

# Validated GC-MS/MS confirmatory method for the EU official control of levels of PCDD/Fs and DL-PCBs in feed material of plant origin

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## Context

### Introduction and strategy

Criteria for sampling and analysis for the official control of dioxins (PCDD and PCDF) and dioxin-like (DL) PCB in feeding stuffs and certain foodstuffs are described in Commission Regulation (EU) No 709/2014 and No 589/2014. They allow the use of GC-QQQ as confirmatory method in addition to GC-HRMS.

We present a full validated method using the Agilent GC-QQQ 7000C instrument for the analysis of PCDD/Fs and DL-PCBs in vegetable oil (feed). We assessed individual analytical criteria specified in the above documents and checked that they meet the requirements. In this study we preferred observing performances of the QQQ (and their compliance with the Regulation), starting from basics, rather than simply comparing duplicated results on QQQ and HRMS. We therefore compiled results arising from different criteria and finally assessed the measurement uncertainty based on those.

### Instrumentation & parameters

GC: Agilent 7890B GC equipped with a PTV injector and 7693A automated liquid sampler (ALS).

Column: DB-5ms 60m x 250µm x 0.25µm

MS: Agilent 7000B series GC-QQQ with 7000C electron ionization (EI) source; ion source T=280° C; quads T=150° C; N<sub>2</sub> collision flow=1.5mL/min; He quench flow=2.25mL/min

Oven T program: 120° C (5min); 25° C/min until 250° C (5min); 3° C/min until 285° C (15min).

PTV: solvent vent mode; start at 40° C (3min) and ramp at 720° C/min until 320° C; vent flow=50mL/min (P=5psi) until 2.8min; purge flow=50mL/min at 5min.

## Results and validation

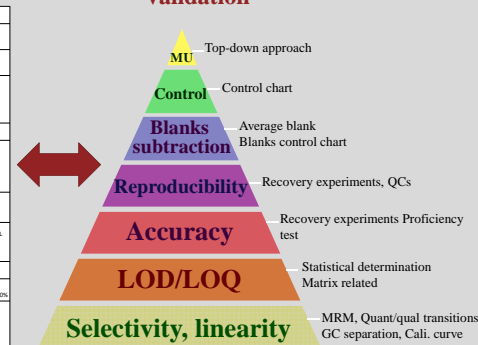
### Targets, sample preparation and method of analysis

29 compounds were investigated including 7 PCDDs, 10 PCDFs, 4 'non-ortho' (NO) PCBs, and 8 'mono-ortho' (MO) PCBs. Pure vegetable oil (sunflower oil) was used as validation matrix. The clean-up is carried out on a Powerprep system using classical column set: mixed bed silica, alumina and carbon. Two fractions are collected from the carbon column. Fraction A, eluted with hexane/Dichloromethane, contains MO-PCBs and fraction B, eluted with toluene, contains PCDD/Fs and NO-PCBs. All analytes are quantified by isotopic dilution against their own <sup>13</sup>C labeled standard, spiked before clean-up. Recovery (syringe) standards are spiked before injection and consist in <sup>13</sup>C<sub>6</sub>-1,2,3,4-TCDD (for tetra/penta dioxins and furans), <sup>13</sup>C<sub>12</sub>-1,2,3,4,7,8,9-HpCDF (for hexa/hepta/octa dioxins and furans), and <sup>13</sup>C<sub>12</sub>-PCB80 (for PCBs). Each compound is defined by a quantifier and a qualifier MRM transition whose collision energy (CE) has been optimized. Recovery experiments for accuracy and reproducibility tests are performed using fortified (with all congeners) sunflower oil.

### Criteria in Regulation

Criteria	PCDD/Fs and DL-PCBs (GC-MS/MS/MS/MS)	NO-PCBs (GC-MS/MS/MS/MS)
Detectable quantity	PCDD/F upper limit (µg/kg) (10 <sup>-12</sup> ) MO-PCBs lower limit (µg/kg) (10 <sup>-12</sup> ) MO-PCBs upper limit (µg/kg) (10 <sup>-12</sup> )	NO-PCBs range (µg/kg)
Selectivity	Chromatographic separation of: 2,3,4,7,8-HxCDF and 2,3,4,7,8-HxCDF 2,3,4,7,8-HxCDF and 2,3,4,7,8-HxCDF	Relative RT ±0.2% (5 vs analyte)
MRM transitions	Monitoring 2 specific precursors with each specific product ion transition for all labeled and unlabeled analytes Relative ion intensities max. ±15% Resolution MS/MS (qualifier vs quant)	Monitoring at least 2 precursor ions and 2 product ions Tolerance ratio ±10% of rel. intens. >50% Tolerance ratio ±25% of rel. intens. 20-50% Resolution MS/MS (qualifier vs quant)
Blank	Used for LOD calculation	Used for LOD calculation Blank value = 50% of maximum level ML
iLOQ	iLOQ calculated from lowest cali. point (lowest concentration point on call. must give acceptable and consistent deviation to the average RRF) Average RRF calculated for all points Deviation to average RRF < 10%	ditto
LOQ	LOQ calculated from average blank level LOQ < 1% of maximum level ML Off-line up and 8h level < 200% ML	ditto Off-line up and 8h level < 20% ML
Accuracy	Demonstrate performance at 0.5ML, ML, 2ML Precision (recovery) ± 20% Within-lab reproducibility (RSD) < 15%	Demonstrate performance at 0.5ML, ML, 2ML Precision for spiked PCB (RSD) < 20% Within-lab reproducibility (RSD) < 20%
Control	QC chart for blanks QC chart control sample Individual internal std in range 60-120% Out of range if < 60% or > 120%	QC chart for blanks QC chart control sample Individual internal std in range 50-120% Out of range if < 50% or > 120%
Recovery	Expanded measurement uncertainty Coverage factor = 2 (k=95%) If user data dispersion of congeners, make sum of separate uncertainty for each of PCDD/F and DL-PCBs.	Expanded measurement uncertainty Coverage factor = 2 (k=95%)
Measurement uncertainty		Expanded measurement uncertainty Coverage factor = 2 (k=95%) GC separation, Cali. curve

### Validation



### Selectivity, linearity

Control of 3 criteria to be verified during analysis: 1) retention time (RT) of targets must be within a +3s window from the internal standard. 2) MRM transition ratio (quant/qual), determined experimentally from standard injections, must be within the ±15% tolerance window. 3) Separation valley between HxCDF congeners must be <25% of peak height. Linearity is controlled from standards and is acceptable when calibration curve (built using average response factors from 18 points) correlation coefficient (R<sup>2</sup>) is >0.9900. Example of control is given in Fig. 1.

### Limits of detection and quantitation

Instrumental limits of quantitation (iLOQ) must be calculated in a different way for GC-MS/MS. Unlike for GC-HRMS, signal-to-noise (S/N) ratio is not suitable to identify limits since it gives unrealistic values due to filtration of ions. We define the iLOQ, a 'performance LOQ' from the standard deviation associated to replicate injections of the lowest calibration point. The 'real LOQ' used in upperbound results is defined using replicate independent procedure blanks injections and is representative of the environment and sample preparation.

$$\text{Performance-iLOQ} = 10 \cdot \text{stdev} \text{ (8 replicate injections of lowest cali. point)}$$

$$\text{Real-LOQ} = \text{blank mean} + 6 \cdot \text{stdev} \text{ (12 distinct blanks)}$$

LOQ's vary from 0.02 pg/g fat for 2,3,7,8-TCDD to 49.66 pg/g fat for PCB77 with a median of 0.10 pg/g fat for all congeners. The GC-HRMS method provides similar LOQ's in the range 0.06-64.59 pg/g fat with a median of 0.10 respectively.

### Accuracy and reproducibility

Six series of spiked materials at 0.5 maximum level (ML), ML, and 2ML were injected over 3 days (table 1). Bias and within lab reproducibility (RSD) are respectively <20% and <15% as required in the Regulation. Accuracy was also tested during proficiency test (PT) on vegetable oil (Rikilt, 2013) (Table 2).

Table 1: results for injections of 6 series of fortified vegetable oil in 3 days (2 series injected per day)

PCDD/Fs	Average	Stdev	RSD	Target	Bias
	ng WHO-TEQ/g	ng WHO-TEQ/g	%	ng WHO-TEQ/g	%
Spike level MU2	0.609	0.029	7.1	0.49	2.36
ML	0.778	0.045	5.7	0.79	-1.54
2ML	1.600	0.035	2.2	1.58	1.30
NO-PCBs	Average	Stdev	RSD	Target	Bias
	ng WHO-TEQ/g	ng WHO-TEQ/g	%	ng WHO-TEQ/g	%
Spike level MU2	0.307	0.028	9.0	0.33	-7.00
ML	0.355	0.020	3.4	0.65	-45.3
2ML	1.256	0.021	1.6	1.30	-3.42

Table 2: results of PT test (upperbound) in vegetable oil (2 different materials). All results were within the measurement uncertainty interval and Z-scores were 0.80 and 0.59 for materials 1 and 2 respectively.

	Reported	Target Value	Accuracy
	pg/g TEQ	pg/g TEQ	%
Material 1	1.110±0.20	1.01	8.8%
PCDD/Fs	0.80±0.19	0.89	-10.3%
DL-PCBs	1.90±0.36	1.90	-0.1%
Material 2			
PCDD/Fs	0.55±0.11	0.48	15.6%
DL-PCBs	0.82±0.21	0.85	-3.0%
Total TEQ	1.38±0.26	1.33	3.7%

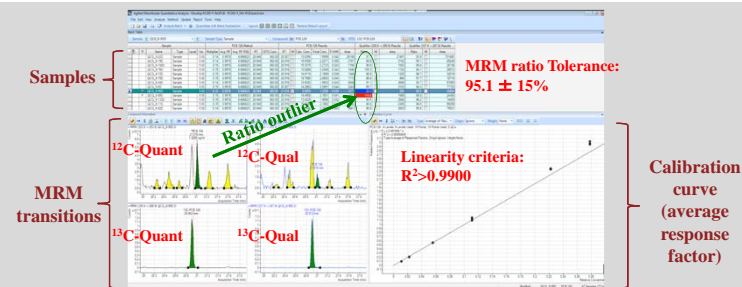


Fig. 1: Mass Hunter program, giving for a sample selected the 4 MRM transitions of a congener (bottom left). The outlier setup highlights out of tolerance MRM ratio, requiring a closer look (here, a wrong integration of <sup>12</sup>C-Quant transition Vs <sup>13</sup>C-Quant internal standard transition). Retention times and linearity can be controlled using outliers setup as well.

### Within lab reproducibility, QC control data, blank subtraction

Quality control (QC) and blank charts were recorded over 6 weeks and over 2 weeks after a 6 months break. A very good control was observed on QC samples (Fig. 2) and slightly worst on blank samples (Fig. 3). For the latter, LOQ's are defined with average blanks levels and they are acceptable outside the classical ±2sdev if they remain below the LOQ dashed line (+6sdev). We subtract from real samples an average blank value as soon as the blanks analyzed as control fall below the LOQ line. The variation below the LOQ level, and therefore the uncertainty on the 'true' blank value will be taken into account in the reported value and in the measurement uncertainty.

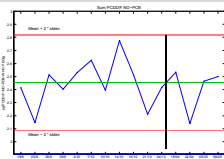


Fig. 2: control chart of QC pork fat over 6 weeks + after 6 months break (black line). Red line is mean + 2 stdev (n=12)

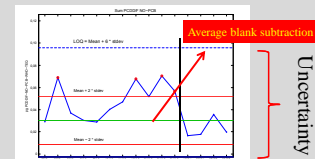


Figure 4: control chart of blank samples. Red line is mean + 2 stdev (n=12). Dashed blue is LOQ defined as mean + 6 stdev.

### Measurement uncertainty

A top-down approach is used to assess measurement uncertainty. Fortified samples used for accuracy test were used to determine the uncertainty on the bias (u<sub>bias</sub>) or systematic error, QC control data were used to determine the contribution of precision (u<sub>RW</sub>) in the uncertainty following Eq. 1.

$$\text{Eq. 1: } \%U = 2 \cdot \sqrt{\%U_{\text{bias}}^2 + \%U_{\text{RW}}^2}$$

We determined a relative measurement uncertainty of 18.5% for the total TEQ, similar than the uncertainty of the HRMS method.

