

Relation between the low temperature stress and catalase activity in gladiolus somaclones (*Gladiolus grandiflorus* Hort.)

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Abstract

In order to evaluate low temperature tolerance of five variable somaclones as well as the cv. Peter Pears of gladiolus (*Gladiolus grandiflorus* Hort.) from which they were originated, catalase (CAT) activity and hydrogen peroxide (H₂O₂) level were measured. Before carrying out the analyses, vitroplants of these clones were exposed, during 21 days, to 8 °C temperature. In response to the stress generated by this low temperature, catalase (CAT) activity revealed on native gel and quantified by spectrophotometry, showed variable levels, being all of them higher compared to cultivar control. For these clones, strong CAT activities were always accompanied by low H₂O₂ levels. Thus, at low temperatures the antioxidants system of adapted clones is more active compared to control.

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1. Introduction

Biotic and abiotic stresses such as drought, salinity, diseases and thermal stresses accelerate the production of active oxygen species (AOS) which then exceeds the capacity of the cells antioxidant system (Foyer et al., 1994; Bowler and Fluhr, 2000). Cells have an enzymatic and non-enzymatic antioxidant system to neutralize these free radicals. In higher plants three major enzymes are implicated in the AOS detoxification: superoxide dismutase (SOD), ascorbate peroxidase (APX) and catalases (Dodet, 1991; Willekens et al., 1995; Scandalios, 2002). In the case of cold sensitive plants, catalases are more implicated than other enzymes (Volk and Feirabend, 1989).

Among AOS, hydrogen peroxide (H₂O₂) seems to be implicated in various stress situations (Bowler and Fluhr, 2000). In fact, H₂O₂ as well as after its transformation into radical OH[•], is the origin of direct cellular damage. In addition to the main role of CAT in the control of H₂O₂ level in plant cells, significant functions in the stress responses are linked to these

enzymes. Wadsworth and Scandalios (1990) indicated that a deficiency in CAT remarkably reduced the germination rate of barley and corn seeds. Willekens et al. (1997) indicated in their work on tobacco lines with partially limited CAT activity, that these enzymes belong to the normal process of the photosynthetic mechanism and are essential for the oxidative stress response. Shikanai et al. (1998) observed an improvement of the tolerance to light stress and drought that over express CAT activity.

Among environment limits, light deficiency and thermal shocks are regarded as the most significant factors that affect CAT activity (Volk and Feirabend, 1989; Willekens et al., 1997; Matsumura et al., 2002; Sairam et al., 2002). In order to establish a possible relation between cold tolerance and certain antioxidant enzymes, CAT activity was analyzed and H₂O₂ level was measured for five gladiolus vitrovariants (R₁, Rb₁, Rb₂, Rib₁, Rib₂) selected in a previous work (Bettaieb, 2003) for their tolerance to low temperatures and adapted to winter culture.

2. Materials and methods

The vitrovariants were regenerated from irradiated callus of 'Peter Pears' with gamma ray (50 Gy). The cultivar 'Peter Pears' was used as control before undertaking analysis, vitroplants were

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placed in 1 l bottles containing a culture medium consisting of rock salt and vitamins of Murashige and Skoog (1962) with 30 g l^{-1} of sucrose and 0.5 mg l^{-1} of IBA (indole-3-butyric acid). The pH of culture medium was adjusted to 5.8 and this medium was solidified with agar (6 g l^{-1}). The bottles remained 21 days at 8°C (to induce the thermal stress), with a photoperiod of 11 h at $36 \mu\text{mol m}^{-2} \text{s}^{-1}$. Frozen plant tissues were ground to a fine powder in a pestle with liquid nitrogen and mixed with 1 volume of extraction buffer (50 mM potassium phosphate pH 7.6, 10 mM sodium metabisulfite, 1 mM ascorbic acid, 1 mM EDTA, 20% (w/v) sorbitol, 2% (w/v) polyvinylpyrrolidone and centrifuged at $12,000 \times g \times 20 \text{ min}$ at 4°C . The supernatant was collected and the protein concentration was determined using Bradford's method (1976).

Soluble protein samples were subjected to non-denaturing PAGE 10% according to Laemmli (1970). Catalase activity was revealed on the gel as follows: the gel was washed three times (15 min each) with water, then incubated for 10 min in 0.88 mM H_2O_2 solution, rinsed again with water, and finally incubated with 1% (w/v) of ferric chloride and potassium ferricyanide solution until yellow bands appeared on a green background). CAT activity was measured essentially as described by Clairbone (1985), with some modifications. The assay contained 15 mM H_2O_2 in 50 mM phosphate buffer (pH 7.0) and $80 \mu\text{g}$ of protein extract in total volume of 1 ml. CAT activity was estimated by a decrease of H_2O_2 absorbance at 240 nm and one unit of CAT was defined as the amount of enzyme dismounting $1 \mu\text{mol}$ of H_2O_2 per minute.

The quantification of H_2O_2 was determined by chemiluminescence (Warm and Laties, 1982), with modifications (Noel, 2001). One gram of plant tissue was ground in liquid nitrogen, mixed to 250 mg of active charcoal and extraction was with 4 ml of ice cold 5% (w/v) trichloroacetic acid. The crude extracts were centrifuged for 30 min at $12,000 \times g$. Two milliliters of supernatant were passed two times through a BioRad column AG1*8 (poly-prep prefilled chromatography columns), in the dark and cold room (8°C). H_2O_2 content was measured by adding $50 \mu\text{l}$ of eluate to $50 \mu\text{l}$ of 0.5 mM luminol and $100 \mu\text{l}$ of 0.5 mM ferricyanide ($\text{K}_3\text{Fe}(\text{CN})_6$). The luminol and the ferricyanide were solubilized in NH_4OH pH 9.5. Recovery estimates (which consisted of adding a known concentration of H_2O_2 to aliquots of the initial extracts that were processed in parallel). Average of 90% was recovered and used as a correction factor for each sample. The experiment was repeated three times and the average was considered. Trials were carried out according to a completely random block model. Each treatment was randomized on three blocks and each experimental unit related to a treatment in a block consisted of six homogeneous vitroplants. The Duncan test has been conducted to check difference between means at the 1% level.

3. Results

Staining gel showed superiority in the catalase activity at the vitrovariants selected for their cold tolerance as compared to the control. Differences between these genotypes were also observed (Fig. 1).

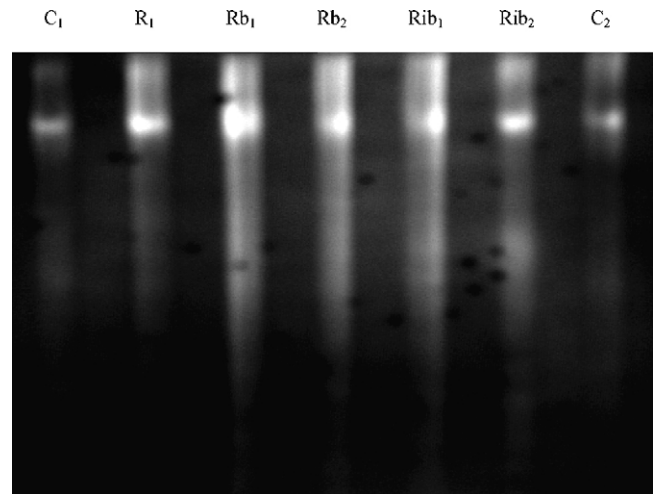


Fig. 1. Zymogram analysis of catalase activity in five clones (R_1 , Rb_1 , Rb_2 , Rib_1 et Rib_2) of gladiolus (*Gladiolus grandiflorus* Hort.) and in two samples of the cv. control Peter Pears (C_1 and C_2) grown at 8°C .

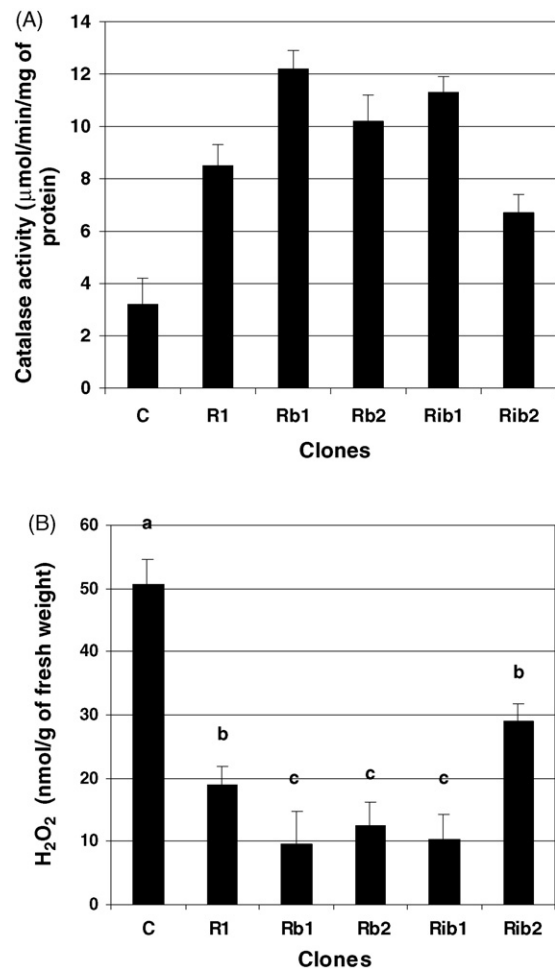


Fig. 2. Catalase activity (A) and hydrogen peroxide content (B) in tissue for five clones (R_1 , Rb_1 , Rb_2 , Rib_1 et Rib_2) of gladiolus (*Gladiolus grandiflorus* Hort.) and of the control cv. Peter Pears grown at 8°C . Indicated values represent the averages of the three repetitions. Bars represent the average of their estimated standard deviation. Means with the same letters are not significant at $p = 0.01$ probability level of Duncan test.

The spectrophotometry quantification of total catalase activity gave results comparable to those observed on non-denaturing gel PAGE with a superiority in all selected genotypes compared to the control (Fig. 2A). The statistical analysis showed highly significant differences between the control and genotypes for total CAT activity. Differences were also observed among genotypes with a superiority of Rb₁, Rb₂ and Rib₁. The H₂O₂ content showed highly significant differences between the control and selected genotypes for their tolerance to low temperatures (Fig. 2B). The control had the lowest level of CAT activity and the highest H₂O₂ content. The lowest level of H₂O₂ was observed in the genotypes Rb₁ which shows the highest level of CAT activity. Indeed, a high correlation ($r^2 = 0.95$) exists between these two parameters.

4. Discussion

The results of our experiments demonstrate that low temperatures induce an accumulation of AOS—who play a role in the photosystem II (PSII) operation (Willekens et al., 1997). The H₂O₂ accumulation was observed in the control (cv. Peter Pears) characterized by its sensitivity to cold (Vidalie, 1978). For the selected genotypes, the low level of H₂O₂ is explained by the important CAT activity that these genotypes exhibit.

Indeed CAT are the first enzymes which take place in H₂O₂ neutralization and thus could avoid cellular damage caused by excessive accumulation of the substrate (Willekens et al., 1994). James et al. (2001) pointed out, in their work on the deficient tobacco plant, the importance of these enzymes under stress conditions and enumerated several roles of H₂O₂ in molecular and biochemical plant mechanisms in response to stress conditions (Foyer and Noctor, 2005).

Feirabend et al. (1992) observed a reduction of 28% of catalase activity and an accumulation of H₂O₂ in rice seedlings exposed to 4 °C. After 24 h of seedlings transfer to 25 °C, these enzyme activity is restored. In addition, Volk and Feirabend (1989) suggested that, among the environmental constraints, thermal shocks are regarded as factors reducing catalase activity more than other enzymes. Puntarol et al. (1988) showed that the catalases are responsible for H₂O₂ elimination and that an over activity exhibition of these enzymes is able to induce hydrogen peroxide detoxification and is considered as a strategy for improving tolerance to the stress for plant cells.

5. Conclusion

The evaluation of the catalase activity and hydrogen peroxide measured in tissues of various selected vitrovariants for their tolerance to low temperatures showed different activities of somaclones. All somaclonal variants obtained in this work exhibited a catalase activity more significant than that of the control. Rb₁ Rb₂ clones and Rib₁ have the most significantly catalase activities and consequently the lowest H₂O₂ contents. These results allow conclude that in the presence of low temperatures, the antioxidant system of adapted clones is more active compared to control. The difference in activity between vitrovariants was also high-

lighted. Considering this behavior, it may be inferred that these clones are becoming tolerant to the low temperature.

References

- Bettaieb, T., 2003. Régénération de vitrovariants de glaïeul (*Gladiolus randiflorus* Hort.) tolérant les basses températures. Ph.D. Institut National agronomique de Tunisie, p. 115.
- Bowler, C., Fluhr, R., 2000. The role of calcium and active oxygen as signals for controlling cross tolerance. Trends Plants Sci. Rev. 5, 241–246.
- Bradford, M.M., 1976. A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. Anal. Biochem. 72, 248–254.
- Clairbone, A., 1985. Catalase activity. In: Greenwald, R.A. (Ed.), Handbook of Methods for Oxygen Radical Research. CRC Press, Boca Raton, pp. 283–284.
- Dodet, B., 1991. La chasse aux radicaux libres oxygénés. Biofutur, May 23–34.
- Feirabend, J., Schaan, C., Hertwig, B., 1992. Photoinactivation of catalase occurs under both high and low temperature stress conditions and accompanies photoinactivation of photosystem II. Plant Physiol. 100, 1554–1556.
- Foyer, C.H., Le Landais, M., Kunert, K.J., 1994. Photooxidative stress in plants. Physiol. Plant. 92, 696–717.
- Foyer, C.H., Noctor, G., 2005. Redox homeostasis and antioxidant signalling: a metabolic interface between stress perception and physiological responses. Plant Cell 17, 1866–1875.
- James, F.D., Inze, D., Van Brensegen, F., 2001. Catalase deficient tobacco plants: tools for in plant studies on the role of hydrogen peroxide. Redox Rep. 6, 37–42.
- Laemmli, U.K., 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature 227, 680–685.
- Matsumura, T., Tabayashi, N., Kamagata, Y., Souma, C., Saruyama, H., 2002. Wheat catalase expressed in transgenic rice can improve tolerance against low temperatures stress. Physiol. Plant. 116, 317–327.
- Murashige, T., Skoog, F., 1962. A revised medium for rapid growth and bioassays with tobacco cultures. Physiol. Plant. 15, 473–497.
- Noel, S., 2001. Rôle du peroxyde d'hydrogène au cours de la dormance du tubercule de pomme de terre (*Solanum tuberosum* L.). Mémoire de fin d'études d'Ingénieur Agronome. Faculté Universitaire des Sciences Agronomiques de Gembloux, Belgium, 71 p.
- Puntarol, S., Sanchez, R.A., Boveris, A., 1988. Hydrogen peroxide metabolism in soybean embryonic axes at the onset of germination. Plant Physiol. 86, 626–630.
- Sairam, K.R., Veerabhadra Rao, K., Srivastava, G.C., 2002. Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. Plant Sci. 163, 1037–1045.
- Scandalios, J.G., 2002. The rise of ROS. Trends Biochem. Sci. 27, 483–486.
- Shikanai, T., Takada, T., Yamauchi, H., Sano, S., Tomizawa, K., Yokota, A., Shigeoka, S., 1998. Inhibition of ascorbate peroxidases under oxidative stress in tobacco having bacterial catalase in chloroplasts. FEBS Lett. 428, 47–51.
- Vidalie, H., 1978. Comportement de différents cultivars de glaïeuls précoces cultivés sous tunnel. Pép. Hort. Mar. 192, 13–18.
- Volk, S., Feirabend, J., 1989. Photoinactivation of catalase at low temperature and its relevance to photosynthetic and peroxide metabolism in leaves. Plant Cell. Env. 12, 701–712.
- Wadsworth, G.L., Scandalios, J.G., 1990. Molecular characterization of a catalase null allele at cat3 locus in maize. Genetics 125, 867–872.
- Warm, E., Laties, G.G., 1982. Quantification of hydrogen peroxide in plant extracts by chemiluminescence reaction with luminol. Phytochemistry 21, 827–831.
- Willekens, H., Langebartels, C., Tiré, C., Van montagu, M., Inzé, D., Van Camp, W., 1994. Differential expression of catalase genes in *Nicotiana plumbaginifolia*. Proc. Natl. Acad. Sci. U.S.A. 91, 10450–10454.
- Willekens, H., Inzé, D., Van Montagu, M., Van Camp, W., 1995. Catalases in plants. Mol. Breed. 1, 207–228.
- Willekens, H., Chamnopol, S., Davey, M., Schravdner, M., Langebartels, C., Van Montagu, C., Inzé, D., Van Camp, W., 1997. Catalase is a sink for H₂O₂ and is indispensable for stress in C₃ plants. EMBO J. 16, 4806–4816.