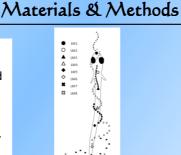
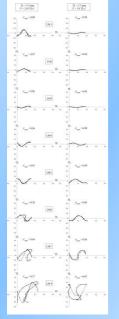
Swimming ontogeny in Dicentrarchus labrax Université de Liège 🗋 **Damien Olivier, Quentin Mauguit and Pierre Vandewalle** Laboratoire de Morphologie Evolutive et Fonctionnelle, Institut de Chimie (B6c) Université de Liège, 4000 Liège, Belgique. Correspondant: dolivier@student.ulg.ac.be Introduction Objectifs Swimming is an essential function involved in many behaviours such as predation, feeding, schooling etc. During growth, a lot of D. labrax Photo by Crocetta F., 2003 (www.fishbase.org) To determine when and parameters can influence swimming ontogeny how the swimming abilities In this study, we observed the as a changing in viscous and inertia forces swimming movements in take place throughout importance, flukes, skeletal, muscular and Dicentrarchus labrax from newly ontogeny. nervous systems developments. hatched larvae to juvenile stages.

Larvae of *D. labrax* were sampled from hatching to 288h (12 days post-hatching). Juveniles were collected on days 40, 79 and 104 post-hatching. Swimming movements were recorded using a high-speed video camera. On each frame, 8 equidistant landmarks were then plotted on fish midline.



The coordinates x-y were measured for tracing the landmarks trajectory during swimming movements. This allowed to calculate several parameters such as swimming speed, tail-beat frequency, amplitude and sinusoidal trajectory of each landmark.



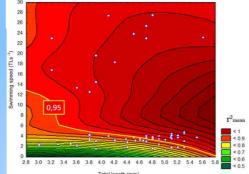
At hatching, only

swimming

movements at high

speed were

established (Fig. 1)



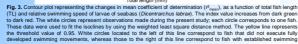


Fig. 2. (on the left) Trajectories of the eight landmarks on two Dicentrarchus labrax larvae, measuring respectively 3.5 and 5.5 mm TL during execution of one complete undiatory swimming movement. UM1 is located at the head-tip and LM8 at the tail-lp. Each continuous lines shows the observed trajectory and dotted line represents the theoretical sinusoid calculated from the observed path (which should correspond to adult motion). The data on the x and y axes are expressed as percentages of the total length of the observed fish. For LM7 on the 3.5 mm fish and for LM8 on both fish, the landmarks trajectories do not approach a sinusoid form. Therefore, it was impossible to determine the theoretical trajectory that should be followed by the landmarks.

During growth, we observed a

more sinusoidal trajectory of the

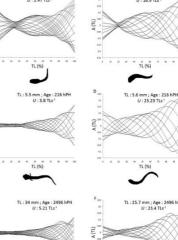
Landmarks (Fig. 2).

At 5.2 mm TL, all swimming

movements were established

(Fig. 1)

Fig. 4. (on the right) Midline kinematics of larvae and adults of seabase (Deartharchic) abravi, during cyclic swimming at different stages of onlegen (A-D. larvae, E-F. juveniles (A,O-E: crucies swimming; B-D.F. burst swimming). The superimposed midlines (time step 4 ms) of one tail-beat cycle show the amplitude envelope of the body wave. Each amplitude envelope of corresponds to the dat represented by the black shape. The amplitude is expressed as a proportion of total fish length (TJ, the x-axis represent the tail faih length (TJ, the x-axis represent the tail faih length (TJ, the x-axis represent the tail faih length (TJ), the x-axis represent the tail faih length (TJ).



π.(%)

Larvae swam with an anguilliform mode while juveniles displayed a subcarangiform mode. In anguilliform mode, the amplitude along fish body linearly increased backwards from the head to the tail. In subcarangiform mode, the increase was more exponential with the amplitude becoming important in the posterior part of fish body (see amplitude enveloppe on figure 4).

All these changes in the swimming movements during growth can be related to anatomical and morphological developments as flukes and skeletal, muscular and nervous systems.

Results & Discussion

Fig. 1. Drawing of a swimming movement executed by a larva of seabase (Dicentrarchus (abray). The eight pat made up of different kind of dos represent the trajectory of every landmarks (UM1, UA2,...) during the execution of