 10 and 20 mU/mL in normal subjects, may decrease somewhat in primary polycythemia, but increase exponentially when an anemia develops below an Hct of 30-35% (59). Therefore, a serum Epo value must always be evaluated in relation to the degree of anemia (Figs.1 and 2) (51). In addition, it should be compared to appropriate reference subjects who should display a normal Epo response to anemia, including patients with iron deficiency or hemolytic anemia (see below). Erythropoietin levels inappropriately low for the degree of anemia are encountered not only in renal failure (60), but also in a number of other conditions, including the anemia of chronic disorders (2,3). Inappropriately high serum Epo levels are often observed in secondary polycythemia, a feature permitting its diagnostic separation from primary polycythemia (61).

Serum Epo levels increase exponentially in proportion to the degree of anemia. We thus constructed reference regressions representing the normal relationships between Hct on

less capable of

gh a feedback kidney, which the capacity of by increasing sensitization. o hypoxia (52) reased rate of (52). Mice made oxia showed an to hypoxic sti- acute hypoxia, l, compared to : for renal but

(51,58). Levels rmal subjects, cythemia, but lops below an o value must ree of anemia e compared to play a normal with iron defi- ropoietin levels re encountered mber of other disorders (2,3). en observed in ; its diagnostic

n proportion to ference regres- etween Hct on

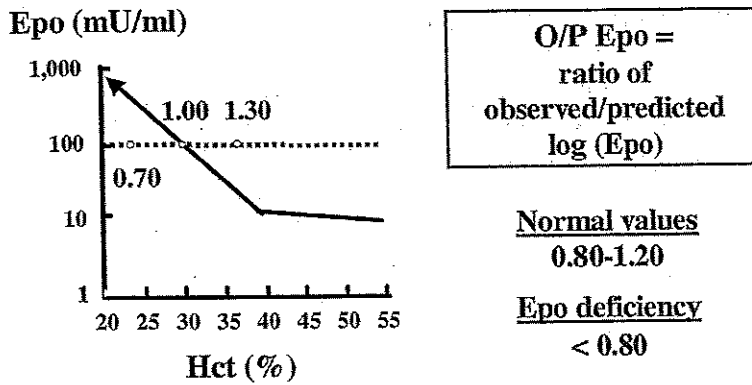


Figure 1 Interpretation of endogenous serum Epo levels. An individual serum Epo value of 100 mU/mL (dotted line) can be interpreted in relation with the degree of anemia through the O/P ratio. For an Hct of 30%, this Epo value is adequate (O/P ratio = 1.00), but for Hct of 23% or 37%, the same absolute Epo value would be defective (O/P ratio = 0.70) or excessive (O/P ratio = 1.30), respectively.

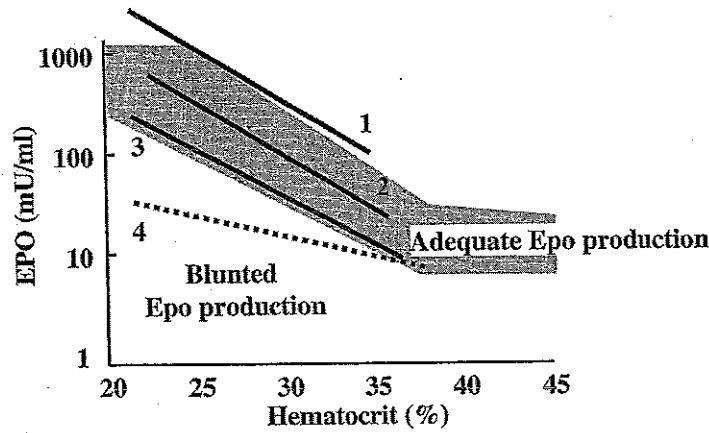


Figure 2 Interpretation of endogenous serum Epo levels. The hatched area represents the 95% confidence limits of the regression of Epo vs. Hct in an appropriate group of reference subjects, e.g., patients with IDA (2). However, serum Epo also depends on erythropoietic activity, with elevated and reduced levels in patients with low [aplastic anemia (1)] or high [thalassemia intermedia (3)] erythropoietic activity, respectively. A group of patients with ACD (4) shows a blunted Epo response to anemia.

one hand and Epo on the other, based on normal subjects and patients with hemolytic anemia (Fig. 2) (62). Two different regression equations were described for Hct $>$ or $<$ 40%. This cutoff Hct was chosen because it allowed for the best correlation for Epo data and because of literature data indicating that beyond such an Hct there is little modification of Epo levels. For Hct below 40%, the following regression ($R = -0.83$, $P = 0.0000$) was obtained between Epo (mU/mL) and Hct (%): $\log(\text{Epo}) = 3.420 - (0.056 \text{ Hct})$. For Hct over 40%, the regression equation ($R = -0.12$, NS) was: $\log(\text{Epo}) = 1.311 - (0.003 \text{ Hct})$. Based on these formulas, predicted $\log(\text{Epo})$ values were derived for each Hct, O/P ratios of observed/predicted $\log(\text{Epo})$ were derived, and 95% confidence limits were obtained in order to define a range of reference values for individual O/P ratios (Fig. 1). These limits are 0.80–1.20 for O/P Epo (62).

The adequacy of Epo production can thus be evaluated by two methods. When investigating a group of patients, this can be achieved by comparing patients and appropriate reference subjects by regression analysis (Fig. 2) (63). In this case, one should ensure that the study group encompasses a range of Hct values similar to that of the reference group; otherwise, the slopes of the regressions may be flawed. When studying an individual patient, the adequacy of Epo production can be evaluated by the O/P ratio (Fig. 1) (62). An O/P ratio below 0.80 indicates inadequate Epo production for the degree of anemia even if the absolute Epo value is high. It should be emphasized that the specific regression equations obtained in our study, on which O/P Epo ratios are based, cannot be automatically transposed to any other study. One must first either ensure that the Epo assay used yields Epo values similar to those measured in our Epo assay or construct one's own reference regressions with appropriate reference subjects.

Serum Epo Levels and Erythropoietic Activity

Many studies have reported higher serum Epo levels in patients with low compared to high erythropoietic activity

nal subjects and
) . Two different
> or < 40%. This
the best correla-
a indicating that
m of Epo levels.
sion ($R = -0.83$,
U/mL) and Hct
over 40%, the
 $\log(\text{Epo}) = 1.311 -$
redicted $\log(\text{Epo})$
ios of observed/
onfidence limits
reference values
ts are 0.80–1.20

be evaluated by
patients, this can
appropriate reference
In this case, one
asses a range of
roup; otherwise,
When studying
roduction can be
O/P ratio below
or the degree of
gh. It should be
iations obtained
ased, cannot be
. One must first
Epo values simi-
struct one's own
nce subjects.

1 Epo levels in
opoietic activity

(Fig. 2). In an early study, urinary Epo secretion was similar in patients with marrow failure or hemolysis (64). The slope of the correlation between Epo and Hb was steeper for patients with iron deficiency anemia (IDA) compared to those with aplastic anemia or transient erythroblastopenia, because Epo values in moderately anemic subjects were higher in the latter group (65). For similar degrees of anemia, patients with aplastic anemia had higher serum Epo levels than patients with iron deficiency or hemolytic anemia (66). At any hemoglobin value, serum Epo levels in patients with pure red cell aplasia were fourfold higher than in those with IDA, and tenfold higher than in patients with megaloblastic or sickle cell anemia (67). In 34 patients with aplastic anemia, serum Epo levels were much higher than in patients with iron deficiency at similar degrees of anemia (68). The same conclusions were obtained in another group of 42 patients with idiopathic aplastic or Fanconi's anemia (69). One log higher serum Epo values were encountered in patients with erythroid hypoplasia or aplasia (erythropoietic activity <0.6 times normal) compared to subjects with thalassemia intermedia (erythroid activity >2 times normal) (70). To account for this effect of erythroid activity, serum Epo levels can be corrected by the ratio of the sTfR (a quantitative marker of erythropoietic activity) value in the patient relative to a normal sTfR value (70). High serum Epo levels are also observed transiently after intensive chemotherapy, whether followed by bone marrow transplantation or not, without concomitant change in hemoglobin or hematocrit (70–76). The peak Epo values are observed 7 days after transplant, i.e., about 14 days after the start of the conditioning regimen, at the time of the nadir of erythropoietic activity. Within 24–72 hr after starting IV iron therapy in patients with IDA, marked decreases in serum Epo were found before any change in Hb (70). Similar observations were obtained with rHuEpo therapy in pure red cell aplasia (70) with vitamin B12, or folate therapy in megaloblastic anemia (70,77–79).

These findings thus point to an inverse relationship between marrow erythropoietic activity and serum Epo levels: the higher the number of erythroid precursors, the lower the

serum Epo value. As Epo exerts its action on target cells after binding to a specific Epo receptor (80), it is tempting to speculate that serum Epo levels may partly depend on the rate of Epo utilization by Epo receptor-bearing cells, primarily erythroid precursors (70,81). Similarly, marrow recovery after autologous stem cell transplantation (ASCT) would restore Epo utilization by erythroid cells, thus progressively returning Epo levels to a range appropriate for the degree of anemia (76). In patients with particularly fast engraftment, the duration of this correction phase is much shorter and may even finally lead to decreased Epo levels (76).

The idea that marrow utilization influences serum Epo levels was initially based on the observation that radiation-induced marrow hypoplasia was associated with a slower decline of serum Epo levels induced by hypoxia (82). However, the rate of Epo disappearance from the plasma of dogs with normal, hypoplastic, or hyperplastic marrow, was later shown to be similar, regardless of the experiment was performed in nephrectomized (83) or unmanipulated (84) animals. Nephrectomy or hepatectomy does not influence the pharmacokinetics of a large dose of native Epo (85) or a tracer dose of rHuEpo (86). Organ accumulation in the kidney and bone marrow of rats was minimal after intravenous injection of a tracer dose of rHuEpo (87,88). Furthermore, erythropoietin life span was similar in normal rats and in rats with bone marrow suppressed by cyclophosphamide or hypertransfusion or stimulated by hemolysis or bleeding (89). Similar conclusions were reached in mice 48 hr after initiation of hemolysis, bleeding or marrow suppression by 5-FU, or 2-24 hr after starting rHuEpo therapy, although the delay between induction of the desired experimental condition and measurement of Epo life span appears to be rather short (90). However, in normal human subjects (91,92) as well as in rats (93), the initial clearance of rHuEpo is decreased when the doses injected are increased, approaching a plateau at high doses. Furthermore, a surge in serum Epo levels after intense phlebotomy translates into decreased clearance of a tracer dose of rHuEpo (94). On the other hand, the pharmacokinetics of rHuEpo in hemodialysis patients was not different before

target cells after emptying to depend on the rate of cell recovery (ASCT) would be progressively faster for the degree of engraftment, shorter and may increase serum Epo in that radiation-damaged mice (82). However, plasma of dogs with anemia was later shown to be depressed after a tracer dose of kidney and bone marrow injection of a radiolabeled erythropoietin in rats with bone marrow hypertransfusion (84). Similar conclusions of hemolysis, or 2–24 hr after induction and measurement (90). However, in rats (93), the plasma at high doses after intense phlebotomy of a tracer dose of radiolabeled erythropoietin pharmacokinetics of the tracer were different before

and after 6 weeks of treatment with rHuEpo (95). In other studies, rHuEpo appeared to be eliminated from the plasma more rapidly after multiple doses than after a single dose in normal volunteers (96), whereas the elimination half-life of rHuEpo was increased on day 8 after two injections of rHuEpo to normal volunteers (97). The clearance of radiolabeled rHuEpo remained unchanged in rats injected with or without previous injections of unlabeled rHuEpo for 19 days (98) but was increased in sheep 8 days after experimental bleeding to Hb levels of 3–4 g/dL, before returning to baseline 4 weeks later (94). It was also progressively decreased in sheep following 5-FU- or busulfan-induced marrow ablation (99). Therefore, variations observed in serum Epo levels after intensive chemotherapy cannot simply be explained by changes in Epo consumption by the bone marrow.

The abnormal persistence of elevated plasma Epo levels in rats after cessation of intensive rHuEpo treatment given for 20 days could relate to suppression of erythroid activity (100). However, this was contradicted by our experiment with hypertransfused rats, in which polycythemia resulted in appropriate reduction rather than elevation of serum Epo levels, with subsequent depression of erythropoietic activity (100). Therefore, it is unlikely that persisting elevated Epo levels were due to nonutilization by a severely depressed erythroid marrow. Alternatively, Bozzini et al. (101) have suggested the existence of a yet unidentified feedback mechanism between Epo-responsive erythroid cells and Epo-producing cells. Cobalt- or hypoxia-induced Epo production in normocythemic mice is increased when erythropoiesis is acutely depressed and reduced when erythropoiesis is recently stimulated (101–103). Plasma Epo levels during hypoxia in mice with 5-FU- or irradiation-induced aplasia were higher than in normal mice (104). On the other hand, hypoxia-induced Epo response in transfused polycythemic mice is much higher when erythropoiesis has been previously stimulated for prolonged periods of time (101–103). These apparently contradictory observations in normal and polycythemic mice may be reconciled if it is a retracting erythron that can induce this Epo-hypersecretory state (101). However,

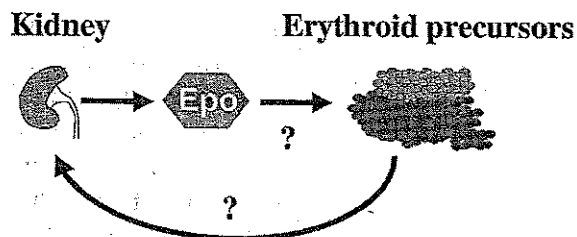


Figure 3 Serum Epo levels are the result of a balance between Epo production in the kidney and Epo utilization by the erythropoietic marrow. It remains to be determined whether the erythroid precursor mass acts directly by utilizing circulating Epo or indirectly by influencing the rate of Epo production.

although the erythron must shrink more after rHuEpo-than transfusion-induced polycythemia, it is unclear how hypoxia-induced Epo production would be relevant to our observed discrepancy in serum Epo levels between the two conditions.

In conclusion, serum Epo levels are the result of a balance between the rate of Epo production and its utilization by the erythroid marrow (Fig. 3). This should also be taken into account when interpreting the adequacy of a serum Epo value in various situations. Whereas it is indisputable that marrow erythropoietic activity independently influences serum Epo levels, it remains to be determined whether the erythroid precursor mass acts directly by utilizing circulating Epo or indirectly by influencing the rate of Epo production. Some other factors linking the erythron to Epo production may also exist. For instance, products resulting from red cell hemolysis may indirectly stimulate marrow erythropoietic activity as well as renal Epo production (105,106).

SERUM ERYTHROPOIETIN IN ANEMIA OF CHRONIC DISORDERS

Serum Epo levels have been examined in a variety of diseases associated with the anemia of chronic disorders. Rather than

producing an exhaustive list of papers encompassing the whole spectrum of diseases that have been investigated for the adequacy of Epo production, we will focus on specific examples that have been particularly well documented. HIV infection will be taken as a model of chronic infection and rheumatoid arthritis as a paradigm for chronic inflammatory diseases. We will then turn to the more complex analysis of the data in the field of cancer.

Serum Epo in HIV Infection

Anemia is a common problem in human immunodeficiency (HIV) infection, being present in 70–95% of patients with AIDS, and frequently exacerbated by therapeutic agents such as zidovudine (107,108). Severe *in vitro* inhibition of erythropoiesis and transient stimulation of granulopoiesis are observed after bone marrow infection with various HIV isolates (109). Several papers have examined the adequacy of endogenous Epo response to anemia in AIDS patients. Serum Epo levels were elevated in HIV-seronegative and HIV-seropositive asymptomatic homosexuals and in patients with lymphadenopathy, AIDS-related complex (ARC) and AIDS, but were normal in asymptomatic HIV-seronegative or HIV-seropositive intravenous drug users (110). However, no attempt was made to correlate these Epo values to Hb or Hct values. Serum Epo levels were higher in HIV-infected subjects compared to normal individuals but again no control anemic group was available for proper evaluation (111). The regression line of serum Epo vs. Hb was quite similar in asymptomatic HIV-infected and uninfected 12-month old infants (112). HIV-infected subjects with AIDS or ARC not receiving zidovudine therapy exhibited a strong inverse relationship between serum Epo and Hb, but there was no comparison with a control group (113). In a group of 82 HIV-positive subjects, 41% of whom were receiving azidothymidine antiviral therapy, the slope of the regression of serum Epo vs. Hb was less steep than in a control group of patients with iron deficiency or aplastic anemia (114). However, few HIV-infected subjects were anemic and no details are

available on the range of Hb values in the controls compared to the study subjects. There are only two papers for which the data can be fully interpreted and both indicate a blunted Epo production in patients with AIDS. Among 152 patients infected with HIV, anemia was present in 18% of asymptomatic, 50% of ARC and 75% of AIDS patients (115). The relationship between serum Epo and Hb disclosed a markedly blunted Epo response to anemia in AIDS patients compared to patients with IDA. The serum Epo-Hb relationship in a group of 42 patients with either ARC or AIDS, including 13 patients on zidovudine, closely resembled that of patients with the anemia of chronic disorders due to chronic infection, and both were considerably blunted compared to the relationship in subjects with iron deficiency (116). In addition, iron metabolism reflected a pattern of ACD with low transferrin saturation and elevated serum ferritin concentration. For any given degree of anemia, patients treated with zidovudine had significantly higher serum Epo concentration than zidovudine-naive patients (111,113,115). Indeed, the anemia associated with zidovudine therapy appeared to be due to red cell hypoplasia or aplasia (117). This occurred in the presence of elevated serum Epo values that again were not evaluated in relation to the degree of anemia in one study (117) but in another investigation even surpassed the Epo response of subjects with IDA (115). In conclusion, although the number of studies is limited, endogenous Epo response appears to be somewhat blunted in AIDS patients, but serum Epo levels are increased by zidovudine therapy.

Serum Epo in Rheumatoid Arthritis

The pathogenesis of anemia in systemic autoimmune diseases, including a possible defect in endogenous Epo production, has been reviewed elsewhere (118). In addition to the effect of cytokines on Epo-producing cells, vascular interstitial damage in the kidney peritubular cell area has been suggested as a cause of Epo deficiency in at least some of these systemic autoimmune disorders (118). There is some evidence for impaired erythropoietin response to anemia in rheumatoid disease (119).

Increa
remain
patien
the stu
Epo le
"unrel
compa
group
signifi
absent
as hav
Among
and 26
lar ave
tion wa
Withir
IDA be
serum
vated f
range
matoi
was ap
ACD a
and co
cantly
evalua
cation
signifi
contro
IDA. T
a cont
Epo re
did no
and th
anothe
deficie
its of t
or hen
tion w

Increased Epo levels were observed in RA patients that remained anemic over the years compared to nonanemic patients, but no appropriate control group was included in the study (120). In another study of 50 RA patients, serum Epo levels were slightly increased over normal values but "unrelated to low Hb concentration," but the data were not compared to an appropriate anemic control group (121). In a group of 14 anemic RA patients, serum Epo levels were significantly higher in those classified as IDA on the basis of absent iron stores in the bone marrow than in those classified as having ACD, but their Hb was a little lower as well (122). Among 58 patients with rheumatoid arthritis, 40 were anemic and 26 were classified as ACD and 14 as IDA (123). With similar average Hb values in the two groups, serum Epo concentration was slightly but not significantly higher in the IDA group. Within a group of 67 RA patients, 20 patients judged to have IDA based on reduced serum ferritin concentration had higher serum Epo levels than 24 other patients with normal or elevated ferritin concentration, while Hb values covered a similar range in the two groups (124). Among 136 patients with rheumatoid arthritis, 75 cases were anemic and a definitive cause was apparent in 65 of them (125). The majority ($n = 43$) had ACD and 15 had iron deficiency. Their Hb values were similar and correlated inversely with serum Epo, but Epo was significantly lower in those with ACD. Yet in another study, an evaluation of stainable bone marrow iron allowed the classification of 35 RA patients into ACD or IDA categories (126). A significant problem with all these studies is the absence of a control group with pure IDA instead of RA patients with IDA. The first of a few studies to compare RA patients with a control group with IDA came up with a relatively blunted Epo response to anemia in RA patients, but the control group did not have the same range of Hb values as the study group and the comparison is therefore not entirely valid (127). In another such study, serum Epo in both iron replete and iron deficient RA patients remained within the 95% confidence limits of the regression obtained in patients with iron deficiency or hemolytic anemia, but there was no clear inverse correlation with the Hb values in either group (128). However, the

rols compared
s for which the
a blunted Epo
152 patients
% of asympto-
115). The rela-
d a markedly
ents compared
ationship in a
, including 13
at of patients
onic infection,
o the relation-
-addition, iron
ow transferrin
centration. For
ith zidovudine
atration than
d, the anemia
to be due to
curred in the
gain were not
in one study
assed the Epo
sion, although
Epo response
nts, but serum

nune diseases,
roduction, has
e effect of cyto-
ial damage in
sted as a cause
temic autoim-
-impaired ery-
disease (119).

range of Hb values was obviously different in the control and RA groups, respectively. In a third study, among 97 anemic RA patients, serum Epo levels were lower in those with serum ferritin concentrations greater than 20 $\mu\text{g/L}$ despite similar Hb values (129). In addition, at comparable Hb levels, serum Epo levels in RA patients with IDA were significantly lower than in IDA controls without RA. In another report, the average serum Epo value was lower in RA patients than in IDA controls at similar average Hb (130). In a final study, the Epo response to anemia was clearly diminished in patients with RA, both iron replete and iron deficient, compared to subjects with pure IDA (131).

On the other hand, in children with systemic-onset juvenile chronic arthritis (JCA), defective iron supply for erythropoiesis rather than inadequate endogenous erythropoietin production appears to be involved in the pathogenesis of anemia (132). Neither O/P Epo ratios nor regression analysis evidenced any defect in endogenous Epo production in this group of children. Indeed, in children with systemic, oligoarticular or polyarticular JCA, serum Epo levels were similar to those of patients with iron deficiency and similar degrees of anemia, while transferrin saturation was low and serum ferritin ranged from iron deficiency to considerably elevated values (133). Whereas severe anemia associated with active systemic-onset juvenile rheumatoid arthritis can be successfully treated with rHuEpo (134), this can also be achieved with IV iron alone (135). Some response to iron has been observed in RA as well (136). In addition, treatment of chronic disease in rheumatoid arthritis with $\text{TNF-}\alpha$ blockade resulted in dose-dependent Hb increments accompanied by a reduction of serum Epo concentration that suggest that $\text{TNF-}\alpha$ directly affected bone marrow precursors rather than suppressed Epo production (137).

In conclusion, rheumatoid arthritis patients often have blunted Epo response to anemia. This is much more prominent in those patients with other biological features of ACD than in those predominantly with IDA. However, these findings are not necessarily transposable to other systemic autoimmune disorders, as, for instance, children with juvenile arthritis have normal Epo response to anemia.

SERUM ERYTHROPOIETIN IN CANCER

Initial Studies

Earlier studies suggested that the anemia of inflammation produced in rats (138) and the anemia of cancer in mice (139) were accompanied by an inappropriate erythropoietin response for the degree of anemia. Similar conclusions were derived from studies measuring serum Epo levels by bioassay in patients with anemia secondary to chronic infection or malignancy, including Hodgkin's and non-Hodgkin's lymphoma, multiple myeloma, and solid tumors (140-142). However, this was not observed in other studies of tumor-bearing rats (143) and other human investigations found that serum Epo levels were diminished relative to expected levels only in patients with infection or inflammation but not in those with malignancies (144). Similarly, normal results were derived from studies of patients with cancer of the uterine cervix (145), renal cell carcinoma (146), and disseminated lung carcinoma (147).

When radioimmunoassays became available, a study showed that, compared to controls suffering from blood losses, iron deficiency, hemolysis or pernicious anemia, patients with hematologic malignancies under treatment with chemotherapy displayed a normal relationship between hematocrit and serum Epo levels (148). Similarly, in a mouse model of experimental melanoma, serum Epo concentrations remained adequate for the degree of anemia until terminal stages of the disease when the animals became severely cachectic (149). However, in a small group of patients with miscellaneous solid tumors, the average serum Epo value was less than in IDA controls with similar average Hb (130). An important study was conducted in 81 anemic patients with solid tumors in which it was found that for any given degree of anemia serum Epo levels were lower as compared to a group of control patients with IDA (150). In addition, the expected inverse relationship between serum Epo and hemoglobin was absent, but this was due to a small group of about 10 patients with inappropriate Epo response while all others were within the normal range.

In addition only 22 patients were untreated and it was shown that the Epo response was further decreased by chemotherapy, often including cisplatin. Adequate Epo production was restored in the presence of hypoxia but the possible role of infections in some patients was not addressed. With all these limitations, this study was taken as a landmark from which it is now widely believed that Epo production is defective in patients with cancer and that this is the major cause of anemia in them. However, the picture is much less clear than that.

Studies in Patients Scheduled for rHuEpo Therapy

Several investigations have been carried out in patients starting rHuEpo therapy. However, inclusion of many patients receiving chemotherapy may yield inaccurate conclusions about the adequacy of Epo production in cancer patients (see below). For instance, the majority of 12 patients with solid tumors selected for rHuEpo therapy, several of them receiving chemotherapy, had inappropriately low serum Epo levels (151). In a large study of anemic cancer patients selected to be treated with rHuEpo, Epo levels for any Hb value were significantly lower in patients receiving cisplatin-based compared to noncisplatin chemotherapy (152,153). In another trial of rHuEpo for cisplatin-associated anemia, serum Epo levels were said to be inappropriately low for the degree of anemia and not to correlate with hemoglobin levels, but no detailed data were available to substantiate this statement (154). A large study of transfusion-dependent chemotherapy-treated patients with multiple myeloma or low-grade non-Hodgkin's lymphoma showed that half of them had inappropriate Epo levels before starting rHuEpo therapy (155). This was also the case in another study of similar patients not requiring transfusions, in which the majority of the patients were found to have inadequately low serum Epo levels before the start of rHuEpo (156). In a multicenter study of patients selected for rHuEpo therapy for nonplatinum chemotherapy-induced anemia (157), serum Epo levels correlated inversely with baseline hemoglobin and

End

app
sm
ina
in e
ing
ure
pat
the

Soli

The
wit
moc
elev
(16
dren
wit
(16
seve
the
35
due
stim
only
pati
app
(164
the
who
Epo
canc
valu
tum
wit
seru
if er
redu
(165

appeared to be inappropriate for the degree of anemia only in a small minority of the patients. Another small study found inadequate Epo production as evidenced by low O/P Epo ratio in eight patients with lymphoma or multiple myeloma receiving chemotherapy, two of whom had mild degrees of renal failure (158). However, another investigation of six similar patients has found no evidence of defective Epo secretion in these disorders (159).

Solid Tumors

There are only few studies examining untreated patients with solid tumors. Among 84 such patients, only 13 were moderately anemic, and their serum Epo levels were slightly elevated but no control group was provided for comparison (160). In a group of 20 moderately anemic or nonanemic children with various solid tumors, serum Epo did not correlate with the degree of anemia but no control group was provided (161). Among 20 women with uterine or ovarian cancer, seven were anemic and their serum Epo relationship with the hematocrit appeared somewhat blunted (162). In 35 untreated patients with lung cancer, anemia was mainly due to impaired erythroid marrow response to erythropoietin stimulation, and a defect in Epo production was operative in only few of them (163). In a large cohort of 232 cancer patients, pretreatment O/P Epo ratios were decreased, apparently indicating defective endogenous Epo production (164). However, these O/E Epo ratios are not valid because the range of Hb values in the group of patients with IDA who served to derive the expected relationship between Epo and Hb was quite different from the one observed in cancer patients, many of them having quite normal Hb values. In a large study of 56 children with miscellaneous solid tumors examined before any treatment, careful comparison with an appropriate pediatric control group showed that serum Epo levels were adequate for the degree of anemia even if erythropoiesis (as assessed by sTfR levels) was significantly reduced, although to a lesser extent than in leukemic subjects (165). Among 92 patients with cirrhosis and hepatocellular

carcinoma, 55 had anemia and 37 a normal Hb value (166). Virtually, all anemic subjects had serum Epo values in the range expected from the 95% confidence limits of iron deficiency controls, whereas only two of nonanemic subjects had inappropriately high serum Epo and polycythemia. Another investigation of 30 patients with hepatocellular carcinoma found no evidence of Epo deficiency (167).

Chronic Myeloid Disorders

The regulation of Epo production in patients with myelodysplastic syndromes (MDS) appears to be extremely variable. In a study of 14 patients, serum Epo levels were markedly elevated, and the slope of the correlation between Epo and hematocrit was similar to that reported for simple IDA (168). In another group of 46 patients, the slope of the regression was closer to that of controls with IDA than to that of controls with aplastic anemia (68). In a larger study of 75 MDS patients, there was also an overall inverse relationship between Epo levels and the degree of anemia (169). However, a wide range of Epo responses was encountered among patients with similar hemoglobin concentrations, and there were many patients with inappropriately low Epo levels as well as many others with inappropriately high values. A similar observation was made in another group of 46 patients with MDS who also had an overall inverse relationship between Epo and hemoglobin levels (170). A wide range of Epo responses between patients with similar hemoglobin concentrations was observed, with the highest values measured in those with less than 10% erythroblasts in the bone marrow. However, another investigation of 20 patients with MDS by the same group found no correlation between serum Epo concentration and total erythroid production, thereby negating any effect of the level of the erythropoietic activity on serum Epo concentration (171). The erythroid abnormality of patients with MDS was further analyzed in 19 nontransfusion-dependent patients (172). Serum Epo concentration was appropriate for the degree of anemia in 15/19 patients and was positively related to the percentage of highly fluorescent reticulocytes

but not to the a controls who ex noon, a circadi observed in MI Epo levels were bin values were this is simply a gen therapy in has been assoc Epo compared t deficiencycont regression was oxysmal noctur that, for any gi to IDA patient (176,177).

Among 61 loid metaplasia, only found in ei observed betwe as well as betw surements of er (179), 65 (180), thrombocythem cantly below no patients were g similar to that c

Leukemia and Ly

Compared to pa titers displayed bin concentrati acute leukemia multiple myeloi in 19 with chro slope of the reg loma patients, clearly had inaj

Hb value (166).
po values in the
its of iron defi-
nic subjects had
themia. Another
lular carcinoma

with myelodys-
remely variable.
re markedly ele-
n Epo and hema-
le IDA (168). In
e regression was
t of controls with
5 MDS patients,
ip between Epo
er, a wide range
tients with simi-
e many patients
as many others
observation was
MDS who also
Epo and hemo-
sponses between
entrations was
n those with less
rrow. However,
DS by the same
po concentration
ting any effect of
um Epo concen-
of patients with
usion-dependent
s appropriate for
d was positively
ent reticulocytes

but not to the absolute reticulocyte count. Contrary to normal controls who exhibit a maximum concentration in the afternoon, a circadian rhythm of serum Epo concentration is not observed in MDS patients (173). Interestingly, higher serum Epo levels were associated with poorer survival but hemoglobin values were not provided, so it cannot be excluded that this is simply an effect of more severe anemia (174). Androgen therapy in MDS (and a few aplastic anemia) patients has been associated with a significant increase in serum Epo compared to untreated patients and even more so to iron deficiency controls, although the slope of the Hct vs. Epo regression was not different (175). Finally, patients with paroxysmal nocturnal hemoglobinuria have serum Epo levels that, for any given degree of anemia, are elevated compared to IDA patients but similar to those with aplastic anemia (176,177).

Among 61 anemic patients with myelofibrosis with myeloid metaplasia, inappropriately low levels of serum Epo were only found in eight patients (178). An inverse correlation was observed between serum Epo concentration and hemoglobin as well as between the O/P Epo ratio and ferrokinetic measurements of erythropoiesis. In four separate reports of 174 (179), 65 (180), 49 (181), and 40 (182) subjects with essential thrombocythemia, serum Epo concentrations were significantly below normal levels in many patients. However, these patients were generally not anemic, and their pattern was similar to that of 343 patients with polycythemia vera (179).

Leukemia and Lymphoma

Compared to patients with iron-deficiency anemia, serum Epo titers displayed similar inverse relationships with hemoglobin concentration in separate analyses of 47 patients with acute leukemia, 54 with non-Hodgkin's lymphoma, 34 with multiple myeloma, 16 with myelofibrosis, but curiously not in 19 with chronic myelogenous leukemia (68). However, the slope of the regression was blunted in lymphoma and myeloma patients, and several patients with multiple myeloma clearly had inappropriately low serum Epo levels. The O/P

Epo ratio was similar in patients with leukemia compared to healthy controls or patients with iron-deficiency anemia, indicating that serum Epo production was appropriate for the degree of anemia.

Other reports have focused on leukemias. Twelve patients with hairy cell leukemia were found to have a normal feedback mechanism for Epo production in response to anemia, but no formal control group was presented (183). The role of Epo in chronic lymphocytic leukemia (CLL) has been reviewed (184). Among 47 patients with CLL, Epo production was found to be adequate for the degree of anemia, and this conclusion was not altered in advanced stages of the disease (185). Inappropriate Epo levels were only found in three patients, two of whom had active infections. When patients with acute leukemia were compared with patients with ulcerative colitis, serum Epo levels were found to be higher for similar degrees of anemia and somewhat less well correlated with hemoglobin (186-188). Although ulcerative colitis represents a form of chronic disorder and therefore does not appear to be an ideal control group, this result at least indicated that there was no evident Epo deficiency in patients with acute leukemia. There are some studies of children with acute leukemia, in which it was also found that serum Epo was considerably increased and inversely related to hemoglobin concentration (189,190). In a large study of 55 children with acute leukemia examined at diagnosis, careful comparison with an appropriate pediatric control group revealed that erythropoiesis (as assessed by sTfR levels) was severely depressed, but serum Epo levels were appropriate for the degree of anemia in virtually all of them (165).

Finally, several papers analyzed Epo levels in patients with lymphoid malignancies. Erythropoietin production in response to anemia was considered normal in 12 children with lymphoma, but no formal control group was presented (161). Others examined the Epo-Hb relationship in 63 untreated patients with Hodgkin's disease and found no evidence for depressed serum Epo levels, as the minority of patients who had anemia responded with adequate Epo production (191). Erythropoietin production has been more

precise
correla
impair
or radi
to be a
tion w
impair
that se
with re
normal
had de
anemic
with r
this ph
by the

Conclu

In con
definit
anemi
either
the pa
patien
chroni
anemi
myelo
sivene
Epo pi
is no
diseas
solid t

SERUM CHEM

Experi

Exper
explor

precisely evaluated in multiple myeloma (192). A negative correlation between erythropoiesis and the degree of renal impairment has been observed (193-195). Using biological or radioimmunological assays, serum Epo levels were found to be appropriate for the degree of anemia when renal function was normal but inadequate when renal function was impaired (195-198). However, in another study, it was shown that serum Epo levels were inadequate not only in patients with renal impairment but also in a number of patients with normal renal function (63). Approximately 25% of all patients had defective Epo production and this increased to 30% of anemic patients, 50% of stage 3 patients, and 60% of those with renal impairment. Plasma viscosity may contribute to this phenomenon by blunting anemia-induced Epo production by the kidney (199).

Conclusions

In conclusion, few studies have been conducted in a way that definitive conclusions can be obtained, i.e., studies in untreated anemic cancer patients with a suitable control group to provide either comparison of regressions of serum Epo vs. Hct or Hb in the patient and control groups or O/P Epo ratios in individual patients. Most studies indicate that patients with leukemia or chronic myeloid disorders have appropriate Epo responses to anemia. A significant proportion of patients with multiple myeloma and possibly lymphoma have impaired Epo responsiveness. There is little evidence for defective endogenous Epo production in patients with solid tumors. However, there is no report specifically addressing metastatic vs. localized disease. Therefore, the overall incidence of Epo deficiency in solid tumor patients remains poorly defined.

SERUM ERYTHROPOIETIN AND CHEMOTHERAPY

Experimental Data

Experiments were conducted in various animal species to explore the effect of chemotherapy and total body irradiation

on the capacity to increase Epo production in response to hypoxia. In rats exposed to hypoxia, neither cyclophosphamide nor sublethal irradiation modified Epo production significantly in the following days (200). Lethal irradiation led to anemia-driven Epo peaks that were not encountered in mice rescued by bone marrow transplantation (201). Administration of nitrogen mustard to sheep suffering from phenylhydrazine-induced hemolytic anemia produced considerably higher titers of serum Epo (202). Administration of vanadium to mice-bearing lymphoma was followed by prolonged enhanced Epo activity (203). Serum Epo levels during continuous exposure to hypoxia in mice with marrow aplasia induced by whole body irradiation or 5-fluorouracil injection were higher than in control mice similarly exposed (104).

These *in vivo* data apparently suggest an enhancing effect of chemotherapy on Epo production. As there are no preformed stores of Epo, this cannot be due to a sudden release of Epo by the kidney, mediated by cytostatic drugs. Some other speculations have been offered as explanation for this phenomenon (73). Cytotoxic therapy could cause a direct injury to the Epo-producing cells mimicking hypoxia. The blood flow to the kidney and/or liver could be altered so as to expose Epo-producing cells to hypoxia. As protein synthesis and gene transcription are necessary for the normal degradation of Epo mRNA, it is also possible that cytotoxic therapy could enhance Epo mRNA stability. However, some experimental data contradict these assumptions. The kidneys of dogs were isolated *in situ* and perfused with blood containing or not containing chlorambucil or thiotepa (204). Cobalt-induced Epo production was markedly suppressed 18–36 hr after the infusion of alkylating agents. *In vitro* studies were conducted to examine the effect of chemotherapeutic agents on Epo synthesis in cultures of the hepatoma cell line, HepG2. The RNA synthesis-inhibiting drugs daunorubicin, cyclophosphamide, ifosfamide, and CDDP, as well as the tubulin-binding agent, vincristine, dose dependently decreased production of erythropoietin. The DNA synthesis-inhibiting drugs methotrexate and cytosine-arabioside did not have inhibitory properties (205,206). Together, these

results indicate Epo production mechanisms in bone marrow.

Cisplatin side effects, in particular associated with its use as a major factor in experimental chemotherapy, inhibit the production of Epo. In a study with HepG2, which is another study on hypoxia- or chemotherapy-induced Epo production, cisplatin damage (208) in rats, hypoxia-affected in another study, and receiving a single high dose of 100 U/kg, resulted in a significantly greater regeneration.

The mechanism of Epo production in the bone marrow is affected by 5-FU-induced anemia, whereas 10, whereas and development of anemia, mainly 7–14. Although the hemoglobin level is low, the early rise in Epo is greater than anemia.

results indicate that chemotherapeutic agents may inhibit Epo production locally but that this effect is offset by other mechanisms, possibly nonutilization by a myelosuppressed bone marrow, leading to increased serum Epo levels.

Cisplatin (CDDP) is associated with a number of serious side effects, including nephrotoxicity and myelosuppression, in particular anemia of long duration (207). As cisplatin is associated with frequent and occasionally severe renal impairment, it has been speculated that Epo deficiency could be a major factor in the development of CDDP-induced anemia. Experimental data support this concept. RNA synthesis-inhibiting drugs, including CDDP, produced a dose-dependent decrease of Epo production by the human hepatoma cell line, HepG2, which partly correlated with cytotoxicity (205). In another study, CDDP also had a strong inhibitory effect on hypoxia- or cobalt-induced Epo mRNA expression and protein production in the Hep3B cell line, with no apparent cell damage (208). Five days after injection of CDDP to mice or rats, hypoxia-induced Epo production was not adversely affected in spite of severe tubular necrosis (209). However, another study reported a significant drop of serum Epo concentration and kidney Epo mRNA content in rats 4–14 days after receiving a bolus injection of cisplatin (210). Rats injected with a single high dose of CDDP developed acute renal failure and anemia that could be prevented or corrected by daily injections of 100 U/kg rHuEpo (211,212). In addition, there was a significantly greater recovery of renal function with increased tubular regeneration.

The most informative study was conducted by Matsmoto who compared the effect of 5-FU and CDDP on erythropoiesis in rats and the role of rHuEpo in this setting (213). 5-FU-induced anemia developed rapidly with a nadir at day 10, whereas the anemia caused by CDDP was less prominent and developed later with a nadir at day 21. In 5-FU-induced anemia, marked serum Epo elevation was observed at days 7–14. Although serum Epo levels correlated negatively with the hemoglobin, they fell rapidly afterwards, indicating that the early rise could be an effect of chemotherapy itself rather than anemia. This was followed by an increase of spleen but

not marrow CFU-E and a rise in reticulocytes, followed by rapid correction of the anemia. In contrast, CDDP-induced anemia was not associated with changes in serum Epo or CFU-E values. As no animal decreased its hemoglobin below 13 g/dL, it is not surprising that serum Epo levels were not elevated around day 20. On the other hand, CDDP did not produce the early release of Epo into the circulation as observed with 5-FU. These results with CDDP were confirmed in another study (214). After injection of 5-FU, treatment with rHuEpo did not prevent the fall of hemoglobin but somewhat accelerated recovery in a dose-dependent fashion (213). Anemia could be completely prevented if rHuEpo was started one week before administration of 5-FU. After CDDP treatment, rHuEpo was very effective in correcting the anemia in a dose-dependent manner, even when started only 2 weeks after CDDP had been given (213,214).

Nonplatinum Chemotherapy in Patients

Several studies have been conducted in cancer patients. In six patients receiving intensive chemotherapy for acute leukemia, serum Epo levels increased substantially after treatment and gradually returned to baseline, often at the time of marrow recovery (71). Intensive chemotherapy given for induction of acute leukemia resulted in marked elevation of serum Epo concentration starting one or two days later and peaking after about 7 days, before normalizing later on (72). High serum Epo levels are also observed transiently after intensive conditioning before bone marrow transplantation without concomitant change in hemoglobin or hematocrit (70-76). Another small study observed a large increment of serum Epo soon after the initiation of chemotherapy for leukemia, which reached values of aplastic anemia patients at similar Hb levels (215). The same group reported the repeated postchemotherapy elevation of serum Epo levels in leukemic patients, pinpointing a nice reciprocal relationship with serum iron (216), and obtained similar findings in patients with lung cancer (217). After treatment with high-dose methotrexate, serum Epo increased in some children despite

Endogenous Erythro

unchanged or after treatment Epo increased hemoglobin (189 administered a after the removal progressively increased hemoglobin level chemotherapy (Anemia on hydroxy therapy had in patients (180). with acute prom a transient increase inversely with relationship between TPO and sphamide also in vasculitis-associated of chemotherapy (221). Whole body Epo response to results powerful Epo values during

Serum Epo chemotherapy for relation to a small with insulin-like reported the evolution patients with number of chemotherapy cases (164). While anemia developed decreased until 1 However, the relationship fully maintained up at the end of nance chemotherapy levels) was further (165). Identical c

unchanged or even increased hemoglobin values, whereas after treatment with high-dose arabinoside cytosine, serum Epo increased markedly in all in response to decreasing hemoglobin (189). Similar observations were made in adults administered a 5-day course of 5-fluorouracil and leucovorin after the removal of colon cancer (218). Serum Epo levels progressively increased for 15 days in the presence of constant hemoglobin levels. Urinary Epo excretion also increases after chemotherapy (219). Patients with essential thrombocythemia on hydroxyurea, α -interferon or radioactive phosphorus therapy had increased Epo levels compared to untreated patients (180). All-trans retinoid acid treatment in patients with acute promyelocytic leukemia was also associated with a transient increase in serum Epo values that correlated inversely with reticulocyte counts, similar to the relationship between TPO and platelets (220). A single dose of cyclophosphamide also increases serum Epo levels in patients with vasculitis-associated hypertension, implying that the effect of chemotherapeutic agents is not limited to cancer patients (221). Whole body hyperthermia does not affect the serum Epo response to chemotherapy (222). Taken together, these results powerfully demonstrate a transient surge in serum Epo values during 1–2 weeks after chemotherapy.

Serum Epo levels after six cycles of non-nephrotoxic chemotherapy for stage 2 breast cancer increased slightly in relation to a small decrease in Hct and correlated negatively with insulin-like growth factor-1 (223,224). A large study reported the evolution of serum Epo and O/P ratios in 232 patients with miscellaneous tumors receiving a variable number of chemotherapy cycles, including cisplatin in 65% of the cases (164). While serum Epo increased progressively as an anemia developed in the majority of them, the O/P Epo ratio decreased until the fourth cycle and recovered at cycle 6. However, the relationship between serum Epo and Hb was fully maintained in 55 children with acute leukemia followed up at the end of induction and during the course of maintenance chemotherapy, whereas erythropoietic activity (sTfR levels) was further reduced compared to pretreatment levels (165). Identical conclusions were derived from the follow-up

of 56 children with solid tumors (165). Pediatric patients investigated at various stages of induction, consolidation, and maintenance chemotherapy for acute leukemia maintained a significant inverse correlation between serum Epo and Hb that was particularly close in those with Hb less than 10 g/dL (225). These data suggest that nonplatinum chemotherapy in general does not induce Epo deficiency in the mid- or long-term.

Cisplatin Chemotherapy in Patients

In a study of 24 patients with gynecologic malignancies, there was a significant decrease of serum Epo levels between 2 and 6 hr after chemotherapy with cisplatin and cyclophosphamide, followed by a return to baseline values after 12 hr (226). Combination chemotherapy regimens based on cisplatin (100 mg/m²) or carboplatin (300 mg/m²) were associated with the usual peak of serum Epo levels observed 1–2 weeks after chemotherapy (227,228). Plasma Epo concentration increased similarly in advanced cancer patients 15 days after chemotherapy did or did not contain cisplatin (229). In seven patients with ovarian carcinoma undergoing cisplatin chemotherapy, serum Epo was increased 24 hr and 7 days later independent of concomitant anemia (230). In another small study, serum Epo in solid tumor patients receiving cisplatin was higher than in similarly anemic patients treated without cisplatin (231). Therefore, apart from a possible very early inhibition of Epo secretion, cisplatin is no exception to the development of a serum Epo peak 1–2 weeks after chemotherapy.

In patients with gynecologic cancer receiving multiple courses of combination chemotherapy including 50 mg/m² cisplatin, prechemotherapy serum Epo values were progressively elevated in relation with the degree of anemia achieved, although a comparison with only eight anemic controls is of little value (232). Few among head and neck as well as other cancer patients receiving cisplatin (100 mg/m²)-based chemotherapy developed inappropriately low Epo levels, and there was no correlation with the amount of cisplatin administered or the degree of renal impairment (233). A

Endogenous

linear rel.
after trea
but no cor
trols was p
ovarian or
(60 mg/m²
anemia co
O/P Epo
degree of
CDDP the
despite pe
there is so
platinum-l
a universa

SERUM ER RESPONSE

Based on o
cancer, it i
underlying
are regula
rHuEpo m
erythropoi
ment of AC
ing HIV
cancer wit

Theor
produce E
those with
mia. As E
degree of a
(O/P ratio
Epo produ
cies, it has
(238) or d
significant
confirmed:

linear relationship between $\log(\text{Epo})$ and Hb was retained after treatment of 12 children with various solid tumors, but no comparison with pretreatment values or normal controls was provided (234). A longitudinal study of patients with ovarian or bladder cancer treated with nine courses of CDDP (60 mg/m^2) and doxorubicin (60 mg/m^2) showed progressive anemia correlating with renal tubular dysfunction (214). O/P Epo ratios declined progressively in proportion to the degree of renal dysfunction and recovered after cessation of CDDP therapy along with restoration of tubular function despite persistently depressed creatinine clearance. Overall, there is some evidence for Epo deficiency after completion of platinum-based chemotherapy, although this is certainly not a universal finding.

SERUM ERYTHROPOIETIN AS PREDICTOR OF RESPONSE TO rHuEpo

Based on our knowledge of the pathophysiology of the ACD and cancer, it is clear that the most useful approach is to treat the underlying disorder (2,14,15). However, red cell transfusions are regularly needed in patients with ACD. In this context, rHuEpo may be of particular value in stimulating endogenous erythropoiesis, and has now been widely tested in the treatment of ACD patients with a variety of diseases (235), including HIV infection (236), rheumatoid arthritis (237), and cancer with or without concomitant chemotherapy (155,156).

Theoretically, patients with a defect in the capacity to produce Epo would be more likely to respond to rHuEpo than those with adequate serum Epo levels for their degree of anemia. As Epo levels must be interpreted in relation to the degree of anemia, the ratio of observed-to-predicted Epo levels (O/P ratio) represents a better assessment of the adequacy of Epo production (62). In patients with hematologic malignancies, it has been observed that low baseline serum Epo levels (238) or decreased O/P ratios (158) were associated with a significantly higher probability of response. This has been confirmed in large multicenter trials in patients with multiple

patients
olidation,
ia main-
erum Epo
less than
num che-
cy in the

cies, there
een 2 and
sphamide,
2 hr (226).

cisplatin
iated with
weeks after
t increased
hemother-
n patients
notherapy,
dependent
dy, serum
higher than
latin (231).
tion of Epo
pment of a

ng multiple
 mg/m^2 cis-
re progres-
of anemia
anemic con-
eck as well
 $.00 \text{ mg/m}^2$ -
y low Epo
nt of cispla-
ent (233). A

myeloma or non-Hodgkin's lymphoma (155,156). An O/P ratio < 0.9 was found to be associated with high response rates, whereas patients with an O/P ratio > 0.9 rarely benefited from therapy (239). Studies in patients with solid tumors have failed to confirm such a consistent predictive value of baseline Epo even when Epo deficiency was demonstrated in part of the patients (151,153,154,157,240). However, a study aiming at preventing anemia in patients with ovarian carcinoma undergoing platinum-based chemotherapy showed a trend for lower transfusion needs in those with an O/P ratio < 0.8 (241). In addition, a small study in patients with a variety of solid tumors suggested that the ratio of baseline Epo/corrected reticulocyte count could provide some predictive information (242).

A combination of baseline parameters and early changes observed after 2 weeks of rHuEpo may provide another useful approach. Among evaluable patients treated in a large multicenter study (156), the failure rate was almost 90% when baseline serum O/P Epo was higher than 0.9 or when serum O/P Epo was less than 0.9 but the hemoglobin increment by week 2 was < 0.3 g/dL. On the other hand, the success rate was around 90% when baseline serum O/P Epo was less than 0.9 and hemoglobin increased by ≥ 0.3 g/dL. Similar findings were obtained in a smaller study in children with solid tumors: an O/P ratio < 1.0 at baseline and a hemoglobin increment > 0.5 g/dL after 2 weeks were associated with higher response rates (243). In another large single center study (239), the combined use of baseline serum Epo and the 2-week increment of sTfR proved to be very powerful. Only 18% of patients with a baseline serum Epo greater than 100 mU/mL responded to treatment, and only 29% responded when the baseline serum Epo was < 100 mU/mL but the 2-week sTfR increment was less than 25%. On the other hand, the response rate was 96% among patients with a low baseline serum Epo and a substantial sTfR elevation.

In conclusion, baseline serum Epo should be measured at baseline in patients with hematologic malignancies: treatment should not be initiated if endogenous serum Epo is above 100 mU/mL (or 200 mU/ml in severely anemic patients) or the

Endogenous Erythropo.

O/P ratio is > 0.9 . be those combinin, ous Epo producti together with som response (changes patients treated v evaluated just pri be valid. Indeed, Epo may be inap chemotherapy cor probably because zation by target excluded that the patients may just l Epo sampling. Wh tion may be relev interest in subject to prevent an anem better tumor oxyge fetal hemoglobin i universal Epo defic

ACKNOWLEDGMENT

This work was supported by the National Research Fund for Scientific Research, FNRS.

REFERENCES

1. Cartwright GL. Hematol 1966;
2. Beguin Y. Eryt Belg 1996; 51:3
3. Means RT Jr, pathogenesis o 80:1639-1647.

O/P ratio is >0.9 . In addition, the best algorithms appear to be those combining an assessment of the adequacy of endogenous Epo production (at least in hematologic malignancies) together with some early indicators of erythropoietic marrow response (changes in hemoglobin or sTfR). Of importance, in patients treated with chemotherapy, serum Epo should be evaluated just prior to chemotherapy for its interpretation to be valid. Indeed, without any change in hematocrit, serum Epo may be inappropriately elevated in the 2 weeks after chemotherapy compared to prechemotherapy values, most probably because myelosuppression then decreases Epo utilization by target cells (see above). Therefore, it cannot be excluded that the failure to predict response in solid tumor patients may just be related to an inadequate timing of serum Epo sampling. While evaluation of endogenous Epo production may be relevant in various forms of anemia, it is of no interest in subjects in whom the aim of rHuEpo therapy is to prevent an anemia that is not yet present, in those in whom better tumor oxygenation before radiotherapy or induction of fetal hemoglobin is sought, or in disorders characterized by universal Epo deficiency.

ACKNOWLEDGMENTS

This work was supported in part by grants from the National Fund for Scientific Research (Fonds National de la Recherche Scientifique, FNRS), Belgium.

REFERENCES

1. Cartwright GE. The anemia of chronic disorders. *Semin Hematol* 1966; 3:351-375.
2. Beguin Y. Erythropoietin and the anemia of cancer. *Acta Clin Belg* 1996; 51:36-52.
3. Means RT Jr, Krantz SB. Progress in understanding the pathogenesis of the anemia of chronic disease. *Blood* 1992; 80:1639-1647.

4. Means RT Jr. Advances in the anemia of chronic disease. *Int J Hematol* 1999; 70:7-12.
5. Weiss G. Pathogenesis and treatment of anaemia of chronic disease. *Blood Rev* 2002; 16:87-96.
6. Lee GR. The anemia of chronic disease. *Semin Hematol* 1983; 20:61-80.
7. Sears D. Anemia of chronic disease. *Med Clin North Am* 1992; 76:567-579.
8. Torti FM, Torti SV. Cytokines, iron homeostasis, and cancer. *Adv Exp Med Biol* 1994; 354:161-170.
9. Konijn AM. Iron metabolism in inflammation. *Clin Haematol* 1994; 7:829-849.
10. Weiss G, Wachter H, Fuchs D. Linkage of cell-mediated immunity to iron metabolism. *Immunol Today* 1995; 16:495-500.
11. Weiss G. Iron and anemia of chronic disease. *Kidney Int Suppl* 1999; 69:S12-S17.
12. Means RT Jr. Pathogenesis of the anemia of chronic disease: a cytokine-mediated anemia. *Stem Cells* 1995; 13:32-37.
13. Trey JE, Kushner I. The acute phase response and the hematopoietic system: the role of cytokines. *Crit Rev Oncol Hematol* 1995; 21:1-18.
14. Spivak JL. Cancer-related anemia: its causes and characteristics. *Semin Oncol* 1994; 21(suppl 3):3-8.
15. Moliterno AR, Spivak JL. Anemia of cancer. *Hematol Oncol Clin North Am* 1996; 10:345-363.
16. Doll DC, Weiss RB. Neoplasia and the erythron. *J Clin Oncol* 1985; 3:429-446.
17. Tchekmedyian NS. Anemia in cancer patients: significance, epidemiology, and current therapy. *Oncology (Huntingt)* 2002; 16:17-24.
18. Groopman JE, Itri LM. Chemotherapy-induced anemia in adults: incidence and treatment. *J Natl Cancer Inst* 1999; 91:1616-1634.

19. Micho
Euro
Oncol
20. Hedde
D, Ro
cell tr
under
86:239
21. Skillir
of red
chemo
22. Benois
tive fa
resecti
212 pa
23. Hensle
R, Sab
tion of
chemo
485-48
24. Cassid
Robsor
with ch
thropo:
9:775-
25. Mara I
fova A
metabo
postpa:
142-14
26. Faquin
tory cy
Blood
27. Vannu
Rossi-F
vitro b
87:18-

19. Michon J. Incidence of anemia in pediatric cancer patients in Europe: results of a large, international survey. *Med Pediatr Oncol* 2002; 39:448-450.
20. Heddens D, Alberts DS, Hannigan EV, Williams SD, Garcia D, Roe DJ, Bell J, Alvarez RD. Prediction of the need for red cell transfusion in newly diagnosed ovarian cancer patients undergoing platinum-based treatment. *Gynecol Oncol* 2002; 86:239-243.
21. Skillings JR, Sridhar FG, Wong C, Paddock L. The frequency of red cell transfusion for anemia in patients receiving chemotherapy. *Am J Clin Oncol* 1993; 16:22-25.
22. Benoist S, Panis Y, Pannegeon V, Alves A, Valleur P. Predictive factors for perioperative blood transfusions in rectal resection for cancer: a multivariate analysis of a group of 212 patients. *Surgery* 2001; 129:433-439.
23. Hensley ML, Lebeau D, Leon LF, Venkatraman E, Waltzman R, Sabbatini P, Almadrone L, Chi D, Spriggs D. Identification of risk factors for requiring transfusion during front-line chemotherapy for ovarian cancer. *Gynecol Oncol* 2001; 81:485-489.
24. Cassidy MJD, De Jager C, Ebrahim O, Camachio P, Robson R. Peripheral blood mononuclear cells from patients with chronic renal failure release factors which suppress erythropoietin secretion in vitro. *Nephrol Dial Transplant* 1994; 9:775-779.
25. Mara M, Zivny J, Eretova V, Kvasnicka J, Kuzel D, Umlaufova A, Marova E. Changes in markers of anemia and iron metabolism and how they are influenced by antianemics in postpartum period. *Acta Obstet Gynecol Scand* 2001; 80:142-148.
26. Faquin WC, Schneider TJ, Goldberg MA. Effect of inflammatory cytokines on hypoxia-induced erythropoietin production. *Blood* 1992; 79:1987-1994.
27. Vannucchi AM, Grossi A, Rafanelli D, Statello M, Cinotti S, Rossi-Ferrini P. Inhibition of erythropoietin production in vitro by human interferon gamma. *Br J Haematol* 1994; 87:18-23.

28. Fandrey J, Jelkmann WE. Interleukin-1 and tumor necrosis factor-alpha inhibit erythropoietin production in vitro. *Ann N Y Acad Sci* 1991; 628:250-255.
29. Jelkmann W, Pagel H, Wolff M, Fandrey J. Monokines inhibiting erythropoietin production in human hepatoma cultures and in isolated perfused rat kidneys. *Life Sci* 1992; 50:301-308.
30. Jelkmann W, Wolff M, Fandrey J. Modulation of the production of erythropoietin by cytokines: in vitro studies and their clinical implications. *Contrib Nephrol* 1990; 87:68-77.
31. Fandrey J, Huwiler A, Frede S, Pfeilschifter J, Jelkmann W. Distinct signaling pathways mediate phorbol-ester-induced and cytokine-induced inhibition of erythropoietin gene expression. *Eur J Biochem* 1994; 226:335-340.
32. Jelkmann W, Hellwig-Buergel T. Tumor necrosis factor p55 receptor (TNF-RI) mediates the in vitro inhibition of hepatic erythropoietin production. *Exp Hematol* 1999; 27:224-228.
33. Hellwig-Burgel T, Rutkowski K, Metzen E, Fandrey J, Jelkmann W. Interleukin-1 beta and tumor necrosis factor-alpha stimulate DNA binding of hypoxia-inducible factor-1. *Blood* 1999; 94:1561-1567.
34. Schobersberger W, Hoffmann G, Fandrey J. Nitric oxide donors suppress erythropoietin production in vitro. *Pflugers Arch* 1996; 432:980-985.
35. Fandrey J, Frede S, Jelkmann W. Role of hydrogen peroxide in hypoxia-induced erythropoietin production. *Biochem J* 1994; 303:507-510.
36. Fandrey J, Frede S, Ehleben W, Porwol T, Acker H, Jelkmann W. Cobalt chloride and desferrioxamine antagonize the inhibition of erythropoietin production by reactive oxygen species. *Kidney Int* 1997; 51:492-496.
37. Jelkmann W, Pagel H, Hellwig T, Fandrey J. Effects of antioxidant vitamins on renal and hepatic erythropoietin production. *Kidney Int* 1997; 51:497-501.
38. Leng HM, Kidson SH, Keraan MM, Randall GW, Folb PI. Cytokine-mediated inhibition of erythropoietin synthesis by dexamethasone. *J Pharm Pharmacol* 1996; 48:971-974.

39. Frede S, Jelkmann W. Erythropoietin production in vitro. *Ann N Y Acad Sci* 1991; 628:250-255.
40. Chung J, Coffey J. Erythropoietin production in vitro. *Life Sci* 1992; 50:301-308.
41. Miller J. Erythropoietin production in vitro. *Contrib Nephrol* 1990; 87:68-77.
42. Niekerk J, Piers J. Erythropoietin production in vitro. *Eur J Biochem* 1994; 226:335-340.
43. Oczko J, Hellwig-Buergel T. Erythropoietin production in vitro. *Exp Hematol* 1999; 27:224-228.
44. Johnson J, Furman S. Erythropoietin production in vitro. *Blood* 1999; 94:1561-1567.
45. Johnson J, Furman S. Erythropoietin production in vitro. *Pflugers Arch* 1996; 432:980-985.
46. Johnson J, Furman S. Erythropoietin production in vitro. *Biochem J* 1994; 303:507-510.
47. Johnson J, Furman S. Erythropoietin production in vitro. *Kidney Int* 1997; 51:492-496.

39. Frede S, Fandrey J, Pagel H, Hellwig T, Jelkmann W. Erythropoietin gene expression is suppressed after lipopolysaccharide or interleukin-1 beta injections in rats. *Am J Physiol* 1997; 273:R1067-R1071.
40. Chuncharunee S, Carter CD, Studtmann KE, Caro J, Coffey RJ, Dessypris EN. Chronic administration of transforming growth factor-beta suppresses erythropoietin-dependent erythropoiesis and induces tumour necrosis factor in vivo. *Br J Haematol* 1993; 84:374-380.
41. Miller KL, Carlino JA, Ogawa Y, Avis PD, Carroll KG. Alterations in erythropoiesis in TGF-beta 1-treated mice. *Exp Hematol* 1992; 20:951-956.
42. Nieken J, Mulder NH, Buter J, Vellenga E, Limburg PC, Piers DA, de Vries EG. Recombinant human interleukin-6 induces a rapid and reversible anemia in cancer patients. *Blood* 1995; 86:900-905.
43. Oczko-Grzesik B, Wiecek A, Kokot F. Influence of IFN-alpha on plasma erythropoietin levels in patients with hepatitis B virus-associated chronic active hepatitis. *J Interferon Cytokine Res* 2001; 21:669-676.
44. Johnson CS, Keckler DJ, Topper MI, Braunschweiger PG, Furmanski P. In vivo hematopoietic effects of recombinant interleukin-1 alpha in mice: stimulation of granulocytic, monocytic, megakaryocytic, and early erythroid progenitors, suppression of late-stage erythropoiesis, and reversal of erythroid suppression with erythropoietin. *Blood* 1989; 73:678-683.
45. Furmanski P, Johnson CS. Macrophage control of normal and leukemic erythropoiesis: identification of the macrophage-derived erythroid suppressing activity as interleukin-1 and the mediator of its in vivo action as tumor necrosis factor. *Blood* 1990; 75:2328-2334.
46. de Haan G, Dontje B, Loeffler M, Nijhof W. Microenvironmentally dependent effects on murine haematopoiesis by a prolonged interleukin-1 treatment. *Br J Haematol* 1993; 85:15-19.
47. Johnson CS, Cook CA, Furmanski P. In vivo suppression of erythropoiesis by tumor necrosis factor-alpha (TNF-alpha):

- reversal with exogenous erythropoietin (EPO). *Exp Hematol* 1990; 18:109-113.
48. Clibon U, Bonewald L, Caro J, Roodman GD. Erythropoietin fails to reverse the anemia in mice continuously exposed to tumor necrosis factor-alpha in vivo. *Exp Hematol* 1990; 18:438-441.
 49. Roodman GD, Johnson RA, Clibon U. Tumor necrosis factor alpha and the anemia of chronic disease: effects of chronic exposure to TNF on erythropoiesis in vivo. *Adv Exp Med Biol* 1989; 271:185-196.
 50. Jelkmann W. Erythropoietin: structure, control of production, and function. *Physiol Rev* 1992; 72:449-489.
 51. Barosi G. Inadequate erythropoietin response to anemia: definition and clinical relevance. *Ann Hematol* 1994; 68:215-223.
 52. Bozzini CE, Alippi RM, Barcelo AC. Enhanced effect of increased erythrocyte production rate on plasma erythropoietin levels of mice during subsequent exposure to hypobaria. *Adv Exp Med Biol* 1989; 271:23-27.
 53. Alippi RM, Boyer P, Leal T, Barcelo AC, Martinez MP, Bozzini CE. Higher erythropoietin secretion in response to cobaltous chloride in post-hypoxic than in hypertransfused polycythemic mice. *Haematologica* 1992; 77:446-449.
 54. Alippi RM, Barcelo AC, Bozzini CE. Enhanced erythropoiesis induced by hypoxia in hypertransfused, post-hypoxic mice. *Exp Hematol* 1983; 11:878-883.
 55. Alippi RM, Barcelo AC, Bozzini CE. Erythropoietic response to hypoxia in mice with polycythemia induced by hypoxia or transfusion. *Exp Hematol* 1983; 11:122-128.
 56. Alippi RM, Barcelo AC, Bozzini CE. Comparison of erythropoietic response to erythropoietin-secreting stimuli in mice following polycythemia induced by transfusion or hypoxia. *Exp Hematol* 1985; 13:159-162.
 57. Alippi RM, Barcelo AC, Kofoed JA, Bozzini CE. Hypoxia-induced renal and extrarenal erythropoietin production in posthypoxic or hypertransfused polycythemic rats. *Exp Hematol* 1986; 14:329-332.

58. Erslev AJ. 1339-1344.
59. Garcia JF, El Cronkite EP. ing levels in 1 Clin Med 198.
60. Caro J, Brown tin levels in u Med 1979; 93:
61. Cotes PM, De Pearson TC, R erythropoietin Med 1986; 31:
62. Beguin Y, Cle assessment of anemia based and erythropo
63. Beguin Y, Yerr in multiple m inappropriate 1992; 82:648-6
64. Alexanian R. E and hemolytic
65. Bray GL, Tayl poietin respons erythroblastop 120:528-532.
66. Schrezenmeier Heimpel H, K transferrin rec tol 1994; 88:28
67. de Klerk G, I Serum erythro 58:1164-1170.
68. Urabe A, Mitar shi Y, Takaku

Hematol

ropoietin
posed to
tol 1990;

sis factor
f chronic
Med Biol

f produc-

mia: defi-
:215-223.

effect of
thropoie-
yopbaria.

MP, Boz-
to cobal-
ansfused
9.

ropoiesis
xic mice.

response
ypoxia or

f erythro-
i in mice
hypoxia.

Hypoxia-
uction in
ats. Exp

58. Erslev AJ. Erythropoietin. *N Engl J Med* 1991; 324: 1339-1344.
59. Garcia JF, Ebbe SN, Hollander L, Cutting HO, Miller MO, Cronkite EP. Radioimmunoassay of erythropoietin: circulating levels in normal and polycythemic human beings. *J Lab Clin Med* 1982; 99:624-635.
60. Caro J, Brown S, Miller O, Murray T, Erslev AJ. Erythropoietin levels in uremic nephric and anephric patients. *J Lab Clin Med* 1979; 93:449-458.
61. Cotes PM, Doré CJ, Liu Yin JA, Lewis SM, Messinezy M, Pearson TC, Reid C. Determination of serum immunoreactive erythropoietin in the investigation of erythrocytosis. *N Engl J Med* 1986; 315:283-287.
62. Beguin Y, Clemons G, Pootrakul P, Fillet G. Quantitative assessment of erythropoiesis and functional classification of anemia based on measurements of serum transferrin receptor and erythropoietin. *Blood* 1993; 81:1067-1076.
63. Beguin Y, Yerna M, Loo M, Weber M, Fillet G. Erythropoiesis in multiple myeloma: defective red cell production due to inappropriate erythropoietin production. *Br J Haematol* 1992; 82:648-653.
64. Alexanian R. Erythropoietin excretion in bone marrow failure and hemolytic anemia. *J Lab Clin Med* 1973; 82:438-445.
65. Bray GL, Taylor B, O'Donnell R. Comparison of the erythropoietin response in children with aplastic anemia, transient erythroblastopenia, and iron deficiency. *J Pediatr* 1992; 120:528-532.
66. Schrezenmeier H, Noe G, Raghavachar A, Rich IN, Heimpel H, Kubanek B. Serum erythropoietin and serum transferrin receptor levels in aplastic anaemia. *Br J Haematol* 1994; 88:286-294.
67. de Klerk G, Rosengarten PCJ, Vet RJWM, Goudsmit R. Serum erythropoietin (ESF) titers in anemia. *Blood* 1981; 58:1164-1170.
68. Urabe A, Mitani K, Yoshinaga K, Iki S, Yagisawa M, Ohbayashi Y, Takaku F. Serum erythropoietin titers in hematologi-

- cal malignancies and related diseases. *Int J Cell Cloning* 1992; 10:333-337.
69. Gaines Das RE, Milne A, Rowley M, Smith EC, Cotes PM. Serum immunoreactive erythropoietin in patients with idiopathic aplastic and Fanconi's anaemias. *Br J Haematol* 1992; 82:601-607.
70. Cazzola M, Guarnone R, Cerani P, Centenara E, Rovati A, Beguin Y. Red blood cell precursor mass as an independent determinant of serum erythropoietin level. *Blood* 1998; 91:2139-2145.
71. Piroso E, Erslev AJ, Caro J. Inappropriate increase in erythropoietin titers during chemotherapy. *Am J Hematol* 1989; 32:248-254.
72. Birgegard G, Wide L, Simonsson B. Marked erythropoietin increase before fall in Hb after treatment with cytostatic drugs suggests mechanism other than anaemia for stimulation. *Br J Haematol* 1989; 72:462-466.
73. Schapira L, Antin JH, Ransil BJ, Antman KH, Eder JP, McGarigle CJ, Goldberg MA. Serum erythropoietin levels in patients receiving intensive chemotherapy and radiotherapy. *Blood* 1990; 76:2354-2359.
74. Beguin Y, Clemons GK, Oris R, Fillet G. Circulating erythropoietin levels after bone marrow transplantation: inappropriate response to anemia in allogeneic transplants. *Blood* 1991; 77:868-873.
75. Beguin Y, Oris R, Fillet G. Dynamics of erythropoietic recovery after bone marrow transplantation: role of marrow proliferative capacity and erythropoietin production in autologous versus allogeneic transplants. *Bone Marrow Transplant* 1993; 11:285-292.
76. Beguin Y, Baron F, Fillet G. Influence of marrow erythropoietic activity on serum erythropoietin levels after autologous hematopoietic stem cell transplantation. *Haematologica* 1998; 83:1076-1081.
77. Kendall RG, Cavill I, Norfolk DR. Erythropoietin consumption during stimulated erythropoiesis. *Ann N Y Acad Sci* 1994; 718:350-352.

78. Finne
excre
Scand
79. Kend
levels
630-6
80. Youss
ish HF
tin rec
81. Bowen
lowing
372-37
82. Stohlr
esis. R
row ac
83. Naets
tion of
84. Bozzini
row on
in dogs
85. Steinbe
poietin
67:646-
86. Widnes
Kisthar
kinetics
hepatec
1205-12
87. Spivak
nant h
90-99.
88. Spivak
in the re
89. Piroso E
span in
Am J He

78. Finne PH, Skoglund R, Wetterhus S. Urinary erythropoietin excretion during initial treatment of pernicious anaemia. *Scand J Haematol* 1973; 10:62-68.
79. Kendall RG, Cavill I, Norfolk DR. Serum erythropoietin levels during haematinic therapy. *Br J Haematol* 1992; 81: 630-631.
80. Youssoufian H, Longmore G, Neumann D, Yoshimura A, Lodish HF. Structure, function, and activation of the erythropoietin receptor. *Blood* 1993; 81:2223-2236.
81. Bowen DT, Janowska-Wieczorek A. Serum erythropoietin following cytostatic therapy [letter]. *Br J Haematol* 1990; 74: 372-373.
82. Stohlman F Jr, Brecher G. Humoral regulation of erythropoiesis. Relationship of plasma erythropoietin level to bone marrow activity. *Proc Soc Exp Biol Med* 1959; 100:40-43.
83. Naets JP, Wittek M. Effect of erythroid hypoplasia on utilization of erythropoietin. *Nature* 1965; 206:726-727.
84. Bozzini CE. Influence of erythroid activity of the bone marrow on the plasma disappearance of injected erythropoietin in dogs. *Nature* 1966; 209:1140-1141.
85. Steinberg SE, Garcia JF, Matzke GR, Mladenovic J. Erythropoietin kinetics in rats: generation and clearance. *Blood* 1986; 67:646-649.
86. Widness JA, Veng-Pedersen P, Schmidt RL, Lowe LS, Kisthard JA, Peters C. In vivo ¹²⁵I-erythropoietin pharmacokinetics are unchanged after anesthesia, nephrectomy and hepatectomy in sheep. *J Pharmacol Exp Ther* 1996; 279: 1205-1210.
87. Spivak JL, Hogans BB. The in vivo metabolism of recombinant human erythropoietin in the rat. *Blood* 1989; 73: 90-99.
88. Spivak JL. Metabolism of recombinant human erythropoietin in the rat. *Adv Exp Med Biol* 1989; 271:29-38.
89. Piroso E, Erslev AJ, Flaharty KK, Caro J. Erythropoietin life span in rats with hypoplastic and hyperplastic bone marrows. *Am J Hematol* 1991; 36:105-110.

J Cell Cloning
 EC, Cotes PM.
 ents with idio-
 r J Haematol
 a E, Rovati A,
 n independent
 blood 1998; 91:
 in erythropeie-
 89; 32:248-254.
 erythropoietin
 with cytostatic
 ia for stimula-
 KH, Eder JP,
 oietin levels in
 l radiotherapy.
 ating erythro-
 on: inappropri-
 ts. *Blood* 1991;
 opoietic recov-
 marrow prolif-
 in autologous
 w *Transplant*
 w erythropeie-
 er autologous
 Haematologica
 etin consump-
 V Y Acad Sci

90. Lezon CE, Martinez MP, Conti MI, Bozzini CE. Plasma disappearance of exogenous erythropoietin in mice under different experimental conditions. *Endocrine* 1998; 8:331-333.
91. Flaharty KK, Caro J, Erslev A, Whalen JJ, Morris EM, Bjornsson TD, Vlasses PH. Pharmacokinetics and erythropoietic response to human recombinant erythropoietin in healthy men. *Clin Pharmacol Ther* 1990; 47:557-564.
92. Cheung WK, Goon BL, Guilfoyle MC, Wacholtz MC. Pharmacokinetics and pharmacodynamics of recombinant human erythropoietin after single and multiple subcutaneous doses to healthy subjects. *Clin Pharmacol Ther* 1998; 64:412-423.
93. Kato M, Kamiyama H, Okazaki A, Kumaki K, Kato Y, Sugiyama Y. Mechanism for the nonlinear pharmacokinetics of erythropoietin in rats. *J Pharmacol Exp Ther* 1997; 283:520-527.
94. Chapel SH, Veng-Pedersen P, Schmidt RL, Widness JA. Receptor-based model accounts for phlebotomy-induced changes in erythropoietin pharmacokinetics. *Exp Hematol* 2001; 29:425-431.
95. Kampf D, Eckardt KU, Fischer HC, Schmalisch C, Ehmer B, Schostak M. Pharmacokinetics of recombinant human erythropoietin in dialysis patients after single and multiple subcutaneous administrations. *Nephron* 1992; 61:393-398.
96. McMahon FG, Vargas R, Ryan M, Jain AK, Abels RI, Perry B, Smith IL. Pharmacokinetics and effects of recombinant human erythropoietin after intravenous and subcutaneous injections in healthy volunteers. *Blood* 1990; 76:1718-1722.
97. Sans T, Joven J, Vilella E, Masdeu G, Farre M. Pharmacokinetics of several subcutaneous doses of erythropoietin: potential implications for blood transfusion. *Clin Exp Pharmacol Physiol* 2000; 27:179-184.
98. Kinoshita H, Ohishi N, Kato M, Tokura S, Okazaki A. Distribution of recombinant human erythropoietin following multiple intravenous administration and effects of age on the distribution in rats. *Arzneimittelforschung* 1992; 42:579-584.
99. Chapel S, Veng-Pedersen P, Hohl RJ, Schmidt RL, McGuire EM, Widness JA. Changes in erythropoietin

- phar
ablat
eryth
Ther
100. Piron
Cessa
eryth
2001;
101. Bozzir
Lezon
and e
718:83
102. Lezon
Bozzin
tion in
1995; 8
103. Bozzini
Bozzini
thropoi
affectin
1997; 5:
104. Barcelo
hypoxia
tin indu
entia 19
105. Bergama
Erythrop
potential
Exp Hem
106. Erslev A
and eryth
107. Means R
ficiency v
179-186.
108. Moyle G
marker a
13-20.

- pharmacokinetics following busulfan-induced bone marrow ablation in sheep: evidence for bone marrow as a major erythropoietin elimination pathway. *J Pharmacol Exp Ther* 2001; 298:820-824.
100. Piron M, Loo M, Gothot A, Tassin F, Fillet G, Beguin Y. Cessation of intensive treatment with recombinant human erythropoietin is followed by secondary anemia. *Blood* 2001; 97:442-448.
 101. Bozzini CE, Alippi RM, Barcelo AC, Conti MI, Bozzini C, Lezon CE, Olivera MI. The biology of stress erythropoiesis and erythropoietin production. *Ann N Y Acad Sci* 1994; 718:83-92.
 102. Lezon C, Alippi RM, Barcelo AC, Martinez MP, Conti MI, Bozzini CE. Depression of stimulated erythropoietin production in mice with enhanced erythropoiesis. *Haematologica* 1995; 80:491-494.
 103. Bozzini CE, Barcelo AC, Conti MI, Martinez MP, Lezon CE, Bozzini C, Alippi RM. Unexpected hypoxia-dependent erythropoietin secretion during experimental conditions not affecting tissue oxygen supply/demand ratio. *Kidney Int* 1997; 51:413-415.
 104. Barcelo AC, Bozzini CE. Erythropoietin formation during hypoxia in mice with impaired responsiveness to erythropoietin induced by irradiation or 5-fluorouracil injection. *Experientia* 1982; 38:504-505.
 105. Bergamaschi G, Recalde HH, Ponchio L, Rosti V, Cazzola M. Erythrophagocytosis increases the expression of erythroid potentiating activity mRNA in human monocyte-macrophages. *Exp Hematol* 1993; 21:70-73.
 106. Erslev AJ. The effect of hemolysates on red cell production and erythropoietin release. *J Lab Clin Med* 1971; 78:1-7.
 107. Means RT Jr. Cytokines and anaemia in human immunodeficiency virus infection. *Cytokines Cell Mol Ther* 1997; 3: 179-186.
 108. Moyle G. Anaemia in persons with HIV infection: prognostic marker and contributor to morbidity. *AIDS Rev* 2002; 4: 13-20.

109. Calenda V, Chermann JC. Severe in vitro inhibition of erythropoiesis and transient stimulation of granulopoiesis after bone-marrow infection with eight different HIV-2 isolates. *AIDS* 1992; 6:943-948.
110. Reddy MM, Grieco MH. Erythropoietin and granulocyte-macrophage colony-stimulating factor (GM-CSF) levels in sera of patients with HIV infection. *Int J STD AIDS* 1991; 2:128-132.
111. Allen UD, King SM, Gomez MP, Lapointe N, Forbes JC, Thorne A, Kirby MA, Bowker J, Raboud J, Singer J, Mukwaya G, Tobin J, Read SE. Serum immunoreactive erythropoietin levels and associated factors amongst HIV-infected children. *AIDS* 1998; 12:1785-1791.
112. Semba RD, Broadhead R, Taha TE, Totin D, Ricks MO, Kumwenda N. Erythropoietin response to anemia among human immunodeficiency virus-infected infants in Malawi. *Haematologica* 2001; 86:1221-1222.
113. Rarick MU, Loureiro C, Groshen S, Sullivan Halley J, Gill PS, Bernstein Singer M, Levine AM. Serum erythropoietin titers in patients with human immunodeficiency virus (HIV) infection and anemia. *J Acquir Immune Defic Syndr* 1991; 4:593-597.
114. Kreuzer KA, Rockstroh JK, Jelkmann W, Theisen A, Spengler U, Sauerbruch T. Inadequate erythropoietin response to anaemia in HIV patients: relationship to serum levels of tumour necrosis factor-alpha, interleukin-6 and their soluble receptors. *Br J Haematol* 1997; 96:235-239.
115. Spivak JL, Barnes DC, Fuchs E, Quinn TC. Serum immunoreactive erythropoietin in HIV-infected patients. *JAMA* 1989; 261:3104-3107.
116. Camacho J, Poveda F, Zamorano AF, Valencia ME, Vazquez JJ, Arnalich F. Serum erythropoietin levels in anaemic patients with advanced human immunodeficiency virus infection. *Br J Haematol* 1992; 82:608-614.
117. Walker RE, Parker RI, Kovacs JA, Masur H, Lane HC, Carleton S, Kirk LE, Gralnik HR, Fauci AS. Anemia and erythropoiesis in patients with the acquired immunodeficiency

- syndr
dine.
118. Berter
in sy
82:375
119. Vreug
anaer
Rheum
120. Graud
of eryt
norma
Scand
121. Chijiw.
recept
correl
2001; 2
122. Vreug
mia in
and fol
Ann Rl
123. Giorda
Morozz
vitamin
rheum
201-20
124. Takash
erythro
binant
J Rheu
125. Remack
Diaz C,
arthriti
1687-16
126. Nielsen
Hansen
arthriti

syndrome (AIDS) and Kaposi sarcoma treated with zidovudine. *Ann Intern Med* 1988; 108:372-376.

118. Bertero MT, Caligaris-Cappio F. Anemia of chronic disorders in systemic autoimmune diseases. *Haematologica* 1997; 82:375-381.
119. Vreugdenhil G, Swaak AJ. The role of erythropoietin in the anaemia of chronic disease in rheumatoid arthritis. *Clin Rheumatol* 1990; 9:22-27.
120. Graudal N, Nielsen OJ, Galloe AM, Graudal HN. The course of erythropoietin in patients with rheumatoid arthritis with normal and low blood-hemoglobin. A longitudinal study. *Scand J Rheumatol* 1993; 22:86-89.
121. Chijiwa T, Nishiya K, Hashimoto K. Serum transferrin receptor levels in patients with rheumatoid arthritis are correlated with indicators for anaemia. *Clin Rheumatol* 2001; 20:307-313.
122. Vreugdenhil G, Wognum AW, van Eijk HG, Swaak AJ. Anaemia in rheumatoid arthritis: the role of iron, vitamin B12, and folic acid deficiency, and erythropoietin responsiveness. *Ann Rheum Dis* 1990; 49:93-98.
123. Giordano N, Cecconami L, Marcucci P, Battisti E, Magaro L, Morozzi G, Mariano A, Marcolongo R. The role of iron, vitamin B12, folic acid and erythropoietin in the anemia of rheumatoid arthritis. *Clin Exp Rheumatol* 1992; 10: 201-202.
124. Takashina N, Kondo H, Kashiwazaki S. Suppressed serum erythropoietin response to anemia and the efficacy of recombinant erythropoietin in the anemia of rheumatoid arthritis. *J Rheumatol* 1990; 17:885-887.
125. Remacha AF, Rodriguez de la Serna A, Garcia Die F, Geli C, Diaz C, Gimferrer E. Erythroid abnormalities in rheumatoid arthritis: the role of erythropoietin. *J Rheumatol* 1992; 19: 1687-1691.
126. Nielsen OJ, Andersen LS, Ludwigsen E, Bouchelouche P, Hansen TM, Birgens H, Hansen NE. Anaemia of rheumatoid arthritis: serum erythropoietin concentrations and red cell

hibition of
ulopoiesis
at HIV-2

anulocyte-
levels in
IDS 1991;

orbes JC,
r J, Muk-
e erythro-
V-infected

icks MO,
ia among
n Malawi.

Halley J,
ythropoie-
ency virus
efic Syndr

heisen A,
thropoietin
p to serum
kin-6 and
35-239.

immunor-
AMA 1989;

encia ME,
els in anae-
iency virus

ie HC, Car-
nd erythro-
iodeficiency

- distribution width in relation to iron status. *Ann Rheum Dis* 1990; 49:349-353.
127. Hochberg MC, Arnold CM, Hogans BB, Spivak JL. Serum immunoreactive erythropoietin in rheumatoid arthritis: impaired response to anemia. *Arthritis Rheum* 1988; 31: 1318-1321.
 128. Noe G, Augustin J, Hausdorf S, Rich IN, Kubanek B. Serum erythropoietin and transferrin receptor levels in patients with rheumatoid arthritis. *Clin Exp Rheumatol* 1995; 13: 445-451.
 129. Boyd HK, Lappin TR, Bell AL. Evidence for impaired erythropoietin response to anaemia in rheumatoid disease. *Br J Rheumatol* 1991; 30:255-259.
 130. Boyd HK, Lappin TR. Erythropoietin deficiency in the anaemia of chronic disorders. *Eur J Haematol* 1991; 46:198-201.
 131. Baer AN, Dessypris EN, Goldwasser E, Krantz SB. Blunted erythropoietin response to anaemia in rheumatoid arthritis. *Br J Haematol* 1987; 66:559-564.
 132. Cazzola M, Ponchio L, de Benedetti F, Ravelli A, Rosti V, Beguin Y, Invernizzi R, Barosi G, Martini A. Defective iron supply for erythropoiesis and adequate endogenous erythropoietin production in the anemia associated with systemic-onset juvenile chronic arthritis. *Blood* 1996; 87:4824-4830.
 133. Kirel B, Yetgin S, Saatci U, Ozen S, Bakkaloglu A, Besbas N. Anaemia in juvenile chronic arthritis. *Clin Rheumatol* 1996; 15:236-241.
 134. Fantini F, Gattinara M, Gerloni V, Bergomi P, Cirila E. Severe anemia associated with active systemic-onset juvenile rheumatoid arthritis successfully treated with recombinant human erythropoietin: a pilot study. *Arthritis Rheum* 1992; 35:724-726.
 135. Martini A, Ravelli A, Di Fuccia G, Rosti V, Cazzola M, Barosi G. Intravenous iron therapy for severe anaemia in systemic-onset juvenile chronic arthritis. *Lancet* 1994; 344:1052-1054.
 136. Cimmino MA, Parisi M, Querci G, Ghio R, Accardo S. Intravenous iron is effective in treating the anaemia of rheumatoid arthritis and is not associated with flares of synovitis [letter]. *Clin Rheumatol* 1997; 16:215-216.

137. Davis D, Elliott M. Erythropoietin: in vivo effects in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
138. Zucker S. Erythropoietin in rheumatoid arthritis. *Arthritis Rheum* 1988; 31:1318-1321.
139. DeGowirion. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
140. Dainiak. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
141. Ward HF. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
142. Cox R. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
143. Zarrabi. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
144. Zucker S. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
145. Lockner. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
146. Nseyo U. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.
147. Schreuder. Erythropoietin in rheumatoid arthritis. *Br J Rheumatol* 1992; 31:1318-1321.

ann Rheum Dis

vak JL. Serum
toid arthritis:
eum 1988; 31:

anek B. Serum
els in patients
atol 1995; 13:

· impaired ery-
oid disease. Br

cy in the anaem-
1; 46:198-201.

itz SB. Blunted
atoid arthritis.

elli A, Rosti V,
. Defective iron
genous erythro-
with systemic-
37:4824-4830.

glu A, Besbas N.
heumatol 1996;

, Cirila E. Severe
uvenile rheuma-
nant human ery-
92; 35:724-726.

cola M, Barosi G.
n systemic-onset
52-1054.

ccardo S. Intra-
ia of rheumatoid
ynovitis [letter].

137. Davis D, Charles PJ, Potter A, Feldmann M, Maini RN, Elliott MJ. Anaemia of chronic disease in rheumatoid arthritis: in vivo effects of tumour necrosis factor alpha blockade. *Br J Rheumatol* 1997; 36:950-956.
138. Zucker S, Lysik R. Bone marrow erythropoiesis in anemia of inflammation. *J Lab Clin Med* 1974; 84:620-631.
139. DeGowin RL, Gibson DP. Erythropoietin and the anemia of mice bearing extramedullary tumor. *J Lab Clin Med* 1979; 94:303-311.
140. Dainiak N, Kulkarni V, Howard D, Kalmanti M, Dewey MC, Hoffman R. Mechanisms of abnormal erythropoiesis in malignancy. *Cancer* 1983; 51:1101-1106.
141. Ward HP, Kurnick JE, Pisarczyk MJ. Serum level of erythropoietin in anemias associated with chronic infection, malignancy, and primary hematopoietic disease. *J Clin Invest* 1971; 50:332-335.
142. Cox R, Musial T, Gyde OH. Reduced erythropoietin levels as a cause of anaemia in patients with lung cancer. *Eur J Cancer Clin Oncol* 1986; 22:511-514.
143. Zarrabi MH, Lysik R, DiStefano J, Zucker S. The anaemia of chronic disorders: studies of iron reutilization in the anaemia of experimental malignancy and chronic inflammation. *Br J Haematol* 1977; 35:647-658.
144. Zucker S, Friedman S, Lysik RM. Bone marrow erythropoiesis in the anemia of infection, inflammation, and malignancy. *J Clin Invest* 1974; 53:1132-1138.
145. Lockner D. Iron absorption and serum erythropoietin concentration in patients with cancer of the uterine cervix. *Acta Med Scand* 1966; 180:651-655.
146. Nseyo UO, Williams PD, Murphy GP. Clinical significance of erythropoietin levels in renal carcinoma. *Urology* 1986; 28:301-306.
147. Schreuder WO, Ting WC, Smith S, Jacobs A. Testosterone, erythropoietin and anaemia in patients with disseminated bronchial cancer. *Br J Haematol* 1984; 57:521-526.

148. Erslev AJ, Wilson J, Caro J. Erythropoietin titers in anemic, nonuremic patients. *J Lab Clin Med* 1987; 109:429-433.
149. Leng HM, Albrecht CF, Kidson SH, Folb PI. Erythropoietin production in anemia associated with experimental cancer. *Exp Hematol* 1999; 27:806-810.
150. Miller CB, Jones RJ, Piantadosi S, Abeloff MD, Spivak JL. Decreased erythropoietin response in patients with the anemia of cancer. *N Engl J Med* 1990; 322:1689-1692.
151. Ponchio L, Beguin Y, Farina G, Pedrazzoli P, Pedrotti C, Poggi G, Rosti V, Bergamaschi G, Battistel V, Cazzola M. Evaluation of erythroid marrow response to recombinant human erythropoietin in patients with cancer anaemia. *Haematologica* 1992; 77:494-501.
152. Abels RI. Recombinant human erythropoietin in the treatment of the anaemia of cancer. *Acta Haematol* 1992; 87(suppl 1):4-11.
153. Abels RI. Use of recombinant human erythropoietin in the treatment of anemia in patients who have cancer. *Semin Oncol* 1992; 19:29-35.
154. Cascinu S, Fedeli A, Del Ferro E, Luzi Fedeli S, Catalano G. Recombinant human erythropoietin treatment in cisplatin-associated anemia: a randomized, double-blind trial with placebo. *J Clin Oncol* 1994; 12:1058-1062.
155. Osterborg A, Boogaerts MA, Cimino R, Essers U, Holowiecki J, Juliusson G, Jager G, Najman A, Peest D. Recombinant human erythropoietin in transfusion-dependent anemic patients with multiple myeloma and non-Hodgkin's lymphoma. A randomized multicenter study. *Blood* 1996; 87:2675-2682.
156. Cazzola M, Messinger D, Battistel V, Bron D, Cimino R, Enller-Ziegler L, Essers U, Greil R, Grossi A, Jager G, LeMevel A, Najman A, Silingardi V, Spriano M, van Hoof A, Ehmer B. Recombinant human erythropoietin in the anemia associated with multiple myeloma or non-Hodgkin's lymphoma: dose finding and identification of predictors of response. *Blood* 1995; 86:4446-4453.
157. Plataniias LC, Miller CB, Mick R, Hart RD, Ozer H, McEvelly JM, Jones RJ, Ratain MJ. Treatment of chemotherapy-

- induc
cance.
158. Cazzo
Libera
taneon
in hen
Blood
159. Oster
ler G,
for the
neopla
956-96
160. Kettel
thropoi
cer 199
161. Kalma
and ery
and tur
162. Chudek
zer J. I
uterine
163. Dowlati
cancer i
thropoi
erythro
164. Lee SJ,
inadequ
J Hema
165. Corazza
Devalck
with car
ity and
1998; 92
166. Sawabe
poietin r
immuno
noma an

titers in anemic,
109:429-433.

I. Erythropoietin
experimental cancer.

MD, Spivak JL.
patients with the
1689-1692.

li P, Pedrotti C,
el V, Cazzola M.
to recombinant
cancer anaemia.

in the treatment
87(suppl 1):4-11.

ropoietin in the
e cancer. Semin

li S, Catalano G.
ent in cisplatin-
blind trial with

rs U, Holowiecki
D. Recombinant
pendent anemic
-Hodgkin's lym-
Blood 1996; 87:

n D, Cimino R,
ssi A, Jager G,
no M, van Hoof
ropoietin in the
or non-Hodgkin's
m of predictors

Dzer H, McEvelly
f chemotherapy-

induced anemia with recombinant human erythropoietin in
cancer patients. *J Clin Oncol* 1991; 9:2021-2026.

158. Cazzola M, Ponchio L, Beguin Y, Rosti V, Bergamaschi G, Liberato NL, Fregoni V, Nalli G, Barosi G, Ascari E. Subcutaneous erythropoietin for treatment of refractory anemia in hematologic disorders. Results of a phase I/II clinical trial. *Blood* 1992; 79:29-37.
159. Oster W, Herrmann F, Gamm H, Zeile G, Lindemann A, Müller G, Brune T, Kraemer HP, Mertelsmann R. Erythropoietin for the treatment of anemia of malignancy associated with neoplastic bone marrow infiltration. *J Clin Oncol* 1990; 8: 956-962.
160. Kettelhack C, Schoter D, Matthias D, Schlag PM. Serum erythropoietin levels in patients with solid tumours. *Eur J Cancer* 1994; 30A:1289-1291.
161. Kalmanti M, Kalmantis T. Committed erythroid progenitors and erythropoietin levels in anemic children with lymphomas and tumors. *Pediatr Hematol Oncol* 1989; 6:85-93.
162. Chudek J, Wiecek A, Kokot F, Grochal A, Switala J, Kanwiszer J. Serum erythropoietin concentration in women with uterine or ovarian tumors. *Przegl Lek* 1998; 55:47-50.
163. Dowlati A, R'Zik S, Fillet G, Beguin Y. Anaemia of lung cancer is due to impaired erythroid marrow response to erythropoietin stimulation as well as relative inadequacy of erythropoietin production. *Br J Haematol* 1997; 97:297-299.
164. Lee SJ, Kwon JH, Jung CW. Erythropoietin response is inadequate in cancer patients receiving chemotherapy. *Int J Hematol* 2001; 74:416-420.
165. Corazza F, Beguin Y, Bergmann P, Andre M, Ferster A, Devalck C, Fondu P, Buyse M, Sariban E. Anemia in children with cancer is associated with decreased erythropoietic activity and not with inadequate erythropoietin production. *Blood* 1998; 92:1793-1798.
166. Sawabe Y, Iida S, Tabata Y, Yonemitsu H. Serum erythropoietin measurements by a one-step sandwich enzyme linked immunosorbent assay in patients with hepatocellular carcinoma and liver cirrhosis. *Jpn J Clin Oncol* 1993; 23:273-277.

167. Malaguarnera M, Bentivegna P, Di Fazio I, Laurino A, Romano M, Trovato BA. Erythropoietin in hepatocellular carcinoma. *Bull Cancer* 1996; 83:977-980.
168. Vadhan-Raj S, Hittelman WN, Lepe-Zuniga JL, Gutterman JU, Broxmeyer HE. Regulation of endogenous erythropoietin levels in anemia associated with myelodysplastic syndromes [letter]. *Blood* 1990; 75:1749-1750.
169. Verhoef GE, De Schouwer P, Ceuppens JL, Van Damme J, Goossens W, Boogaerts MA. Measurement of serum cytokine levels in patients with myelodysplastic syndromes. *Leukemia* 1992; 6:1268-1272.
170. Jacobs A, Janowska-Wieczorek A, Caro J, Bowen DT, Lewis T. Circulating erythropoietin in patients with myelodysplastic syndromes. *Br J Haematol* 1989; 73:36-39.
171. Bowen DT, Jacobs A, Cotes PM, Lewis TC. Serum erythropoietin and erythropoiesis in patients with myelodysplastic syndromes. *Eur J Haematol* 1990; 44:30-32.
172. Bowen DT, Culligan D, Beguin Y, Kendall R, Willis N. Estimation of effective and total erythropoiesis in myelodysplasia using serum transferrin receptor and erythropoietin concentrations, with automated reticulocyte parameters. *Leukemia* 1994; 8:151-155.
173. Pasqualetti P, Collacciani A, Casale R. Circadian rhythm of serum erythropoietin in myelodysplastic syndromes. *Eur Rev Med Pharmacol Sci* 2000; 4:111-115.
174. Wallvik J, Stenke L, Bernell P, Nordahl G, Hippe E, Hast R. Serum erythropoietin (EPO) levels correlate with survival and independently predict response to EPO treatment in patients with myelodysplastic syndromes. *Eur J Haematol* 2002; 68:180-185.
175. Piedras J, Hernandez G, Lopez-Karpovitch X. Effect of androgen therapy and anemia on serum erythropoietin levels in patients with aplastic anemia and myelodysplastic syndromes. *Am J Hematol* 1998; 57:113-118.
176. McMullin MF, Hillmen P, Elder GE, Lappin TR, Luzzatto L. Serum erythropoietin levels in paroxysmal nocturnal haemo-

- globinuria: im
Haematol 199
177. Nakakuma H, Iwamoto N, Ka
erythropoietin
levels in pati
nuria. *Int J H*
178. Barosi G, Libe
patients with
Haematol 199
179. Najean Y, Sch
poietin conce
primary thron
55:272-273.
180. Andreasson B,
in essential th
to myelosuppr
113-120.
181. Messinezy M,
Sherwood RS,
in erythrocyto
Haematol 200
182. Griesshammer
Bangerter M,
thropoietin an
tial thrombo
533-538.
183. Jansen JH, Wi
den Ottolande
monocyte color
leukin-6 in re
Leukemia 199
184. Mauro FR, Ge
phocytic leuke
185. Beguin Y, Lan
tin in chronic
93:154-156.

- globinuria: implications for therapy [see comments]. *Br J Haematol* 1996; 92:815-817.
177. Nakakuma H, Nagakura S, Kawaguchi T, Horikawa K, Iwamoto N, Kagimoto T, Takatsuki K. Markedly high plasma erythropoietin and granulocyte-colony stimulating factor levels in patients with paroxysmal nocturnal hemoglobinuria. *Int J Hematol* 1997; 66:451-457.
178. Barosi G, Liberato LN, Guarnone R. Serum erythropoietin in patients with myelofibrosis with myeloid metaplasia. *Br J Haematol* 1993; 83:365-369.
179. Najean Y, Schlageter MH, Toubert ME, Rain JD. Erythropoietin concentration in the serum from patients with primary thrombocythaemia [letter]. *Eur J Haematol* 1995; 55:272-273.
180. Andreasson B, Lindstedt G, Kutti J. Plasma erythropoietin in essential thrombocythaemia: at diagnosis and in response to myelosuppressive treatment. *Leuk Lymphoma* 2000; 38:113-120.
181. Messinezy M, Westwood NB, El Hemaidi I, Marsden JT, Sherwood RS, Pearson TC. Serum erythropoietin values in erythrocytoses and in primary thrombocythaemia. *Br J Haematol* 2002; 117:47-53.
182. Griesshammer M, Kubanek B, Beneke H, Heimpel H, Bangerter M, Bergmann L, Schrezenmeier H. Serum erythropoietin and thrombopoietin levels in patients with essential thrombocythaemia. *Leuk Lymphoma* 2000; 36:533-538.
183. Jansen JH, Wientjens GJ, Fibbe WE, Ralph P, Van Damme J, den Ottolander GJ, Willemze R, Kluin Nelemans JC. Serum monocyte colony-stimulating factor, erythropoietin and interleukin-6 in relation to pancytopenia in hairy cell leukemia. *Leukemia* 1992; 6:735-737.
184. Mauro FR, Gentile M, Foa R. Erythropoietin and chronic lymphocytic leukemia. *Rev Clin Exp Hematol Suppl* 2002; 1:21-31.
185. Beguin Y, Lampertz S, Bron D, Fillet G. Serum erythropoietin in chronic lymphocytic leukemia. *Br J Haematol* 1996; 93:154-156.

186. Johannsen H, Jelkmann W, Wiedemann G, Otte M, Wagner T. Erythropoietin/haemoglobin relationship in leukaemia and ulcerative colitis. *Eur J Haematol* 1989; 43:201-206.
187. Jelkmann W, Johannsen H, Wiedemann G, Otte M, Wagner T. Dependence of serum erythropoietin level on erythropoiesis in leukemia. *Hamatol Bluttransfus* 1990; 33:83-86.
188. Jelkmann W, Wiedemann G. High serum immunoreactive erythropoietin in leukaemic patients with bone marrow insufficiency of erythropoiesis [letter]. *Eur J Haematol* 1990; 45:271-272.
189. Hellebostad M, Marstrander J, Slrdahl SH, Cotes PM, Refsum HE. Serum immunoreactive erythropoietin in children with acute leukaemia at various stages of disease—and the effects of treatment. *Eur J Haematol* 1990; 44:159-164.
190. Kivivuori SM, Viinikka L, Teppo AM, Siimes MA. Serum transferrin receptor and erythropoiesis in children with newly diagnosed acute lymphoblastic leukemia. *Leuk Res* 1994; 18:823-828.
191. Pohl C, Schobert I, Moter A, Woll EM, Schwonzen M, Hiersche A, Diehl V. Serum erythropoietin levels in patients with Hodgkin's lymphoma at the time of diagnosis. *Ann Oncol* 1992; 3:172-173.
192. Beguín Y. Erythropoiesis and erythropoietin in multiple myeloma. *Leuk Lymphoma* 1995; 18:413-421.
193. Paaske Hansen O, Drivsholm A. Interrelationships between blood volume, venous hematocrit and renal failure in myelomatosis. *Scand J Haematol* 1978; 20:461-466.
194. Birgens HS, Paaske Hansen O, Henriksen JH, Wantzin P. Quantitation of erythropoiesis in myelomatosis. *Scand J Haematol* 1979; 22:357-363.
195. Nielsen OJ, Brandt M, Drivsholm A. The secretory erythropoietin response in patients with multiple myeloma and Waldenstrom's macroglobulinaemia. *Scand J Clin Lab Invest* 1990; 50:697-703.
196. Majumdar G, Westwood NB, Bell-Witter C, Muggleston D, Phillips J, Pearson TC. Serum erythropoietin and circulating

- BFU-E without
197. Ariad S. Erythro myelom. bone ma
198. Paaske. erythrop 106-110
199. Singh A, Ratcliffe. ity as a J Clin I
200. Host H, mide an in rats. :
201. McDona. hypoxia, erythrop
202. Magid F. from sh Lab Inve
203. Chakrab. suppress in murir Neoplas
204. Fisher J. erythrop
205. Wolff M. immuno: tin in h 27-31.
206. Jelkman. tin produ: bute to Biol 199:

Endogenous Erythropoietin

- BFU-E in patients with multiple myeloma and anaemia but without renal failure. *Leuk Lymphoma* 1993; 9:173-176.
197. Ariad S, Clifford D, Penfold G, MacPhail AP, Bezwoda WR. Erythropoietin response in anaemic patients with multiple myeloma and other lymphoid malignancies infiltrating the bone marrow. *Eur J Haematol* 1992; 49:59-62.
 198. Paaske Hansen O, Thorling EB, Drivsholm A. Serum erythropoietin in myelomatosis. *Scand J Haematol* 1977; 19:106-110.
 199. Singh A, Eckardt KU, Zimmermann A, Gotz KH, Hamann M, Ratcliffe PJ, Kurtz A, Reinhart WH. Increased plasma viscosity as a reason for inappropriate erythropoietin formation. *J Clin Invest* 1993; 91:251-256.
 200. Host H, Skjaelaaen P. Comparative effects of cyclophosphamide and total body irradiation on erythropoietin production in rats. *Scand J Haematol* 1966; 3:154-157.
 201. McDonald TP, Lange RD, Congdon CC, Toya RE. Effect of hypoxia, irradiation, and bone marrow transplantation on erythropoietin levels in mice. *Radiat Res* 1970; 42:151-163.
 202. Magid E, Hansen P. The erythropoietin content of plasma from sheep treated with nitrogen mustard. *Scand J Clin Lab Invest* 1966; 18:347-352.
 203. Chakraborty A, Chatterjee M. Enhanced erythropoietin and suppression of gamma-glutamyl transpeptidase (ggt) activity in murine lymphoma following administration of vanadium. *Neoplasma* 1994; 41:291-296.
 204. Fisher JW, Roh BL. Influence of alkylating agents on kidney erythropoietin production. *Cancer Res* 1964; 24:983-988.
 205. Wolff M, Jelkmann W. Effects of chemotherapeutic and immunosuppressive drugs on the production of erythropoietin in human hepatoma cultures. *Ann Hematol* 1993; 66:27-31.
 206. Jelkmann W, Wolff M, Fandrey J. Inhibition of erythropoietin production by cytokines and chemotherapy may contribute to the anemia in malignant diseases. *Adv Exp Med Biol* 1994; 345:525-530.

207. Von Hoff DD, Schilsky R, Reichert CM, Reddick RL, Rozenzweig M, Young RC, Muggia FM. Toxic effects of *cis*-dichlorodiammineplatinum(II) in man. *Cancer Treat Rep* 1979; 63:1527-1531.
208. Horiguchi H, Kayama F, Oguma E, Willmore WG, Hradecky P, Bunn HF. Cadmium and platinum suppression of erythropoietin production in cell culture: clinical implications. *Blood* 2000; 96:3743-3747.
209. Rothman SA, Paul P, Weick JK, McIntyre WR, Fantelli F. Effect of *cis*-diamminedichloroplatinum on erythropoietin production and hematopoietic progenitor cells. *Int J Cell Cloning* 1985; 3:415-423.
210. Unami A, Nishina N, Terai T, Sato S, Tamura T, Noda K, Mine Y. Effects of cisplatin on erythropoietin production in rats. *J Toxicol Sci* 1996; 21:157-165.
211. Vaziri ND, Zhou XJ, Liao SY. Erythropoietin enhances recovery from cisplatin-induced acute renal failure. *Am J Physiol* 1994; 266:F360-F366.
212. Baldwin MD, Zhou XJ, Ing TS, Vaziri ND. Erythropoietin ameliorates anemia of cisplatin induced acute renal failure. *ASAIO J* 1998; 44:44-47.
213. Matsumoto T, Endoh K, Kamisango K, Akamatsu K, Koizumi K, Higuchi M, Imai N, Mitsui H, Kawaguchi T. Effect of recombinant human erythropoietin on anticancer drug-induced anaemia. *Br J Haematol* 1990; 75:463-468.
214. Wood PA, Hrushesky WJ. Cisplatin-associated anemia: an erythropoietin deficiency syndrome. *J Clin Invest* 1995; 95:1650-1659.
215. Sawabe Y, Kikuno K, Iseki T, Lida S, Tabata Y, Yonemitsu H. Changes in serum erythropoietin and the reticulocyte count during chemotherapy for leukemias. *Eur J Haematol* 1996; 57:384-388.
216. Sawabe Y, Kikuno K, Iseki T, Iida S, Yonemitsu H. Serum erythropoietin values and serum iron status during chemotherapy for leukemia. *Eur J Haematol* 1998; 60:315-316.

217. Sav
iyar
poi
dur
199
218. Cer
nac
ing
Exj
219. Mi
Gu
un
Ph
220. Ki
Ok
th
ac
tre
221. Fr
A
th
to
222. K:
M
se
gc
aj
223. S:
K
ol
(I
r
C
224. S
E
h
r
1

Reddick RL, et al. Effects of cisplatin on erythropoiesis. *Cancer Treat Rep* 1980; 63:163-168.

Wang H, Hradecky P, et al. Effect of erythropoietin on erythropoiesis. *Blood* 2000; 95:163-168.

Wang H, Fantelli F, et al. Erythropoietin increases erythropoiesis. *Int J Cell Physiol* 1998; 174:1-6.

Wang H, Noda K, et al. Erythropoietin increases erythropoiesis. *Am J Physiol* 1998; 275:R1000-R1005.

Wang H, et al. Erythropoietin increases erythropoiesis. *Am J Physiol* 1998; 275:R1000-R1005.

Wang H, et al. Erythropoietin increases erythropoiesis. *Am J Physiol* 1998; 275:R1000-R1005.

Wang H, Koizumi T, et al. Effect of erythropoietin on erythropoiesis. *Int J Cell Physiol* 1998; 174:1-6.

Wang H, et al. Erythropoietin increases erythropoiesis. *Am J Physiol* 1998; 275:R1000-R1005.

Wang H, Yonemitsu H, et al. Erythropoietin increases erythropoiesis. *Am J Physiol* 1998; 275:R1000-R1005.

Wang H, Yonemitsu H, et al. Erythropoietin increases erythropoiesis. *Am J Physiol* 1998; 275:R1000-R1005.

217. Sawabe Y, Takiguchi Y, Kikuno K, Iseki T, Ito J, Iida S, Kuriyama T, Yonemitsu H. Changes in levels of serum erythropoietin, serum iron and unsaturated iron binding capacity during chemotherapy for lung cancer. *Jpn J Clin Oncol* 1998; 28:182-186.
218. Cerruti A, Castello G, Balleari E, Bogliolo G, Lerza R, Pannaccioli I. Serum erythropoietin increase in patients receiving adjuvant therapy with 5-fluorouracil and leucovorin. *Exp Hematol* 1994; 22:1261-1263.
219. Miranda CE, Scaro JL, Buys MC, Torrejon I, Martin B, Guerra L. Urinary elimination of erythropoietin in patients under treatment with cytostatics drugs. *Acta Physiol Pharmacol Ther Latinoam* 1998; 48:207-210.
220. Kinjo K, Kizaki M, Takayama N, Michikawa N, Oda A, Okamoto S, Tahara T, Kato T, Miyazaki H, Ikeda Y. Serum thrombopoietin and erythropoietin levels in patients with acute promyelocytic leukaemia during all-trans retinoic acid treatment. *Br J Haematol* 1999; 105:382-387.
221. Franek E, Marcinkowski W, Kokot F, Wiecek A, Nowicki M. A single dose of cyclophosphamide (Cph) does increase erythropoietin concentration in patients with hypertension due to vasculitis [letter]. *Clin Nephrol* 1994; 42:139-140.
222. Katschinski DM, Jelkmann W, Wiedemann GJ, Mentzel M, Mulkerin DL, Touhidi R, Robins HI. Dynamic changes in serum erythropoietin levels in solid tumour patients undergoing 41.8°C whole body hyperthermia and/or chemotherapy. *Int J Hyperthermia* 1997; 13:563-569.
223. Shamseddine A, Medawar W, Seoud M, Ibrahim K, Habbal Z, Kahwaji S, Khalil A. The relationship between serum levels of erythropoietin (EPO) and insulin-like growth factor-1 (ILGF-1) and hematocrit (HCT) in breast cancer patients receiving non-nephrotoxic chemotherapy. *Eur J Gynaecol Oncol* 1998; 19:591-593.
224. Shamseddine A, Khalil A, Seoud M, Kahwaji S, Taher A, Bizri AR, Medawar W. The relation between erythropoietin, hematocrit and hemoglobin in breast cancer patients on non-nephrotoxic chemotherapy. *Eur J Gynaecol Oncol* 1998; 19:577-579.

225. Dowd MD, Morgan ER, Langman CB, Murphy S. Serum erythropoietin levels in children with leukemia. *Med Pediatr Oncol* 1997; 28:259-267.
226. Lechner W, Artner Dworzak E, Solder E, Sachsenmaier M, Kolle D, Moncayo H, Reitsamer R. Influence on erythropoietin levels of treatment with cisplatinum-endoxan. *Arch Gynecol Obstet* 1992; 252:49-53.
227. Saijo Y, Nakai Y, Saito J, Sugawara S, Suzuki S, Numata Y, Motomiya M. Changes in serum erythropoietin levels during chemotherapy for lung cancer. *Chemotherapy* 1992; 38:281-285.
228. Onat H, Inanc SE, Dalay N, Karaloglu D, Erturk N, Yasasever V. Effect of cisplatin on erythropoietin and iron changes [letter]. *Eur J Cancer* 1993; 29A:777.
229. Canaparo R, Casale F, Muntoni E, Zara GP, Della PC, Berno E, Pons N, Fornari G, Eandi M. Plasma erythropoietin concentrations in patients receiving intensive platinum or nonplatinum chemotherapy. *Br J Clin Pharmacol* 2000; 50:146-153.
230. Pedain C, Herrero J, Kunzel W. Serum erythropoietin levels in ovarian cancer patients receiving chemotherapy. *Eur J Obstet Gynecol Reprod Biol* 2001; 98:224-230.
231. Orhan B, Yalcin S, Evrensel T, Kurt E, Manavoglu O, Erbas T. Does cisplatin stimulate erythropoietin secretion from the peritubular cells of the kidney? [letter]. *Clin Nephrol* 1998; 50:202-203.
232. Hasegawa I, Tanaka K. Serum erythropoietin levels in gynecologic cancer patients during cisplatin combination chemotherapy. *Gynecol Oncol* 1992; 46:65-68.
233. Smith DH, Goldwasser E, Vokes EE. Serum immunoerythropoietin levels in patients with cancer receiving cisplatin-based chemotherapy. *Cancer* 1991; 68:1101-1105.
234. Bray GL, Reaman GH. Erythropoietin deficiency: a complication of cisplatin therapy and its treatment with recombinant human erythropoietin. *Am J Pediatr Hematol Oncol* 1991; 13:426-430.

235. Cazzola M, human eryt 1997; 89:42.
236. Fischl M, G Kennedy P, RI, Tsai HC tin for patie J Med 1990
237. Pincus T, O TJ, Boccagr nant huma rheumatoid
238. Ludwig H, J Schuster : treatment : 1056-1063.
239. Cazzola M, Lucotti C, N diction of r (rHuEpo) ir 434-441.
240. Oberhoff C Rebmann U Armand J Mathieu N, in the treat vention of tumors: a Ann Oncol
241. ten Bokkel Morack G, Reed NS, F GB, van Tin of the influe poietin on : with ovari chemothera

phy S. Serum eryth-
mia. *Med Pediatr*

Sachsenmaier M,
ice on erythropoie-
doxan. *Arch Gynecol*

zuki S, Numata Y,
poietin levels during
therapy 1992; 38:

glu D, Erturk N,
ropoietin and iron
777.

, Della PC, Berno E,
ropoietin concentra-
um or nonplatinum
50:146-153.

rythropoietin levels
emotherapy. *Eur J*
-230.

navoglu O, Erbas T.
secretion from the
Clin Nephrol 1998;

poietin levels in gynecol-
1 combination che-
8.

um immunoerythro-
receiving cisplatin-
01-1105.

iciency: a complicat-
at with recombinant
ematol Oncol 1991;

235. Cazzola M, Mercuriali F, Brugnara C. Use of recombinant human erythropoietin outside the setting of uremia. *Blood* 1997; 89:4248-4267.
236. Fischl M, Galpin JE, Levine JD, Groopman JE, Henry DH, Kennedy P, Miles S, Robbins W, Starrett B, Zalusky R, Abels RI, Tsai HC, Rudnick SA. Recombinant human erythropoietin for patients with AIDS treated with zidovudine. *N Engl J Med* 1990; 322:1488-1493.
237. Pincus T, Olsen NJ, Russell IJ, Wolfe F, Harris ER, Schnitzer TJ, Boccagno JA, Krantz SB. Multicenter study of recombinant human erythropoietin in correction of anemia in rheumatoid arthritis. *Am J Med* 1990; 89:161-168.
238. Ludwig H, Fritz E, Leitgeb C, Pecherstorfer M, Samonigg L, Schuster J. Prediction of response to erythropoietin treatment in chronic anemia of cancer. *Blood* 1994; 84:1056-1063.
239. Cazzola M, Ponchio L, Pedrotti C, Farina G, Cerani P, Lucotti C, Novella A, Rovati A, Bergamaschi G, Beguin Y. Prediction of response to recombinant human erythropoietin (rHuEpo) in anemia of malignancy. *Haematologica* 1996; 81:434-441.
240. Oberhoff C, Neri B, Amadori D, Petry KU, Gamucci T, Rebmann U, Nowrousian MR, Voigtmann R, Monfardini S, Armand JP, Herrmann R, Netter-Pinon J, Tubiana-Mathieu N, Zwierzina H. Recombinant human erythropoietin in the treatment of chemotherapy-induced anemia and prevention of transfusion requirement associated with solid tumors: a randomized, controlled study [see comments]. *Ann Oncol* 1998; 9:255-260.
241. ten Bokkel Huinink WW, de Swart CA, van Toorn DW, Morack G, Breed WP, Hillen HF, van der Hoeven JJ, Reed NS, Fairlamb DJ, Chan SY, Godfrey KA, Kristensen GB, van Tinteren H, Ehmer B. Controlled multicentre study of the influence of subcutaneous recombinant human erythropoietin on anaemia and transfusion dependency in patients with ovarian carcinoma treated with platinum-based chemotherapy [see comments]. *Med Oncol* 1998; 15:174-182.

242. Charuruks N, Voravud N, Limpanasithikul W. Ratio of baseline erythropoietin (EPO) level and corrected reticulocyte count as an indicator for a favourable response to recombinant human erythropoietin (rhEPO) therapy in anaemic cancer patients. *J Clin Lab Anal* 2001; 15:260-266.
243. Leon MP, Jimenez MM, Barona ZP, Riol DM, Castro PL, Sierasesumaga AL. Recombinant human erythropoietin in anemia associated with pediatric cancer: study of the identification of predictors of response. *An Esp Pediatr* 1998; 49:17-22.