

## Introduction

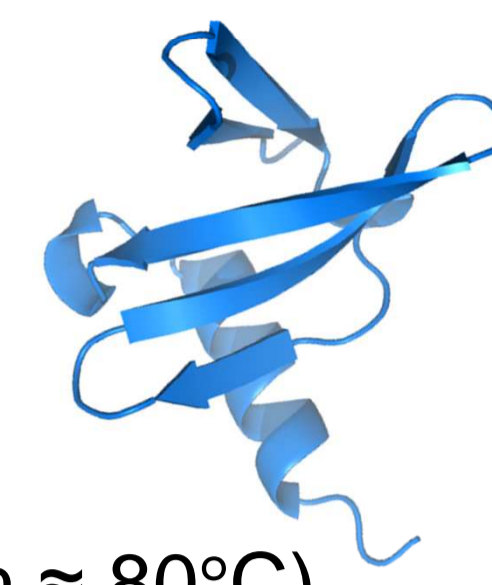
The formation of a C- $^{18}\text{F}$  bond requiring hard conditions, an alternative method developed few decades ago consists in incorporating the  $^{18}\text{F}$  on a prosthetic group and then coupling it to the biomolecule by bioorthogonal reactions.[1-2] Following a similar strategy, a single cysteine has been incorporated in a Nanofitin(NF) model and its regioselective radiolabeling has been performed with  $[^{18}\text{F}]$ -4-fluorobenzamido-N-ethylamino-maleimide ( $[^{18}\text{F}]$ -FBEM). Coupling with the  $[^{18}\text{F}]$ -FBEM has been achieved on Ni magnetic beads system. PET-MRI studies on mice were conducted after injection of  $[^{18}\text{F}]$ -FBEM-Cys-NF in order to know the biodistribution of this NF model.



Figure 1 Scheme of radiosynthesis of  $[^{18}\text{F}]$ -FBEM-Cys-NF

## Nanofitins

- Small protein (+/-10kDa)
- pH stability (0-13)
- Temperature stability ( $T_m \approx 80^\circ\text{C}$ )
- Produced by Sac7D bacteria
- Alternative to antibodies
- Cysteine-free protein



## Methods

We first synthesized the labeling precursor **1** (Figure 2) as previously reported in literature.[3] The three-step synthetic pathway synthesis was implemented on a GE Healthcare FastLab<sup>TM</sup>.

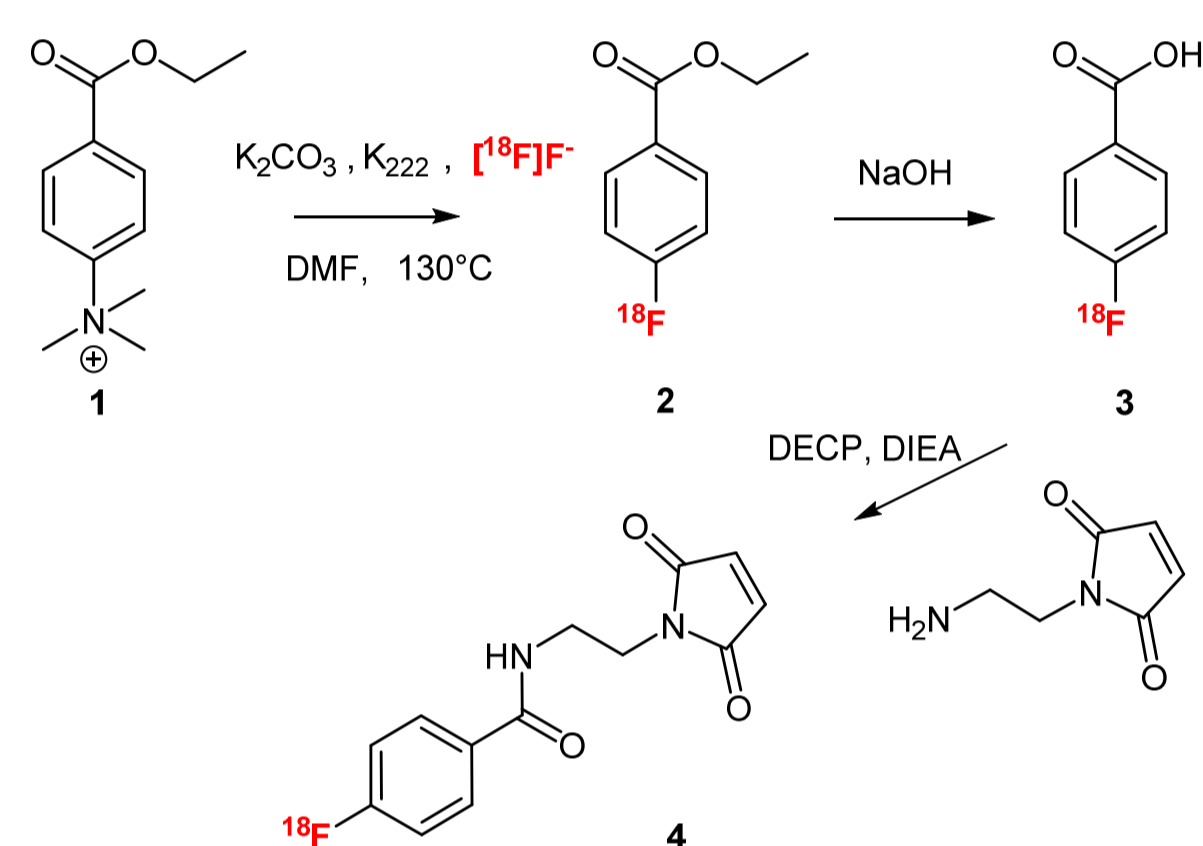


Figure 2 Radiosynthesis of  $[^{18}\text{F}]$ -FBEM

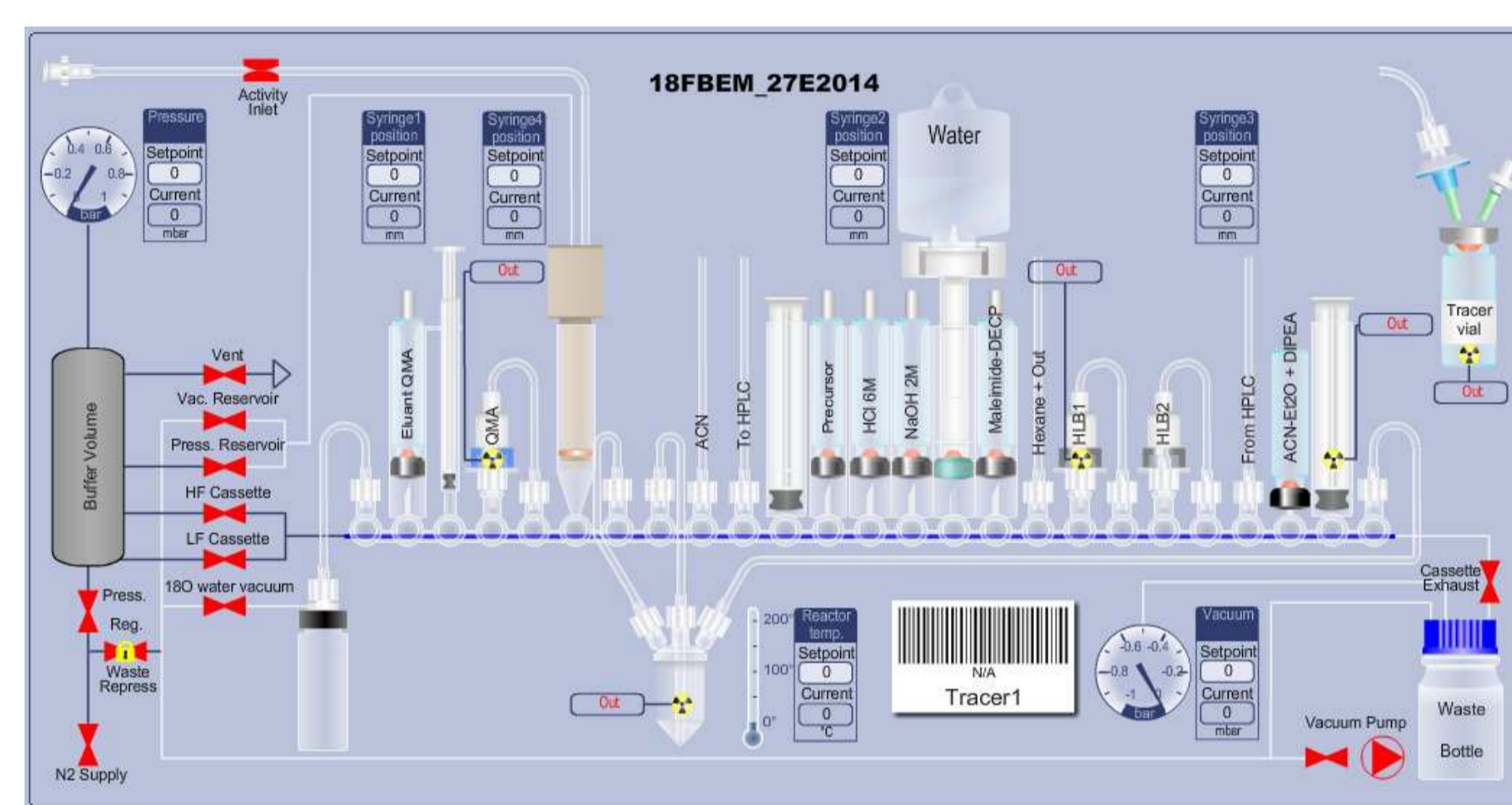


Figure 3 Cassette layout for  $[^{18}\text{F}]$ -FBEM preparation

The labeling was then done on a selected model histagged NF that has been reengineered to bear a unique cysteine. This cysteine leads to the dimerization of the NF via a disulfide bridge which needs to be reduced before the coupling. To overcome the reduction of the maleimide moiety of the  $[^{18}\text{F}]$ -FBEM in presence of reducing agent, the NF was first trapped on Ni-NTA magnetic beads and then reduction and labeling were proceeded successively on-bead (Figure 4).

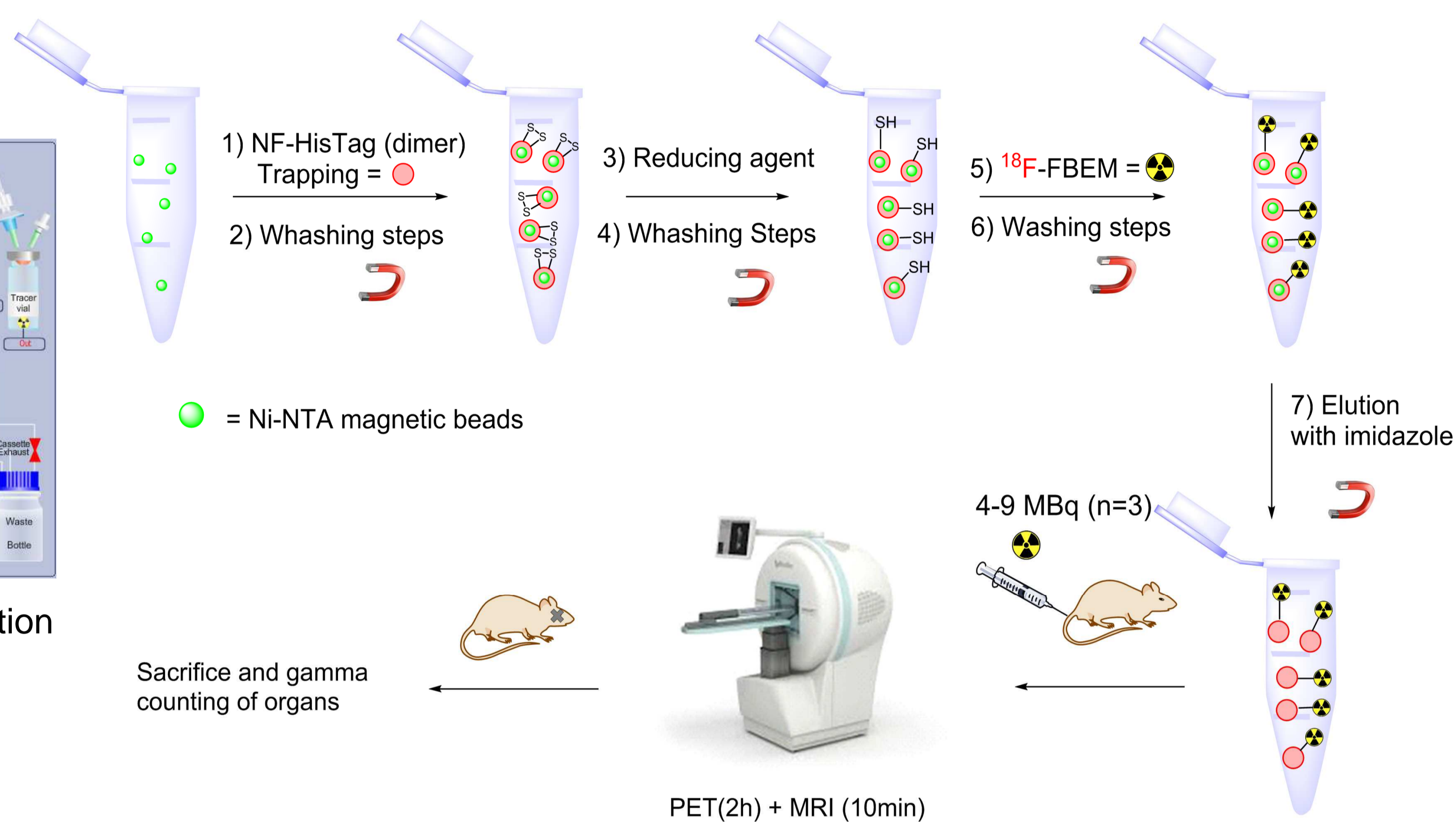


Figure 4 Radiolabeling of Nanofitin with  $[^{18}\text{F}]$ -FBEM on Ni-NTA magnetic beads

## Results

Radiochemical yield(RCY) of  $[^{18}\text{F}]$ -FBEM synthesis was  $28.7 \pm 3.2\%$  ( $n=10$ ) and lasted 90 min including purification.

The RCY obtained for NF labeling is  $53.8 \pm 7.3\%$  ( $n=9$ ) with a radiochemical purity  $>95\%$ .

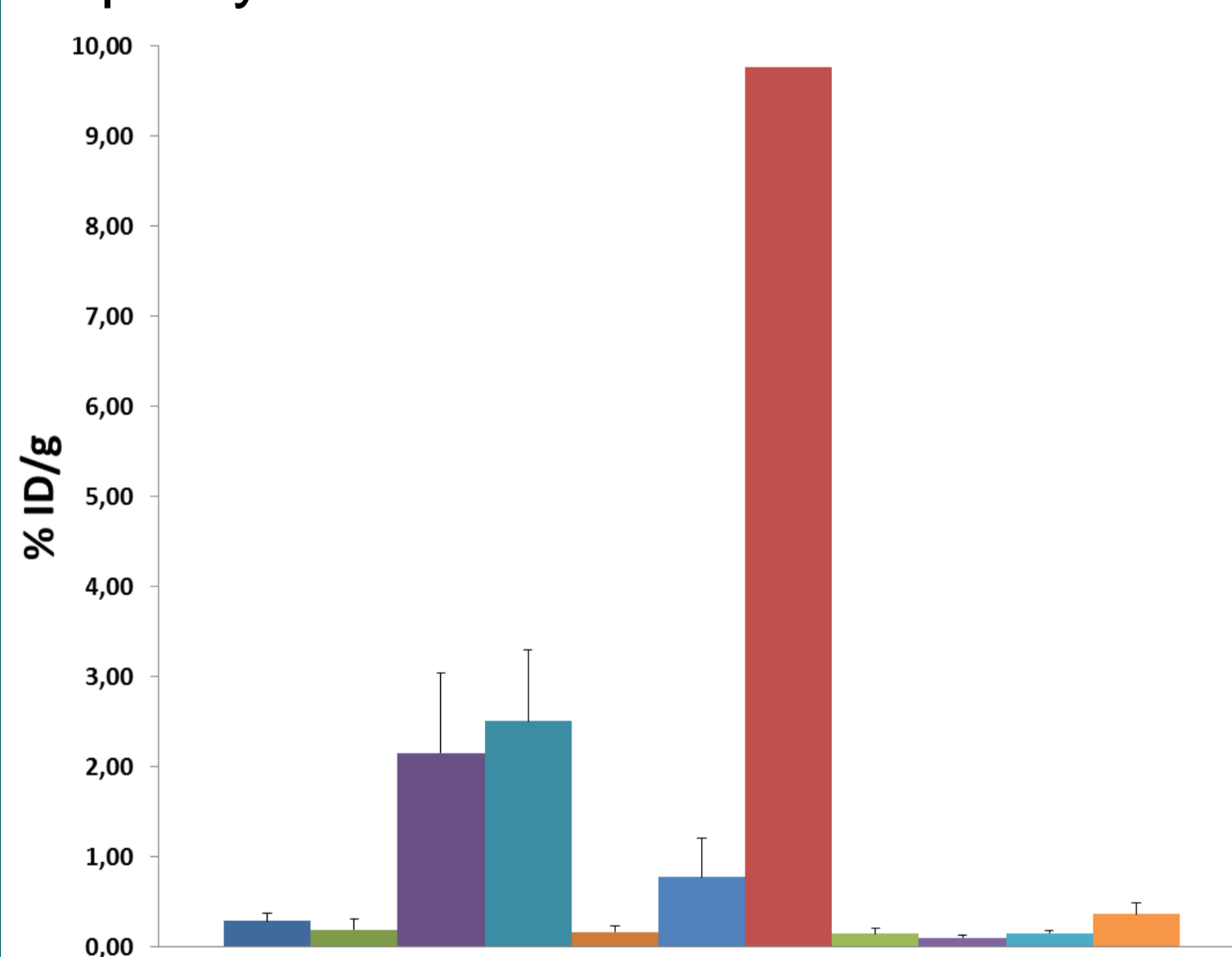


Figure 6 %ID/g for each organ 3h after injection

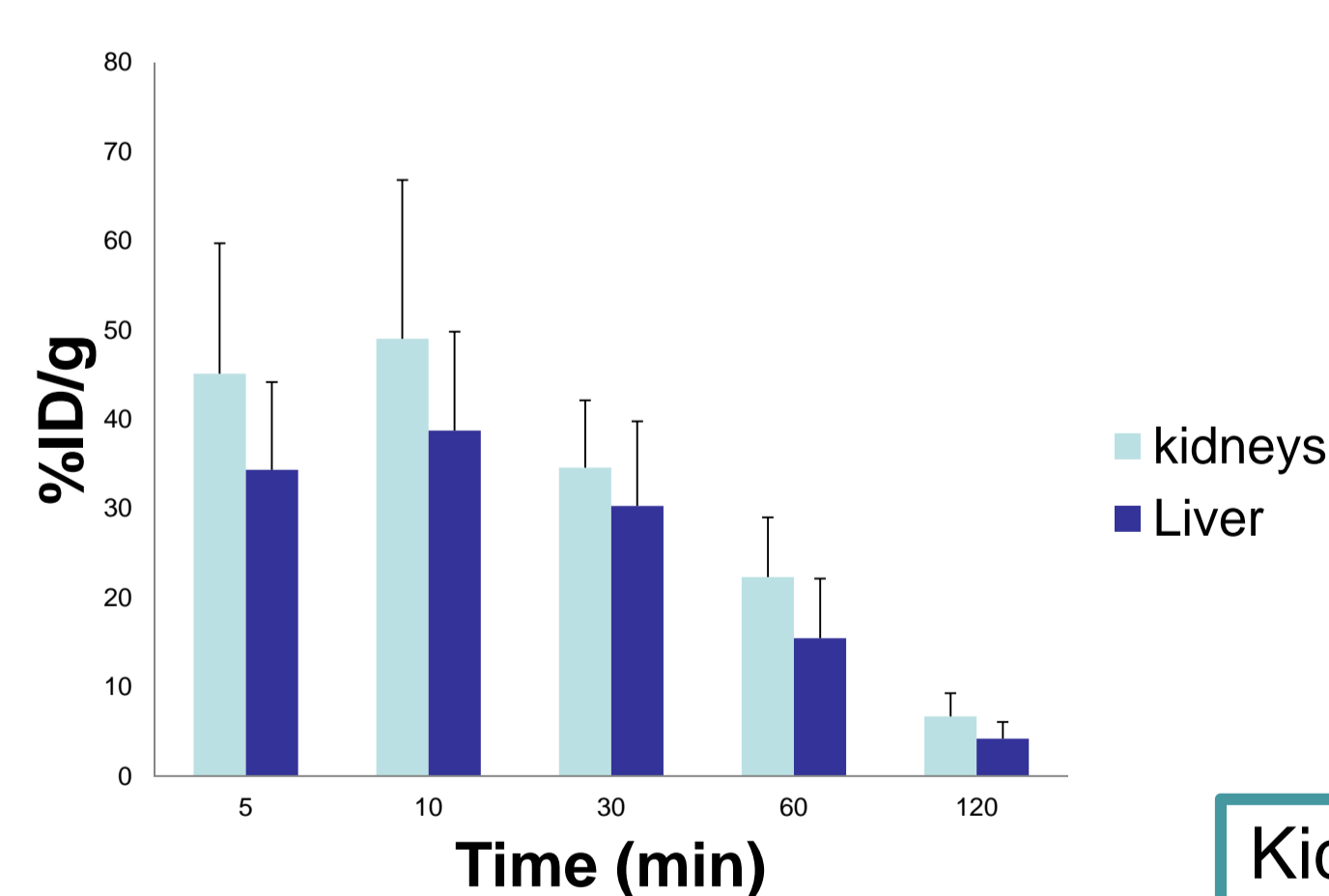


Figure 7 Evolution of the %ID/g for the kidneys and the liver

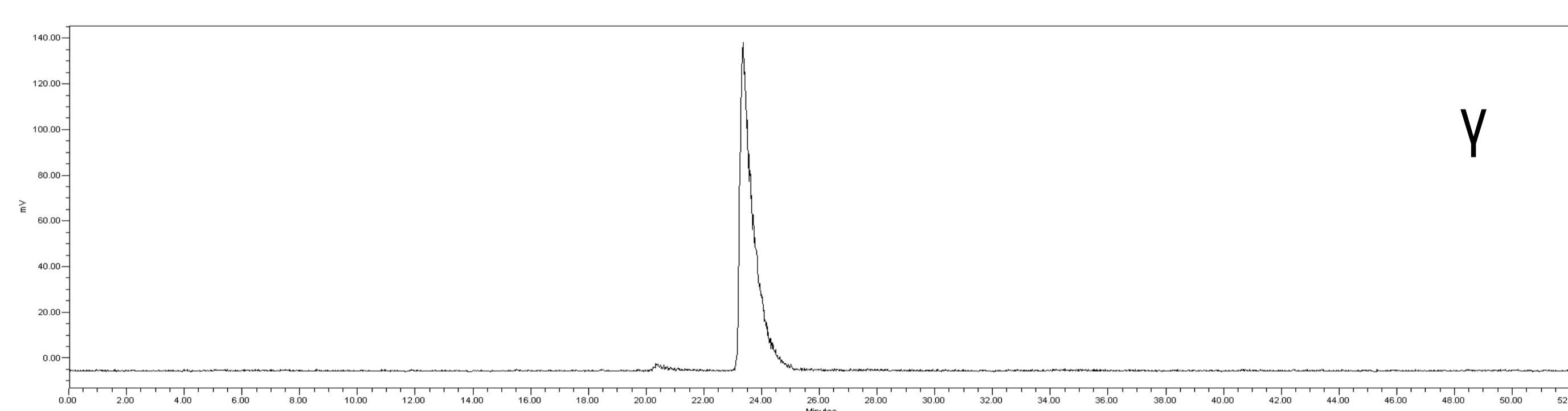


Figure 5 Analytical C8-HPLC of the  $[^{18}\text{F}]$ -NF

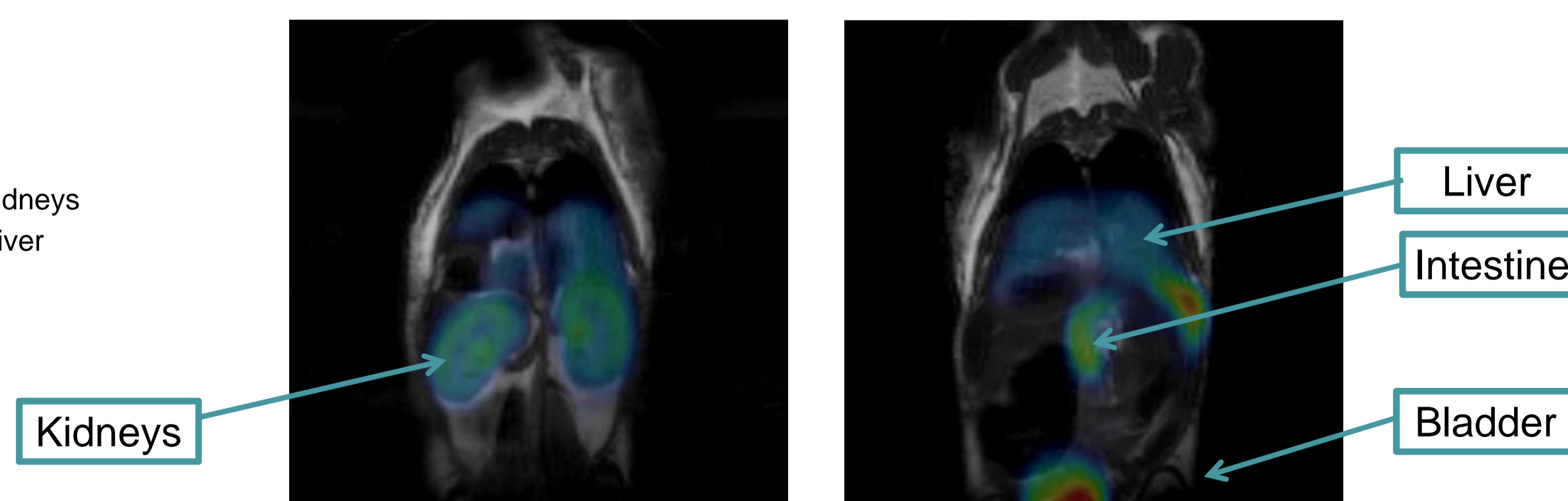


Figure 8 PET-MRI images of the biodistribution of the NF

## Conclusions

We successfully radiolabeled a Cys-tagged Nanofitin with  $[^{18}\text{F}]$ -FBEM automated synthesized at high activity. A simple and new method for the radiolabeling of biomolecule with high yield (up to 61%) on Ni magnetic beads was developed. PET-MRI studies on mice were conducted after injection of  $[^{18}\text{F}]$ -NF in order to know the biodistribution of this Nanofitin model. The results show a fast renal clearance and an uptake in the liver. The metabolites are excreted by kidneys through urine and by the liver via biliary excretion to the intestine.

## References

- [1] Kiesewetter D.O. et al., *Eur J Nucl Med Mol Imaging*. **2012**, 39(2), 300–308
- [2] Cheng Z. et al., *J Nucl Med*. **2012**, 53, 1110-1118
- [3] Garg, S. et al., *Bioconjugate Chemistry* **2009**, 20, 583–590

## Acknowledgements

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