



Recent decline of roach *Rutilus rutilus* stock in a large river ecosystem in relation with its population dynamics

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With 3 figures and 6 tables

Abstract: Studies dealing with quantitative decline of freshwater fish species with long-time series data are rather scarce and primarily limited to migratory species. We present an original work integrating over two decades investigations on roach (*Rutilus rutilus* L.) stock, one of the dominant cyprinid fish species in the River Meuse (Belgium). This study aimed to quantify changes in the stock of roach after recent indicators (captures from sport fishermen) suggested a sharp decline. Three methods were used: capture-mark-recapture (CMR) estimation, catch per unit of effort by gillnetting (CPUE) and long-term monitoring of fish-passes. Population dynamics of roach was also investigated. CMR method demonstrated a decrease of 91 % of the roach stock compared to earlier data (1993–2003), with current densities close to 300 roach ha⁻¹. This was also confirmed by a decrease of 95 % in CPUEs and a decrease between 93 and 98 % in two fish-passes during the same period. On the other hand, growth and mortality did not indicate radical changes before and after the decline was found. As roach stock decline is generalizable to the whole Belgian's Meuse following estimations in two other sites, further investigations are needed to explain the reasons underlying this decline in order to take conservation measures. First, the uptake of primary production (i.e. phytoplankton) by invasive clams is supposed. And secondly, an increase in predation pressure by the Great Cormorant is effective as this avian predator was found in high densities from 2000 until 2006.

Key words: catch per unit of effort; fish pass; gillnet, large river; mark-recapture model; roach decline.

Introduction

Understanding changes affecting community structure and ecosystem functioning is one of the main challenges of modern ecology (Poff et al. 1997). Compilation of large datasets, involving time series observations, is extremely useful to relate observed changes to disturbances of an ecosystem (Daufresne et al. 2003).

Estimates of fish stocks dynamics are frequently used to evaluate ecological interactions in aquatic communities. These estimates are also important in fisheries management to control exploitation of a

stock to avoid its collapse in a near future (Peltonen et al. 1999). Results of fish stock assessment should be of sufficient quality. In fisheries management, serious errors have been made following inaccuracy in stock assessment. The exactness of an estimate with a particular method is rarely known (Bailey 1984). Although contradictory estimates can be made by different assessment methods (Schnute & Hilborn 1993), conclusions are more secure if different methods give coherent estimates (Foote & Stefansson 1993). To highlight abundance and structure of a particular species population is a real challenge, mainly in large riv-

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ers. All sampling techniques are biased to some degree when estimates of abundance, richness or distribution are needed (Olin et al. 2009). Stock evaluations rely on active capture of fish such as electrofishing or passive capture of fish such as gillnetting (Murphy & Willis 1996; Goffaux et al. 2005), but these methods have their own limits. Gillnetting is size selective (Hamley 1975), tends to underestimate young individuals of a given species or species being more sedentary (Rulifson 1991). Size selectivity can be reduced by using gillnets with different mesh sizes (Goffaux et al. 2005). Electrofishing is widely considered as efficient for describing population structure but is not appropriate for large and deep rivers (Pusey et al. 1998). Moreover electrofishing efficiency is influenced by physical and chemical parameters, swimming capacities of fish (Casselmann et al. 1990) and response to electric field may vary between species and size class (Pusey et al. 1998). In large rivers, combination of these two techniques allows both sampling of the main channel and of the banks (Goffaux et al. 2005).

Mark-recapture method first developed by Petersen (1896) is an alternative to the removal method (Thompson et al. 1998) and the most used technique for stock estimates in inland water (Gatz & Loar 1988; Donkers et al. 2011). The assumptions of this model are a closed population, random distribution and same catch probability between mark and unmarked individuals of the same species (White et al. 1982) and no mark loss. The mark-recapture model is generally used for abundance estimates in small (Gresswell et al. 1997) to large lakes (Donkers et al. 2011; Dahm et al. 1992). This model is more appropriate for low capture efficiency or low stock (Laurent & Lamarque 1975). Since the pioneer work of Petersen, several models based on mark-recapture have been developed (Begon 1979).

Catches per unit of effort (CPUE), instead of giving estimate densities, offer a view on proportional changes which occurred in an exploited fish population. Comparison of CPUE information and stock density assessment is more useful if changes occur in fish population can be detected by both methods (Peltonen et al. 1999).

In addition, monitoring of fish passes enables data collection on population dynamics taking into account the migratory part of the fish populations. The most common monitoring method is to catch fish in a trap disposed within or at the outflows of the pass (Larinier 1998). When performed during consecutive years, monitoring of fish pass is an efficient tool to quantify changes in fish population in a river such as

a decline of a particular species (Mallen-Cooper & Brand 2007).

Roach *Rutilus rutilus* (L., 1758) is one of the most commonly and widely distributed fish in European rivers (Kottelat & Freyhof 2007) but is also considered as a potential pest where it has been introduced and became the dominant species. In these environments, its high abundance creates concern for the other species present (Giannetto et al. 2014). Native in Belgium, roach stock in the River Meuse has been investigated by mark-recapture using the Jackson method in 1993, in order to evaluate the need of a fish restocking programme to support a roach population subject to capture by sport fishermen. Its density was estimated at 3695 roach ha⁻¹ (Didier & Micha 1996). Based on this stock assessment, it was decided to stop the roach restocking programme in different reaches of the River Meuse. The study was repeated 7 years later from 2000 to 2002 and the density was estimated between 3035 and 3145 roach ha⁻¹ indicating a stable roach population in the River Meuse despite the absence of restocking (Evrard & Micha 2003). The restocking programme was definitively abandoned. However, since these studies, according to indirect indicators (captures from sport fishermen), roach stock was decreasing.

This study aimed to quantify roach stock using three different methods (estimation by mark-recapture, catch per unit effort and fish pass monitoring) over the last decade in the River Meuse (Belgium). Results between different sites and years are presented and compared with previous investigations in the same area. The assessment methods were also confronted to secure conclusion about the importance of the decline. Growth and mortality parameters were also investigated to check if changes in population abundance have impacted vital population parameters.

Study area

The River Meuse is 905 km long and its total catchment area is 34584 km² (Descy et al. 2009). The river rises in Eastern France and flows throughout Belgium and The Netherlands before meeting the Lower Rhine and forming the Dutch delta. It debouches in the North Sea (Fig. 1) (Descy 1987). The average slope of the River Meuse is 0.23 ‰ (Micha & Pilette 1988). The River Meuse is a regulated river but management has been conducted with few considerations for aquatic fauna. As a result, plant and animal biodiversity has decreased (Descy et al. 2009). In Belgium, the river looks like a bream zone (deep water and slow current) according to Huet zonation (Huet 1949) as a consequence of heavily canalisation during the last 150 years. Naturally, the river in Belgium is a barbel zone characterised by a greater current and a lower depth (Micha & Pilette 1988). Fif-

Table 1. Annual means of debit, temperature, suspended matter, nitrates and orthophosphates at Tailfer during the studied period.

Years	1993	2000	2001	2002	2010	2011
Debit (m ³ s ⁻¹)	171	239	224	143	167	146
Temperature (°C)	11	12.8	12.9	14.5	13.1	13.8
Suspended matter (mg l ⁻¹)	18	25	20	27.2	12.2	12.8
Nitrates (mg l ⁻¹)	2.6	2.7	2.6	2.5	2.7	2.9
Orthophosphates (mg l ⁻¹)	0.06	0.05	0.04	0.03	0.05	0.05

**Fig. 1.** Location of the River Meuse basin with the three sampling sites (bold names) and the two fish passes monitored.

teen dams have been built on its course in Belgium which allow shipping of 1350 tons upstream Namur (Tans 2000) and 9000 tons downstream Namur but disturb flow rate, migration of fish species and natural zonation patterns (Micha & Pilette 1988). Natural banks are relatively rare as a result of canalisation. Over the past three decades, water temperature has increased by 0.16–0.89 °C and discharge tends to increase. Eutrophication is widespread with organic pollution high in some parts of the river (Descy et al. 2009). In addition, invasive species, mostly macroinvertebrates, are present and widespread (Descy et al. 2009). During the study period at Tailfer (Belgium, 518 km from source), chlorophyll-*a* and zooplankton have decreased drastically, presumably in response to invasive molluscs spread (*Corbicula*, *Dreissena polymorpha*, *Dreissena rostriformis*) (Pigneur et al. 2014). Extinction coefficient, inversely related to phytoplankton biomass, has increased. Soluble reactive phosphate presents a slight decrease. Nitrate and ammonium, temperature and discharge remain in the same range (Table 1) (Pigneur et al. 2014). In the Belgian's section of the River Meuse, 29 fish species were found during sampling by gillnetting and electrofishing from 1996 to 2012 (Table 2) with roach being the most abundant (Goffaux et al. 2005).

Table 2. Species list found in the River Meuse during sampling

Species	English name
<i>Abramis brama</i>	Bream
<i>Alburnus alburnus</i>	Bleak
<i>Alburnus bipunctatus</i>	Stream bleak
<i>Anguilla anguilla</i>	Eel
<i>Aspius aspius</i>	Asp
<i>Barbatula barbatula</i>	Stone loach
<i>Barbus barbus</i>	Barbel
<i>Blicca bjoerkna</i>	Silver bream
<i>Carassius carassius</i>	Crucian carp
<i>Chondrostoma nasus</i>	Nase
<i>Cottus gobio</i>	Bullhead
<i>Cyprinus carpio</i>	Carp
<i>Esox lucius</i>	Pike
<i>Gasterosteus aculeatus</i>	Three-spined stickleback
<i>Gobio gobio</i>	Gudgeon
<i>Gymnocephalus cernuus</i>	Ruffe
<i>Lepomis gibbosus</i>	Pumpkinseed
<i>Leuciscus idus</i>	Ide
<i>Leuciscus leuciscus</i>	Dace
<i>Perca fluviatilis</i>	Perch
<i>Phoxinus phoxinus</i>	Minnow
<i>Rhodeus sericeus</i>	Bitterling
<i>Rutilus rutilus</i>	Roach
<i>Salmo trutta fario</i>	Trout
<i>Sander lucioperca</i>	Pikeperch
<i>Scardinius erythrophthalmus</i>	Rudd
<i>Silurus glanis</i>	European catfish
<i>Squalus cephalus</i>	Chub
<i>Tinca tinca</i>	Tench

The present study was carried on three reaches in Belgium: Hastière, Tailfer and Visé located 488, 518 and 611 km respectively from the source (Fig. 1). The reach of Tailfer was previously studied by Didier & Micha (1996) and Evrard & Micha (2003).

Material and methods

Fish sampling

Estimates of roach stocks in the three sites have been based on the protocol (see below) described in Didier & Micha (1996) and applied in the study of Evrard & Micha (2003). Before es-

timations, roach grown in captivity (Total length: 101–190 mm) in a fish farm were marked by clipping off the half right pelvic fin and restocked homogeneously immediately after marking.

The option to mark roach grown in fish farm for the mark-recapture operations is due to several reasons: i) there were no other possibilities to catch and mark a large number of fish in a short period of time (one day); ii) the most efficient technique of roach capture in large rivers is gillnet, but this method is invasive and largely affects the survival rate of captured fish (Didier & Micha 1996).

For restocking, marked roach stock were divided in several small tanks (500–1000 roach) placed at 200–300 m intervals along the banks. In the reach of Tailfer, 10,100 marked roach were stocked in 2010 and 10,000 in 2011. In the reaches of Hastière and Visé, 7,500 and 8,500 marked roach were stocked in 2012. In the reach of Tailfer, restocking was done at the end of March and sampling was conducted between April and Mai. In 2012, restocking and sampling were done in November.

To meet general assumptions of mark-recapture models, marking and recapture periods must be short (Krebs 1999 in Donkers et al. 2011). In the present study, samplings were completed within 4 days on each reach, less than a week after restocking of marked roach. This short period of time reduces the risk of method violations due to emigration and immigration. It's assumed that neither growth nor recruitment occurred during the sampling period (Ricker 1975). The closed population assumption appears to be met by dams delimiting each reach. Further, estimates have been conducted out of the reproductive period in order to avoid upstream migration of roach through fish passes. The assumptions of zero mark loss were addressed by fin clipping and correct reporting of mark by the use of three observers. Fin clipping was chosen for its facility and rapidity of marking and because of non-regeneration possible during the short period of sampling. Short period of time between marking and sampling should have reduced the probability of unequal mortality among unmarked and marked roach (Gresswell et al. 1997). Survival after marking was verified by maintaining in a tank placed in the River Meuse 30 marked and 30 unmarked roach for one month in 1993. No difference in survival rate was observed suggesting that marking didn't affect roach survival (Didier & Micha 1996). We assumed an equal capture probability between marked and unmarked fish of the same size.

In 2010, 11 samples by gillnetting and 10 samples by electrofishing were done in the reach of Tailfer. In 2011, 12 samples by gillnetting and 8 by electrofishing were taken. In the reach of Hastière, 13 samples (5 by gillnetting and 5 by boat electrofishing) were collected in 2012. In the reach of Visé, 7 samples by gillnetting and only 3 by electrofishing were done because of low availability of bank less than 2 m deep.

Gillnetting was performed with a set of 3 to 6 gillnets (length: 50 m; height: 2 m) with different mesh sizes (20, 30 and 40 mm knot to knot). Gillnet 20 mm was preferred to other gillnets because its ability to catch fish of the size class restocked. Two to four gillnets were used per set. Gillnet 30 mm was used as an intermediate gillnet with one to two gillnets in the set. Gillnet 40 mm was used to a lesser extent (one gillnet per set). Samples were taken at different locations along reaches. Location selection was done randomly. Gillnets were exposed in the main channel for 2 to 6 h during daytime. This large difference is due to the high abundance of fish caught in gillnets. During the first 2 h of capture by gillnet, an electrofishing was carried out along shallow banks (<2 m) by boat (Zodiac®, 4.2 m long) in an upstream direction over a distance of 200–800 m. All type

of habitat found in the reaches were investigated proportionally to their abundance. The generator consists in a 3 KW alternator delivering a continuous current (300 V at 4 A, DEKA 7000). The cathode was floating on the rear side of the boat. Three persons were used, one boat driver, one dip netter and one hand-held anode. After sampling, fishes were identified. Roach were measured to the nearest mm (Total Length, TL) and weighted to the nearest g individually. Identification as marked or unmarked roach was done by three persons for a correct reporting of marks.

Stocks estimates

Estimates have been investigated with the Jackson's positive method (1) (Jackson 1939). With this method, marking occurs on one occasion followed by multiple recaptures events (Begon 1979):

$$N_0 = \frac{r_0}{q_0} \quad (1)$$

where

N_0 = total population estimated on day 0

q_0 = marked proportion of a hypothetical random sample taken on day 0

r_0 = number of individuals marked and released on day 0

Methods for estimating q_0 and confidence intervals of N_0 are from Begon (1979).

Estimations of N_0 were done with number of roach unmarked with size comprised between minimum and maximum size of roach marked. The total population was estimated by adding to N_0 a proportional estimate of roach outside of minimum and maximum size class defined by marked roach.

Estimate of total biomass B (kg) is calculated by the formula (2) using N_0 (1) and its confidence limits determined following method of Begon (1979):

$$B = N * \bar{P} \quad (2)$$

where \bar{P} = Average weight (kg)

Estimates of density (roach ha⁻¹) and biomass (kg ha⁻¹) per hectare are calculated by dividing estimates of total population and its total biomass by the surface of each reach.

Capture per unit of effort by gillnet

In addition to stock estimates, stocks dynamics was also investigated with gillnet catches per unit of effort (CPUE) performed during stock evaluation. Even if CPUE is not an absolute estimate of density, it can show proportional changes in stocks (Peltonen et al. 1999). Data from gillnets with mesh size 20 and 30 mm were used following sufficient data. CPUEs are reported in number of fish caught per net per 30 min of fishing effort.

Because of unequal size samples and variance heterogeneity, we used Welch's Test F ratio (Welch 1951) to compare CPUEs. Multiple Welch paired-tests were performed to test significant differences between years in CPUEs with the same mesh size. Analysis of year samples will reinforce our confidence in changes revealed by mark-recapture estimates. Corrections for multiple paired-tests following the sequentially rejective multiple test procedure developed by Holm (1979). Comparisons with the same conclusion were grouped for facility and the F value, degrees of freedom and probabilities given referred to the highest statically ($p < 0.05$) significant difference observed.

Fish pass analysis

In order to use different methods to reveal and quantify roach stock, two fish passes were monitored, at Tailfer dam on the Middle Meuse and at Lixhe dam on the Lower Meuse. At Tailfer pass, fish were captured by trap at the following years: 1989 to 1994, 2006 and 2009 to 2011. This fish pass was monitored from March until December in 1989 to 1994 and from March until September in 2006 and 2009–2011. Fourteen pools composed the fish pass (1.95 × 1.50 × 0.75 m) each dropping in elevation by 0.13–0.15 m. Discharge at the outlet is closed to 0.24 m³ s⁻¹ and water velocity over the traverse is about 0.9 m s⁻¹ (Prignon et al. 1998). The only trap was placed in the upper pool of the pass and had a cone at the entrance and a steel grid (1 cm wide between bars) at the exit. All fish were counted except young of the year and common bleak (*Alburnus alburnus* L.) because of high abundance during the nineties. During migration peaks (April–June), fish were collected daily. Outside migration period, the trap was checked twice a week.

Lixhe dam has two fish passes. The first was put into operation in 1980 and the second in 1998. Monitoring was done from January until June. The older is a pool and weir pass of 48 m length composed of 56 pools (1.5 × 0.9 × 0.4 m) with 2 cones trap at the upper pool. The new one is a pool and vertical slot pass of 305 m length composed of 26 pools (1.4 × 1.1 × 1.5 m) with a cage at the upper pool. The mean discharges were 0.135 and 1.0 m³ s⁻¹ respectively. The older fish pass was monitored from 1992 to 2012, except years 1994 and 1996 and the new one from 1998 to 2012. Both old and new fish passes were monitored at least twice a week and all fish were counted except young of the year.

Roach abundance was expressed as catch in number per month. For each year, we divided fish abundance by the period of sampling (in month). At Tailfer pass, we compared the two periods of monitoring ranging from 1989 to 1994 (6 years) and from 2006 and 2009 to 2011 (4 years). As no great gap existed in the monitoring of the Lixhe pass, periods were determined to correspond closer to the periods of the Tailfer pass. More, data were ordered to check if chronology and decreasing were concordant. Periods compared were one before year 2000 (7 years), a second period from 2001–2006 (6 years) and a third period from 2007–2012 (6 years). Following high variance heterogeneity and unequal samples size, we used Welch's Test F ratio to compare CPUEs. Corrections for multiple paired-tests following the sequentially rejective multiple test procedure developed by Holm (1979).

Dynamics of roach populations

Age and growth rate were read from scales (Mann 1973). Between 3 and 10 scales were collected from 56 specimens caught in each reach. Scales were taken above the lateral line up the dorsal fin. After removing mucus, scales were placed between 2 microscope slides (@VWR). Annual radii were determined as the point where circuli became closely spaced followed by areas of widely spaced circuli. Annual radii were recorded only when structures could be viewed around the circumference of the scale (Cragg-Hine & Jones 1969). Age reading was carried out independently by two persons. After age accordance, measures of radii (to the nearest mm) were carried out on the posterior field using a binocular. Total length at different ages was back-calculated using the Lee (1920) formula based on measures of radii.

$$L_t = a + (L_n - a) * \frac{R_t}{R_n} \quad (3)$$

where

L_t = back-calculated fish body length at age t

a = intercept from the regression of body length on mean scale length

L_n = fish body length at capture

R_t = mean scale length at annulus t

R_n = mean scale total length at capture

Growth was described by the Von Bertalanffy growth curve (VBGC) model (Von Bertalanffy 1938) described by the equation (4):

$$L_t = L_\infty * (1 - e^{-K*(t-t_0)}) \quad (4)$$

where

L_t = Total length at age t

L_∞ = Asymptotic length or Maximal size

K = Growth coefficient

t_0 = Hypothetical age when length equal 0

The method of Ford-Walford was used to determine asymptotic length L_∞ which correspond to the intercept of the right $x = y$ and the regression of body length at time $t+1$ and body length at time t . Growth coefficient K and theoretical age t_0 were determined using the Von Bertalanffy method (Sparre & Venema 1996). We used Excel software to construct the VBGC model.

The index \emptyset' of growth performance (Pauly & Munro 1984, Sparre & Venema 1996, Naddafi et al. 2005) calculated by formula (5) was employed in order to compare growth between years and sites:

$$\emptyset' = \ln(K) + 2 * \ln(L_\infty) \quad (5)$$

The instantaneous rate of total mortality Z was estimated using length-converted age catch curve using the equation of Von Bertalanffy and by reporting age on the X-axis and natural logarithm of number of individuals by cohort on the Y-axis. Cohorts under-estimated following a low sampling effort due to gears used were rejected for this analysis. The regression is equal to (6):

$$\ln(N_t) = a + b * t \quad (6)$$

Total mortality Z is equal to $-b$ (Ricker 1975; Pauly 1997).

Instantaneous rate of natural mortality M was estimated using the empirical equation of Pauly (Pauly 1980) (7):

$$\log(M) = -0,006 - 0,279 * \log(L_\infty) + 0,6543 * \log(K) + 0,4634 * \log(T) \quad (7)$$

where L_∞ and K are from the VBGC (5)

T = Mean annual water temperature (°C)

Instantaneous rate of fishing mortality F was calculated as the difference between instantaneous rates of total and natural mortality (8) (Sparre & Venema 1996):

$$F = Z - M \quad (8)$$

The ratio of instantaneous rate of fishing and total mortality was calculated to estimate exploitation E (9) (Sparre & Venema 1996):

$$E = F/M \quad (9)$$

Results

Density estimates

In the reach of Tailfer, 761 unmarked roach were caught by electrofishing and gillnet in 2010 and 303 in 2011. In 2010, 222 marked roach were caught (Recapture rate of 2.2 %) and 104 in 2011 (Recapture rate of 1.04 %). Gillnet catches represented for this site 49 % of the captures. A total of 462 unmarked roach were caught in the reach of Visé in 2012. Gillnet catches represented 99.8 % of the captures. On 8500 marked roach, 71 have been recaptured during the sampling period. Recapture rate was 0.84 %. On the reach of Hastière, a total of 241 unmarked roach were caught, mostly by gillnet sampling (97%). Recapture rate was 3.75 % with a total of 281 marked roach caught. Most of the recaptures in this site were due to electrofishing (62 %) along the banks. Lengths of roach caught by electrofishing were relatively constant between 1993 and 2002. For years 2010–2012, size class length was reduced following low capture. Lengths of fish caught by gillnetting were constant along the two decades reflecting the selectivity of this method (Table 3).

Between 1993 and 2002, estimates by Jackson method reveal a steady stock of roach. According to

this method, roach stock was estimated between 3695 [2554–5346] and 3035 [1950–4719] roach ha⁻¹ (Table 4). Moreover, CPUEs obtained between 1993 and 2002 from gillnets with mesh size of 20 and 30 mm did not show any significant differences during that period (Fig. 2). In 2010 and 2011, roach stock has undergone a strong decrease in less than 10 years with estimated densities around 300 roach ha⁻¹. Even if no historical data exists for the reaches of Hastière and Visé, roach stocks appeared close to the results obtained in the reach of Tailfer between 2010 and 2011. Density is estimated at 225 [205–248] roach ha⁻¹ in Visé and 109 [60–197] roach ha⁻¹ in Hastière.

Gillnet surveys

Total lengths of roach caught by 20 mm mesh size gillnets ranged between 101 and 250 mm. For gillnet 30 mm, total lengths of roach ranged between 121 and 340 mm (Table 2). Values and confidence intervals (95 %) of CPUEs are shown in Figure 2. CPUEs for 2010 were not used because fishing time was not reported. CPUEs obtained in 2011 and 2012 on both sites were not significantly different ($F_{\text{Welch}}(1,19) = 2.28$; $p > 0.147$ for gillnet of 20 mm and $F_{\text{Welch}}(1,21) = 63.47$; $p > 0.625$ for gillnet 30 mm). On the other hand CPUEs

Table 3. Date, site, area, recapture rate and minimum and maximum total length of roach restocked, caught by sampling methods and for stock estimations.

Year	Site	Area (ha)	Recapture rate (%)	Restocking	Electrofishing	Gillnet 20 mm	Gillnet 30 mm	Stock estimations
1993	Tailfer	76.4	1.73	111–260	51–270	131–230	171–310	51–310
2000	Tailfer	76.4	2.16	81–190	81–300	141–250	171–320	81–370
2001	Tailfer	76.4	0.65	101–220	71–360	121–250	121–310	71–340
2002	Tailfer	76.4	1.31	91–250	71–360	121–250	121–310	71–370
2010	Tailfer	76.4	2.20	81–190	151–160	101–240	121–340	81–340
2011	Tailfer	76.4	1.04	111–240	71–180	101–240	151–300	71–320
2012	Visé	230.0	0.84	81–180	141–150	111–240	191–300	81–330
2012	Hastière	45.8	3.75	81–180	61–160	131–220	201–280	61–340

Table 4. Jackson estimates and resulted density and biomass obtained. Data from 1993 and 2000–2002 are reworked from Didier & Micha (1996) and Evrard & Micha (2003). N_0 , roach stock estimation (roach comprised between minimum and maximum length of marked roach). Density and biomass include also estimation of roach outside length class defined by marked roach.

Year	Site	N_0	Density (roach ha ⁻¹)			Biomass (kg ha ⁻¹)		
			Lower 95 %	Mean	Upper 95 %	Lower 95 %	Mean	Upper 95 %
1993	Tailfer	273445	2554	3695	5346	138.6	200.6	290.2
2000	Tailfer	240248	2021	3145	4892	112.3	174.7	271.8
2002	Tailfer	180025	1950	3035	4719	108.3	168.6	262.2
2010	Tailfer	27885	380	473	590	16.8	20.9	26.0
2011	Tailfer	10584	91	149	243	5.7	9.3	15.1
2012	Visé	433752	205	225	248	10.3	11.3	12.5
2012	Hastière	3609	60	109	197	2.9	5.3	9.5

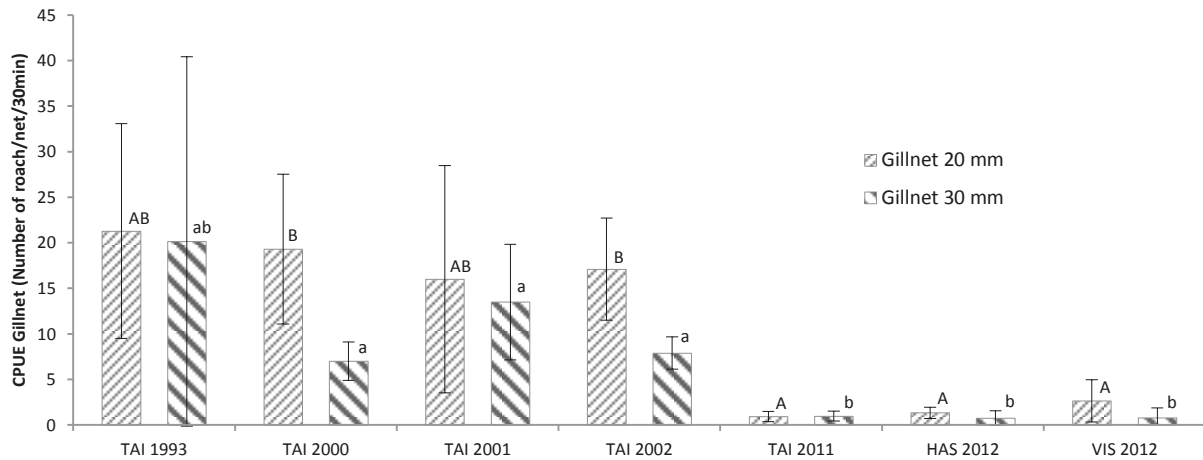


Fig. 2. CPUE for gillnet 20 and 30 mm obtained in the reaches of Tailfer (TAI), Hastière (HAS) and Visé (VIS), errors bars: 95 % interval of confidence around estimated mean values based on specified number of observation per group represented by each bar. Items with the same letter (uppercase for gillnet with mesh size of 20 mm, lowercase for 30 mm) are not significantly different. Data from 1993 and 2000–2002 are reworked from Didier & Micha (1996) and Evrard & Micha (2003).

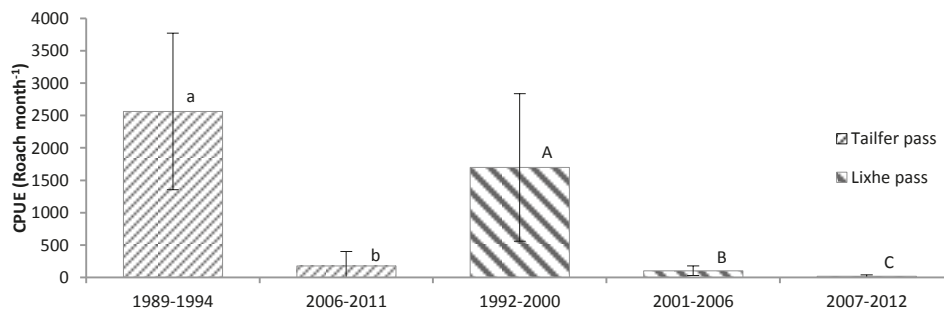


Fig. 3. Evolution of the catch per unit effort (Roach month⁻¹) at the Tailfer and Lixhe passes, errors bars: 95 % interval of confidence around estimated mean values based on specified number of observations (year) per group represented by each bar. Items with the same letter (lowercase for Tailfer, uppercase for Lixhe) are not significantly different

in 2011 and 2012 for gillnets with mesh size of 20 mm significantly lower from those calculated for 2000 and 2002 ($F_{\text{Welch}(1,23)} = 16.56$; $p < 0.008$) but were not significantly lower for CPUEs from 1993 and 2001 ($F_{\text{Welch}(1,6)} = 17.72$; $p > 0.084$) after correction with sequentially rejective Bonferroni test and following the high variability for these years. CPUEs in 2011 and 2012 for gillnets of 30 mm were highly different from 2000, 2001 and 2002 ($F_{\text{Welch}(1,15)} = 17.61$; $p < 0.0008$) but not significantly from 1993 ($F_{\text{Welch}(1,6)} = 5.44$; $p > 0.06$). CPUEs values for gillnets of 20 mm did not significantly differ between years 1993, 2000, 2001 and 2002 ($F_{\text{Welch}(1,19)} = 0.48$; $p > 0.494$). Captures with gillnets of 30 mm were similar when comparing the years 1993, 2000, 2001 and 2002 ($F_{\text{Welch}(1,18)} = 4.27$; $p > 0.05$) (Fig. 2).

Fish pass analysis

From 1989 to 1994, a mean of 2565 roach per month were captured at Tailfer pass. During last years (2006, 2009–2011), we noticed on average 179 roach per month (Fig. 3). Comparison of the two periods indicates a highly significant decrease in the number of roach passing through the pass ($F_{\text{Welch}(1,5)} = 25.20$; $p = 0.004$). Between these periods, we noticed a decrease of 93 % in roach passages. At Lixhe pass, during the first period (1992–2000), on average 1699 roach were counted per month, 103 during the second period (2001–2006) and only 19 during the third one (2007–2012) (Fig. 3). Comparison of the three periods reveals a significant decrease ($F_{\text{Welch}(1,6)} = 11.70$; $p < 0.014$) of 94 % between the first two periods, 81 %

Table 5. Growth parameters of VBGC, performance index (θ'), mortalities and exploitation of roach. L_∞ , asymptotic total length; K , growth rate; t_0 , hypothetical age when length equals 0; Z , total mortality; M , natural mortality; F , fishing mortality; E , exploitation. R^2 is the coefficient of determination of the regression of natural logarithm of number of individuals by cohorts on age.

Sites	Year	L_∞	K	t_0	θ'	Z	M	F	E	$R^2 (Z)$
Tailfer	2010	302	0.20	-1.0	9.8	0.92	0.45	0.47	0.51	0.94
Tailfer	2011	316	0.18	-0.6	9.8	0.60	0.40	0.20	0.33	0.96
Hastière	2012	438	0.15	-0.8	10.3	0.82	0.33	0.49	0.60	0.96
Visé	2012	385	0.21	-0.6	10.3	0.78	0.45	0.33	0.42	0.92

between the last two periods and 98 % between the first and last periods.

Dynamics of roach populations

Following low capture of young roach due to the gears (gillnet more precisely) used, dynamic analysis was performed with roach older than 1+ until 8+. Scales were taken on 56 fish taken during sampling taking account of length frequency. It appears that roach populations in the reaches of Hastière, Tailfer and Visé have a growth K comprised between 0.15 and 0.21 year⁻¹. Asymptotic lengths L_∞ estimated in 2012 are higher to those estimated at Tailfer in 2010 and 2011 (Table 5). For the reach of Tailfer, asymptotic length tends to decrease between estimates of 2010/11 and 1993/2002 while growth remains stable (Tables 5 & 6). Growth performance index θ' reveals a low variability between estimates. The highest values are found in the reaches of Visé and Hastière (Table 5).

Exploitation, which reflects all types of mortalities, was high in the reach of Hastière. In the reach of Visé, exploitation was lower and comprised between estimates for the reach of Tailfer (Table 5).

Discussion

A strong decline in roach stock has been observed in the River Meuse in the past decade according to direct indicators such as stock estimates by mark-recapture, catch per unit effort with gillnet and roach passages at two fish passes. According to mark-recapture model, roach stock declined by 91 % in the reach of Tailfer reaching densities of 300 fish ha⁻¹. Size class investigated are of the same order. At the same time, a decrease of 95 % in CPUE by gillnetting was observed. The study of Tailfer pass provided the same conclusion with a decrease of 93 % in roach passages. Other stock estimates conducted in the reaches of Hastière and Visé showed that the decline observed at Tailfer can be extrapolated to the whole Belgian Meuse as stocks at Hastière and Visé reaches were low too. In addition, study of the Lixhe pass (adjacent to the reach of

Visé) has shown a similar decrease, higher than 96 % of roach passages. Growth and mortality presented no radical changes suggesting that the decrease in density did not influence vital population parameters.

Estimates of roach density

Choice of mark-recapture method instead of removal method was due to high complexity for applying removal method in the River Meuse, which is a large (> 100 m) and deep (> 2 m) river. As Kelso and Shuter (1989) doubt on the efficiency of removal method for lake populations, same conclusions can be drawn for great rivers. For such environments, the mark-recapture is the most used method (Donkers et al. 2011; Gresswell et al. 1997). The principle of the Jackson method is that marking occurs on one occasion only and followed by several recaptures (Begon 1979). As mentioned before, all roach were marked on a single day. This single mass-marking was followed by multiple recapture events by a trained team. These two aspects make the use of the Jackson's positive method efficient and extremely useful. This method is an extension of the Petersen method which allows loss but no gain (Begon 1979).

The choice we made of fish grown in fish farm can have some biases due to the behaviour of these fish when released in the wild. For example, in the reach of Hastière, a lot of marked roach stayed hidden in aquatic vegetation more present in this reach (personal observation) which can lead to underestimate the natural population by increasing marked fish capture. Recapture rate in this reach was the highest in this study. Moreover, these fish can suffer higher mortality rate than wild fish of the same age (Brown & Laland 2001; Phillipart 1995).

Although the analysis of fish pass data is an indirect and selective fish inventory method, inherently unique, it is an important tool in river management to control fish populations (Roscoe & Hinch 2010). Furthermore, in great rivers, there are no methods which can offer a perfect sample fully representative of fish fauna. Electrofishing and gillnetting have also their

own limits (Pusey et al. 1998; Casselman et al. 1990), as demonstrated by Goffaux et al. (2005).

Our results indicate that abundance of roach estimated by mark-recapture in the River Meuse in 2010–2012 was very low compared with earlier data (1993–2003) obtained with the same protocol. In the three monitoring sites, recent estimates were below 500 roach ha⁻¹ (61–340 mm). The lowest density was found in the reach of Hastière with 109 roach ha⁻¹ (61–340 mm). According to previous estimates based on the Jackson method, a density relatively stable comprised between 3000 and 3700 roach ha⁻¹ (51–370 mm) was found.

In the River Thames (England), roach stock has been investigated by mark-recapture by Williams (1965) and its density was estimated at 10000 roach ha⁻¹ for fish over 10 cm fork length. In Lake Tjeukemeer (The Netherlands), roach density was estimated at 585 roach ha⁻¹. This low density was attributed to scarcity of zoobenthos and competition from other species (Goldspink 1979). In Lake Årungen, a eutrophic lake in Norway, biomass of roach was estimated at 550 kg ha⁻¹ in the 1980s (Eie & Borgstørn 1981). Eutrophication can have profound effects on fish communities, with cyprinid fishes such as roach dominating communities in these environments (Willemssen 1980, Winfield 1992). The River Meuse is a eutrophic river that corroborates well with the high density of roach found in the 1990s and beginning of 2000s. Recent studies reported that nutrient concentrations, which could explain such decline in fish stocks,

didn't show any sharp decrease during the study period (Pigneur et al. 2014), suggesting that other causes such as bivalves or a high pressure predation should be responsible of such drastic decline in roach density.

Gillnet CPUE has been widely used as an important parameter in monitoring abundance changes or like an index of fish abundance (Olin et al. 2002; Olin et al. 2009; Mehner et al. 2005). Statistical comparisons of CPUEs corroborate the conclusion about the decrease in estimated densities. CPUEs obtained in 2011 and 2012 by gillnets are significantly lower than those obtained in previous studies except in 1993 (gillnets of 20 and 30 mm) and 2001 (gillnets of 20 mm). As roach stocks are low, CPUEs are low too.

We can assert that the use of fish pass data confirms what other methods revealed. Time-series study of fish pass indicated a decrease of 93 % in roach passages, in accordance with the decrease of 95 % revealed by CPUE with gillnet and with the decrease of 91 % in roach stock estimates at Tailfer reach. Based on these observations and considering also the marked decline in roach passages in the Lixhe pass (reach of Visé), it appears that roach population in the Belgian section of the River Meuse decreased by almost 90 % since the beginning of the 2000s.

Growth and mortality of roach

Growth of roach can be considered as normal in the monitoring sites. The parameter K is comprised between 0.15 and 0.21 year⁻¹, in line with past estimates in

Table 6. Values reported in literature of growth parameters of VBGC, performance index (θ'), mortalities and exploitation of roach. L_∞ , asymptotic total length; K , growth rate; t_0 , hypothetical age when length equals 0; Z , total mortality; M , natural mortality; F , fishing mortality; E , exploitation. R^2 is the coefficient of determination of the regression of natural logarithm of number of individuals by cohorts on age.

Sites	Reference	Year	L_∞	K	θ'	Z	M	F	E	$R^2 (Z)$
Meuse at Waulsort		1989	421	0.15	10.2					
Meuse at Waulsort		1990	451	0.12	10.1					
Meuse at Tailfer	Didier & Micha 1996	1993	352	0.20	10.1	0.71	0.40	0.31	0.43	0.99
Meuse at Tailfer	Evrard & Micha 2003	2000	431	0.12	10.0					
Meuse at Tailfer	Evrard & Micha 2003	2001	322	0.22	10.0					
Meuse at Tailfer	Evrard & Micha 2003	2002	371	0.19	10.2					
Meuse at Tailfer	Evrard & Micha 2003	2000–2002	368	0.18	10.1	0.69	0.38	0.31	0.45	0.96
Lake Sapanca (Turkey)	Okgerman et al. 2009		318–472	0.11–0.19	9.9–10.1					
River Watra (Poland)	Przybylski 1996			0.13–0.27						
Lake Maggiore (Italy)	Volta & Jepsen 2008		357	0.29	10.5					
Lake Piediluco (Italy)	Giannetto et al. 2014		478	0.24	10.9					
Lake Geneva (France)	Ponton & Gerdeaux 1987			0.21	10.1					
Lake Lugano (Switzerland)	Guthruf 2002			0.30	10.6					
England	Wyatt 1988		425							
Netherlands	Goldspink 1979		206							

the reaches of Tailfer and Waulsort (Table 6). In other countries, growth rates range from 0.11 (Okgerman et al. 2009) to 0.30 (Guthruf 2002). Regarding asymptotic length, populations examined in the present study display maximal lengths of 438 and 385 mm which are close to the values previously reported in Tailfer and Waulsort. These values are greater than those reported in the literature (see Table 6). The \emptyset values are 10.3 at Hastière and Visé, slightly greater than values reported in previous estimates in Tailfer and Waulsort. Current values at Tailfer are lower than the past ones. Values reported in literature are between 9.9 (Okgerman et al. 2009) and 10.9 (Giannetto et al. 2014). The obtained \emptyset values of roach were very close to reported values from other countries. Regarding values reported in literature for roach growth rate, maximal length and the index of growth performance, values reported in this study are in line with the biology of this species. More, decline of roach population in the River Meuse had no detectable impact on vital parameter of this species.

Current estimates of total mortality (0.78 to 0.92 year⁻¹) are higher than past values (0.69 to 0.71 year⁻¹) except for the year 2011 (0.60 year⁻¹). In terms of survival, current estimates are between 0.40 and 0.55 and past values are between 0.49 and 0.50. Vøllstad & L'Abée-Lund (1987) found that survival of mature male and female was 0.30 and 0.52 and did not vary with age. An et al. (2009) found for mature female and male values of survival of 0.53 and 0.50, respectively.

Assumptions of the method used for estimating total mortality Z are a constant mortality for all ages considered in the evaluation and a large sample covering a maximum number of cohorts. Each cohort must be recruited in equal quantities and, up to a critical length, vulnerability of each group to capture is supposed constant (Pauly 1997). First and second assumptions can be considered as respected. Populations of roach in the reaches studied exhibited relatively constant total mortality over the entire range of ages sampled which is reflected by a high coefficient of determination (Table 5). For the evaluation of Z , 7 to 8 cohorts were used. Third and fourth assumptions have been respected by rejecting from the analysis cohorts that were under-estimated following a low sampling effort due to gears used.

In an exploited stock, natural mortality is the most difficult parameter to estimate but is important for models of stock management. *A priori*, this mortality must be estimated in unexploited stocks (Pauly 1997). Due to the stock exploitation in the River Meuse and insufficient catch curve, we used the equation devel-

oped by Pauly (1980) to estimate natural mortality. In the present study natural mortality was very close to previous estimates in the reach of Tailfer. Fishing mortality was high in roach stock of Hastière, leading to an over exploitation (60%). In the reach of Visé, exploitation was similar to past estimates at Tailfer. In the reach of Tailfer, exploitation remained constant between 1993 and 2002. A slight increase is observed for the year 2010. In 2011, exploitation was low (33%). The use of parameters in Pauly's equation has been revisited by Griffiths and Harrod (2007). The use of K , L_{∞} and T for determining natural mortality is a good agreement but habitat and taxon are also important factors which have to be taken into account (Griffiths & Harrod 2007). Estimating natural mortality can also be based on capture-recapture analyses of tagged individuals (Quinn & Deriso 1999; Donkers et al. 2011) but this method is time consuming due to tagging of fish and recapture.

Potential causes of decline

Different reasons can be proposed to explain the observed drastic change of the most common fish in the River Meuse. Alteration of habitat heterogeneity for navigation has conducted to a decrease of biodiversity in the River Meuse (Descy et al. 2009). However, channelization, deepening of the river bed and stabilisation of banks were done in the last century (Micha & Borlée 1989) and thus cannot be considered as the main causes of the recent decline observed for roach. But these perturbations can facilitate the establishment of invasive species potentially harmful for freshwater fauna.

Since several years, water quality of the River Meuse tended to improve following the construction of many sewage treatment plants (Descy et al. 2009). But nutrient levels (C, P and N) still remained rather stable during the last decade (Latli et al. unpublished). However, from the mid-2000s, an unexpected drastic decline in phytoplankton biomass has been reported while nutrients are still sufficient for plankton growth. As phytoplankton growth conditions have improved rather than degraded, Pigneur et al. (2014) hypothesized an increase of losses linked to the spectacular invasion of Asian clams, *Corbicula* sp. The decline in plankton biomass can lead to a "bottom-up" effect on fish populations following the 70% loss of primary production (Pigneur et al. 2014) which cannot be replaced by macrophytes because of river channelization. For example, in the San Francisco Bay, the invasion by the Asiatic clam *Pomatocorbicula amurensis* resulted in the decline of plankton (Alpine & Cloern

1992; Kimmerer et al. 1994) and, consequently, of plankton-feeding fish (Moyle et al. 1992). On the other hand, spread of invasive molluscs can increase the biomass of benthophagous fish, as shown by Karatayev et al. (1997) in Eastern Europe.

Population of Great Cormorant (*Phalacrocorax carbo*) has established a wintering population in the River Meuse valley since 1991 (Clotuche & Schaeken 1991) and colonized reach of Tailfer in 1994. More generally, after a constant period of growth (16 % per year) until 2001, the Great Cormorant population along the Meuse River stood high until 2004. Since 2004, the wintering population declined progressively by 67 % per year until 2009 while birds started colonizing more intensively small tributaries (Paquet 2007, 2011). Since 2010, the population along the River Meuse remained very low and stable. This observation can suggest a “top-down” effect of Great Cormorant on fish stock during its period of outbreak as roach is one of its favourite preys. In 2002, it was found that roach represent 33 % of prey found in the diet of Great Cormorant in the reach of Tailfer (Evrard & Tarbe 2002). Evrard et al. (2005) estimated a consumption of roach by cormorant between 21.9 and 24 kg ha⁻¹ year⁻¹.

These changes in predation pressure and primary production can be highlighted as potential causes of roach population decline and more investigations are needed in order to determine which ones are the most impacting on fish population and communities in the River Meuse to make conservation measures efficient.

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