

Rates of microbial sulfur oxidation in low oxygen environments

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As one of the great biogeochemical cycles, the S cycle intimately couples with the C and N cycles [Garrels and Perry, 1974]. Microbial sulfide oxidation, in particular, likely represents a globally important yet poorly constrained source of biological carbon production and fixed nitrogen loss in many low oxygen environments [Walsh et al. 2009, Lavik et al. 2009, and Canfield et al. 2010]. Rates of microbial sulfide oxidation, however, are infrequently measured, likely due to the lack of standardized methods and protocols for these measurements. Bottle experiments yield information on the sulfide oxidation activity [Zopfi et al. 2001, Canfield et al. 2010], yet their coarse temporal resolution limits the extent to which reliable kinetic data can be extracted. We present here two novel methods for the determination of microbial sulfide oxidation rates in environmental systems: 1) based on the incubation of water samples in gas-tight glass syringes with sulfide determinations by the standard spectrophotometric technique; and 2) near real time measurement of sulfide oxidation using voltammetric techniques with Au/Hg amalgam microelectrodes. We have used these techniques individually and in concert to probe sulfide oxidation rates in diverse environments ranging from stratified East African lakes to marine oxygen minimum zones to laboratory cultures. Our measurements indicate that microbial sulfide oxidation is widespread in low oxygen environments, and is characterized by sulfide uptake with high affinity kinetics.

Garrels RM, Perry EA (1974) Cycling of carbon, sulfur, and oxygen through geologic time. *The Sea*, ed Goldberg ED (Wiley, New York), Vol 5, pp 303–336.

Walsh DA, Zaikova E, Howes CG, Song YC, Wright JJ, Tringe SG, Tortell PD, and Hallam SJ (2009), *Science*, 326, 578-582

Lavik G, Stuhrmann T, Bruchert V, Van der Plas A, Mohrholz V, Lam P, Mußmann M, Fuchs B, Amann R, Lass U, and Kuypers MMM (2009) Detoxification of sulphidic African shelf waters by blooming chemolithotrophs, *Nature*, 457, 581-584

Canfield DE, Stewart FJ, Thamdrup B, De Brabandere L, Dalsgaard T, Delong EF, Revsbech NP, and Ulloa O (2010) A Cryptic Sulfur Cycle in Oxygen-Minimum-Zone Waters off the Chilean Coast, *Science*, 330, 1375-1378

Zopfi J, Ferdelman TG, Jorgensen BB, Teske A, and Thamdrup B (2001) Influence of water column sulfide dynamics on sulfide oxidation and other major biogeochemical processes in the chemocline of Mariager Fjord (Denmark), *Marine Chemistry*, 74, 29-51