

Journal of Feline Medicine and Surgery

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Journal of Feline Medicine and Surgery 2013 15: 563
DOI: 10.1177/1098612X13489214

The online version of this article can be found at:

<http://jfm.sagepub.com/content/15/7/563>

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BARTONELLA SPECIES INFECTION IN CATS

ABCD guidelines on prevention and management

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Bacterial properties

Bartonella (previously named *Rochalimaea*) species are small, vector-transmitted Gram-negative intracellular bacteria that are well adapted to one or more mammalian reservoir hosts.

To date, over 22 *Bartonella* species have been described, but their role as pathogens of humans and domestic animals is the subject of ongoing investigations (Table 1).¹ The most common species in both cats and humans is *B henselae*, the agent of cat scratch disease as well as of other potentially fatal disorders in immunocompromised people.

B henselae is naturally transmitted among cats by the flea *Ctenocephalides felis felis*, or by flea faeces. In the infected cat, *Bartonella* inhabits red blood cells, which are ingested by the flea, and the bacterium survives in the flea's gut. Contaminated flea faeces deposited on the skin of the cat end up under the cat's claws

European Advisory Board on Cat Diseases
The European Advisory Board on Cat Diseases (ABCD) is a body of experts in immunology, vaccinology and clinical feline medicine that issues guidelines on prevention and management of feline infectious diseases in Europe, for the benefit of the health and welfare of cats. The guidelines are based on current scientific knowledge of the diseases and available vaccines concerned.

The latest version of the *Bartonella* species infection in cats guidelines is available at www.abcd-vets.org

due to self-scratching. A cat scratch is the common mode of transmission of the organism to other animals, including humans.²

B henselae has been experimentally transmitted among cats by transferring fleas fed on naturally infected cats to specific pathogen-free (SPF) cats, and by intradermal inoculation of excrement collected from fleas fed on *B henselae*-infected cats.² This has demonstrated that both the vector and the cat – through scratches – may transmit the organism. Infection is amplified in the flea hindgut, and *B henselae* can persist in the environment in flea faeces for at least 9 days.³ Blood transfusion also represents a risk: cats have been experimentally infected with *B henselae* and *B clarridgeiae* by intravenous or intramuscular inoculation with infected cat blood.⁴

Overview: Over 22 *Bartonella* species have been described in mammals, and *Bartonella henselae* is most common worldwide. Cats are the main reservoir for this bacterium. *B henselae* is the causative agent of cat scratch disease in man, a self-limiting regional lymphadenopathy, but also of other potentially fatal disorders in immunocompromised people.

Infection: *B henselae* is naturally transmitted among cats by the flea *Ctenocephalides felis felis*, or by flea faeces. A cat scratch is the common mode of transmission of the organism to other animals, including humans. Blood transfusion also represents a risk.

Disease signs: Most cats naturally infected by *B henselae* do not show clinical signs but cardiac (endocarditis, myocarditis) or ocular (uveitis) signs may be found in sporadic cases. *B vinsonii* subspecies *berkhoffii* infection has reportedly caused lameness in a cat affected by recurrent osteomyelitis and polyarthritis.

Diagnosis: Isolation of the bacterium is the gold standard, but because of the high prevalence of infection in healthy cats in endemic areas, a positive culture (or polymerase chain reaction) is not confirmatory. Other compatible diagnoses must be ruled out and response to therapy gives a definitive diagnosis. Serology (IFAT or ELISA) is more useful for exclusion of the infection because of the low positive predictive value (39–46%) compared with the good negative predictive value (87–97%). Laboratory testing is required for blood donors.

Disease management: Treatment is recommended in the rare cases where *Bartonella* actually causes disease.

Table 1 Species and subspecies of *Bartonella* that are confirmed or potential human pathogens¹

<i>Bartonella</i> species	Primary reservoir	Vector	Accidental host
<i>B bacilliformis</i>	Human	<i>Lutzomyia verrucarum</i>	None
<i>B quintana</i> *	Human	<i>Pediculus humanus</i>	Cat, dog, monkey
<i>B elizabethae</i>	<i>Rattus norvegicus</i>	<i>Xenopsylla cheopis</i>	Human, dog
<i>B grahamii</i>	Several species of wild mice	Rodent fleas	Human
<i>B henselae</i> *	Cat	<i>Ctenocephalides felis felis</i> (<i>C felis</i>) Other vectors?	Human, dog
<i>B clarridgeiae</i> *	Cat	<i>C felis</i>	Human, dog
<i>B koehlerae</i> *	Cat	<i>C felis</i>	Human
<i>B vinsonii</i> subspecies <i>berkhoffii</i> *	Coyote, dog	Ticks?	Human, cat
<i>B vinsonii</i> subspecies <i>arupensis</i>	<i>Peromyscus leucopus</i>	Ticks? Fleas?	Human
<i>B washoensis</i>	<i>Spermophilus beecheyii</i>	Fleas?	Human, dog
<i>B asiatica</i>	Rabbit	Fleas?	Human

*Zoonotic species

B henselae can persist in the environment in flea faeces for at least 9 days.

B henselae transmission did not occur when infected cats lived together with uninfected cats in a flea-free environment. Transmission consequently does not occur through bites, scratches (in the absence of fleas), grooming, or sharing of litter boxes and food dishes. Furthermore, transmission could not be demonstrated between bacteraemic female cats and uninfected males during mating, or to the kittens of infected females either during gestation or in the neonatal period, again in flea-free environments.⁵

Ticks may also act as vectors for transmission among cats, human beings, dogs and other mammalian hosts: trans-stadial transmission of *B henselae* was demonstrated in *Ixodes ricinus*.⁶

Epidemiological evidence and experimental studies have demonstrated the important role of fleas in the transmission of *B henselae* and *B clarridgeiae* among cats. Three other species, *B koehlerae*, *B bovis* and *B quintana*, have been isolated from cat blood, but the modes of transmission and the reservoir potential of these species in felids have not been established. In addition, *B vinsonii* subspecies *berkhoffii* DNA was detected in the blood of a cat.⁷



Epidemiology

Bartonella species have a worldwide distribution, with highest prevalences in areas where conditions are most favourable for arthropod vectors, mainly fleas. In Europe, many studies have been carried out and the antibody prevalence in cats has ranged from 8–53% (Table 2).

Pathogenesis

Chronic bacteraemia mainly occurs in young cats, under the age of 2 years.²⁰ Young experimentally infected cats maintained relapsing *B henselae* or *B clarridgeiae* bacteraemia for as long as 454 days.²¹ Immune system avoidance via intracellular location, frequent genetic rearrangements and alteration of outer membrane proteins are considered important factors for the maintenance of persistent bacteraemia. The location within erythrocytes and vascular endothelial cells is believed to protect *Bartonella* also from antimicrobial agents.

As the host-adapted reservoir of *B henselae*, cats display minimal pathogenic effects after experimental infection. Gross necropsy findings were unremarkable in experimentally infected cats but some histopathological changes emerged: follicular hyperplasia of lymph nodes and spleen, lymphocyte and plasma cell infiltrates in liver, heart, kidney and eye, and pyogranulomatous inflammation in liver, spleen, kidney, heart and lymph nodes.^{5,21}

Immunity

The antibody response to *B henselae* has been investigated for the identification of vaccine candidates. The kinetics in response to *B henselae* antigens in chronically infected experimen-

Table 2 Antibody prevalence of *Bartonella* infection in the feline population in European countries

Country	Number of cats	Prevalence (%)	Reference
Netherlands	163 (stray)	52	Bergmans et al (1997) ⁸
Austria	96	33	Allerberger et al (1995) ⁹
Switzerland	728	8	Glaus et al (1997) ¹⁰
Germany	713 245	15 37.1	Haimerl et al (1999) ¹¹ Morgenthal et al (2012) ¹²
France	64 94 179	36 53 41	Chomel et al (1995) ¹³ Heller et al (1997) ¹⁴ Gurfield et al (2001) ¹⁵
Spain	680	23.8	Ayllon et al (2012) ¹⁶
Italy	540 1300 (stray)	38 23.1	Fabbi et al (2004) ¹⁷ Brunetti et al (2013) ¹⁸
Scotland	78	15.3	Bennett et al (2011) ¹⁹

tal cats is highly variable in degree and duration.^{2,21,22} Reinfection by a different strain of *B henselae* is possible, as supported by the isolation of unrelated bacterial clones from the same cat at different times.²³ Antibodies are, therefore, considered not protective, and *Bartonella* species antibody positive cats may be infected.¹⁷ A cell-mediated response was not evident in investigated experimentally infected cats.²¹

Clinical signs

Cats naturally infected with *Bartonella* species usually do not show clinical signs. Both experimental and natural infection studies have tried to establish an association between clinical signs and infection, but a link has not been unequivocally proven.

Experimental infection

Exposure to infected fleas does not result in clinical signs [EBM grade II].^{2,24} In some cases of experimental inoculation, a self-limiting febrile disease, transient mild anaemia, localised or generalised lymphadenopathy, mild neurological signs and reproductive failure have been reported [EBM grade III].²¹

Natural infection

The role of *Bartonella* as a cause of clinical signs is even more unclear after natural infection despite a plethora of studies. Studies based on antibody detection are of limited value because antibody only proves exposure, and not necessarily an active infection. Moreover, there is cross-reactivity between different *Bartonella* species and strains that may or may not cause clinical signs. Because of the high percentage of infected healthy cats in endemic areas an association between clinical signs and *B henselae* infection is not easy to demonstrate.

It has been suggested that *Bartonella* infection could play a role in chronic gingivostomatitis, but the prevalence of antibodies or organisms was not higher in diseased cats than in control populations [EBM grade II].^{10,25–30}

Cats positive for both feline immunodeficiency virus (FIV) and *Bartonella* antibodies had in one study an increased risk of lymphadenopathy [EBM grade III].²⁵

An association between *Bartonella* antibodies and urinary tract disease or haematuria was found in two studies [EBM grade III].^{10,31}

Pearce et al did not find any difference in antibody prevalence between healthy cats and cats with seizures or other neurological conditions.³² A non-controlled retrospective study reported *Bartonella* DNA and antibodies in cerebrospinal fluid from cats with CNS disease [EBM grade III].³³

No difference in *Bartonella* antibody preva-

Because of the high percentage of infected healthy cats in endemic areas, an association between clinical signs and *B henselae* infection is not easy to demonstrate.



lence was found between healthy cats and those affected by uveitis, but in some cases evidence of *Bartonella* species exposure was reported in cats with uveitis responsive to drugs considered effective against *Bartonella* [EBM grade IV].^{34–36}

No difference in *Bartonella* antibody prevalence was found in cats affected by anaemia, but in cats positive for *Bartonella* antibodies a significant association with hyperglobulinaemia was seen [EBM grade I].^{37,38} Lappin et al demonstrated no association between fever and positivity to *Bartonella* antibodies or DNA.³⁹

A study based on serology and culture did not find an association between *Bartonella* infection and chronic rhinosinusitis.⁴⁰ Also no link was reported between *Bartonella* infection and pancreatitis, based on the finding that cats with normal feline pancreatic lipase immunoreactivity values and cats with elevated values did not show any difference in *Bartonella* antibody prevalence.⁴¹

A few case reports concern *B henselae*-associated endocarditis or myocarditis. Fatal aortic and mitral valve *B henselae*-associated endocarditis was reported in two cats in the USA.^{42,43} Moreover, *B henselae* anterior mitral valve leaflet vegetative endocarditis was successfully treated in a cat presenting with a grade III/IV systolic heart murmur and signs of aortic embolisation (lethargy and weakness in the hind limbs, weak femoral pulses, pelvic pain, increased serum creatine kinase activity).⁴⁴ This case report confirms that *Bartonella* species may be a cause of blood culture-negative endocarditis, as suspected.⁴⁵ *Bartonella*-associated myocarditis was suspected in a cat presenting with supraventricular tachycardia responsive to antibiotic therapy.⁴⁶

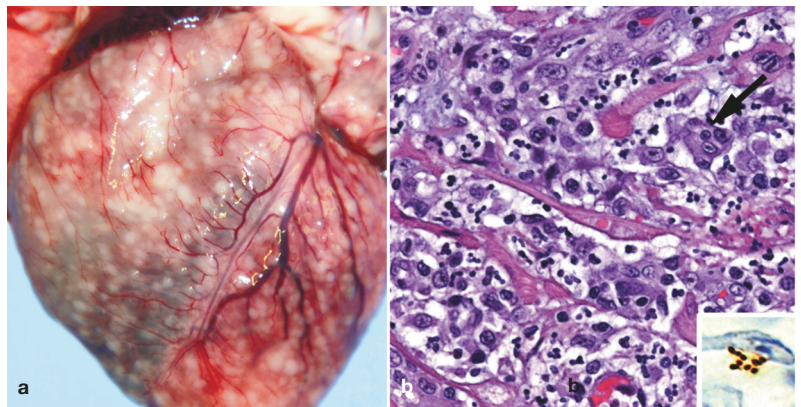


Figure 1 Gross and histological findings in two cats from a North Carolina shelter that died after a litter of flea-infested kittens was introduced to the shelter. (a) Coalescing granulomas distributed throughout the myocardium of an approximately 9-week-old female kitten. (b) Pyogranulomatous myocarditis in an 8-month-old castrated male cat, which had been co-housed with the flea-infested kittens. Macrophages, with a rare multinucleated giant cell (arrow), are particularly numerous towards the upper left of the image; haematoxylin and eosin stain. Inset: Cluster of short bacilli in an inflammatory focus are immunoreactive (brown) for *B henselae*-specific monoclonal antibody; immunohistochemistry with diaminobenzidine chromogen and haematoxylin counterstain. Reproduced, with permission, from Varanat et al (2012)⁴⁷

Disease in humans

B. henselae is the causative agent of cat scratch disease (Figure 2). This is a usually self-limiting regional lymphadenopathy that develops after a primary papular lesion and lasts for a few weeks to several months.⁴⁸ Abscessation of the lymph node and systemic signs are occasionally reported. Atypical forms and an expanding spectrum of clinical conditions are being associated with *B. henselae* infection, such as neuroretinitis, uveitis, endocarditis and neurological disorders.^{48–52} An unusual case of cat scratch disease has been reported

in a veterinarian affected by persistent fever and back pain after an accidental needle puncture.⁵³ The occupational risk of exposure to arthropod vectors, as well as the reservoir host, should be also considered by veterinarians for this vector-borne disease.⁵⁴ Immunocompetent individuals may even experience subclinical *Bartonella* infection; conversely, immunocompromised persons

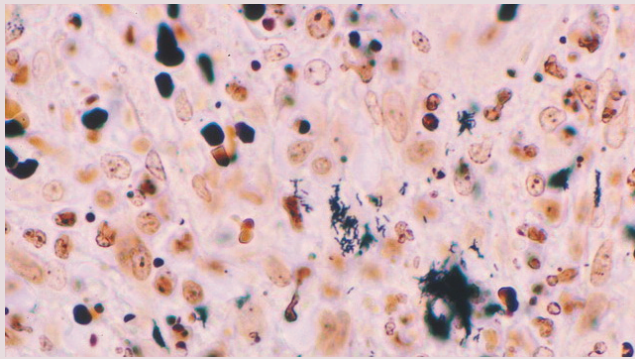


Figure 2 Immunohistochemical identification of *B. henselae* in a case of cat scratch disease. With Warthin-Starry silver stain, these coccobacillary pathogens can be seen singly, in small clumps or in chains in necrotic foci. Courtesy of Dharam Ramnani, Webpathology.com

suffer from clinical forms of a disseminated infection that may be fatal if untreated.^{55,56}

Zoonosis
B. henselae is the causative agent of cat scratch disease.



Post mortem evidence of pyogranulomatous myocarditis and diaphragmatic myositis associated with *B. henselae* infection was also obtained in two cats (Figure 1).⁴⁷

Lameness and pain during limb palpation were observed in a cat affected by recurrent osteomyelitis and polyarthrits associated with *B. vinsonii* subspecies *berkhoffii* infection and bacteraemia.⁷

In conclusion, most cats naturally infected by *B. henselae* do not show clinical signs. The identification of *Bartonella* infection in cats with disease should prompt a critical assessment of the role of the infection in the causation of the clinical signs and the exclusion of other compatible diagnoses.

Diagnosis

Bartonella laboratory testing is required for feline blood donors, for pet cats belonging to immunosuppressed persons, or when a human *Bartonella*-related disease is diagnosed in a cat's home.

Isolation of the bacterium is the gold standard, but because of the high prevalence of infection in healthy cats in endemic areas, a positive culture is not confirmatory, and other compatible diagnoses must be ruled out.

The disease is, therefore, diagnosed on the basis of exclusion, and by assessing the response to therapy. The *ex juvantibus* inference about disease causation from the observed response to a treatment may apply to uveitis, endocarditis, myocarditis, osteoarthritis and multifocal central nervous system (CNS) disease, which are considered compatible with feline bartonellosis.

PCR may be used on samples of blood,

aqueous humour, cerebrospinal fluid or tissues, and several gene targets have been studied. To reduce false-negative test results, repeated blood cultures are required or PCR performed on more than one kind of biological sample (blood, lymph node, oral swab).

Serology (IFAT or ELISA) is more useful for exclusion than for confirmation of the infection because of the low positive predictive value (39–46%) compared with the good negative predictive value (87–97%) [EBM grade III].^{13,15,17,20} *Bartonella* IgM antibodies are found in experimentally and naturally infected cats but their occurrence seems not to be related to bacteraemia [EBM grade II].⁵⁷

Treatment

Treatment is recommended in the rare cases where *Bartonella* has actually caused disease (eg, endocarditis).

EBM grades
The ranking system for grading the level of evidence of various statements within this article is described on page 533 of this Special Issue.

Table 3 Suggested treatment for bartonellosis in cats				
Drug	Dose	Duration	Reference(s)	EBM grade
Doxycycline	10 mg/kg PO q12–24 h	2–4 weeks	Lappin and Black (1999) ³⁵	IV
Azithromycin	10 mg/kg PO q24h (q48h)	For 7 days followed by every other day for 6–12 weeks	Ketring et al (2004) ³⁶ Varanat et al (2009) ⁷	IV
Marbofloxacin	5 mg/kg PO q24h	6 weeks	Perez et al (2010) ⁴⁴	IV
Amoxicillin–clavulanate (with azithromycin)	62.5 mg PO q12h	2 months	Varanat et al (2009) ⁷	IV

Public health considerations

Cats are the main reservoir for *B. henselae*, the agent of cat scratch disease and more severe human diseases observed in severely immunosuppressed persons such as bacillary angiomatosis and peliosis hepatis. Recognised risk factors for bacteraemia in cats are young age, infestation with fleas, outdoor lifestyle and a multicat environment.^{13,15,20,48}

There is no evidence of benefit to cats or their owners from routine testing of the pet for *Bartonella* infection or from antibiotic treatment of healthy antibody positive cats [EBM grade IV].⁶¹ Infection does not always lead to clinical signs in healthy persons and many of them have antibodies.⁵⁵

Owner education by practitioners about *Bartonella* transmission is essential to reduce the zoonotic risk, and the presence of immunosuppressed persons in the cat family should be specifically considered. It is recommended that immunosuppressed persons are allowed to keep their pet cat or to adopt a new one and a few simple tips summarised on the right can minimise their exposure risk.^{61,62}

Recommendations for prevention of *B. henselae* infection in immunocompromised persons^{61,62}

- ✦ Immunocompromised owners should adopt cats that are older than 1 year, flea-free, in good health, not from shelters or multicat households, and have no history of contact with cats of unknown health status
- ✦ Strict flea control should be exercised under veterinary supervision
- ✦ Rough play should be avoided, and the cat's claws kept trimmed
- ✦ Any wound should promptly be cleaned with soap and water, and medical advice sought
- ✦ Cats should be kept indoors to avoid exposure to fleas and other possible vectors, and also to prevent other zoonotic risks

Current therapeutic strategies in cats (Table 3) are based on in vitro studies and human bartonellosis. Data from controlled efficacy studies in cats are lacking. A cat affected by recurrent osteomyelitis and polyarthritides associated with *B. vinsonii* subspecies *berkhoffii* genotype II infection and bacteraemia recovered after therapy with azithromycin (10 mg/kg PO q48h for 3 months) and amoxicillin-clavulanate (62.5 mg PO q12 for 2 months) [EBM grade IV].⁷

After natural or experimental infection with *B. henselae* or *B. clarridgeiae*, healthy cats have been treated to eliminate bacteraemia and many drugs have been evaluated: doxycycline, amoxicillin, amoxicillin-clavulanate, enrofloxacin, erythromycin and rifampicin

[EBM grade II].⁵⁸⁻⁶⁰ Based on these results, clearance of bacteraemia cannot be guaranteed and, in the case of failure, there is the risk of inducing antimicrobial resistance. Treatment of healthy carriers, therefore, cannot be considered an effective measure for eliminating the zoonotic risk, as is sometimes requested in human cases of cat scratch disease or where other *Bartonella*-related disease occurs in a family member.

Prevention

Based on transmission studies to date, strict flea (and tick) control is the only successful preventive measure. There is no vaccine available against *Bartonella* infection.

KEY POINTS

- ✦ *B. henselae* is the most common *Bartonella* species infecting cats, which are the main reservoir for this bacterium.
- ✦ *B. henselae* is the causative agent of cat scratch disease in man, a self-limiting regional lymphadenopathy.
- ✦ *B. henselae* is naturally transmitted among cats by the faeces of *Ctenocephalides felis felis* fleas; contact infections do not occur. Blood transfusion also represents a risk.
- ✦ Most cats naturally infected by *B. henselae* do not show clinical signs but sporadic cases of endocardial, myocardial or ocular feline disease have been reported.
- ✦ Other *Bartonella* species, for which cats are accidental hosts, may have more pathogenic potential.
- ✦ Antibodies are not protective, and seropositive cats may be reinfected.
- ✦ Bartonellosis is diagnosed on the basis of exclusion, and by assessing the response to antibiotic therapy.
- ✦ Testing healthy cats or people is of no benefit.
- ✦ Treatment of healthy carriers is no measure for eliminating the zoonotic risk.
- ✦ Strict flea and tick control is the only effective preventive measure.



Funding

The authors received no specific grant from any funding agency in the public, commercial or not-for-profit sectors for the preparation of this article. The ABCD is supported by Merial, but is a scientifically independent body.

Conflict of interest

The authors do not have any potential conflicts of interest to declare.

References

- Chomel BB, Boulouis H-J, Maruyama S and Breitschwerdt EB. *Bartonella* spp. in pets and effect on human health. *Emerg Infect Dis* 2006; 12: 389–394.
- Chomel BB, Kasten RW, Floyd-Hawkins KA, Chi B, Yamamoto K, Roberts-Wilson J, et al. Experimental transmission of *Bartonella henselae* by the cat flea. *J Clin Microbiol* 1996; 34: 1952–1956.
- Finkelstein JL, Brown TP, O'Reilly KL, Wedincamp J Jr and Foil LD. Studies on the growth of *Bartonella henselae* in the cat flea (*Siphonaptera: Pulicidae*). *J Med Entomol* 2002; 39: 915–919.
- Abbott RC, Chomel BB, Kasten RW, Floyd-Hawkins KA, Kikuchi Y, Koehler JE, et al. Experimental and natural infection with *Bartonella henselae* in cats. *Comp Immunol Microbiol Infect Dis* 1997; 20: 41–57.
- Guptill L, Slater L, Wu C-C, Lin T-L, Glickman LT, Welch DF, et al. Experimental infection of young specific pathogen-free cats with *Bartonella henselae*. *J Infect Dis* 1997; 176: 206–216.
- Cotté V, Bonnet S, Le Rhun D, Le Naour E, Chauvin A, Boulouis H-J, et al. Transmission of *Bartonella henselae* by *Ixodes ricinus*. *Emerg Infect Dis* 2008; 14: 1074–1080.
- Varanat A, Travis A, Lee W, Maggi RG, Bissett SA, Linder KE, et al. Recurrent osteomyelitis in a cat due to infection with *Bartonella vinsonii* subsp. *berkhoffii* genotype II. *J Vet Intern Med* 2009; 23: 1273–1277.
- Bergmans AMC, de Jong CMA, van Amerongen G, Schot CS and Schouls LM. Prevalence of *Bartonella* species in domestic cats in The Netherlands. *J Clin Microbiol* 1997; 35: 2256–2261.
- Allerberger F, Schonbauer M, Zangerle R and Dierich M. Prevalence of antibody to *Rochalimaea henselae* among Austrian cats. *Eur J Ped* 1995; 154: 165.
- Glaus T, Hofmann-Lehmann R, Greene C, Glaus B, Wolfensberger C and Lutz H. Seroprevalence of *Bartonella henselae* infection and correlation with disease status in cats in Switzerland. *J Clin Microbiol* 1997; 35: 2883–2885.
- Haimerl M, Tenter AM, Simon K, Rommel M, Hilger J and Autenrieth IB. Seroprevalence of *Bartonella henselae* in cats in Germany. *J Med Microbiol* 1999; 48: 849–856.
- Morgenthal D, Hamel D, Arndt G, Silaghi C, Pfister K, Kempf VA, et al. Prevalence of haemotropic *Mycoplasma* spp, *Bartonella* spp and *Anaplasma phagocytophilum* in cats in Berlin/Brandenburg (Northeast Germany). *Berl Munch Tierarztl Wochenschr* 2012; 125: 418–427.
- Chomel BB, Gurfield AN, Boulouis HJ, Kasten RW and Piemont Y. Réservoir félin de l'agent de la maladie des griffes du chat, *Bartonella henselae*, en région parisienne: resultants préliminaires. *Rec Med Vet* 1995; 171: 841–845.
- Heller R, Artois M, Xemar V, de Briel D, Gehin H, Jaulhac B, et al. Prevalence of *Bartonella henselae* and *Bartonella clarridgeiae* in stray cats. *J Clin Microbiol* 1997; 35: 1327–1331.
- Gurfield AN, Boulouis H-J, Chomel BB, Kasten RW, Heller R, Bouillin C, et al. Epidemiology of *Bartonella* infection in domestic cats in France. *Vet Microbiol* 2001; 80: 185–198.
- Ayllon T, Diniz PP, Breitschwerdt EB, Villaescusa A, Rodriguez-Franco F and Sainz A. Vector-borne diseases in client-owned and stray cats from Madrid, Spain. *Vector Borne Zoonotic Dis* 2012; 12: 143–150.
- Fabbi M, De Giuli L, Tranquillo M, Bragoni R, Casiraghi M and Genchi C. Prevalence of *Bartonella henselae* in Italian stray cats: evaluation of serology to assess the risk of transmission of *Bartonella* to humans. *J Clin Microbiol* 2004; 42: 264–268.
- Brunetti E, Fabbi M, Ferraioli G, Prati P, Filice C, Sasseria D, et al. Cat-scratch disease in Northern Italy: atypical clinical manifestations in humans and prevalence of *Bartonella* infection in cats. *Eur J Clin Microbiol Infect Dis* 2013; 32: 531–534.
- Bennett AD, Gunn-Moore DA, Brewer M and Lappin MR. Prevalence of *Bartonella* species, haemoplasmas and *Toxoplasma gondii* in cats in Scotland. *J Feline Med Surg* 2011; 13: 553–557.
- Guptill L, Wu CC, HogenEsch H, Slater LN, Glickman N, Dunham A, et al. Prevalence, risk factors, and genetic diversity of *Bartonella henselae* infections in pet cats in four regions of the United States. *J Clin Microbiol* 2004; 42: 652–659.
- Kordick D, Brown TT, Shin K and Breitschwerdt EB. Clinical and pathologic evaluation of chronic *Bartonella henselae* or *Bartonella clarridgeiae* infection in cats. *J Clin Microbiol* 1999; 37: 1536–1547.
- Yamamoto K, Chomel BB, Kasten RW, Hew CM, Weber DK and Lee WI. Experimental infection of specific pathogen free (SPF) cats with two different strains of *Bartonella henselae* type I: a comparative study. *Vet Res* 2002; 33: 669–684.
- Arvand M, Viezens J and Berghoff J. Prolonged *Bartonella henselae* bacteremia caused by reinfection in cats. *Emerg Infect Dis* 2008; 14: 152–154.
- Bradbury CA and Lappin MR. Evaluation of topical application of 10% imidacloprid-1% moxidectin to prevent *Bartonella henselae* transmission from cat fleas. *J Am Vet Med Assoc* 2009; 236: 869–873.
- Ueno H, Hohdatsu T, Muramatsu Y, Koyama H and Morita C. Does coinfection of *Bartonella henselae* and FIV induce clinical disorders in cats? *Microbiol Immunol* 1996; 40: 617–620.
- Quimby J, Elston T, Hawley J, Brewer M, Miller A and Lappin MR. Evaluation of the association of *Bartonella* species, feline herpesvirus 1, feline calicivirus, feline leukemia virus and feline immunodeficiency virus with chronic feline gingivostomatitis. *J Feline Med Surg* 2008; 10: 66–72.
- Dowers KL, Hawley JR, Brewer MM, Morris AK, Radecki SV and Lappin MR. Association of *Bartonella* species, feline calicivirus, and feline herpesvirus 1 infection with gingivostomatitis in cats. *J Feline Med Surg* 2010; 12: 314–321.
- Pennisi MG, La Camera E, Giacobbe L, Orlandella BM, Lentini V, Zummo S, et al. Molecular detection of *Bartonella henselae* and *Bartonella clarridgeiae* in clinical samples of pet cats from Southern Italy. *Res Vet Sci* 2010; 88: 379–384.
- Belgard S, Truyen U, Thibault JC, Sauter-Louis C and Hartmann K. Relevance of feline calicivirus, feline immunodeficiency virus, feline leukemia virus, feline herpesvirus and *Bartonella henselae* in cats with chronic gingivostomatitis. *Berl Munch Tierarztl Wochenschr* 2010; 123: 369–376.
- Namekata DY, Kasten RW, Boman DA, Straub MH, Siperstein-Cook L, Couvelaire K, et al. Oral shedding of *Bartonella* in cats: correlation with bacteremia and seropositivity. *Vet Microbiol* 2010; 146: 371–375.

- 31 Breitschwerdt EB, Levine JF, Radulovich S, Hanby SB, Kordick DL and La Perle KMD. *Bartonella henselae* and *Rickettsia seroreactivity* in a sick cat population from North Carolina. *Int J Appl Res Vet Med* 2005; 3: 287–302.
- 32 Pearce LK, Radecki SV, Brewer M and Lappin MR. **Prevalence of *Bartonella henselae* antibodies in serum of cats with and without clinical signs of central nervous system disease.** *J Feline Med Surg* 2006; 8: 315–320.
- 33 Leibovitz K, Pearce L, Brewer M and Lappin MR. *Bartonella* species antibodies and DNA in cerebral spinal fluid of cats with central nervous system disease. *J Feline Med Surg* 2008; 10: 332–337.
- 34 Fontenelle JP, Powell CC, Hill AE and Radecki SV. **Prevalence of serum antibodies against *Bartonella* species in the serum of cats with or without uveitis.** *J Feline Med Surg* 2008; 10: 41–46.
- 35 Lappin MR and Black JC. *Bartonella* spp infection as a possible cause of uveitis in a cat. *J Am Vet Med Assoc* 1999; 214: 1205–1207.
- 36 Ketring KL, Zuckerman EE and Hardy Jr WD. *Bartonella*: a new etiologic agent of feline ocular disease. *J Am Anim Hosp Assoc* 2004; 40: 6–12.
- 37 Ishak AM, Radecki S and Lappin MR. **Prevalence of *Mycoplasma haemofelis*, 'Candidatus *Mycoplasma haemominutum*', *Bartonella* species, *Ehrlichia* species, and *Anaplasma phagocytophilum* DNA in the blood of cats with anemia.** *J Feline Med Surg* 2007; 9: 1–7.
- 38 Whittmore JC, Hawley JR, Radecki SV, Steinberg JD and Lappin MR. *Bartonella* species antibodies and hyperglobulinemia in privately owned cats. *J Vet Intern Med* 2012; 26: 639–644.
- 39 Lappin MR, Breitschwerdt E, Brewer M, Hawley J and Hegarty B. **Prevalence of *Bartonella* species antibodies and *Bartonella* species DNA in the blood of cats with and without fever.** *J Feline Med Surg* 2009; 11: 141–148.
- 40 Berryessa NA, Johnson LR, Kasten RW and Chomel BB. **Microbial culture of blood samples and serologic testing for bartonellosis in cats with chronic rhinosinusitis.** *J Am Vet Med Assoc* 2008; 233: 1084–1089.
- 41 Bayliss DB, Steiner JM, Sucholdolski JS, Radecki SV, Brewer MM, Morris AK, et al. **Serum feline pancreatic lipase immunoreactivity concentration and seroprevalences of antibodies against *Toxoplasma gondii* and *Bartonella* species in client-owned cats.** *J Feline Med Surg* 2009; 11: 663–667.
- 42 Chomel B, Wey AC, Kasten RW, Stacy BA and Labelle P. **Fatal case of endocarditis associated with *Bartonella henselae* type I infection in a domestic cat.** *J Clin Microbiol* 2003; 41: 5337–5339.
- 43 Chomel BB, Kasten RW, Williams C, Wey AC, Henn JB, Maggi R, et al. *Bartonella* endocarditis: a pathology shared by animal reservoirs and patients. *Ann N Y Acad Sci* 2009; 1166: 120–126.
- 44 Perez C, Hummel JB, Keene BW, Maggi RG, Diniz PPVP and Breitschwerdt EB. **Successful treatment of *Bartonella henselae* endocarditis in a cat.** *J Feline Med Surg* 2010; 12: 483–486.
- 45 Malik R, Barrs VR, Church DB, Zahn A, Allan GS, Martin P, et al. **Vegetative endocarditis in six cats.** *J Feline Med Surg* 1999; 1: 171–180.
- 46 Nakamura RK, Zimmerman SA and Lesser MB. **Suspected *Bartonella*-associated myocarditis and supraventricular tachycardia in a cat.** *J Vet Cardiol* 2011; 13: 277–281.
- 47 Varanat M, Broadhurst J, Linder KE, Maggi RG and Breitschwerdt EB. **Identification of *Bartonella henselae* in two cats with pyogranulomatous myocarditis and diaphragmatic myositis.** *Vet Pathol* 2012; 49: 608–611.
- 48 Boulouis HJ, Chang CC, Henn JB, Kasten RW and Chomel BB. **Factors associated with the rapid emergence of zoonotic *Bartonella* infections.** *Vet Res* 2005; 36: 383–410.
- 49 Fonollosa A, Galdos M, Artaraz J, Perez-Irezabal J and Martinez-Alday N. **Occlusive vasculitis and optic disk neovascularization associated with neuroretinitis.** *Ocul Immunol Inflamm* 2011; 19: 62–64.
- 50 Tsuneoka H, Yanagihara M, Otani S, Katayama Y, Fujinami H, Nagafuji H, et al. **A first Japanese case of *Bartonella henselae*-induced endocarditis diagnosed by prolonged culture of a specimen from the excised valve.** *Diagn Microbiol Infect Dis* 2010; 68: 174–176.
- 51 Mascarelli PE, Maggi RG, Hopkins S, Mozayeni BR, Trull CL, Bradley JM, et al. ***Bartonella henselae* infection in a family experiencing neurological and neurocognitive abnormalities after woodlouse hunter spider bites.** *Parasit Vectors* 2013; 6: 98. DOI: 10.1186/1756-3305-6-98.
- 52 Maggi RG, Ericson M, Mascarelli PE, Bradley JM and Breitschwerdt EB. ***Bartonella henselae* bacteremia in a mother and son potentially associated with tick exposure.** *Parasit Vectors* 2013; 6: 101. DOI: 10.1186/1756-3305-6-101.
- 53 Lin JW, Chen CM and Chang CC. **Unknown fever and back pain caused by *Bartonella henselae* in a veterinarian after a needle puncture: a case report and literature review.** *Vector Borne Zoonotic Dis* 2011; 11: 589–591.
- 54 Maggi RM, Mascarelli PE, Havenga LN, Naidoo V and Breitschwerdt EB. **Co-infection with *Anaplasma platys*, *Bartonella henselae* and *Candidatus Mycoplasma haematoparvum* in a veterinarian.** *Parasit Vectors* 2013; 6: 103. DOI: 10.1186/1756-3305-6-103.
- 55 Massei F, Messina F, Gori L, Macchia P and Maggiore G. **High prevalence of antibodies to *Bartonella henselae* among Italian children without evidence of cat scratch disease.** *Clin Infect Dis* 2004; 38: 145–148.
- 56 Lange D, Oeder C, Waltermann K, Mueller A, Oehme A, Rohrberg R, et al. **Bacillary angiomatosis.** *J Dtsch Dermatol Ges* 2009; 7: 767–769.
- 57 Ficociello J, Bradbury C, Morris A and Lappin MR. **Detection of *Bartonella henselae* IgM in serum of experimentally infected and naturally exposed cats.** *J Vet Intern Med* 2011; 25: 1264–1269.
- 58 Greene CE, McDermott M, Jameson PH, Atkins CL and Marks AM. ***Bartonella henselae* infection in cats: evaluation during primary infection, treatment, and rechallenge infection.** *J Clin Microbiol* 1996; 34: 1682–1685.
- 59 Regnery RL, Rooney JA, Johnson AM, Nesby SL, Manzwitsch P, Beaver K, et al. **Experimentally induced *Bartonella henselae* infections followed by challenge exposure and antimicrobial therapy in cats.** *Am J Vet Res* 1996; 57: 1714. Erratum in: *Am J Vet Res* 1997; 58: 803.
- 60 Kordick DL, Papich MG and Breitschwerdt EB. **Efficacy of enrofloxacin or doxycycline for treatment of *Bartonella henselae* or *Bartonella clarridgeiae* infection in cats.** *Antimicrob Agents Chemother* 1997; 41: 2448–2455.
- 61 Kaplan JE, Benson C, Holmes KH, Brooks JT, Pau A, Masur H, et al. **Guidelines for prevention and treatment of opportunistic infections in HIV-infected adults and adolescents: recommendations from CDC, the National Institute of Health, and the HIV Medicine Association of the Infectious Diseases Society of America.** *MMWR Recomm Rep* 2009; 58: 1–207.
- 62 Brunt J, Guptill L, Kordick DL, Kudrak S and Lappin MR. **American Association Feline Practitioners 2006 Panel report on diagnosis, treatment, and prevention of *Bartonella* spp infections.** *J Feline Med Surg* 2006; 8: 213–226.