

SIMULATION AND ANALYSIS OF RECEPTOR DYNAMICS IN A BMP REGULATORY NETWORK

Morgan Germain^{1,2}, Johanna Bolander^{2,3}, Wei Ji^{2,3}, Liesbet Geris^{1,2,4}

¹Biomechanics Research Unit, Université de Liège, Belgium ; ²Prometheus, Division of Skeletal Tissue Engineering, KU Leuven, Belgium ; ³Skeletal Biology and Engineering Research Center, KU Leuven, Belgium ; ⁴Biomechanics Section, KU Leuven, Belgium

KU LEUVEN

Université de Liège



INTRODUCTION

BMPs

- *Bone Morphogenetic Proteins* (BMPs) are crucial for bone formation
- BMPs signal through two transmembrane receptors: type 1 (R1) and type 2 (R2) receptor
- Two pathways : Canonical (C) and Non-canonical (NC)
- The final complex ligand-receptors (LR1R2) can be formed in two different ways
- Way of recruiting the receptors determines the activated pathway [1,2]

FOP

- *Fibrodysplasia Ossificans Progressiva* (FOP) is a disease that causes heterotopic ossification of soft tissues [3,4]
- FOP is linked to a constitutive activation of Alk2 (type 2 BMP receptor)
- The effect is coupled
 - ✓ Non-enzymatic cooperation with R2 receptors
 - ✓ Disruption of the non-canonical pathway

Aim of this study

- Simulation and analysis of a complete BMP model :
 - ✓ receptor traffic
 - ✓ both signaling pathways (C and NC)
- Simulation of FOP conditions to identify the key factors in this disease

MATERIALS & METHODS

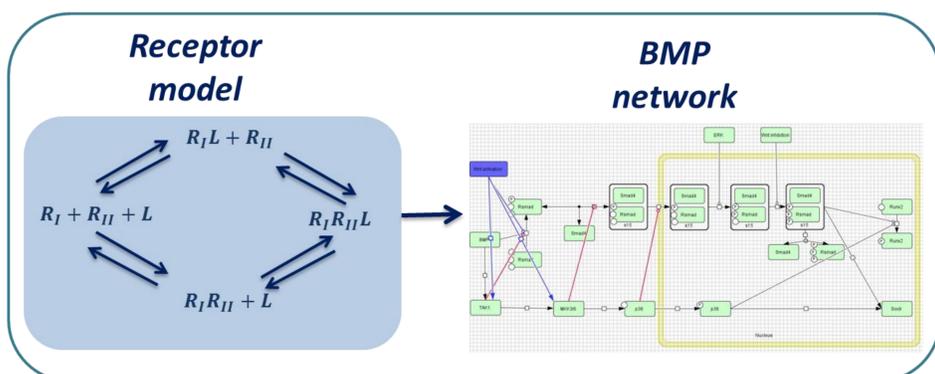


Figure 1 : Schematic representation of the model

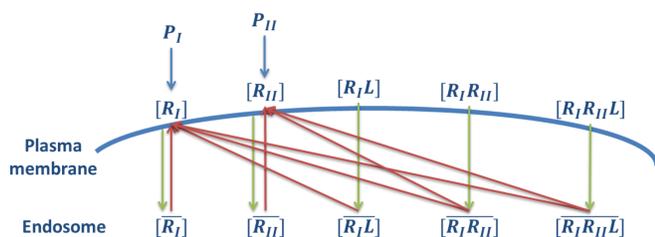


Figure 2 : Schematic representation of the traffic between plasma membrane and endosomes

Two literature-based mathematical models :

- ✓ **Receptors model** : How receptors binds to BMP and amongst themselves [2]
 - Two ways of binding modeled
 - 10 variables, 17 parameters
 - Traffic between plasma membrane and the endosomes also investigated (Figure 2) [5]
- ✓ **BMP network** : The non-canonical and the canonical pathways [6,7]
 - Various crosstalks between both pathways modeled
 - 12 variables, 23 parameters

- **Parameter values** derived from previous models [2], based on experiments
- **Ordinary Differential Equations (ODEs)** describe the temporal evolution of the various model constituents, e.g.:

$$\frac{d[LR1R2]}{dt} = k_1[LR1][R2] + k_2L[R1R2] - k_3[LR1R2]$$

where k_3 regroups different terms (degradation, internalization, recycling,...).

- Model implemented and simulated in **Matlab**

RESULTS

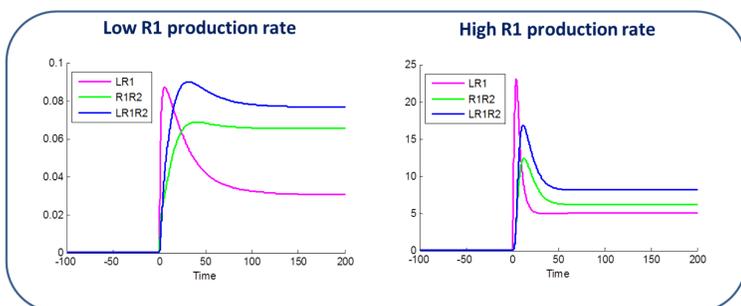


Figure 3 : Evolution over time of relative quantities of three key factors of the receptor dynamics model : LR1 (the NC pathway indicator), R1R2 (the C pathway indicator) and LR1R2 (the final receptor-ligand complex) for low (left) and high (right) R1 production rates (PR1)

Dynamics simulation by adding ligand (BMP) at t=0

- **FOP** :
 - ✓ simulated by the over-expression or higher activation of the receptor R1
 - ✓ disruption of the NC pathway experimentally observed
- **The model predicts that**
 - ✓ the rate of R1 production determines the preference for either the canonical or the non-canonical pathway
 - ✓ the ratio of the peak of the NC pathway to the peak of the C pathway is inversely related to the PR1
 - ✓ the ratio of their steady-states is bigger when PR1 is higher

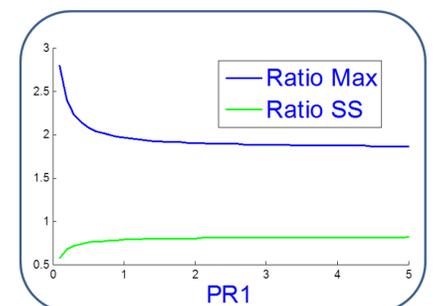


Figure 4 : Evolution of the ratio LR1/R1R2 (indicative for the ratio NC/C) for different values of R1 production rates. Ratios are shown for both peak values and steady-state values

DISCUSSION

- In absence of quantitative parameter information, the presented ODE model provides qualitative predictions on changes in the concentrations of all modelled components
- The model is able to capture experimental observations such as the choice of different BMP pathways BUT is not able to correctly reproduce the disruption of the non-canonical pathway in FOP cells suggesting that back-up pathways are activated (hypothesized in the field)
- Experimental work is underway to guide further model development

REFERENCES

[1] Sieber et al., 2009, Cytokine & Growth Factor Reviews, 20:343-355 [2] Heinig et al., 2011, Plos ONE, 6(10):25163 [3] Wang et al., 2014, Genes & Diseases, 1, 87-105 [4] Shi et al., 2013, Cell. Mol. Life Sci., 70:407-423 [5] Vilar et al., 2006, Plos Computational Biology, 2(1): e3 [6] Clarke et al., 2006, Syst Biol (Stevenage), 153(6):412-24. [7] Chen et al., 2012, Int. J. Biol. Sci., 8(2): 272-288

CONTACT

Université de Liège, Belgium,
M. Germain: morgan.germain@ulg.ac.be ; L. Geris: liesbet.geris@ulg.ac.be