Trace Elements in Marine Environments: Occurrence, Threats and Monitoring with Special Focus on the Coastal Mediterranean

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Abstract

Trace elements, as building blocks of matter, are naturally present in the environment. However, their extraction, production, use and release by men can lead to the increase of their environmental levels to concentrations that may be toxic for both men and the biota. The overall aim of this review is therefore to recall that trace elements remain contaminants of concern that still require scientific attention. Because marine coastal systems (and transitional environments in general) are particularly vulnerable to contamination processes, they deserve to be accurately monitored with quality indicator species. As an example, the 2 most widely used water and sediment measurements to assess the health status of the marine environment (3), and more particularly as regards the accurate and complementary indicators that are seagrasses (4) and mussels (5).

Keywords: Trace element; Toxicity; Aquatic environment; Monitoring; Seagrass; Mussel; Mediterranean

Introduction

Such as terrestrial ecosystems, marine ecosystems are submitted to increasing anthropogenic disturbances [1]. On the basis of expert judgment, Halpern et al. [2,3] mapped the impact of 17 anthropogenic drivers of ecological change (e.g., pollution, fishing, ocean acidification, species invasion etc.) on marine ecosystems. Their analysis indicated that no area remained unaffected by human activities, that a large fraction of the oceans was strongly affected by those drivers (41%), but that some large, less-impacted oceanic areas still remained (3.7%, particularly near the poles). The ecologic, economic and social importance of marine ecosystems being irrefutable [1,4], a well-planned approach of managing the marine space is essential to achieve sustainability [1]. Otherwise, entire ecosystems will stop functioning under their actual form, as is the case for the highly productive hotspots of biodiversity that are coral reefs [5], which is likely to lead to the complete loss of goods and services derived from these ecosystems [6].

The Mediterranean is one of the richest regions of Europe in terms of diversity of marine species with a high rate of endemism [7]. In the Mediterranean, coastal ecosystems are dominated by macrophytes (magnoliophytes and algae) [8], a globally net autotrophic system displaying many ecological benefits (e.g., primary production, habitats, source of food and oxygen, carbon well, stabilization of sediments etc.) [9]. Despite their environmental, economic and social importance, a growing number of reports document the occurring regression and/or ecofunctional changes of these coastal ecosystems (e.g., Ref. [10-12]). Pressures suffered by the Mediterranean make it a vulnerable ecological unit, in particular because this sea is of too small dimensions to ecologically self-counterbalance. Thus, the point of saturation of the contaminants discharged in the Mediterranean will be more quickly achieved than in the oceans [13]. And as regards the specific chemical contamination by trace elements, the high levels currently measured in the Mediterranean indicate non-stationary geochemical cycles which result from an increase of external inputs [14]. In addition, the almost total absence of tide does not allow the dilution of contaminants and prevents the natural phenomena of depuration as encountered in larger water bodies (i.e., in oceans). The Mediterranean also shows a deficiency in the movement of its deep water masses and of its surface currents which "turn in circles" in this almost closed basin. The consequence of these specific features is that the answer of the Mediterranean to environmental disturbances is more rapid than in the larger oceans [15,16] which makes of this sea a privileged laboratory to study environmental changes resulting from anthropogenic pressures [17].

Trace elements

Definition

According to the International Union of Pure and Applied Chemistry [18], trace elements are any element having an average concentration of less than about 100 parts per million atoms (ppma) or less than 100 µg g\(^{-1}\). Such a precise definition does not exist in earth sciences because the concentration of an element in a given phase can be so low that it is considered a trace element, whereas the same element can constitute a main part of another phase (e.g., Fe and Al) [19]. Previously, scientists used the generic term "heavy metals" when referring to trace elements. Today this appellation is discussed. Effectively, some metals are not particularly "heavy" (e.g., Al, Ni). In addition, some elements are not metals (e.g., As, Se). For these reasons, the majority of researchers prefer today the name "metallic trace elements" (if it is indeed metals) to the appellation "heavy metals", or the formula "trace elements" when they are not metals (e.g., As, Se, B) [20]. In the present review, we will consider trace elements (acronym TEs used throughout the paper) as any element, metallic or not, other
than the few major constitutive ones (i.e., C, H, N, S, O, P, Si, Cl, K, Na, Ca and Mg) forming the bulk of living and mineral (except Fe and Al) matter, whose concentrations are mostly below but sometimes above 100 ppma according to the matrix analyzed (Figure 1) [21,22].

TEs can either be essential or non-essential (Figure 1). The essential elements recognized by the World Health Organization (WHO) are I, Zn, Se, Fe, Cu, Cr and Mo, the latter playing an important role in biological systems [23,24]. Others TEs may/could also be essential, such as Mn, Co, As, Ni or V. For these elements among others, the essentiality is a characteristic which evolves according to our knowledge and to the sensitivity of the authors who have a propensity more or less strong to classify an element among the essential or not [25]. Non-essential TEs such as Hg, Pb or Cd play no physiological role, and are often toxic even in very small quantity [26]. For these non-essential TEs, only a threshold of toxicity exists, while essential TEs can be either deficient in too small quantities, either toxic when they are absorbed in high concentrations [25].

Production and uses

The world refinery and mine production of most TEs except a few (e.g., As, Cd, Pb, Sn) have substantially increased these last decades (e.g., Fe, Al, Mo) and particularly since the beginning of years 2000 (Table 1). World demand for minerals is affected by 3 general factors: (i) uses for mineral commodities, (ii) the level of population that will consume these mineral commodities and (iii) the standard of living that will determine just how much each person consumes [27]. Today, with the integration of India, the People’s Republic of China and other populous developing and emerging countries (e.g., Brazil and Russia) into the world economy, more than 50% of the world’s population (instead of the previous 20%) account for the largest part of raw materials consumption [28,29]. The increasing demand for mineral raw materials further concerns numerous “emerging elements”. These elements can be “truly emerging” because they have just gained entry to the environment (new commercial uses and industrial releases) or have nowadays become contaminants “of emerging concern” while they were not in the past (new advances in analytical chemistry, new knowledge on their toxicity, new environmental compartments explored) [30,31].

The use of these emerging chemicals are multiple and diverse. For example, V is regarded as one of the hardest of all metals. This new knowledge on their toxicity, new environmental compartments [25].

Essential micronutrients may also show toxic effects when ingested at too high levels, as reviewed by Goldhaber [46] and widely detailed in Nordberg et al. [26]. The ingestion of very high doses of Cr causes liver and kidney problems. Abdominal pain, cramps, nausea, diarrhea, and vomiting have been caused by the consumption of beverages containing high levels of Cu and liver damage has been seen in individuals with diseases of Cu metabolism. The consumption of Mn-containing well water has caused lethargy, tremor, and mental disturbances in Japan risk of mortality from lung, bladder and kidney cancer and an increased [38]. For example, populations exposed to inorganic As via drinking water showed excess populations exposed to inorganic As via drinking water showed excess risk of mortality from lung, bladder and kidney cancer and an increased risk of skin cancer and other skin lesions such as hyperkeratosis and pigmentation changes [43]. Japanese who consumed Cd polluted rice and river water accumulated in their bodies a large amount of Cd that lead to a serious osteoporosis-like bone disease, the “itai-itai byo” or “ouch–ouch disease” [44] and in some of the great Cairo cities, Egypt, renal failure was related to Pb and Cd contamination of ground drinking water [45].

Sources of TEs in the sea

Continental runoff and atmospheric deposition are the primary natural inputs of TEs in the marine environment [47]: crustal material is either weathered on (dissolved) and eroded from (particulate) the Earth’s surface or injected into the atmosphere by volcanic activity. Forest fires and biogenic sources are of a lesser importance [48,49]. In addition to these natural sources, there exists a multitude of anthropogenic emission sources, the major ones resulting from mining and smelting activities [47]. Other important land-based anthropogenic sources of TEs result from the growth of industrial, agricultural and urban activities since the early-60s (e.g., Ref. [50-52]). According to the United Nations [53], more than 80% of the pollution of the seas comes from inland via the rivers or through runoff and discharges from the coastal areas. As at least 60% of the world’s population live within 100 km of the coast, the contamination of coastal waters may pose serious risks to human health as well as marine ecosystems [52].

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![Figure 1: Some essential and non-essential elements, abundant or in traces (modified after Ref. [25]).](image-url)

<table>
<thead>
<tr>
<th>Trace element</th>
<th>Symbol</th>
<th>1990</th>
<th>2000</th>
<th>△ (1%)</th>
<th>2010</th>
<th>△ (1%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aluminum</td>
<td>Al</td>
<td>17,817</td>
<td>24,400</td>
<td>37%</td>
<td>40,800</td>
<td>129%</td>
</tr>
<tr>
<td>Antimony</td>
<td>Sb</td>
<td>83.2</td>
<td>122</td>
<td>47%</td>
<td>167</td>
<td>101%</td>
</tr>
<tr>
<td>Arsenic</td>
<td>As</td>
<td>47.6</td>
<td>36.9</td>
<td>-23%</td>
<td>52.8</td>
<td>11%</td>
</tr>
<tr>
<td>Beryllium</td>
<td>Be</td>
<td>0.286</td>
<td>0.226</td>
<td>-21%</td>
<td>0.203</td>
<td>-29%</td>
</tr>
<tr>
<td>Bismuth</td>
<td>Bi</td>
<td>3.33</td>
<td>3.75</td>
<td>13%</td>
<td>8.47</td>
<td>154%</td>
</tr>
<tr>
<td>Cadmium</td>
<td>Cd</td>
<td>20.2</td>
<td>20.2</td>
<td>0%</td>
<td>21.4</td>
<td>6%</td>
</tr>
<tr>
<td>Chromium</td>
<td>Cr</td>
<td>12,846</td>
<td>4,320</td>
<td>-66%</td>
<td>7,290</td>
<td>-43%</td>
</tr>
<tr>
<td>Cobalt</td>
<td>Co</td>
<td>37.1</td>
<td>33.3</td>
<td>-10%</td>
<td>89.5</td>
<td>141%</td>
</tr>
<tr>
<td>Copper</td>
<td>Cu</td>
<td>8,815</td>
<td>13,200</td>
<td>50%</td>
<td>16,000</td>
<td>82%</td>
</tr>
<tr>
<td>Iron</td>
<td>Fe</td>
<td>543,000</td>
<td>1,061,148</td>
<td>95%</td>
<td>2,590,000</td>
<td>377%</td>
</tr>
<tr>
<td>Lead</td>
<td>Pb</td>
<td>3,367</td>
<td>3,100</td>
<td>-8%</td>
<td>4,140</td>
<td>23%</td>
</tr>
<tr>
<td>Manganese</td>
<td>Mn</td>
<td>27.2</td>
<td>20.2</td>
<td>-26%</td>
<td>42.7</td>
<td>57%</td>
</tr>
<tr>
<td>Molybdenum</td>
<td>Mo</td>
<td>112</td>
<td>129</td>
<td>16%</td>
<td>242</td>
<td>117%</td>
</tr>
<tr>
<td>Nickel</td>
<td>Ni</td>
<td>1,029</td>
<td>1,250</td>
<td>21%</td>
<td>1,590</td>
<td>54%</td>
</tr>
<tr>
<td>Selenium</td>
<td>Se</td>
<td>1,789</td>
<td>1,460</td>
<td>-18%</td>
<td>2,120</td>
<td>19%</td>
</tr>
<tr>
<td>Silver</td>
<td>Ag</td>
<td>17.7</td>
<td>18.4</td>
<td>4%</td>
<td>23.1</td>
<td>31%</td>
</tr>
<tr>
<td>Tin</td>
<td>Sn</td>
<td>219</td>
<td>238</td>
<td>9%</td>
<td>265</td>
<td>21%</td>
</tr>
<tr>
<td>Vanadium</td>
<td>V</td>
<td>31.0</td>
<td>43.0</td>
<td>39%</td>
<td>57.6</td>
<td>86%</td>
</tr>
<tr>
<td>Zinc</td>
<td>Zn</td>
<td>7,325</td>
<td>8,730</td>
<td>19%</td>
<td>12,000</td>
<td>64%</td>
</tr>
</tbody>
</table>

### Potential human health effects

**Al**: WHO: 0.9 (<0.1 in conventional treatment plants; <0.2 in other treatment types) ppm from the WHO, EPA, HC. Erosion of natural deposits; Al salts used as flocculents during the treatment of drinking water. Little indication that orally ingested Al is acutely toxic to humans; no adverse health effect at levels found in drinking water; Al exposure is a risk factor for the development or acceleration of onset of Alzheimer disease.

**Sb**: WHO: 0.020; EPA, HC: 0.006 ppm. Erosion of natural deposits; discharge from petroleum refineries; fire retardants; ceramics; electronics; solders; contaminants from pipes and fittings. Increase in blood cholesterol; decrease in blood sugar; microscopic changes in organs and tissues (thyroid, kidney, liver, spleen, thyroid).

**As**: WHO, EPA: 0.01 ppm. Erosion of natural deposits (erosion and weathering of soils, minerals, ores); runoff from orchards; runoff from glass and electronic production wastes. Skin damage or problems with the circulatory system; increased risk of getting cancer (lung, bladder, liver, skin - classified as human carcinogen); neurological effects (numbness and tingling of extremities).

**Be**: WHO: 0.010; EPA: 0.004 ppm. Discharge from metal refineries and coalburning factories; discharge from electrical, aerospace and defense industries. Intestinal lesions; rarely found in drinking-water at concentrations of health concern.

**Bi**: No guideline value. Concentrations of Bi in drinking water have not been reported. Doses used in medicines are very much larger than the estimated dietary exposure; dietary exposures to Bi are unlikely to be of toxicological concern.

**Cd**: WHO: 0.003; EPA, HC: 0.005 ppm. Erosion of natural deposits; corrosion of galvanized pipes; discharge from metal refineries; runoff from waste batteries and paints; leaching from solids or black polyethylene pipes; industrial and municipal wastes. Kidney damage; softening of bone; classified as human carcinogen.

**Cr**: WHO, HC: 0.05; EPA: 0.01 ppm. Erosion of natural deposits; releases or spills from industrial uses (steel and pulp mills). Enlarged liver; irritation of the skin, respiratory and gastrointestinal tracts; kidney problems.

**Co**: No guideline value. Drinking water has a low-Co content, usually between 0.0001 and 0.0050 ppm. Cardiovascular effects (cardiogenic shock, sinus tachycardia, left ventricular failure, and enlarged hearts) observed in people who consumed large amounts of beer over several years time containing Co sulfate as a foam stabilizer; gastrointestinal effects (nausea, vomiting, and diarrhea), effects on the blood, liver injury, and allergic dermatitis have also been reported in humans from oral exposure to Co.

**Cu**: WHO: 2; EPA: TT Action Level=1.3; HC<1.0 ppm. Erosion of natural deposits (erosion and weathering of rocks and minerals); corrosion of household plumbing systems; contaminants from pipes and fittings; acidic mine water drainage; landfill leachates; sewage effluents; iron-related industries. Short term exposure: gastrointestinal distress; long-term exposure: liver or kidney damages. Cu is an essential element in human metabolism; adverse health effects occur at levels much higher than the aesthetic objectives.

**Fe**: HC: aesthetic objectiveness0.3 ppm. Erosion of natural deposits (erosion and weathering of rocks and minerals); use of Fe coagulants; corrosion of steel and cast iron pipes. Not of health concern at levels causing acceptability problems in drinking-water.
Removal processes from seawater

Unlike organic pollutants that can be degraded to less harmful components by biological or chemical processes, TEs are considered as non-degradable pollutants [19,54]. This persistent character of TEs can alter, sometimes quite strongly, their natural biogeochemical balance in contaminated environments [55]. Processes removing TEs from seawater firstly include active biological uptake processes [56]. TEs are mainly transported into biological cells in ionic form through ionic channels. In addition, specific transport mechanisms cross the membrane barrier like binding with membrane carrier proteins or transport through hydrophilic membrane channels. Lipid-soluble (non-polar) metal forms including alkyl-TE compounds and neutral, lipophilic, inorganically complexed TE species can cross biological membrane by diffusion. TEs bound to very fine particles can also be engulfed by endocytosis [57,58]. Following absorption, TEs are transported to internal organs for utilization, storage, toxic effects, and possibly release [57].

TEs can further be removed from seawater through passive scavenging, i.e., the combined process of surface adsorption onto a wide variety of relatively high affinity surface sites on both living and dead particulate material followed by particle settling [56,59,60]. Much of this particulate material (along with its associated TEs) is recycled either in the water column or in superficial sediments. Labile bound TEs can desorb from settling particles and resupply free TEs to the dissolved pool [57]. Marine sediments can also act as a source of TEs by releasing chemicals back to the overlying water column [54,61]. The primary flux processes between sediments and the water column are resuspension and deposition, bioturbation, advection, upwelling/downwelling, diagenesis reactions, and diffusion [61]. Because of these remobilization processes, the effects of metal pollution on local environments and organisms can be substantial and long lasting in spite of years of restoration efforts [54].

**TE toxicity on aquatic biota**

TE metabolism and toxicity testing: The potential impact of a contaminant on aquatic biota depends on the total concentration of the contaminant, its speciation, interactions at receptors sites (e.g., at a fish gill membrane or on an algal cell surface) and uptake into the organism/cell, with either subsequent adverse effects or intracellular detoxification [62]. The toxicity of each TE will therefore be related to an organism-specific, metabolically available threshold concentration [63]. TEs accumulated into an organism occur in a bioreactive fraction that is metabolically active and available and a fraction that has been

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**Table 2:** Trace element guideline values (ppm) for drinking water, common sources of trace elements in drinking water and potential effects on human health. Data compiled from the World Health Organization (WHO) [37], the United States Environmental Protection Agency (EPA) [38], Health Canada (HC) [40], Nordberg et al. [26] and the Committee on Toxicity of Chemicals in Food, Consumer Products and the Environment (COT) [41]. Treatment Technique (TT) Action Level: US drinking water requiring a treatment process in order to reduce the level of a contaminant.

<table>
<thead>
<tr>
<th>Element</th>
<th>WHO</th>
<th>EPA</th>
<th>Action Level</th>
<th>Drinking-water source</th>
<th>Health concern</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pb</td>
<td>0.010</td>
<td>0.015</td>
<td>Erosion of natural deposits; corrosion of plumbing systems (pipes, solder, brass fittings and lead service lines); contaminants from pipes and fittings.</td>
<td>Infants and children (under 6 years); delays in physical or mental development; neurobehavioural effects; children could show slight deficits in attention span and learning abilities. Adults: kidney problems; high blood pressure. Others: anaemia; central nervous system effects; in pregnant women, can affect the unborn child; classified as probably carcinogenic to humans.</td>
<td></td>
</tr>
<tr>
<td>Mn</td>
<td>0.4</td>
<td>0.015</td>
<td>Erosion and weathering of rocks and minerals; naturally occurring in many surface water and groundwater sources, particularly in anoxic or low oxidation conditions (the most important source for drinking-water).</td>
<td>Not of health concern at levels causing acceptability problems in drinking-water.</td>
<td></td>
</tr>
<tr>
<td>Ni</td>
<td>0.07</td>
<td>0.002</td>
<td>Ni levels range from 0.002 to 0.010 ppm in fresh and tapwater; water is generally a minor contributor to the total daily oral intake; the Ni contribution from water may be significant where there is heavy pollution, in areas where Ni occurs naturally in groundwater, where there is use of certain types of kettles, of non-resistant material in wells or when water has come into contact with Ni-plated taps; released from fittings; released from industrial Ni deposits.</td>
<td>Occurs in drinking-water at concentrations well below those of health concern.</td>
<td></td>
</tr>
<tr>
<td>Se</td>
<td>0.04</td>
<td>0.05</td>
<td>Naturally occurring (erosion and weathering of rocks and soils); discharge from petroleum and metal refineries; discharge from mines.</td>
<td>Toxic effects: hair or fingernail losses at extremely high levels of exposure; numbness in fingers or toes; circulatory problems.</td>
<td></td>
</tr>
<tr>
<td>Ag</td>
<td>Available data inadequate to permit derivation of health-based guideline value; EPA: 0.1</td>
<td>Water-soluble Ag compounds such as the nitrate have a local corrosive effect and may cause fatal poisoning if swallowed accidentally.</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>V</td>
<td>No guideline value</td>
<td>Drinking-water is not a significant source of Sn; increasing use of Sn in solder, which may be used in domestic plumbing, and proposed for use as a corrosion inhibitor.</td>
<td>Occurs in drinking-water at concentrations well below those of health concern; main adverse effect of excessive levels of Sn in canned beverages or other canned foods has been acute gastric irritation; no evidence of adverse effects in humans associated with chronic exposure to Sn.</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Zn</td>
<td>HC: aesthetic objective &lt; 0.05</td>
<td>Naturally occurring; industrial and domestic emissions; leaching may occur from galvanized pipes, hot water tanks and brass fittings; Zn concentrations in water from active or inactive mines can be substantial.</td>
<td>Not of health concern at levels found in drinking-water; effects on human health by contamination on water supplies must be rare.</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

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detoxified and is unavailable (Figure 2) [64]. TEs bound to inducible TE-binding proteins such as metallothioneins or phytocelatins or precipitated into insoluble concretions consisting of TE-rich granules are biologically detoxified; on the contrary, TEs bound to sensitive fractions such as organelles and heat-sensitive proteins can be metabolically active. The higher the proportion of TE-sensitive fractions, the greater the vulnerability to TE toxicity [65,66]. The TE-sensitive fractions and TEs bound to metallothioneins are available for trophic transfer to predators [66]. This compartmentalization of TEs as defined by Wallace and Luoma [66] and Wallace et al. [65] is a useful tool to interpret multiple ecotoxicological consequences of the subcellular partitioning of TEs within organisms.

Mayer-Pinto et al. [67] critically reviewed studies on the effects of TEs on aquatic assemblages and/or populations of invertebrates. They pointed out that most studies in the field had been descriptive: they generally demonstrated that the diversity of an assemblage tended to decrease with an increase of environmental TE pollution and that there were differences in the structure of assemblages facing high pollution levels. Such descriptive studies are, however, unable to demonstrate any causal relationship between the environmental pollution and the changes observed [67]. Toxicity testing methods are therefore required as a tool for predicting and assessing the impacts of anthropogenic environmental stressors on organisms and ecosystems [25]. Laboratory-based toxicity testing has the obvious advantage to be unaffected by habitat or natural disturbances. This allows for experimental evaluation of various parameters and conditions such as temperature, toxicant threshold effect levels, mixture interactions, life stages or exposure duration under strictly controlled conditions [61]. Despite laboratory studies show lethal and sub-lethal effects of TEs on organisms, extrapolating such findings to the field is however little reliable [68]. Indeed, the exposure of aquatic organisms to contaminants is mostly episodic due to changing water and sediment quality. In addition, physicochemical characteristics of aquatic ecosystems (e.g., temperature, pH, water hardness, dissolved organic carbon) greatly influence contaminant bioavailability, toxicity and bioaccumulation [69-71]. The naturally occurring variability of water and sediment properties affecting TE chemical bioavailability cannot be easily simulated in the laboratory [72]. Additional field experiments are therefore necessary to validate laboratory results under relevant environmental conditions.

Complementary results from laboratory and field toxicity tests are useful for decision making, particularly if the responses of the test organisms are severe and occur in multiple species [61]. Within this perspective, detailed reviews on tested organisms from different taxonomic groupings allow to identify species that may be suitable candidates in a suite of toxicity test protocols [73] and to highlight toxicity knowledge gaps that require to be addressed before using the retained species in routine toxicity test procedures [74]. Such a work of synthesis was performed for the Australian coastal waters [74,75]. Thus, Van Dam et al. [74] reviewed the toxicity testing methods for water column contaminants, summarizing data available for 16 taxonomic groupings, among them vascular plants (seagrasses and mangroves) and bivalve molluscs. Moreover, Adams and Stauber [75] reviewed the whole sediment toxicity tests developed with numerous species of 7 taxonomic groupings, of which bivalve molluscs. These authors reported that bivalve molluscs were particularly relevant test organisms for water and sediment toxicity tests and that seagrasses, as strong accumulator of TEs, could act as integrated markers for environmental TE exposure.

**TE toxic effects on bivalve molluscs:** Bivalve molluscs show many physiological attributes (sensitive to contaminants, tolerant of a wide range of abiotic factors, easy to grow and maintain in a laboratory etc.) that make them appropriate bioassay organisms for toxicity testing [76]. They are moreover good accumulators of organometallic contaminants and TEs due to their behaviour and mode of feeding [77]. A number of standardized toxicity test protocols have been developed for determining toxicity of single chemicals, complex effluents and ambient samples of water or sediments to marine bivalves (e.g., Ref. [61]; detailed guidance manuals available from the US Environmental Protection Agency (EPA) and the American Society for Testing and Materials (ASTM) [78]). Toxicity tests established on bivalve embryo-larval developments are among the most sensitive in the EPA’s national toxicity dataset, which is used to derive water quality criteria [79]. Among bivalve molluscs, mussels from the genus Mytilus have been largely used. TE toxicity on bivalve molluscs can be determined at different structural levels, from genes to individuals, and response parameters have included, in addition to larval-embryo developments, changes in growth rates, clearance rates and survival rates, DNA-damages, or changes in tissue morphology and in specific component immunoreactivity (Table 3) [79-88].

Toxicity varies greatly between TEs. The median effect concentrations (EC50) for *Meretrix meretrix* embryogenesis is 188 times higher for Cd than for Hg. Moreover, this difference in toxicity directly relies upon the response parameter of interest. Thus, the EC50 for *M. meretrix* larval growth is only 6 times higher for Cd than for Hg [83]. TE toxicity also fluctuates spatially and over time with water properties, as demonstrated for Cu. The EC50 obtained by Cu spiking of ambient water samples for mussel embryo development was lower at sites located near the mouth of the San Diego Bay (California, USA) compared to sites at the back of that Bay. This increase was consistent with similar increasing trends in dissolved organic carbon (DOC) and total suspended solids [79]. This protective effect of DOC on Cu toxicity, experimentally demonstrated with contaminated filtered seawater spiked with DOC, appeared related to higher fulvic acid and lower humic acid contents [80].

**TE toxic effects on seagrasses:** Current knowledge on aqueous toxicity of TEs in seagrasses consists mainly of single TE exposures
tested on 8 species (Table 4) [89-96]. Experimental designs have varied considerably due, in part, to the lack of standardized toxicity tests for marine vascular plants contrary to bivalve molluscs. Test durations were between 6 hours and 51 days and response parameters included photosynthetic activity, amino acid concentrations, tissue growth, pigment contents or leaf cell mortality. Cu, Cd, Pb, and Zn were the most commonly tested compounds with Cu toxicity being particularly high, leading to seagrass leaf necrosis and decay (e.g., Ref. [93,94]). Interspecific differences in sensitivity to the same TE have been reported. Prange and Dennison [91] incubated 5 seagrass species (Halophila ovalis, Halophila spinulosa, Halodule uninervis, Zostera capricorni and Cymodocea serrulata) with Fe or Cu (1 ppm); seagrass responses to pollutants were assessed by changes in PSII photochemical efficiency and free amino acid content. Fe additions only affected Halophila spp. while Cu additions affected the other seagrass species as well. TE sensitivities can even differ between populations of a same species. Macinnis-Ng and Ralph [94] in situ monitored the effects of Cu and Zn on 3 geographically isolated populations of Z. capricorni in Sydney area (Australia). Photosynthetic efficiency and chlorophyll pigment concentrations showed different sensitivities to Cu exposures at the 3 sites, with seagrasses from the least developed estuary being the most sensitive to Cu.

Marine vascular plants are still rarely used in ecotoxicological testing, primarily because of difficulties in culturing/adapting and testing with such large, slow growing organisms [74]. To overcome these difficulties, and for greater environmental relevance, more recent toxicity tests involved the in situ measurement of photosynthetic endpoints (using PAM fluorometry) on wild plants in chamber experiments (e.g., Ref. [93,94]). Field measurements of photosynthetic efficiency can moreover be easily used as an efficient overall ecocriterion of seagrass health (e.g., Ref. [97]). In addition, phytotoxic effect levels for sediment-bound chemicals, spiked or in a whole sediment matrix, are relatively unknown for seagrasses. But because concentrations of several anthropogenic chemicals in rooted sediments exceed sediment quality guidelines, there is a need to better understand the bioavailability

<table>
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<tr>
<th>Species</th>
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<th>Measured parameters</th>
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<tr>
<td>Mytilus galloprovincialis (laboratory)</td>
<td>water exposure: Cu; San Diego Bay (CA, USA)</td>
<td>embryo-larval development</td>
<td>48 h.</td>
<td>unfiltered seawater; nominal tested concentration range (µg L⁻¹): 0-50; filtered seawater used for reference toxicant tests</td>
<td>absence of ambient toxicity to bivalve embryos; reference toxicant test EC50 value: 6.43 ± 1.36 µg L⁻¹; EC50 of Cu spiked ambient water samples: 1.7 to 3.4 times lower at sites located near the mouth of the Bay compared to sites at the back of the Bay.</td>
<td>Normally developed bivalve larvae possess a hinged D-shaped shell (prodissoconch); differences between unfiltered Cu spiked water samples indicate a gradient in complexation capacity increasing from the mouth to the back of the Bay (consistent with similar increasing trends in DOC and TSS).</td>
<td>[79]</td>
</tr>
<tr>
<td>Mytilus trossolus (laboratory)</td>
<td>water exposure: Cu, Zn, Ni, Cd (tested separately)</td>
<td>embryo-larval development</td>
<td>48 h.</td>
<td>filtered seawater; measured concentration ranges (µg L⁻¹): Cu=1.1-71.0, Zn=5-576, Ni=2-760, Cd=DL-1-200; effect of DOC addition on metal toxicity tested for Cu</td>
<td>EC50 (in µg L⁻¹): 9.6 for Cu, 99 for Zn, 150 for Ni, and 502 for Cd; experimental addition of DOC reduced Cu toxicity.</td>
<td>Normally developed bivalve larvae possess a hinged D-shaped shell (prodissoconch); protective effects of DOC on Cu toxicity are influenced by their distinct physicochemical properties: protection appears to be related to higher fulvic acid and lower humic acid contents.</td>
<td>[80]</td>
</tr>
<tr>
<td>Mytilus galloprovincialis (laboratory)</td>
<td>water exposure: Cr</td>
<td>gill morphology and immunoreactivity to components involved in gill motility; total glutathione content; activities of GSH-related enzymes, of catalase, and of key glycolytic enzymes; mRNA expression of selected genes</td>
<td>96 h.</td>
<td>artificial seawater; nominal tested concentrations (µg L⁻¹): 0.1, 1, 10</td>
<td>Morphological, biochemical and molecular changes in mussel gills when exposed to concentrations ranging from 0.1 to 10 µg L⁻¹.</td>
<td>Progressive changes in gill morphology and in immunoreactivity to components involved in neurotransmission; increased activities of GSH-related enzymes and total glutathione content suggesting Cr detoxication/reduction at the site of metal entry; increased activity of glycolytic enzymes, indicating modulation of carbohydrate metabolism; significant changes in transcription of different genes (sex- and concentration-related differences).</td>
<td>[81]</td>
</tr>
<tr>
<td>Mytilus edulis (laboratory)</td>
<td>water exposure: Ni</td>
<td>clearance rate; haemolymph genotoxicity and cytotoxicity</td>
<td>120 h.</td>
<td>filtered seawater; nominal tested concentrations (µg L⁻¹): 4.6 (control), 18, 56, 180</td>
<td>56 µg L⁻¹: clearance rate decreases. 180 µg L⁻¹: NRR decreases and % tail DNA increases in mussel haemocytes.</td>
<td>Clearance rate is Ni concentration-dependent (decreased by 30 % at the highest concentration); NRR assays, designed to assess the viability of the cells based on the penetration of a weakly cationic dye across lysosomal membranes, indicate a cytotoxic response; Ni has a genotoxic effect on the integrity of the DNA in haemocytes.</td>
<td>[82]</td>
</tr>
<tr>
<td><strong>Meretrix meretrix</strong> (laboratory)</td>
<td><strong>Tellina deltoidalis</strong> (laboratory)</td>
<td><strong>Mytilus galloprovincialis</strong> (field)</td>
<td><strong>Mytilus spp.</strong> (field)</td>
<td><strong>Mytilus trossulus</strong> (field)</td>
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<tr>
<td>water exposure: Cd, Pb, Hg (tested separately)</td>
<td>metal spiked sediment exposure: Cd, Cu, Ni, Pb, Zn (tested separately); water exposure: Cu and Zn (tested separately)</td>
<td>sediment exposure: Ni, Kaštela and Trogir Bays (Croatia)</td>
<td>sediment elutrate (water soluble fraction) exposure: Zn, Ni, Cr, Cu, As, Pb, Hg, Ag, Cd, Se, San Francisco Bay (USA)</td>
<td>sediment exposure: As, Cr, Cu, Pb, Hg, Zn, Sn_{sub}^{1}, Puget Sound (WA, USA)</td>
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<tr>
<td>embryogenesis; survival, growth and metamorphosis of larvae</td>
<td>10 d.</td>
<td>MN test (toxin induced heightened MN frequency) and Comet assay (tail DNA) with mussel haemocytes</td>
<td>% normal larval development</td>
<td>juvenile mussel growth rate</td>
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<tr>
<td>24 h.: embryogenesis; 48 h.: metamorphosis; 96 h.: growth and survival</td>
<td>30 d.</td>
<td>na</td>
<td>48 h.</td>
<td>82 d.</td>
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<tr>
<td>unfiltered seawater; measured concentration ranges (µg L^{-1}): Hg=2-17.977; Cd=1-10.167; Pb=2-7.158</td>
<td>nominal metal concentrations in sediment tests (mg kg^{-1}): 75, 1, 300, 420, 1,000, and 4,000 for Cd, Cu, Ni, Pb, and Zn, respectively; measured metal ranges in seawater tests: 0-710 µg L^{-1} for Cu; 0-33 mg L^{-1} for Zn</td>
<td>na</td>
<td>Sediment quality guidelines used to evaluate the potential toxicity of sediments: concentrations &lt;ERL, between the ERL and ERM or &gt;ERM are rarely (&lt;11%), occasionally (16-18%) or frequently (48-52%) associated with toxicity, respectively; Ni usually above the ERM; As, Cr, Cu, and Hg often exceed their respective ERLs</td>
<td>Trace element ranges in sediments (µg g^{-1}): As: 0-57; Cr: 44-93; Cu: 66-965; Pb: 22-297; Hg: 0.13-1.95; Zn: 107-592; Sn_{sub}^{1}: 0.16-18.41.</td>
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<td>EC50 for embryogenesis (µg L^{-1}): 7.4 for Hg, 1.014 for Cd and 297 for Pb; 96 h. LC50 for D-shaped larvae (µg L^{-1}): 14 for Hg, 68 for Cd and 353 for Pb; growth retardation (µg L^{-1}): 18.5 for Hg, 104 for Cd and 197 for Pb; EC50 for metamorphosis: similar to 48 h. LC50; higher than 96 h. LC50.</td>
<td>Sediment exposure: absence of toxicity. Water exposure: at Cu concentrations of 50 and 230 µg L^{-1}, time to LC50=7 d. and 5 d. respectively; at Zn concentration of 13 mg L^{-1}, time to LC50=4 d.</td>
<td>na</td>
<td>Sediment quality toxicity varied spatially (decreasing from sites located at the back of the Bay near 2 main river mouths - no survival of mussel larvae - to sites located near the Bay entrance - no toxicity), no significant trends in larval development over time at most sites; larval bivalve toxicity was associated with metals in bulk sediments.</td>
<td>In contaminated sites, mussels had lower growth rates than the reference site mussels; Sn_{sub}^{1}, Cu: contaminants of greatest concern; Pb and Zn: contaminants of additional concern; statistically significant inverse relationship between growth rate and toxicity-normalized sediment contamination.</td>
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Embryo toxicity: the % of normal D-shaped larvae decreases when Hg, Cd, and Pb concentrations increase. Larval growth: dead larval with extruded velum and granulated tissues in the more toxic treatments; injury to the velum and swimming inhibition at lower concentrations; reduction in growth rate following exposure to most concentrations from 24 h. Concentration-dependent survival inhibition of larvae. Hg most toxic and Pb least toxic to metamorphosing larvae.

Survival to metal spiked sediments: 88–100 %; the absence of toxicity is consistent with the low sensitivity of T. deltoidalis to these metals in the dissolved intertidal phase and indicated that the exposure from the ingestion of metal-contaminated particles was not sufficient to cause toxicity over the 10-days period. Survival to water exposure: concentration-dependent survival; Cu toxicity higher than Zn toxicity.

Increased % of MN (defined as small round structures in the cytoplasm smaller than 1/3 of the nucleus diameter) compared to mussels from the reference location evident for most of the contaminated locations; increase in % of tail DNA in individuals collected from mostly all the polluted sites; the 2 methods complement each other and it is desirable to use them both in monitoring the impacts of genotoxic pollution.

In contaminated sites, mussels had lower growth rates than the reference site mussels; Sn_{sub}^{1}, Cu: contaminants of greatest concern; Pb and Zn: contaminants of additional concern; statistically significant inverse relationship between growth rate and toxicity-normalized sediment contamination.

**Table 3**: Biological responses in bivalves exposed to dissolved and sediment trace elements under laboratory or field conditions. h.: hour; d.: day; na: not applicable; EC50: the median effect concentration; LC50: the median lethal concentration; DOC: dissolved organic carbon; TSS: total suspended solids; DL: detection limit; Glutathione (GSH)-related enzymes; NRR: neutral red retention; MN: micronucleus; ERL: effects range-low; ERM: effects range-mediated.
and phytotoxicity of sediment-bound contaminants [72].

The Monitoring of the Marine Environment

Biomonitoring

Until the early 70s, the monitoring of terrestrial and marine environments mainly relied on the detection and quantification of pollutants in physical environments - air, water, soils and sediments [25,98]. During the years 80s, almost all environmental monitoring networks stopped monitoring the water itself to assess the quality of aquatic ecosystems. Reasons for this abandonment were diverse, among them: the measured concentrations of chemicals are generally low (in the order of a few ng L$^{-1}$ for most TEs) and often close to the limits of detection of the analytical techniques; the risk of secondary accidental contamination makes the measurements sensitive; the concentrations of dissolved substances may vary considerably over time (e.g., with tidal cycles, water run-off, seasons etc.) and episodic pollution events can be missed; the measurement of dissolved pollutants do not provide an assessment of the portion which is available for uptake and accumulation by aquatic organisms [25,99]. The analysis of sediments overcomes some of these disadvantages. Contaminants accumulated in sediments, particularly in organically rich sediments, are more easy to monitor and much less susceptible to accidental contaminations. Sediments also offer a degree of time integration. However, sediment accumulation of contaminants is much affected by sediment characteristics (particle size, mineralogy, organic carbon content) and measured concentrations again do not provide an assessment of their bioavailable fractions [25,99].

When compared with the conventional chemical analysis of water and sediments, the monitoring relying upon the biota exhibits obvious predominance. Biomonitoring reveals the biological changes of organisms affected by chemicals and the integrated effects of multiple pollutants on these latest; has high sensitivity because of the rapid responses induced in organisms exposed to pollutants; realizes the monitoring of pollutants at low levels because of their chronic toxicities under long-term exposure; allows widely sampling even at remote areas; avoids the limits of the conventional chemical analysis such as continuous sampling, needs of expensive instruments etc. [100].

Bioindicators

Since the ultimate purpose of pollution monitoring is the protection of ecosystems and human beings, the main interest of the use of quantitative sentinel organisms with regard to water or sediments is their capacity to give information on the bioavailability of pollutants [101]. Currently, the term “bioindicator” is a deeply ambiguous term which has different meanings in different contexts [102]. To prevent problems due to different interpretations of this term, we use the definition of Blandin [103]: “a biological indicator (or bioindicator) is an organism or a set of organisms that allows, by reference to biochemical, cytological, physiological, ecological or ethological variables, in a practical and safe way, to characterise the status of an ecosystem or an eco-complex and to highlight as early as possible their changes, natural or caused”. Bioindicators therefore allow to accurately assess the effects of anthropogenic activities on ecosystems.

To be considered as a good bioindicator of environmental contamination, the selected species must meet a number of criteria, as listed by Cossa [101] or Rainbow [104]: the sentinel organism should be a net strong accumulator of contaminants and should not regulate the total concentration of a contaminant in its body tissues; it should be sedentary and reasonably abundant; it should have a sufficiently long life to permit sampling of more than one-year class; it should be large enough to provide sufficient tissue for analyses and should bioaccumulate sufficiently to allow direct measurement without preconcentration; it should be resistant to handling stresses; a correlation should exist between the level of contaminants in the organism and in the surrounding environment; it should be tolerant of exposure to environmental variations in physicochemical parameters and the effects on the organism of these variations should be known. No single species however combines all these qualities, and a compromise must be found [101].

For marine pollution monitoring, the bioindicator species used belong to numerous taxonomic groupings of which micro- and macro-algae, seagrasses, ascidians, sponges, bivalve and gastropod molluscs, polychetes, crustaceans, fishes, seabirds, marine reptiles and mammals [105,106], each bioindicator showing some special merits when compared to the others [100]. In the two next sections, we will present two bioindicator species widely used in the monitoring of the health status of the coastal Mediterranean, from punctual surveys to international monitoring programs: the Neptune grass Posidonia oceanica and the Mediterranean mussel Mytilus galloprovincialis. These two bioindicators respond appreciably and quantitatively to the coastal pollution by TEs and complement one another: the two species accumulate contaminants dissolved in the water column; $P.$ oceanica, deeply rooted in sediments, also reflects the contamination of this compartment; mussels, as a filter feeder, accumulate contaminants from their particulate phase. Together, they give an estimate of the overall pollution (water, sediments, suspended matter) of Mediterranean coastal environments [107].

Posidonia oceanica

Biology

$P.$ oceanica is a marine magnoliphyte endemic to the Mediterranean [108,109]. It grows on sandy and rocky bottoms and forms patchy and continuous meadows regarded as one of the climax communities of the Mediterranean [110]. It colonizes large areas of the infralittoral floor from the surface to maximal depths of 45 m [110,111]. $P.$ oceanica beds cover a surface estimated between 25,000 and 50,000 km$^2$, i.e., between 1 and 2% of the Mediterranean [112], and is only missing in zones under the influence of large estuaries (Po, Rhone, Nile - diminution of salinity and increase of turbidity) [113]. The light and the transparency of the water are determining factors for its growth [114]. $P.$ oceanica has the same morphology as the other marine magnoliphytes: below-ground parts consist of roots for anchoring and rhizomes for mechanical support; above-ground parts consist of shoots bearing several leaves [115]. Rhizomes grow horizontally (competition for space: plagiotropic rhizome) or vertically (competition for access to the light: orthotropic rhizome). The progressive silting and the two types of rhizome growth result in a typical terraced formation called “matte” consisting of the intertwining of various strata of rhizomes, roots, and sediments [113].

$P.$ oceanica plays various ecological and functional roles (reviewed in Ref. [12,16,116]). First of all, $P.$ oceanica meadows are considered to be among the most productive ecosystems of our planet. This ecosystem is made up by the juxtaposition of two types of primary production: the net primary production of $P.$ oceanica which is on average of 420 g$\text{C}$ m$^{-2}$ year$^{-1}$ and can reach 1,300 g$\text{C}$ m$^{-2}$ year$^{-1}$; the net primary production of the epiphytes which is between 100 and 500 g$\text{C}$ m$^{-2}$ year$^{-1}$. On a global scale, only seagrass ecosystems display this specific feature. $P.$ oceanica exhibits structural roles: $P.$ oceanica leaf
### Trace Elements in Marine Environments: Occurrence, Threats and Monitoring with Special Focus on the Coastal Mediterranean


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<tr>
<td><em>Zostera marina</em> (laboratory)</td>
<td>Cu, Cr, Cd, Hg, Zn, Pb (tested separately)</td>
<td>growth rate</td>
<td>0.5, 2, 5, 8, 12 and 19 d.</td>
<td>0.1, 0.5, 5 and 50 µM</td>
<td>Growth rate inhibition - Cd: after 12 d. at 5 µM and after 8 d. at 50 µM; Cu: after 5 d. at 5 µM and after 2 d. at 50 µM; Hg: rapid effect at all concentrations; Zn: after 2 d. at 50 µM.</td>
<td>Growth rate inhibited by Cd, Cu, Hg, Zn; no effect of Cr and Pb exposures; toxicity of metals decreases in the order: Hg ≥ Cu &gt; Cd ≥ Zn &gt; Cr and Pb; cellular substances are leached to the water and plants turned black in the Cu 50 µM experiment and in the Hg 5 µM and 50 µM experiments.</td>
<td>[99]</td>
</tr>
<tr>
<td><em>Halophila stipulacea</em> (laboratory)</td>
<td>Al</td>
<td>leaf cell viability</td>
<td>12 d.</td>
<td>from 10^{-4} to 10^{-8} mol L^{-1}</td>
<td>Cellular damages from 10^{-4} to 10^{-8} mol L^{-1}.</td>
<td>Protoplast necrosis in all cell categories (except in the mid-rib cells); plasmatic resistance decreases in the order mid-rib, mesophyll, epidermal, teeth cells.</td>
<td>[90]</td>
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<tr>
<td><em>Halophila ovalis</em>, <em>Halophila spinulosa</em>, <em>Halodule uninervis</em>, <em>Zostera capricorni</em> and <em>Cymodocea serrulata</em> (laboratory)</td>
<td>Fe, Cu (tested separately)</td>
<td>changes in PSII photochemical efficiency (Fv/Fm) and free amino acid content</td>
<td>12 d. of exposure; 5 d. of recovery</td>
<td>1 mg L^{-1} + EDTA</td>
<td>Fe and Cu: 1 mg L^{-1}; decline in PSII photochemical efficiency and in amino acid contents; effects are species-specific.</td>
<td>Fe addition experiments: declines in PSII photochemical efficiency in <em>H. ovalis</em> and <em>H. spinulosa</em> correspond with the replacement of fresh seawater (12 days), suggesting that these species became acclimated to the new environmental conditions; <em>Z. capricorni</em> exhibited a decline in total free amino acid contents (could be a precursor signal of Fe induced stress). Cu addition experiments: Fv/Fm ratio response was highly variable between the 5 seagrass species (death of <em>H. spinulosa</em>); decline in amino acid concentrations in <em>Z. capricorni</em> and <em>H. uninervis</em>.</td>
<td>[91]</td>
</tr>
<tr>
<td><em>Halophila ovalis</em> (laboratory)</td>
<td>Cu, Cd, Pb, Zn (tested separately)</td>
<td>chlorophyll a fluorescence; pigments (chlorophyll a, b, and carotenoids)</td>
<td>4 d.</td>
<td>1, 5 and 10 mg L^{-1}</td>
<td>Cd - 1 to 10 mg L^{-1}; limited stress. Cu - 5 and 10 mg L^{-1}; lethal effect. Pb - 1 to 10 mg L^{-1}; limited effect on fluorescence; chl. a, b decrease. Zn - 1 to 10 mg L^{-1}; changes to the chl. a fluorescence responses; various effects on pigment contents.</td>
<td>Variety of effects on the photosynthetic processes, with Cu and Zn having greater effects than Pb and Cd; quantum yield is the most sensitive measure of the photosynthetic processes; pigment contents generally confirm the chlorophyll a fluorescence responses.</td>
<td>[92]</td>
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<tr>
<td><em>Zostera capricorni</em> (field)</td>
<td>Cd, Cu, Pb, Zn (tested separately); 1 reasonably pristine site at Pittwater (NSW, Australia)</td>
<td>photosynthetic efficiency (ΔF/Fm′); pigments (chlorophyll a, b, and carotenoids)</td>
<td>10 h. of exposure; 3 d. of recovery</td>
<td>0.1 and 1 mg L^{-1}</td>
<td>Cu, Zn - 0.1 and 1 mg L^{-1}; photosynthetic efficiency decreases during the exposure period; Cu - 0.1 and 1 mg L^{-1}; after 96 h., carotenoid pigments decline, the chlorophyll a/b ratio is depressed and the chlorophyll a carotenoid ratio is elevated.</td>
<td>Samples exposed to Zn recover to pre-exposure levels but those exposed to Cu do not; browning of leaves and some leaf loss occurred due to exposure to Cu; Cd and Pb do not have impact on the chlorophyll a fluorescence and the pigment data support these findings.</td>
<td>[93]</td>
</tr>
<tr>
<td>3 isolated populations of <em>Zostera capricorni</em> (field)</td>
<td>Cu, Zn (tested separately); 1 semi-pristine and 2 impacted sites in Sydney area (Australia)</td>
<td>photosynthetic efficiency (ΔF/Fm′); pigments (chlorophyll a, b, and carotenoids)</td>
<td>10 h. of exposure; 3 d. of recovery</td>
<td>0.1 and 1 mg L^{-1}</td>
<td>Cu in the semi-pristine site - 0.1 mg L^{-1}; fluorescence decreases; 1 mg L^{-1}; chlorophyll concentration decreases. Cu in the 2 impacted sites - 1 mg L^{-1}; fluorescence decreases.</td>
<td>Lack of response due to Zn exposure; different sensitivities to Cu: greater impact of Cu on the more naive population (higher decrease of fluorescence during exposure and lower recovery).</td>
<td>[94]</td>
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</tbody>
</table>
canopy acts as a sediment trap; *P. oceanica* meadows reduce the coastal erosion and stabilize coastlines through direct effect on wave motion and through the formation of "banquettes" (wedge-shaped deposits of *P. oceanica* leaf litter). *P. oceanica* shoots build up structurally complex ecosystems, providing adequate life conditions and ecological niches for an important number of organisms. *P. oceanica* plays a crucial role in the coastal biogeochemical cycles: it modifies chemical properties (nutrient, oxygen, organic matter and dissolved inorganic carbon concentrations) of both water column and sediments; the high diurnal rates of oxygen production support respiration of a significant amount of heterotroph organisms; its presence enhances the nutrient recycling by the heterotroph bacteria; the important formation of matte, and associated long-term burial of organic matter, is a significant carbon sink. *P. oceanica* is also an important food supplier; direct herbivory, generally regarded as limited (only 10% - up to 70% locally - of living *P. oceanica* organic matter would enter the food webs); detritivory (detritivores feeding on litter rely on micro-organisms colonizing detritus to achieve nutritional balance); epiphyte consumers (epiphytes can represent more than 40% of the total foliar biomass of *P. oceanica*).

### Seagrasses as bioindicator species

*Posidonia oceanica* descriptors: *P. oceanica* is used since decades as a powerful integrator of the overall quality of Mediterranean marine coastal waters (e.g., Ref. [117-120]). This large-size long-living species, sedentary and abundant, colonizes the major part of the Mediterranean coasts where it can be easily collected. It is sensitive to chemical pollution and mechanical disturbances [12,117,121] and makes account, by its presence and its vitality (or its regression materialized by dead matte), of the quality of the Mediterranean coastal waters [12]. The footprint of the quality of waters on *P. oceanica* is permanent. Consequently, many environmental parameters may be recorded by *P. oceanica* meadows [12]. In 2005, 39 scientists from as many as 23 Research Centres synthetized in a common publication the descriptors of *P. oceanica* in order to better define the respective advantages of each one to assess the good ecological status of coastal zones [121]. Descriptors covered all the levels of organization of *P. oceanica* meadows, from the biochemical composition of the seagrass to the structure of the entire ecosystem.

In summary: chemical and biochemical composition of *P. oceanica* can provide information about the level of plant stress, and seems in adequacy with the level and impact of human activities; levels of contaminants provide information about the overall pollution of the meadow; leaf biometry is indicative of the environmental conditions (anthropisation, water motion, action of grazers etc.) and the dynamics and vegetative growth of the meadow; lepidochronology (cyclic changes along rhizomes) [122,123] and plastochnorale interval index (an interpolation method used to estimate leaf age) [124] provide information about the temporal evolution of primary production, sedimentation rates, sexual reproduction and dynamics of the meadow; structural characteristics of the meadow (shoot density, bottom cover, speed of growth of rhizomes and matte structure) provide information on the vitality, the macrostructure and dynamics of the meadow, as well as information on sedimentation, hydrodynamism, currents or human impacts; lower and upper limits provide pertinent information about the quality of the meadow and environmental changes, both natural and human-induced (e.g., water transparency, hydrodynamism, sedimentary balance, coastal developments, anchorage etc.); disappearance of the seagrass bed is indicative of the freshening at the outlets of coastal rivers or groundwater; associated species (fauna and flora) supply relevant information concerning the biodiversity of the meadow and the interactions meadow-species; epiphytes also provide information on water quality, especially data on nutrient inputs [12,121].

<table>
<thead>
<tr>
<th><em>Posidonia oceanica</em> (laboratory)</th>
<th>Cd</th>
<th>DNA methylation and chromatin reconfiguration: expression of PoMT2k and PoCMT1; nuclear chromatin ultrastructure</th>
<th>6 h., 2 d., 4 d.</th>
<th>10 and 50 µM</th>
<th>PoMT2k expression - Cd 50 µM: increase in PoMT2k expression after 6 h. in apical tips and leaves; Cd 10 µM: increase in PoMT2k expression after 2 d. in leaves. Changes in DNA methylation and in PoCMT1 expression: Cd 10 µM after 6 h. Chromatin reconfiguration: Cd 50 µM after 2 d.</th>
<th>Cd treatment induces a DNA hypermethylation (time- and dose-dependent) as well as an up-regulation of CMT, indicating that de novo methylation occurs; a high dose of Cd leads to a progressive heterochromatinization of interphase nuclei and apoptotic figures are observed after long-term treatment; Cd perturbs the DNA methylation status through the involvement of a specific metaltransferase; such changes are linked to nuclear chromatin reconfiguration likely to establish a new balance of expressed/repressed chromatin; the data show an epigenetic basis to the mechanism underlying Cd toxicity in plants.</th>
</tr>
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<tbody>
<tr>
<td>Halophila ovalis (laboratory)</td>
<td>Pb, Cu (tested separately)</td>
<td>growth rate; leaf fluctuating asymmetry and dimension</td>
<td>51 d.</td>
<td>Pb: 10 and 50 mg L⁻¹; Cu: 0.5, 2 and 4 mg L⁻¹</td>
<td>Growth rate decrease - Cu: 0.5 mg L⁻¹; Pb: 10 mg L⁻¹; Reduced leaf dimension - Cu: 0.5 mg L⁻¹; Pb: 50 mg L⁻¹; Increased leaf asymmetry - Cu: 2 mg L⁻¹</td>
<td>Reduced growth rate of the seagrass observed both in Pb and Cu treatments; leaf size of the plant reduces as the metal concentrations increase and when the plants are exposed to the metals for longer duration; increased leaf asymmetry more apparent at the 2 ppm Cu treatment; no increase in fluctuating asymmetry in Pb treatments; the mortality of leaves is especially high in Cu treatments.</td>
</tr>
</tbody>
</table>

Table 4: Biological responses in seagrasses exposed to dissolved trace elements under laboratory or field conditions. EDTA: ethylenediaminetetraacetic acid; PSII: photosystem II; PoMT2k: *Posidonia oceanica* Metallothionein (MT) 2k, an important metal tolerance gene; PoCMT1: one member of the *Posidonia oceanica* chromomethylase (CMT) family, a DNA methyltransferase. Fv: variable fluorescence; Fm: maximum fluorescence; ΔF: fluorescence yield; Fm'/Fm: light-adjusted maximal fluorescence.

**References:**

Natural and human-induced disturbances experimented by *Posidonia oceanica* meadows (e.g., eutrophication, decrease of water transparency, water motion) and the time of answer of their various related descriptors (hours to decades) vary greatly. A better understanding of the complexity and interactions of these disturbances thus requires a global approach combining different descriptors [121].

The choice of proper descriptors: Martínez-Crego et al. [125] identified from the literature 59 seagrass descriptors sensitive to environmental changes at different levels of biological organisation, for the most identical to the ones summarized in Pergent-Martini et al. [121]. Authors validated these descriptors on deep (15 m) and shallow (5 m) *P. oceanica* meadows from the Catalan coast (Spain) covering a wide anthropogenic gradient ranging from undisturbed to severely disturbed sites. Numerous descriptors were discarded: either because they failed to detect large scale (i.e., between-site) variability due to the masking effect of high spatial heterogeneity at smaller scales (i.e., variability between zones; e.g., number of leaves, amino acid contents in rhizomes etc.); either because they were influenced by natural sources of variability such as herbivory or physical settings and showed a low efficiency in ordering deep meadows (e.g., herbivore bite marks, leaf length and width etc.); or because, although they seemed to be linked to environmental status or to specific pollutants, responses of these indicators appeared to be influenced by interactions between different sources of pollution (e.g., interactions between metals and nutrients, interactions between different sources of anthropogenic nitrogen etc.). Among the 59 seagrass descriptors assessed, only 16 were finally unequivocally related to the environmental status gradient under study (among them, 7 concerned TEs); they were representative of physiological (e.g., carbohydrates), biochemical (e.g., TEs), individual (e.g., shoot necrosis), and population (e.g., meadow cover) levels of biotic organisation. Their combination was necessary to cover the entire environmental gradient and to reflect the multiple anthropogenic disturbances causing the gradient [125].

The choice of an adequate suite of indicators appears thenceforth decisive to ensure the consistency of multimetric indices and to provide an ecologically relevant interpretation of the response of biota to multiple stressors [125]. Such multimetric indices, however simplified (i.e., less descriptors) for convenience of application, have been experimentally assessed to experiment the ecological status of *P. oceanica* meadows, as for example: the POMI index (*Posidonia oceanica* multivariate index) [126,127], the BiPo index (Biotic index using *Posidonia oceanica*) [128], or the PREE index (*Posidonia oceanica* Rapid Easy Index) [129]. These indices need the destructive sampling of shoots from the meadow in order to measure some of the indicators required for their calculation. However, *P. oceanica* being a protected species, the monitoring of the state of conservation of the Mediterranean coastline using *P. oceanica* must therefore evolve toward the implementation of less invasive methods [130] and toward the development of non-destructive index, as proposed by Gobert et al. [131].

**TE bioaccumulation in seagrasses**

*Posidonia oceanica as a model:* In the Mediterranean, *P. oceanica* is regarded as a powerful indicator of bioavailable TEs since it highly bioaccumulates these chemicals. Thus, biocentration factors from filtered seawater to *P. oceanica* leaves ranged between 2,000 and 36,000 for Cd, Cr, Cu, Pb and Zn in two uncontaminated Italian areas [132,133]. *P. oceanica* further accumulates TEs at levels reflecting the contaminantion status of its environment (water and sediments) as shown from experiments and field studies (e.g., Ref. [134-139]). In addition, *P. oceanica* ability to record the past levels of contaminants coupled to the dating possibilities offered by the lepidochronology provide relevant biological archives about the temporal evolution of the coastal contamination in the Mediterranean (e.g., Ref. [140-143]).

Seagrass meadows can be conceptualized as the juxtaposition of 5 separate components, i.e., seagrass shoots, epiphytes, associated algae and animals, and detritus, exchanging flows of TEs between themselves and with their environment, i.e., water and sediments. This conceptualization can be drawn in energy circuit language (Figure 3). Each symbol (production, consumption, storage and flow) of the energy circuit language is rigorously and mathematically defined. By writing such a diagram, one, in essence, is writing equations describing a system. Varying the size of symbols further indicates their physical size and their importance [144,145]. No detailed study has so far modelled the cycling of TEs within *P. oceanica* meadows. Given that the exact quantification of flows and distribution of TEs between *P. oceanica* bed components remains partly poorly known, a similar size was given to symbols modelling their cycling (Figure 3) [146]. An ecological relevant size of the symbols could be gained by combining elemental analysis [147,148], TE uptake experiments [148] and mass balance analysis [147]. The model given here for *P. oceanica* community is derived from Schroeder and Thorhaug [148] work, the only detailed study to our knowledge of the global cycling of TEs within a seagrass (*Thalassia testudinum*) ecosystem. Since providing details on all the components of a *P. oceanica* meadow and their interconnecting flows would exceed the frame of this paper, particular attention will therefore be delivered to the processes directly related to the sole seagrass and its associated epiphytes. TE flows discussed in the next section will be referred to their corresponding lettered-numbered label given in Figure 3 model for clarity purpose.

**TE balance within Posidonia oceanica:** Sanz-Lázaro et al. [147] recently demonstrated the key role played by *P. oceanica* (as a species, not as an ecosystem) in the cycling of TEs in Mediterranean coastal waters. These authors calculated the TE incorporation rates in *P. oceanica* rhizomes, roots and new leaves from mean tissue concentrations and tissue production rates, subtracted TE loss rates through leaf shedding, mechanical breakage (CF1) and grazing (FF1), and extrapolated the balances obtained for their reference meadow to the whole Mediterranean (Table 5, according to the estimates of the total cover of 50,000 km² of *P. oceanica* meadows) [112]. Depending on the plant compartment where TEs were mainly accumulated and on their incorporation and loss dynamics, Sanz-Lázaro et al. [147] calculated that *P. oceanica* could act either as a sink (positive balance) or as a source (negative balance; TEs given back accessible to other components of the system or exported) for these chemicals.

Seagrass leaves provide an expanded area to sorb and sequester chemicals (UF1, UF2), and their root-rhizome system facilitates the absorption and accumulation of sediment contaminants (UF6) [68,72]. The extent to which these uptake processes are passive or subject to active physiological regulation will determine the final accumulation behaviour of seagrass compartments relative to the TE levels they are exposed to [149]. TEs accumulated by the leaf canopy and the root-rhizome system may afterward be translocated to below- (basipetal, TF2) or above-ground (acropetal, TF1) tissues, respectively [68,137]. TE concentrations in both above- and below-ground compartments further follow an annual cycle (e.g., Ref. [107,136,149,150]). This seasonality had been initially only attributed to variations in the plant growth dynamics that induced a dilution of the accumulated TEs [151]; but climatic patterns (seasonal rainfalls and storm frequency) leading
to changes in chemical loads in the water and sediments show an equal or greater influence on this seasonality [91,149].

The only few studies that have so far monitored the levels of TEs in *P. oceanica* epiphytes reported a more important accumulation of many TEs (e.g., V, Cr, Fe, As, Pb etc.) in these latest compared to the other seagrass compartments [137,147,149]. In addition to the direct uptake of TEs from the water column (UF2), it has been suggested that their leaching from the seagrass leaves could be an important route of transfert from the plant to its epiphytes (TF4) [152]. Seagrass leaves and more certainly epiphytes (as preferential food resource for numerous grazers) [153] further represent sources of potentially toxic chemicals to the grazer community (FF1, FF2). Since epiphytes are a key component of *P. oceanica* meadows and since they are ubiquitous on the leaves of seagrass species, they should be taken more into consideration when studying TE cycling in seagrass meadows [147].

Seagrass-accumulated chemicals and those associated with the epiphytic layer can be lost to the surrounding water in a dissolved form (LF1, LF2) or be exported bound to blade fragments (CF1, CF2) at senescence [72]. Some TEs like Zn, Cd, Sr or Rb show high release rates through decomposition of *P. oceanica* detritus and are expected to be released in the meadow (LF5); a contrario, others like Ca, Ti or Bi show low release rates through decomposition and are more likely to be buried (MF1) or exported to adjacent ecosystems [147]. Finally, the well-developed belowground system of *P. oceanica* roots and rhizomes forming mats can persist for thousands of years [154], thenceforth sequestering a fraction of the accumulated and potentially toxic TEs and reducing the total amount bioavailable to other organisms [147,155]. Seagrasses therefore act not only as biological filters but also as storage compartments, thereby favouring the decrease of environmental toxic substances [156].

Mytilus spp.

**Biology**

It is considered that there exist three species of mussels within the genus *Mytilus* – *M. edulis*, *M. galloprovincialis* and *M. trossulus*. *M. edulis* is distributed along the Atlantic coasts of Europe till high northern latitudes; *M. galloprovincialis* has a more southern repartition along the Atlantic coasts, from Britain to Morocco, and is distributed along the Mediterranean coasts. *M. trossulus* is distributed in the Baltic Sea [157]. In parts of Europe where species are sympatric, both hybridisation and introgression occur [157,158]. *M. galloprovincialis* has moreover become invasive in many parts of the world, including in Australia, Asia, California and the Puget Sound in the United States, and in South Africa [159]. Under optimal conditions, *M. edulis* and *M. galloprovincialis* attain a shell length of 100-130 mm, whereas in marginal conditions, mussels may measure as little as 20-30 mm. Feeding and respiration are carried out via currents of water directed across the gills. Food particles, trapped by cilia, are conveyed towards the labial palps and mouth. Mussels can also actively transport DOM across their gill membrane and utilise it as a nutritional supplement [160].

Reproduction in *Mytilus* spp. follows an annual seasonal cycle. Gametogenesis and energy storage occur in the mantle tissue, where a large shift in cell types (adipogranular cells and vesicular connective tissue vs. gametes) is evident throughout the annual cycle [161]. Sex of mature individuals can be told by the colour of their gonads: pink to orange for females and creamy-white to yellow for males [162]. After spawning, this sex segregation based on mantle colour is not possible anymore as mantle becomes thin and translucent [162,163]. In *M. galloprovincialis* collected in Galicia (Spain), spawning occurs from spring until late August or September [164]; along the French Mediterranean coasts, preponderant spawning occurs in January/February and partial spawning is observed in April/June [165,166]. As most studies regarding the uptake and elimination kinetics of TEs implicitly assume steady-state conditions for physiological processes, i.e., without a reproductive period [167], the knowledge of mussel reproductive cycle in the area under study is a necessary prerequisite to their use as bioindicator species.

**Mytilus spp. as bioindicator species**

**Quality indicators:** It is the unique combination of biomonitoring features *Mytilus* spp. exhibit that make them particularly well suited as indicator species [101,168]. They have a broad geographical distribution, ranging from temperate to subarctic regions, and colonize, as euryhalin, estuarine to fully marine waters. Because they are keystone species, it is likely that a pollutant that affects a mussel population will also exhibit a negative impact for the entire ecosystem. As long-living sessile organisms they can integrate the contaminations of their environment over long time periods. Mussels are relatively large, easy to handle, and can be used under laboratory and field conditions. They accumulate chemicals both from their diet (via the gastro-intestinal tract) and from the ambient water (via the tissue membranes in contact with water), exhibit only a limited ability to eliminate pollutants and reach higher bioaccumulation factors than other systematic groupings for many toxicants. Consequently, pollutants might exhibit negative

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**Figure 3:** Trace element (TE) cycling between the different components of a *P. oceanica* meadow. Model is drawn in energy circuit language (left) and symbol meaning is given (right) [145]. In such schematic representations, varying the size of symbols indicates their physical size and their importance [144]. Given that the quantification of flows and distribution of TEs between *P. oceanica* bed components remains partly poorly known, a similar size was given to each symbol. The 29 lettered-numbered TE flows between *P. oceanica* bed components are the following: Uptake Flow (UF) leaves (UF1), epiphytes (UF2), algae (UF3) animals (UF4) and detritus (UF5) from water, roots/rhizomes (UF6) and algae (UF7) from sediments, and sediments from water (UF8); Flow exchanges with adjacent ecosystems (FF1), epiphytes (FF2), algae (FF3) animals (FF4) and detritus (FF5) to sediments, and sediments from water (FF6); Feeding Flow (FF) leaves (FF1), epiphytes (FF2), algae (FF3) and animals (FF4) to detritus; Conversion Flow (CF) leaves (CF1), epiphytes (CF2), algae (CF3) and animals (CF4) to detritus; Mineralization Flow (MF) detritus to sediments (MF1). Flow exchanges with adjacent ecosystems are not shown (modified after Ref. [145]).
impacts on mussels at lower environmental concentrations, facilitating their use as an ecological early warning system. The histomorphology and the physiology of *Mytilus* spp. being well characterised, effects of environmental stresses can be measured at various levels of biological organisation (from molecules to communities). They further are non-controversial as organisms for ecotoxicological research. Finally, as shellfish products, they are a potential source of contaminant exposure for humans. For all these reasons, mussels are very widely used in programs monitoring the chemical contamination of the marine coastal environment (e.g., Ref. [169-172]). Furthermore, consensus methods developed for optimizing the use of *Mytilus* spp. as comparative quantitative indicators make their wide use as environmental monitoring tool strongly relevant [101,173].

**Pollution monitoring:** The level of contaminants in the flesh of mussels reflects the level of bioavailable contaminants in the water [174]. It results from a balance between the concentration in the organism and its environment, which depends on the processes of absorption, excretion and accumulation [101]. Based on the use of these molluscs for monitoring purposes, two types of monitoring strategies have been adopted: passive or active biomonitoring. Passive biomonitoring surveys rely on the use of native populations of wild or cultivated mussels (e.g., the Mussel Watch Program in the USA, Ref. [171], the RNO program in France, Ref. [175]); active biomonitoring surveys resort to transplants of individuals (e.g., RINBIO and MYTILOS programs in the Mediterranean, Ref. [176,177]). In the latter case, caged mussels are immersed on-site during their sexual dormancy for several months so they can accumulate contaminants to balance with their transplantation environment.

The biomonitoring of contaminants in coastal waters is mostly carried out by direct quantification of the accumulated pollutants within individuals (e.g., Ref. [176,178-182]). However, the monitoring of biological responses to pollutants (or biomarkers) at the molecular and cellular level has been shown to be a useful complementary tool in environmental quality evaluation and risk assessment [183,184]. Alteration in specific biomarkers reflect the type of pollution organisms are facing. For example, metallothioneins and the enzyme d-amino-n-valeric acid dehydratase are indicative of metal contamination, the mixed function oxidase system, glutathione S-transferase and acetylcholinesterase are indicative of organic contamination and superoxide dismutase, catalase, glutathione peroxidase and lipid peroxidation are biomarkers of oxidative stress. A multibiomarker approach therefore turns out to be a useful approach as complex mixtures of contaminants usually occur in the environment [185]. But the use of mussels as bioindicator species is not limited to the chemical contaminants. Thus, Kačar et al. [185] showed that *M. galloprovincialis* bioconcentrated microbial pollution indicators (heterotrophic bacteria and faecal coliforms) and pathogens (*Salmonella* spp.) and could therefore be used to prevent potentially harmful adverse effects of microorganisms from polluted waters and shellfish. Browne et al. [186] experimentally demonstrated that *M. edulis* bioaccumulated microplastics in its flesh. And Lassauque et al. [187] successfully traced sewage and natural freshwater inputs in a Northwest Mediterranean bay from carbon and nitrogen isotopic ratio measurements in *M. galloprovincialis*.

**Seafood and human health:** Mussels are commercially important seafood species. The determination of accumulated concentrations of contaminants in their flesh is essential because of the potential adverse effects of their consumption on human health [188,189]. In the particular case of metals, the maximum permissible limits fixed by the European Commission [190] in edible tissues of mussels are 0.5 mg kg\(^{-1}\) for Hg and 1 mg kg\(^{-1}\) for Cd and Pb (related to fresh weight). Comparison of the concentrations determined in the soft tissues of *M. galloprovincialis* sampled worldwide with the European legislation showed that the levels of these metals generally did not exceed the existing limits (detailed dataset in Ref. [159]). Nevertheless, these toxic elements might also pose some sanitary risks to consumers of shellfish purchased from contaminated hot spots that were identified in all the investigated seas [159,189].

**Trace element bioaccumulation in *Mytilus* spp.**

**Driving factors:** Since the mid-70s, *Mytilus* spp. have been widely used to monitor the chemical contamination of coastal and estuarine ecosystems (e.g., Ref. [101,171,176,177,191,192]). It is well known that TE levels in aquatic organisms represent a time-integrated response to bioavailable pollutants in food and water [193,194]. But these levels do not depend solely on their bioavailability in the environment, since biotic and abiotic factors are further acting [101]. In *Mytilus* spp., biotic factors involved in determining the levels of accumulated TEs are related to the organism growth (age, size, soft tissue weight) and reproduction (sex and gametogenesis). Environmental factors essentially revolve around seasonal cycles (temperature, primary production, salinity etc.), although other parameters may also be involved (e.g., properties of TEs and their interactions, the position of mussels in the intertidal etc.). The influence of these factors must be identified and quantified so that they may be taken into account during sampling, validation of results and monitoring data interpretation (Figure 4) [101]. The environmental biomonitoring is thenceforth made complex due to the diversity of contaminant characteristics (nature, speciation, concentrations and interactions), the diversity of ecological factors (abiotic and biotic) and their variations and interactions in space and time [195].

**Physiological correction:** According to Amiard et al. [169], the seasonal variability of body TE concentrations can be explained primarily by the fluctuations of the mussel body mass. Variations of the mussel flesh weight are related to the availability of food in the environment but also to their ecophysiological cycle including

<table>
<thead>
<tr>
<th>Source</th>
<th>Trace element</th>
<th>World prod.</th>
<th>Equivalence</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fe</td>
<td>1,891</td>
<td>2,590,000</td>
<td>0.07%</td>
</tr>
<tr>
<td>Ni</td>
<td>175</td>
<td>1,590</td>
<td>11%</td>
</tr>
<tr>
<td>Cr</td>
<td>30</td>
<td>7,290</td>
<td>0.41%</td>
</tr>
<tr>
<td>As</td>
<td>4.6</td>
<td>52.8</td>
<td>8.70%</td>
</tr>
<tr>
<td>Ag</td>
<td>3.6</td>
<td>23.1</td>
<td>16%</td>
</tr>
<tr>
<td>Pb</td>
<td>-8</td>
<td>4,140</td>
<td>0.19%</td>
</tr>
<tr>
<td>Cd</td>
<td>-11</td>
<td>21</td>
<td>53%</td>
</tr>
<tr>
<td>Co</td>
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<td>V</td>
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<td>58</td>
<td>67%</td>
</tr>
<tr>
<td>Cu</td>
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<td>16,000</td>
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</tr>
<tr>
<td>Mn</td>
<td>-587</td>
<td>43</td>
<td>1375%</td>
</tr>
<tr>
<td>Zn</td>
<td>-1,459</td>
<td>12,000</td>
<td>12%</td>
</tr>
</tbody>
</table>

Table 5: Annual balances of trace element (TE) amounts in *Posidonia oceanica* (tons y\(^{-1}\)) for the whole Mediterranean [147]. Positive or negative amounts indicate either incorporation or release by *P. oceanica*, respectively. These balances, expressed in equivalent % to the 2010 mean world production (tons y\(^{-1}\), see Table 1), reflect the quantitative importance of the role played by this species in the cycling of TEs.
gametogenesis and constitution of reserve substances. The gonad development leads to an increase in the body mass and, during this period, TEs present in their flesh are in some way "diluted", while they are "concentrated" when their body mass decreases at the release of the breeding material. The same goes for an increase/decrease in body mass linked to a high/low availability of food (e.g., Ref. [99,196-198]). A comparison of the raw concentrations between mussels from sites with different trophic and physicochemical characteristics is therefore not possible and an adjustment of the data is requested [176]. For this reason, the French Research Institute for Exploitation of the Sea (the IFREMER, that monitors the coastal contamination of France, its overseas departments and territories and the whole Mediterranean littoral) adjusts the concentrations of pollutants measured in caged mussels in the frame of their Mussel Watch Programs with a condition index corresponding to the ratio of the dry flesh weight on the shell dry weight. This condition index has the advantage of being easy to measure, of being a global index comprising several physiological factors (nutrition, reproduction, etc.) [199], and seems to be the biometric variable which is the more closely related to tissue concentrations for a large panel of contaminants [176]. Thus, for TEs, the tissue concentration is inversely proportional to the condition index. Once "adjusted", monitored concentrations can be accurately intercompared between sites and studies [176].

Conclusion

Unlike organic pollutants that can be degraded to less harmful components by biological or chemical processes, TEs are considered as non-degradable pollutants that will accumulate in coastal environments (mainly in sediments) to levels that might be toxic to the biota. Their toxic properties result from complex interactions between numerous biotic and abiotic factors; the understanding of this complex mechanism therefore requires both laboratory and field toxicological studies with adequate model organisms. In addition, because of the remobilization processes of chemicals between environmental compartments (e.g., from sediments to the water column), the adverse effects of TE pollution in contaminated environments can remain substantial and long lasting in spite of years of restoration efforts. For these reasons, the monitoring of the contamination by TEs still remains an environmental issue of great concern for the biota, but also for human beings. When compared with the conventional chemical analysis of physical environments (i.e., air, water, soils and sediments), the monitoring relying upon the biota exhibits obvious predominance, mainly because it provides an assessment of the portions of contaminants that are available for uptake and accumulation by aquatic organisms. The election of an appropriate bioindicator organism must meet a set of objective criteria, the first being to accumulate chemicals to levels representative of the contamination status of its environment. For the coastal Mediterranean, we confirm that the two main bioindicator species, i.e., the seagrass Posidonia oceanica and the mussel Mytilus galloprovincialis, are indeed appropriate indicators species for TE contamination monitoring [107,137-139,198,200]; and globally, we consider that seagrasses and bivalve mollusks have to be considered among the most suitable environmental indicator organisms because of their large geographical distribution, but also because of their complementarity (according to their lifestyle) as bioindicator species of chemical contamination. In addition, seagrasses form productive and biologically complex coastal ecosystems; the study of specific seagrass descriptors from the molecular to the ecosystem level thus allow to monitor the impact of anthropogenic activities at all levels of biological organization. Finally, bivalve mollusks are commercially important seafood products. Their use as bioindictor firstly provides relevant information on the quality of water bodies where fish and shellfish products are grown, but also provides important phytosanitary data on the safety of seafood consumption and potential threats to human health.

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