Dual Blockade of the Homeostatic Chemokine CXCL12 and the Proinflammatory Chemokine CCL2 Has Additive Protective Effects on Diabetic Kidney Disease

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Monocyte/chemoattractant protein-1/chemokine ligand (CCL) 2 and stromal cell–derived factor-1/CXCL12 both contribute to glomerulosclerosis in mice with type 2 diabetes mellitus, through different mechanisms. CCL2 mediates macrophage-related inflammation, whereas CXCL12 contributes to podocyte loss. Therefore, we hypothesized that dual antagonism of these chemokines might have additive protective effects on the progression of diabetic nephropathy. We used chemokine antagonists based on structured l-enantiomeric RNA (so-called Spiegelmers) ie, the CCL2-specific mNOX-E36 and the CXCL12-specific NOX-A12. Male db/db mice, uninephrectomized at the age of 6 weeks, received injections of Spiegelmer, both Spiegelmers, nonfunctional control Spiegelmer, or vehicle from the age of 4 months for 8 weeks. Dual blockade was significantly more effective than monotherapy in preventing glomerulosclerosis. CXCL2 blockade reduced glomerular leukocyte counts and renal-inducible nitric oxide synthase or IL-6 mRNA expression. CXCL12 blockade maintained podocyte numbers and renal nephrin and podocin mRNA expression. Consistently, CXCL12 blockade suppressed nephrin mRNA up-regulation in primary cultures of human glomerular progenitors induced to differentiate toward the podocyte lineage. All previously mentioned parameters were significantly improved in the dual-blockade group, which also suppressed proteinuria and was associated with the highest levels of glomerular filtration rate. Blood glucose levels and body weight were identical in all treatment groups. Dual chemokine blockade can have additive effects on the progression of diabetic kidney disease when the respective chemokine targets mediate different pathomechanisms of disease (ie, inflammation and progenitor differentiation toward the podocyte lineage). (Am J Pathol 2011, 179:116–124; DOI: 10.1016/j.ajpath.2011.03.004)
tory chemokines promote tissue inflammation and remodeling by recruiting and activating immune cells in DN, as in other types of kidney diseases.\textsuperscript{7–10} For example, targeted deletion or inhibition of monocyte chemoattractant protein-1 [renamed chemokine ligand (CCL) 2] signaling can prevent glomerulosclerosis by blocking macrophage recruitment to the glomeruli of mice with type 1 or 2 diabetes mellitus.\textsuperscript{11,12} In fact, CCL2 may represent a promising therapeutic target in DN because delayed onset of CCL2 blockade was able to prevent diabetic glomerulosclerosis and restore the glomerular filtration rate (GFR) by preventing glomerular macrophage recruitment in late-stage DN of uninephrectomized db/db mice with type 2 diabetes.\textsuperscript{13}

Although structurally related, a subgroup of the chemokine superfamily, known as “homeostatic” chemokines, displays functions independent of tissue inflammation. Homeostatic chemokines are rather constitutively expressed because they contribute to the physiological homing and migration of immune cells in the bone marrow or lymphoid organs.\textsuperscript{14} For example, recently, the homeostatic chemokine stromal cell–derived factor-1, also named CXCL12, was constitutively expressed by podocytes and CXCL12 blockade prevents diabetic glomerulosclerosis in a way that was independent of glomerular macrophage recruitment.\textsuperscript{15} The mechanism underlying the protective effect of CXCL12 blockade on DN remains unclear, but a profound effect on podocyte counts and proteinuria was documented. Because we could not detect increased bone marrow–derived progenitor cells in kidneys of treated db/db mice, an enhancer effect on the capacity of intraglomerular podocyte progenitors to restore podocytes in DN was suspected.\textsuperscript{15}

Our previous attempts to increase the therapeutic effects of chemokine blockade on the progression of DN were not successful. For example, blockade of CCR2 and CCR5 with a dual antagonist did not reveal any additional effects compared with single CCR blockade (S.G.S and H.J.A., unpublished data). We had attributed this finding to the fact that CCR2 and CCR5 are redundant when coexpressed on the same class of monocytes, which may preclude additive effects on glomerular pathological characteristics in DN.

Therefore, we hypothesized that combining the protective effects of reducing glomerular leukocyte recruitment by blocking the proinflammatory chemokine CCL2 with the protective effects on podocyte loss by blocking the homeostatic chemokine CXCL12 should finally elicit additive protective effects on diabetic glomerulosclerosis. As such, we compared monotherapy of either CCL2 or CXCL12 antagonists with dual blockade in an accelerated model of glomerulosclerosis in db/db mice with type 2 diabetes.

\section*{Materials and Methods}

\textbf{CCL2 and CXCL12 Antagonistic Spiegelmers}

The sequences of the CCL2 antagonistic Spiegelmer mNOX-E36 (5’-GGCGACAUUGGUUGGCAUGAGGC-GAGGCCCUUUAGGAUACCGCGGCCA-3’) and the CXCL12 antagonistic Spiegelmer NOX-A12 (5’-CGCGUCGUGUGAUCAUGAUUGUCCGAUGUCAG-GUACGC-3’) were obtained from in vitro selection experiments that were essentially performed as described.\textsuperscript{16} To distinguish possible target-specific from unspecific substance class effects, a nonfunctional Spiegelmer with the reverse sequence of NOX-A12 (5’-CGCAUGGACUGAUCCAGCGGUUUAGAUCUGUGUGGGGU-GCG-3’) was used. The Spiegelmers were modified with 40-kDa branched polyethylene glycol at the 3’ end (mNOX-E36) or the 5’ end (NOX-A12 and reverse sequence of NOX-A12). mNOX-E36 and NOX-A12 bind to CCL2 and CXCL12, respectively, with subnanomolar affinities. Chemokine inhibition was determined for each Spiegelmer using leukocyte chemotaxis assays, as described.\textsuperscript{16,17} All Spiegelmers were previously tested as single agents in DN by our group, as described.\textsuperscript{13}

\section*{Animal Studies}

Male diabetic C57BLKS db/db mice or nondiabetic C57BL/6 mice, aged 5 weeks, were obtained from Taconic (Ry, Denmark) and housed in filter-top cages with a 12-hour dark-light cycle. All animals had unlimited access to food and water throughout the duration of the study. At the age of 6 weeks, uninephrectomy (1K mice) or sham surgery was performed through a 1-cm flank incision, as previously described.\textsuperscript{18} In the sham surgery group, the kidney was left in situ. At the age of 4 months, 1K db/db mice with documented blood glucose levels $>11$ mmol/L and creatinine/albumin ratios $>3$ (ratio in age-matched wild-type mice, 0.1) were divided into four groups ($n = 10$ to 12); each of these groups received either nil (no injections) or s.c. injections of 50 mg/kg NOX-A12, mNOX-E36, or control Spiegelmer in 5% glucose or 5% glucose itself as a vehicle control, on every other day. This dose corresponds to 13.4 mg/kg NOX-A12 or 14.4 mg/kg mNOX-A36 if based on the oligonucleotide part of the molecule as anhydrous free acid. The treatment dose and injection intervals were comparable to those in previous db/db mouse studies.\textsuperscript{13,15,19} with similar formulations of Spiegelmers targeting these chemokines. Treatment was continued for 8 weeks. Tissues were harvested for histopathological evaluation at the end of the treatment period, 3 to 4 hours after the last Spiegelmer injection. Blood and urine samples were obtained at monthly intervals for the estimations of urinary albumin, by enzyme-linked immunosorbent assay (Bethyl Labs, Montgomery, TX); and serum and urinary creatinine, by Jaffé reaction (DiaSys Diagnostic Systems, Holzheim, Germany). Blood glucose levels were monitored using an Accu check sensor (Roche, Mannheim, Germany). Plasma chemokine levels were determined at the end of the treatment period by enzyme-linked immunosorbent assay (CCL2: BD Biosciences Pharmingen, San Diego, CA; CXCL12: R&D Systems, Wiesbaden, Germany). All animal experiments were approved by the local government authorities.
Histopathological Characteristics

Kidneys were harvested from each mouse and were fixed in 10% formalin in PBS and embedded in paraffin. Sections, 2 μm, were stained with PAS reagent. Glomerular sclerotic lesions were assessed using a semiquantitative score by a blinded observer, as follows: 0 indicates no lesion; 1, <25% sclerotic; 2, 25% to 49% sclerotic; 3, 50% to 74% sclerotic; and 4, 75% to 100% sclerotic. Fifteen glomeruli were analyzed per section. The number of glomerular cells was determined on PAS stains by counting cell nuclei in 15 glomerular tufts per section. Pole-cut glomerular cross sections were ignored. Immunostaining was performed, as described,20 using the following primary antibodies: CD45 (leukocytes; BD Biosciences, Mannheim; 1:10), Wilms tumor (WT) = 1 (podocytes; Santa Cruz, Santa Cruz, CA; 1:200), Ki-67 (proliferating cells; DiaNova, Hamburg, Germany; 1:50), terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling (TUNEL; apoptotic cells; Roche, Mannheim; 1:10), TUNEL- and DAPI-positive cells (Sigma-Aldrich, Steinheim, Germany) were counted by fluorescent microscopy, and all other stains were assessed by light microscopy in 15 glomeruli per section by a blinded observer (H.J.M.).

In Vitro Studies

Normal renal tissues were obtained from four patients who underwent nephrectomy because of renal cell carcinoma, in agreement with the Ethical Committee on Human Experimentation of the Azienda Ospedaliero–Università Car-eggii, Florence, Italy. Human embryonic kidney 293 cells were purchased from American Type Culture Collection (Manassas, VA) and cultured as specified by the supplier. The human keratinocyte cell line was a gift from Dr. Rosa Mancina (Florence, Italy). Human renal progenitors were obtained by immunomagnetic sorting for CD45 after depletion of digested total renal cell suspensions for CD45 and podocalyxin, as previously described.18 Primary cultures of human renal progenitors were >99% positive for CD133 and CD24 coexpression, as checked by fluorescence-activated cell sorter analysis. Podocyte markers were absent. For podocyte differentiation, cells were treated for 2 days with VRAD medium, as previously described.18,21 Nephrin mRNA quantification was performed using Assay on Demand kits (Applied Biosystems, Warrington, UK),20,21 according to the manufacturer’s recommendations; and Taq-Man RT-PCR was performed using a 7900HT Real Time PCR System (Applied Biosystems), as previously described.20,21 To accurately compare samples by real-time RT-PCR, an identical number of cells (n = 10,000) was analyzed in all experiments; and glyceraldehyde-3-phosphate dehydrogenase was used to evaluate mRNA quality and integrity.

RNA Preparation and Real-Time Quantitative (TaqMan) RT-PCR

Total RNA was isolated from kidneys using a Qiagen RNA extraction kit (Hilden, Germany) following the manufacturer’s instructions. After quantification, RNA quality was assessed using agarose gels. From isolated RNA, cDNA was prepared using reverse transcriptase (Superscript II; Invitrogen, Carlsbad, CA). Real-time PCR was performed using SYBR Green PCR master mix, and results were analyzed with a Light Cycler 480 (Roche). All gene expression values were normalized using 18s RNA as a housekeeping gene. All primers used for amplification were from Metabion (Martin-ried, Germany): CCL2 (sense, 5’-CCTGCTGTTCCAGT- TGCC-3’; antisense, 5’-ATTGGGATCATCTTTGCTGTG-3’), CXCL12 (sense, 5’-TTTCAATGCGTGACCTGG-3’; anti-sense, 5’-GCCGCCTGTCCAATGAC-3’), tumor necrosis factor (sense, 5’-AGGGTCTCGGCCCATAGA-3’; anti-sense, 5’-CCACCCACGCTTCTGTGCTAC-3’), inducible nitric oxide synthase (sense, 5’-TAAAGAAAAACCCCTTGTTGCT-3’; antisense, 5’-TTTCTGTGCTGCCTCCAGTGAG-3’), nephrin (sense, 5’-CTCTTTTCTACGCTCAACG-3’; anti-sense, 5’-TACGACACACGGAGACAGG-3’), podocin (sense, 5’-CAGGAAAGAGATGTCCTAG-3’; antisense, 5’-TGACGCTTTCTTTTTCATC-3’), IL-6 (sense, 5’-ACCA- GAGGAATTTTTCAATTGGC-3’; antisense, 5’-TGATGCTTCTGGCAAGAAC-3’), and 18s RNA (sense, 5’-AGGG- CCTCACTAAACCATC-3’; antisense, 5’-GCAATTATTC- CCCATGAACG-3’).

Statistical Analysis

Data are presented as mean ± SEM. A comparison of groups was performed using an analysis of variance, and post hoc Bonferroni’s correction was used for multiple comparisons. P < 0.05 indicated statistical significance.

Results

Dual CCL2-CXCL12 Blockade Has Additive Effects on Glomerulosclerosis in db/db Mice

To test our hypothesis, we initiated s.c. injections with vehicle alone, control Spiegelmer, mNOX-E36 (anti-CCL2 Spiegelmer), NOX-A12 (anti-CXCL12 Spiegelmer), or a combination of the latter two at a dose of 50 mg/kg given every other day from the age of 4 to 6 months in uninephrectomized db/db mice. To ensure Spiegelmer exposure and its biological activity in vivo, we determined plasma CCL2 and CXCL12 levels at the beginning and the end of the study. Plasma CCL2 and CXCL12 levels remained undetectable or low in sham–operated on db/db mice and vehicle- or control Spiegelmer–treated 1K db/db mice. Anti-CXCL12 or anti-CCL2 Spiegelmer injections increased the plasma levels of either chemokine in 1K db/db mice (Figure 1). This finding is consistent with previous observations.13,15,17 with these Spiegelmers, indicating that Spiegelmer antagonists retain their molecular targets in the circulation. Such Spiegelmer-chemokine complexes no longer display chemotactic activity in trans-well migration assays (data not shown). Body weight and blood glucose levels did not differ among the injected 1K db/db mice groups (Figure 2). Renal histomorphological features in 6-month-old db/db mice showed moderate glomerulosclerosis compared
with age-matched nondiabetic mice that underwent sham surgery, which was aggravated to diffuse glomerulosclerosis by early uninephrectomy of db/db mice (Figure 3). CCL2 and CXCL12 inhibition reduced the extent of glomerulosclerosis in uninephrectomized db/db mice to the level of age-matched sham–operated on db/db mice, whereas the control Spiegelmer had no effect. Dual CCL2-CXCL12 blockade further improved glomerular pathological features, with significantly less severe lesions and more normal glomeruli compared with either of the monotherapies. 1K db/db mice with dual blockade displayed even less glomerular pathological features than age-matched db/db mice that underwent sham surgery (Figure 3B). Thus, dual CCL2-CXCL12 blockade has additive protective effects on glomerulosclerosis in 1K db/db mice.

**Only CCL2 Blockade Reduces Leukocyte Numbers in Glomeruli of 1K db/db Mice**

Chemokine-mediated glomerular pathological features in db/db mice can be mediated by leukocyte recruitment. Therefore, we evaluated the number of glomerular leukocytes by immunostaining for CD45. Significant numbers of CD45+ leukocytes were detected in glomeruli of vehicle- or control Spiegelmer–treated 1K db/db mice (Figure 4). Consistent with previous data, the numbers of glomerular CD45+ leukocytes were significantly reduced by CCL2, but not by CXCL12, blockade. The numbers of glomerular leukocytes were also reduced accordingly with dual-chemokine blockade (Figure 4). Therefore, blockade of the proinflammatory chemokine CCL2, but not of the homeostatic chemokine CXCL12, affects the number of glomerular leukocytes in db/db mice.

**Dual CCL2-CXCL12 Blockade Has Additive Effects on Podocyte Numbers in db/db Mice**

Previously, CXCL12 blockade protected 1K db/db mice from podocyte loss as another marker of glomerular pathological features in db/db mice. We questioned whether dual-chemokine blockade would have additive protective effects on podocytes and quantified glomerular podocytes by WT-1 immunostaining in all groups of 1K db/db mice. Compared with an average number of 15 to 20 WT-1–positive podocytes in murine glomerular cross sections (data not shown), the 6-month-old 1K db/db mice revealed only an average of 11 cells per glomerular cross section. CCL2 and, particularly, CXCL12 blockade both significantly increased glomerular podocyte counts (Figure 5). Interestingly, dual CCL2-CXCL12 blockade showed a small, but statistically significant, additive effect up to an average of 17 WT-1–positive cells per glomerular cross section (Figure 5). Thus, dual CCL2-CXCL12 inhibition improves glomerular pathological features in association with more podocytes in db/db mice.

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**Figure 1.** Plasma chemokine levels in Spiegelmer-treated db/db mice. Plasma CCL2 and CXCL12 levels were determined in 14- and 24-week-old uninephrectomized db/db mice by enzyme-linked immunosorbent assay, which represents before and after treatment. Data are given as mean ± SEM. *P < 0.001 versus the 24-week vehicle group.

**Figure 2.** Blood glucose levels and body weight in uninephrectomized db/db mice. Uninephrectomized db/db mice with different treatments were monitored for blood glucose levels (A) and body weight (B) from baseline (at the age of 16 weeks) until the end of the study (at the age of 24 weeks). Data are given as the mean ± SEM. n.s. indicates nonsignificant.
Dual CCL2-CXCL12 Blockade and Renal mRNA Expression in db/db Mice

Next, we determined the mRNA expression levels of leukocyte- and podocyte-related genes in kidneys of 1K db/db mice. Both antagonists did not significantly alter intrarenal expression of their target gene (Figure 6). However, CCL2 blockade, but not CXCL12 blockade, was associated with lower mRNA levels of inducible nitric oxide synthase, a parameter of activated macrophages (Figure 6). The cytokine IL-6 showed a trend toward lower levels in all treatment groups. In contrast, CXCL12 blockade increased the mRNA levels of nephrin and podocin, two podocyte parameters, much more than CCL2 blockade. Dual-chemokine blockade increased these two podocyte markers most significantly (Figure 6). Together, CCL2 and CXCL12 single blockade has different effects on renal mRNA expression profiles; both effects combine with dual blockade.

Dual CCL2-CXCL12 Blockade and Glomerular Cell Counts in db/db Mice

Glomerular pathological features are often related to an altered cell turnover of glomerular cells. Therefore, we determined the total numbers of glomerular cells, Ki-67+ proliferating glomerular cells, and TUNEL+ apoptotic glomerular cells. Total numbers of glomerular cells were determined by counting DAPI-positive nuclei in glomerular tuft cross sections. None of the interventions significantly affect the total number of cell nuclei in glomerular tufts (Figure 7A). The overall numbers of Ki-67+ proliferating cells within glomeruli were low, but CCL2 and CXCL12 blockade both significantly reduced their numbers in glomeruli (Figure 7B). Dual CCL2-CXCL12 blockade had an additive suppressive effect on glomerular cell proliferation (Figure 7B). TUNEL+ apoptotic cells were only occasionally seen and did not differ between all treatment groups (data not shown). We conclude that dual-chemokine blockade has an additive antiproliferative effect on glomerular cells in type 2 diabetic mice; however, given the opposite effects (eg, on glomerular podocyte counts), the total number of glomerular cells was not significantly affected.
Dual CCL2-CXCL12 Blockade Increases GFR and Reduces Proteinuria in db/db Mice

We determined the GFR by assessing fluorescein isothiocyanate insulin clearance kinetics in all groups of mice. Uninephrectomy was associated with a reduced GFR compared with a normal GFR of approximately 350 mL in mice. Either CCL2 or CXCL12 blockade significantly increased GFR in 6-month-old 1K db/db mice (Figure 8A). Dual CCL2-CXCL12 blockade was associated with the highest GFR (Figure 8A). Albuminuria is another clinically important parameter of glomerular dysfunction and damage. We compared urinary albumin/creatinine ratios at the beginning and at the end of the treatment course in all groups. CCL2 and CXCL12 significantly reduced the urinary albumin/creatinine ratio when compared with control Spiegelmer treatment at 6 months (Figure 8B). When compared with the baseline urinary albumin/creatinine ratio, CXCL12 and dual blockade most effectively prevented proteinuria. Therefore, we conclude that dual CCL2-CXCL12 blockade has a potent protective effect on GFR decline. CXCL12 blockade, either alone or in combination, effectively prevents proteinuria in 1K db/db mice, which is consistent with its positive effect on podocytes.

CXCL12 Suppresses Nephrin de Novo Expression of Podocyte Progenitors

How does CXCL12 blockade significantly increase podocyte numbers and nephrin expression without any direct effect on podocytes, which we previously excluded?15

Given the critical role of CXCL12 in the maintenance of...
stem cell niches in multiorgan systems, we evaluated the possibility that podocyte-derived CXCL12 may regulate the potential of renal progenitor cells to differentiate toward podocytes. Several groups\textsuperscript{15–19} have recently reported that a subpopulation of parietal epithelial cells in glomeruli represents progenitor cells and has the capacity to differentiate into mature podocytes by progressively migrating and differentiating from the urinary pole of the Bowman’s capsule toward the glomerular tuft. Therefore, we questioned whether CXCL12 serves as a factor that regulates the differentiation of renal progenitor cells toward the podocyte phenotype. A previous report\textsuperscript{15} indicated that culturing human renal progenitors in the VRAD medium over 2 days resulted in their differentiation into podocytes, as demonstrated by novel expression of nephrin, WT-1, synaptopodin, podocin, and anti-glomerular epithelial protein 1 at both the mRNA and protein levels. Because CXCL12 blockade up-regulated nephrin expression in our \textit{in vivo} models, in this study, we evaluated primary cultures of human renal progenitors that are already known to express receptors for CXCL12\textsuperscript{15} for their potential to up-regulate nephrin mRNA expression in the presence or absence of this chemokine. The human embryonic kidney epithelial cell line 293 and the human keratinocyte cell line were evaluated as additional controls for nephrin mRNA expression before and after treatment with VRAD. As shown in Figure 9A, the effect of VRAD treatment was irrelevant in human embryonic kidney 293 and human keratinocyte cell lines, whereas it induced strong up-regulation of nephrin mRNA expression in primary cultures of human renal progenitors, as assessed by quantitative RT-PCR. Adding recombinant CXCL12 suppressed nephrin mRNA up-regulation in a dose-dependent manner (Figure 9A). When CXCL12 was blocked with anti-CXCL12 Spiegelmer, \textit{de novo} mRNA expression of nephrin was no longer compromised, an effect that was not observed with the unspecific control Spiegelmer (Figure 9B). Thus, CXCL12 suppresses the potential of renal progenitors to express nephrin. Vice versa, CXCL12 antagonism increased levels of nephrin mRNA expression, which may explain the findings on CXCL12 blockade in db/db mice.

\textbf{Discussion}

We hypothesized that dual blockade of CCL2 and CXCL12 was superior compared with single blockade in preventing the progression of diabetic glomerulosclerosis because both approaches target different disease pathomechanisms. Our data support this hypothesis and demonstrate that dual blockade combines the protective effect on CCL2-mediated glomerular leukocyte recruitment with that on CXCL12-mediated loss of podocytes. Thus, our data render dual CCL2-CXCL12 blockade as a novel strategy to more efficiently prevent glomerulosclerosis in type 2 diabetes.

Data of the present study confirm previous reports\textsuperscript{13,15} on CCL2 blockade with Spiegelmers in the same experimental setup. Attempts to increase the therapeutic benefit by initiating treatment earlier were not successful.\textsuperscript{19} In contrast, even a later onset of anti-CCL2 Spiegelmer treatment (from the age of 5 months) was equally effective in preventing glomerulosclerosis in 1K db/db mice.\textsuperscript{19} In the present and in previous studies, CCL2 blockade...
resulted in a 50% to 60% reduction of glomerular leukocytes that entirely represent macrophages in the db/db mouse model. These findings are consistent with data from models of type 1 and 2 diabetes in Ccl2-deficient mice or with CCR2 blockade in 1K db/db mice. CCR2+ macrophages belong to the proinflammatory (M1) phenotype that contributes to intrarenal inflammation and tissue damage via the release of reactive oxygen species or cytokines, such as tumor necrosis factor-α. Thus, anti–CCL2-Spiegelmer–treated 1K db/db mice revealed fewer glomerular leukocytes that were associated with lower intrarenal mRNA levels of inducible nitric oxide synthase, IL-6, or tumor necrosis factor-α at the end of the study. Consistent with previous findings, the protection from glomerulosclerosis with CCL2 blockade correlated with some lower levels of proteinuria and a higher GFR. Thus, CCL2 blockade prevents the progression of DN by interfering with macrophage-driven glomerular inflammation.

The present study also reconfirms previous data on CXCL12 blockade in this model. NOX-A12 prevented the progression of glomerulosclerosis in 1K db/db mice without affecting glomerular leukocyte counts, indicating that the homeostatic chemokine CXCL12 is not involved in glomerular leukocyte recruitment. Recently, CXCL12 was constitutively expressed in podocytes. CXCL12 blockade also increased podocyte numbers in 1K db/db mice in the present study. Furthermore, CXCL12 blockade increased renal mRNA expression levels of nephrin and podocin, two podocyte-slit membrane-related proteins that serve as markers of podocyte differentiation. Podocyte damage is an important pathomechanism promoting the progression of diabetic glomerulosclerosis and seems to result from various triggers that foster podocyte dedifferentiation and podocyte apoptosis. It remains unclear whether the beneficial effect of CXCL12 blockade on podocyte numbers relates to a protective effect on podocyte death or detachment or is the result of enhanced podocyte regeneration from local podocyte progenitors (eg, from parietal epithelial cells). The latter concept may be more likely because CXCL12 blockade also mobilizes hematopoietic progenitor cells from the bone marrow. In fact, our in vitro studies demonstrate that CXCL12 suppressed the induced maturation of renal progenitors toward podocytes, as documented by inhibition of de novo nephrin expression. This effect was specific to renal progenitors and was not observed in other epithelial cells. Our in vitro finding that CXCL12 blockade reverses this inhibitory effect on nephrin expression may correspond to our in vivo observation that CXCL12 blockade increased renal nephrin mRNA expression and podocyte numbers. Thus, CXCL12 blockade may enhance renal progenitor differentiation toward the podocyte lineage, thus enhancing podocyte regeneration.

Thus, one hypothesis is that dual blockade with anti–CCL2- and anti–CXCL12-Spiegelmers could lead to Spiegelmer-Spiegelmer interaction effects, such as complex formation or additive off-target effects. However, the plasma chemokine level analysis confirmed that dual blockade does not significantly affect the biological activity of each Spiegelmer in terms of binding its natural target (in the plasma). This observation is consistent with the finding that 1K db/db mice with dual blockade recapitulate the biological effects of single blockade. For example, 1K db/db mice treated with dual blockade displayed the reduction of glomerular leukocytes and a trend toward lower inducible nitric oxide synthase and tumor necrosis factor-α expression to the same extent as seen with CCL2 blockade. In addition, dual blockade also increased podocyte numbers and nephrin or podocin mRNA expression to the same extent as seen with CXCL12 blockade only. As a result, the combination of these different ways dual blockade was more potent than single blockade in preventing diffuse glomerulosclerosis was evidenced by significantly fewer glomeruli with global glomerulosclerosis and significantly more normal glomeruli. This additive therapeutic effect was less prominent for GFR and proteinuria because CXCL12 blockade by itself already had a profound effect on these two functional end points. Together, these data first document that chemokine antagonist combinations hold a potential for additive preventive effects on (diabetic) glomerulosclerosis when the individual chemokine targets mediate different pathomechanisms in the specific disease process (ie, inflammation and renal progenitor differentiation toward the podocyte lineage).

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References


