

Tandem in time mass spectrometry and immunoassay for PCDD/Fs screening

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Polychlorinated dibenzo-*p*-dioxins (PCDDs) and polychlorinated dibenzofurans (PCDFs) (“dioxins”) are of great concern because of their presence as contaminants (ppt level) in the environment. GC/HRMS is the reference method in the field of trace analysis but doesn’t allow a rapid and low cost screening. Ion trap GC/MS/MS has been presented by March (Splendore et al., 1997) as a valuable physico-chemical screening method for PCDD/Fs. The fragmentation of stored parent yields to the formation of characteristic daughter ions resulting from the loss of COCl^\bullet (Chess and Gross, 1980).

On the other hand, the use of a dioxin enzyme immunoassay (EIA) kit based on polyclonal antibody specific to 2,3,7,8-TCDD and related congeners can also be foreseen (Harrison and Carlson, 1997, 1998). It is presented when used together with MS/MS in screening campaigns.

MS/MS optimisation

Optimisation of the MS/MS method was mainly focused on daughter ions formation, (CID voltage, q_z) couple optimisation, excitation mode and CID time variation.

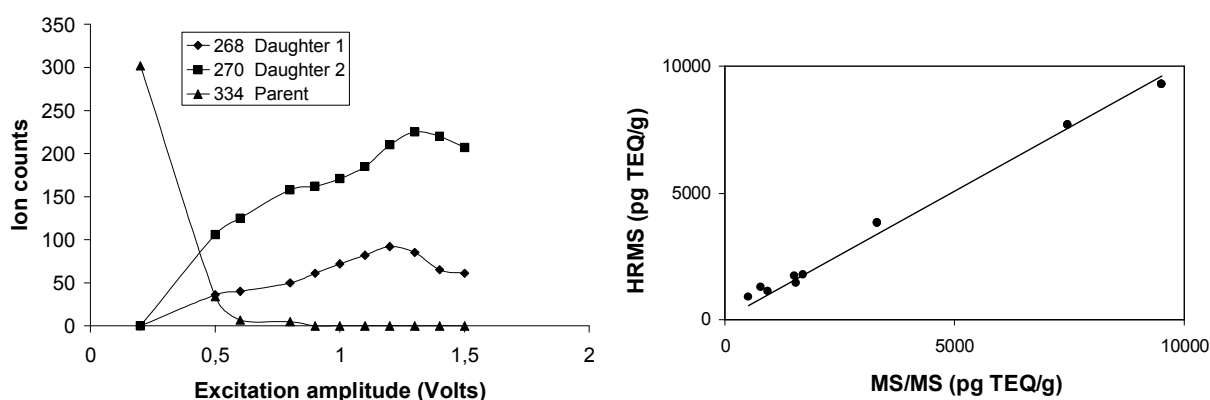


Fig. 1 : Global evolution of the TCDD species following CID voltage (left) and Correlation curve between MS/MS (Saturn 2000 GC/MS/MS) and HRMS (VG-AutoSpec-Q) fly ashes measurements.

Figure 1 (left) indicates the position of the optimum CID voltage for TCDD daughter ions formation; the good correlation (0.99, slope of 1.01) between MS/MS and HRMS is also illustrated.

Immunoassay semi-quantification

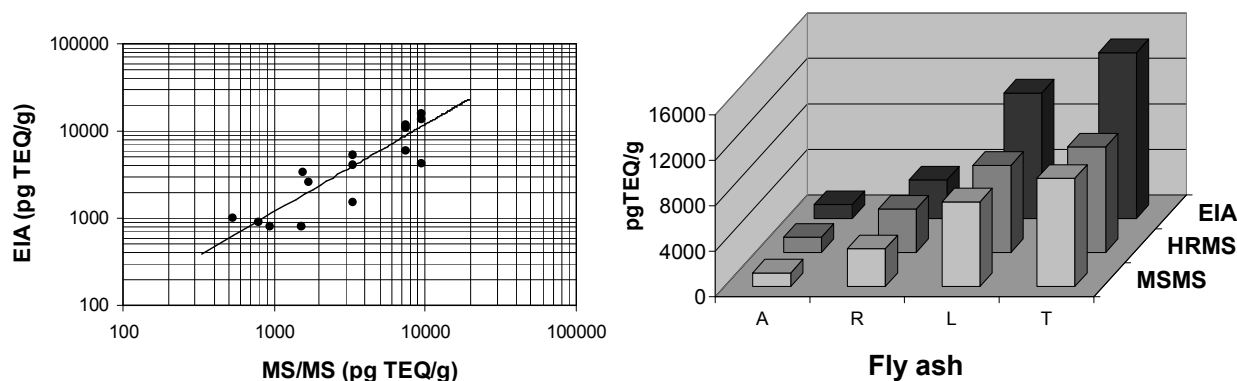


Fig. 2 : EIA response against MS/MS results on fly ashes (left) and overview of measurements (right).

EIA response follows quite well the evolution in the PCDD/Fs concentration (0.84, slope 1.19), EIA generally provides a general overestimation of the PCDD/Fs contents (Fig. 2).

A simple and time effective analytical strategy for the monitoring of the PCDD/Fs TEQ content in environmental samples is possible. This strategy can be described in 3 steps. A first sorting out using EIA can allow to separate negative from positive samples. In a second step, positive samples can be confirmed by GC/MS/MS. Nevertheless, a fully secure strategy should include the GC/MS/MS confirmation of negative EIA samples. Finally, the last step can be described as a confirmation of positive samples by HRGC/HRMS in order to allow precise congener and source identification. This combination of immunoassay and physico-chemical methods will reduce the global time and cost for efficient monitoring.

References

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