

1 **Potential use of milk mid-infrared spectra to predict individual**  
2 **methane emission of dairy cows**

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18 **ABSTRACT**

19         This study investigates the feasibility to predict individual methane (CH<sub>4</sub>)  
20 emissions from dairy cows using milk mid-infrared (MIR) spectra. To have a large  
21 variability of milk composition, two experiments were conducted on 11 lactating  
22 Holstein cows (2 primiparous and 9 multiparous). The first experiment aimed to  
23 induce a large variation in CH<sub>4</sub> emission by giving two different diets: the first one  
24 was mainly composed of fresh grass and sugar beet pulp and the second one of  
25 maize silage and hay. The second experiment consisted of grass and corn silage  
26 with cracked corn, soybean meal and dried pulp. Twice a day, for each milking  
27 period, the milk yields were recorded and a milk sample of 50 mL was collected from  
28 each cow and analyzed by MIR spectrometry. Individual CH<sub>4</sub> emissions were  
29 measured daily using the SF<sub>6</sub> method during a 7-day period. CH<sub>4</sub> daily emissions  
30 ranged from 10.2 to 47.1 g CH<sub>4</sub>/kg of milk. The spectral data were transformed to  
31 represent an average daily milk spectrum (AMS), which was related to the recorded  
32 daily CH<sub>4</sub> data. By assuming a delay before the production of fermentation products  
33 in the rumen and their use to produce milk components, five different calculations  
34 were used: AMS at days 0, 0.5, 1, 1.5, and 2 compared to the CH<sub>4</sub> measurement.  
35 The equations were built using Partial Least Squares regression. From the calculated  
36 R<sup>2</sup><sub>cv</sub>, it appears that the accuracy of CH<sub>4</sub> prediction by MIR changed in function of  
37 the milking days. In our experimental conditions, the AMS at day 1.5 compared to the  
38 measure of CH<sub>4</sub> emissions gave the best results. The R<sup>2</sup> and SE of the cross  
39 validation were equal to 0.79 and 5.14 g of CH<sub>4</sub>/kg of milk. The multiple correlation  
40 analysis done in this study showed the existence of a close relationship between milk  
41 fatty acid (FA) profile and CH<sub>4</sub> emission at day 1.5. The lower square root of the R<sup>2</sup>

42 (R = 0.87) obtained between FA profile and CH<sub>4</sub> emission compared to the one  
43 corresponding to the obtained calibration (R<sub>c</sub> = 0.93) shows the interest to apply  
44 directly the developed CH<sub>4</sub> equation instead of the use of correlations between FA  
45 and CH<sub>4</sub>. In conclusion, our preliminary results suggest the feasibility of direct CH<sub>4</sub>  
46 prediction from milk MIR-spectra. Additional research has the potential to improve the  
47 calibrations even further. This alternative method could be useful to predict the  
48 individual CH<sub>4</sub> emissions at farm level or at regional scale and it also could be used to  
49 identify low-CH<sub>4</sub>-emitting cows.

50 **Keywords:** methane, mid-infrared, milk, spectra, cows

51

## 52 **IMPLICATIONS**

53 **Potential use of milk mid-infrared spectra to predict the individual methane**  
54 **emission of dairy cows** *By Dehareng et al.* The disadvantages of existing methods  
55 for CH<sub>4</sub> measurement are the financial and human costs and/or the practical  
56 difficulties of these methods when they are used directly on field. The mid-infrared  
57 spectrometry used for milk analysis corrects this inconvenient. Therefore, the aim of  
58 this study was to develop an accurate and robust tool to predict the individual CH<sub>4</sub>  
59 emission of dairy cows by using mid-infrared spectrometry.

60 **INTRODUCTION**

61 Livestock farming is right at the forefront of climate change issues. Agriculture is  
62 regarded as the biggest producer of anthropogenic methane (**CH<sub>4</sub>**), mainly through  
63 ruminant gas emissions (FAO, 2010). Enteric CH<sub>4</sub> is produced by ruminants during  
64 the microbial digestion of feed in the rumen. Methane contributes widely to global  
65 warming, accounting for about 52 percent of the green house gases (**GHG**)  
66 emissions in both developing and developed countries (FAO, 2010). Methane  
67 absorbs 25 times as much infrared radiation than CO<sub>2</sub> enlarging global warming  
68 problems (Fuglestvedt, 2009). In addition to those environmental concerns, the  
69 eructed CH<sub>4</sub> induces a significant loss of gross energy intake for the animal. This gas  
70 loss corresponds between 3 and 10% of gross energy intake (Johnson and Johnson,  
71 1995, Jouany, 2008, Beauchemin *et al.*, 2009). These losses are mainly explained by  
72 the animal (age and species), the diet (intake level, composition) and the level of milk  
73 production (Vermorel, 1995). However, individual variations between animals of the  
74 same breed were reported in the range of 30-60% in production of CH<sub>4</sub> /unit of feed  
75 ingested when these animals received the same diet at equivalent ingestion and  
76 production levels (Lassey *et al.*, 1997). They are related to different rumen microbial  
77 populations, coming from different ruminal kinetics salivation; rumination (Demeyer  
78 and Fievez, 2000) and different feeding behaviors indicating interaction between  
79 animals and their microbiota. Recent results suggest therefore a genetic component  
80 in variation of individual CH<sub>4</sub> production (Martin *et al.*, 2010). However, most studies  
81 showed that ruminants that are genetically different for feed efficiency (e.g., Hegarty  
82 *et al.*, 2007) and productivity showed reduced daily CH<sub>4</sub> production (e.g., Bell *et al.*,  
83 2010). Given the complexity of this variation, large scale studies should be

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84 conducted, but no *in vivo* method to measure CH<sub>4</sub> emissions in ruminants is  
85 sufficiently accurate to be applied on a large number of animals (Clark, 2010). Indeed  
86 the available methods such as calorimeter or methane chambers, the tunnel system  
87 and the sulfur hexafluoride (**SF<sub>6</sub>**) tracer technique, are not easy to use on farm and  
88 are laborious to estimate individual methane production of many cows in a short  
89 period. Therefore, a simpler and appropriate method is required to predict individual  
90 CH<sub>4</sub> emissions. For this, the method investigated here was based on the fact that  
91 CH<sub>4</sub> synthesis depends on the kind of rumen fermentation which also influences  
92 many other parameters such as milk composition (Ørskov *et al.*, 1969).

93 Carbohydrates are the most important source of energy and the primary precursors  
94 of fat and lactose in cow's milk. The end products of carbohydrates degradation are  
95 the volatile fatty acids (**VFA**), CO<sub>2</sub>, CH<sub>4</sub> and H<sub>2</sub>. Demeyer and Fievez (2000) have  
96 developed an equation that shows clearly the relationship between VFA and CH<sub>4</sub>.  
97 During the fermentation process acetate and butyrate promote CH<sub>4</sub> production while  
98 propionate maintains a competitive role for the use of hydrogen. According to  
99 Miettinen and Huhtanen (1996), an increased ratio butyrate/propionate decreases the  
100 lactose content and increases the fat content in milk and the rumen CH<sub>4</sub> synthesis.  
101 Thus, a relationship among CH<sub>4</sub>, lactose, and fat contents could be assumed as  
102 suggested by Vlaeminck and Fievez (2005). In addition, the rumen VFA composition  
103 also influences milk fat composition. In ruminants, the milk fatty acids (**FA**) come from  
104 two sources, uptake from circulation and *de novo* synthesis within the mammary  
105 gland. About one-half of the milk FA (molar percent) is derived from *de novo*  
106 synthesis, based on acetate and butyrate (Bauman and Griinari, 2003). Short and  
107 medium-chains FA (4 to 14 carbons) arise totally from *de novo* synthesis. Long-chain

108 FA (>16 carbons) are collected in the circulating lipids, and FA of 16 carbons depend  
109 on these two sources (Bauman and Griinari, 2003). Thus, the proportions of different  
110 FA reflect the ruminal fermentation via VFA and then CH<sub>4</sub> production (Weill *et al.*,  
111 2008, 2009). Moreover, several authors have already reported, for different diets,  
112 relationships between milk FA (measured by gas chromatography) and CH<sub>4</sub> emission  
113 of dairy cows (Chilliard *et al.*, 2009; Dijkstra *et al.*, 2010; Delfosse *et al.*, 2010). These  
114 results suggest that in general, CH<sub>4</sub> emissions are linked to milk composition. As the  
115 Mid-Infrared (**MIR**) spectrum reflects the milk composition (Soyeurt *et al.*, 2006,  
116 2011), it is logical to postulate that the MIR spectrum could predict directly the  
117 individual emission of CH<sub>4</sub>. Therefore, the aim of this study was to have a first  
118 evaluation of the potential use of milk spectra obtained by Fourier Transform  
119 InfraRed analysis (**FTIR**) apparatus directly to predict the quantities of CH<sub>4</sub> eructated  
120 by individual dairy cows. The development of a MIR CH<sub>4</sub> equation will permit to  
121 predict and therefore to study at large scale the methane emission of dairy cows  
122 because the MIR technology is already implemented in milk labs to quantify the major  
123 milk components used for the milk payment and the routine milk recording.

## 124 **MATERIALS AND METHODS**

### 125 *Animals and Diets*

126 Two experiments were carried out with different experimental conditions in  
127 order to maximize the variability of individual CH<sub>4</sub> emission needed to establish a  
128 robust calibration model.

129 In the first experiment, 2 primiparous and 6 multiparous (3 of 2<sup>nd</sup> lactation and  
130 3 of 3<sup>rd</sup> lactation) lactating Holstein cows were divided into two groups of four cows

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131 each. At the beginning of the trial, the groups were similar in terms of lactation stage  
132 (180 days in milk) and of average milk production ( $17.4 \pm 3.9$  kg/d). Two isoenergetic  
133 experimental diets (17 kVEM) were offered according to a 2x2 cross-over design.  
134 Experimental diets were fed twice daily at 0900 and 1600 as total mixed rations  
135 (TMR). The diets were fed at an intake level of 18.5 kg/d and aimed at producing 20  
136 kg/d of milk. Diet 1 consisted of: fresh-cut pasture grass (third cutting, *Holcus*  
137 *lanatus*), 550 g/kg DM; dried beet pulp, 200 g/kg DM; soybean meal, 150 g/kg DM;  
138 and soybean hulls, 100 g/kg DM. Diet 2 consisted of: corn silage, 400 g/kg DM;  
139 meadow hay, 200 g/kg DM; cracked corn, 130 g/kg DM; rapeseed meal, 150 g/kg  
140 DM; palm meal, 55 g/kg DM; soybean meal, 55 g/kg DM; a 50:50 mix of coconut and  
141 flaxseed oil, 10 g/kg DM. Both diets contained a mixture of vitamins and minerals.

142 In the second experiment, 3 multiparous (2 of 2<sup>nd</sup> lactation and 1 of 4<sup>th</sup>  
143 lactation) lactating Holstein cows with a similar milk production ( $26.2 \pm 1.9$  kg/d) were  
144 fed with a same basal diet, with an intake level of 20.3 kg/d. This diet (TMR)  
145 consisted of grass silage, 520 g/kg DM; corn silage; 130 g/kg DM; cracked corn, 130  
146 g/kg DM; soybean meal, 110 g/kg DM; and dried beet pulp 110 g/kg DM.

147 For both experiments, the adaptation period was 21 days and milk and CH<sub>4</sub>  
148 samples were then collected from day 22 to day 28. Fresh water was available at all  
149 times.

150 All the procedures used involving the animals were approved by the regional  
151 ethics committee in animal experimentation (protocol CRAW09/01).

152

153 *Sampling and Analyses*

154 Milk yield was recorded daily at each sampling period at 0730 and at 1630. A  
155 50 mL aliquot of milk, containing sodium azide (0.32 g/L), was stored at 4°C until  
156 infrared analyses. The sample stored at 4°C was analyzed by a FTIR Lactoscope  
157 spectrometer (Delta Instruments, Drachten, the Netherlands). The Lactoscope works  
158 within 925 to 3000 cm<sup>-1</sup> and uses an interferometer. This instrument gave the  
159 spectral data as well as the direct measurement of milk components such as lactose,  
160 protein, fat, non protein nitrogen (**NPN**; considered to be essentially urea). Fatty acid  
161 composition predictions were also obtained based on equations specific to this  
162 machine. **Table 1** presents the statistical parameters of these equations. For each FA  
163 equations, the standard error of cross-validation (SECV) given by the FTIR  
164 Lactoscope equations are included between those given by Soyeurt *et al.* (2006) and  
165 those of Soyeurt *et al.* (2011). A total of 154 samples were collected and analyzed  
166 (spectra were in triplicate).

167 *Quantification of CH<sub>4</sub>*

168 During the 7-day period, the SF<sub>6</sub> gas tracer technique was used according to  
169 the method described by Martin *et al.* (2008) to measure the individual production of  
170 enteric CH<sub>4</sub> by the studied cows. Briefly, a calibrated permeable tube containing ultra  
171 pure SF<sub>6</sub> was placed in the rumen of each cow before the experimental period. The  
172 average release rate of SF<sub>6</sub> from the tubes was 1189.5 ± 168.6 ng/min.

173 A representative of breath gas sample, containing respired and eructated gas,  
174 was collected through a capillary tube situated between the nostril and the mouth of



175 each animal thanks to an halter. This gas sample was stored in a canister located  
 176 near the animal (more or less one meter) in a way that the animal could not touch it.  
 177 The canister was changed daily after morning feeding and the CH<sub>4</sub> and SF<sub>6</sub>  
 178 concentrations were analyzed using gas chromatography (**GC**) (Martin *et al.*, 2008).

179 A GC (Varian-Chrompack, CP-9003, Les Ulis, France) fitted with an electron  
 180 capture detector (**ECD**) (Perkin Elmer instruments; Autosystem XL, Courtaboeuf,  
 181 France) and with a flame ionisation detector (**FID**) was used to determine the  
 182 concentrations of SF<sub>6</sub> and CH<sub>4</sub>, respectively. The samples were run on GCs  
 183 equipped either with a Molecular Sieve 0.5 nm column (3 m × 3.2 mm i.d) maintained  
 184 at 50°C for the SF<sub>6</sub>, or with a Porapak N 80-100 mesh column (3 m × 3.2 mm i.d.)  
 185 maintained at 40°C for the CH<sub>4</sub>. The flow rate of the carrier gas was 30 mL/min of N<sub>2</sub>  
 186 for the SF<sub>6</sub> and 40 mL/min of He for CH<sub>4</sub>. Chromatographic analyses were performed  
 187 after calibration with standard gases (Air Liquide, Mityr-Mory, France) for SF<sub>6</sub> (55 and  
 188 195 ppt) and CH<sub>4</sub> (100 ppm).

189 To determine the released amount of CH<sub>4</sub> (**Q<sub>CH4</sub>**), the concentration of SF<sub>6</sub> and  
 190 CH<sub>4</sub> (**C<sub>CH4</sub>** and **C<sub>SF6</sub>**) in the canister and the pre-determined released rate of SF<sub>6</sub>  
 191 (**Q<sub>SF6</sub>**) were used. In parallel to each daily measurement, the concentration of the  
 192 atmospheric air (**C<sup>b</sup><sub>CH4</sub>** and **C<sup>b</sup><sub>SF6</sub>**) determined with another canister was subtracted  
 193 (Johnson *et al.*, 1994):

$$Q_{CH_4} = \frac{C_{CH_4} - C_{CH_4}^b}{C_{SF_6} - C_{SF_6}^b} Q_{SF_6} \frac{MW_{CH_4}}{MW_{SF_6}}$$

194 where MW<sub>CH<sub>4</sub></sub> and MW<sub>SF<sub>6</sub></sub> are the molecular weight of CH<sub>4</sub> and SF<sub>6</sub>,  
 195 respectively.

196           The CH<sub>4</sub> emission was therefore expressed in g of CH<sub>4</sub>/day and re-expressed  
197 later in g of CH<sub>4</sub>/kg of milk thanks to the recorded milk yield because the global daily  
198 CH<sub>4</sub> production is linked to productivity.

199

### 200 *Spectral Data Treatment*

201           Each cow was milked twice daily providing two individual samples, which were  
202 analyzed by MIR, thus 2 spectra, whereas individual CH<sub>4</sub> emission was measured  
203 once daily. Therefore, to build the calibration model, the recorded spectral data were  
204 transformed to represent one daily spectrum related to one daily CH<sub>4</sub> record. The  
205 methodology used to create the average milk spectra (**AMS**) was tested by  
206 comparing MIR predictions obtained from AMS and the real spectral data obtained  
207 from the infrared analysis of a representative daily milk sample (i.e., 50% of morning  
208 and 50% of evening milks). This ratio was chosen because it corresponds to the  
209 averaging used during the majority of milk recording process. Alternatively, the  
210 weighted average was also tested. It corresponded to the average of the two milk  
211 spectra of the day in proportion to the amount of milk produced by the cow in each  
212 respective milking (AM and PM). The reason for this was that a weighted average  
213 represented better the biological background of the process.

214           A dynamic relationship between milk composition (and, therefore, spectral  
215 data) and CH<sub>4</sub> was considered by assuming that there is a delay between the  
216 production of fermentation products and their use to produce milk components.  
217 Consequently, different ways exist to define the AMS. Indeed, the spectral data used

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218 to create the average milk spectra can be generated by the milk analysis of samples  
219 collected at different times compared to the time of CH<sub>4</sub> measurement. The average  
220 milk spectra were calculated from five methods averaging always two samples which  
221 are illustrated in Figure 1:

- 222 - the same day of CH<sub>4</sub> measurement (day 0),
- 223 - evening of the same day and morning next day (day 0.5),
- 224 - next day (day 1),
- 225 - evening next day and morning 2 days later (day 1.5),
- 226 - and 2 days later (day 2).

227 Always two samples were averaged to reflect a whole test-day in the milk as whole  
228 day was also represented in the CH<sub>4</sub> measurements.

229 Given the stability of diets and the high degree of auto-correlation that can be  
230 therefore assumed across values for the five methods, the objective of this was not  
231 infer a definitive response what the delay between the production of fermentation  
232 products and their use to produce milk components, but to find retrospectively which  
233 definition of the AMS gives the best predictions under the condition of this study.

### 234 *Calibration Model*

235 As mentioned previously, a calibration set with a large spectral variability is  
236 required to develop robust calibration models. Therefore, a principal component  
237 analysis (PCA) was carried out on available spectra at day 0 in order to illustrate the  
238 spectral variation observed from the collected milk samples.

239           The calibration models were developed by Foss WINISI 4 software from the  
240 recorded spectral and CH<sub>4</sub> data using Partial Least Squares regressions (**PLS**). The  
241 spectral regions used for that were: 972-1,589 cm<sup>-1</sup>, 1,720-1,782 cm<sup>-1</sup>, and 2,746-  
242 2,970 cm<sup>-1</sup>. No additional pre-treatment was applied on spectral data because with a  
243 derivative, results were not significantly better probably due to the fact that only one  
244 spectrometer was used in this experiment. The number of factors used was  
245 determined by a full cross-validation (with N observations, create N models by  
246 removing N time one sample which is predicted by the others N-1), which was also  
247 used to estimate its robustness. The accuracy of the resulting calibration models was  
248 evaluated by calculating the calibration coefficient of determination (**R<sup>2</sup>c**), cross-  
249 validation coefficient of determination (**R<sup>2</sup>cv**), standard error of calibration (**SEC**), and  
250 the standard error of cross-validation (**SECV**). The predictability of the calibration was  
251 evaluated through the ratio of performance to deviation (**RPD**; SD/SECV where **SD**  
252 was the standard deviation of the CH<sub>4</sub> measures). The RPD should be as high as  
253 possible; values between 5 and 10 are adequate for quality control, and values > 2.5  
254 are satisfactory for screening breeding programs (Williams and Sobering, 1993).

#### 255 *Correlations with Milk Components*

256           To show the interest of using a new direct MIR equation to quantify the CH<sub>4</sub>  
257 emissions, the best correlation between milk components already predicted by MIR  
258 and CH<sub>4</sub> was calculated. In this way, it was possible to verify whether the predicted  
259 CH<sub>4</sub> production data obtained by the developed calibration equation were due to new  
260 recombination of global spectral information or just to specific milk components  
261 already predicted by MIR and whose the equations are available. If the cross-

262 validation correlation is higher than the correlation between CH<sub>4</sub> and specific MIR  
263 milk component, it can be assumed that the developed equation provides additional  
264 information than the simple correlation between CH<sub>4</sub> and the already predicted milk  
265 components.

## 266 **RESULTS AND DISCUSSION**

### 267 *Spectral Variation*

268 The first two principal components described 56.8% of the total variation  
269 observed in the studied data set (N = 77). These principal components (Figure 2)  
270 were used to illustrate the spectral differences of studied milk samples. Spectra from  
271 cows feeding with different diets (i.e., grass silage, corn silage and fresh pasture) are  
272 separated (Figure 2). Based upon the literature (Collomb *et al.*, 2002), this suggests  
273 that the use of different diets involved a change in milk composition and therefore in  
274 the milk spectral data. Moreover, Table 2, with descriptive statistics, also shows high  
275 variations of milk composition primarily fat, protein, lactose, and saturated fatty acids  
276 (**SAT**). This variability of spectral data is needed to develop a robust calibration  
277 equation.

278 Table 2 also shows the variability of CH<sub>4</sub> expressed in g/day and g/kg of milk.  
279 The coefficients of variation were 29.84% and 33.33%, respectively. This large  
280 variation could be explained partly by the use of three different diets. A slightly larger  
281 variation was observed for the CH<sub>4</sub> emission expressed in g of CH<sub>4</sub>/kg of milk  
282 because the definition of this trait took into account simultaneously the variability of  
283 CH<sub>4</sub> and milk yield.

284

285 *Spectral Data Treatment*

286 As mentioned previously, for each cow, two spectra measurements and one  
287 CH<sub>4</sub> measurement were available. In order to have one spectrum for one measure of  
288 CH<sub>4</sub>, the collected spectral data from morning and evening milk samples were  
289 averaged. To validate the methodology, the contents of major milk components (i.e.,  
290 fat, protein, lactose, true protein, and NPE) predicted from AMS derived from 50%  
291 morning and 50% evening spectral data and from the spectral data obtained from the  
292 infrared analysis of a representative 24h milk sample were compared. Table 3 shows  
293 no significant difference ( $P > 0.005$ ) in milk components predicted by AMS and by the  
294 24h spectral data. Therefore, AMS was considered similar to the spectrum of the  
295 representative 24h milk sample showing the pertinence of the method used to create  
296 the AMS.

297 *Calibration Model*

298 Different equations were built to predict the quantity of CH<sub>4</sub> produced per day  
299 in function of the five different possibilities used to create AMS as shown in Figure 1.  
300 **Erreur ! Source du renvoi introuvable.** presents the statistical parameters obtained for  
301 the 20 developed calibration equations. The prediction of CH<sub>4</sub> by MIR, based on  
302 R<sup>2</sup>cv, was better by considering g of CH<sub>4</sub>/kg of milk instead of g of CH<sub>4</sub>/day  
303 (R<sup>2</sup>cv=0.79 vs. R<sup>2</sup>cv=0.73). This could be explained by the fact that the expression  
304 unit g of CH<sub>4</sub>/kg of milk compared to g of CH<sub>4</sub>/day takes into account the milk  
305 production, which is directly related to the release of CH<sub>4</sub> by cow (Vermorel, 1995)

306 and, therefore, reflected in the spectral data thanks to the dilution effect known for  
307 the majority of milk components. For instance, the literature shows that the contents  
308 of major milk components such as fat, protein, and fatty acids (Soyeurt *et al.*, 2009)  
309 are negatively correlated with the milk yield. **Table 4** shows also that the use of a  
310 ratio taking into account the milk produced during the evening and morning milkings  
311 for the calculation of AMS compared to the ratio 50% of morning and 50% of evening  
312 spectral data gave better results. This could be explained by the relationship between  
313 the milk production and the CH<sub>4</sub> emission by dairy cows (Vermorel, 1995). Therefore,  
314 the equations using the CH<sub>4</sub> content expressed in g/kg of milk and the morning and  
315 evening milk AMS gave the best results.

316 The interval between the measurement of CH<sub>4</sub> by the SF<sub>6</sub> tracer method and  
317 the spectral data used (Figure 1) gave different results (**Table 4**). By observing the  
318 last 5 lines of **Table 4**, the day 1.5 gave the best result for the internal validation (i.e.,  
319 R<sup>2</sup>cv=0.79). The best equation seems to be the one built from the AMS created from  
320 the weighted average taking into account the milk produced, the CH<sub>4</sub> content  
321 expressed in g/kg of milk, and an interval between the measurement of CH<sub>4</sub> and the  
322 spectral data equal to 1.5 day. This equation also seemed to be the most robust  
323 because the difference between R<sup>2</sup>c and R<sup>2</sup>cv was the lowest (0.08). These results  
324 are only first indications of the delayed response of milk composition to rumen  
325 fermentation. More research is needed in other experimental conditions; for instance  
326 by inducing the inhibition of the rumen methanogenesis and observing the milk  
327 spectra modifications in the next few days..

## Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra

328 Even if this study must be validated on a larger data set, the obtained results  
329 suggest a clear indirect link between the CH<sub>4</sub> emission and the milk composition  
330 through the direct use of the MIR spectral data. Chilliard *et al.* (2009) also showed  
331 this relation through the link between the contents of milk FA analyzed by GC and the  
332 production of CH<sub>4</sub> by dairy cows fed with diets supplemented or not with linseeds.  
333 The present study also shows the feasibility of the MIR prediction of CH<sub>4</sub> produced by  
334 dairy cows and a relative robustness of the developed equation. Indeed, despite of  
335 the large variation observed for g of CH<sub>4</sub>/kg of milk produced per cow and per diet  
336 over the study period and the observed variability of milk composition (**Table 2**), the  
337 MIR CH<sub>4</sub> predictions were good and R<sup>2</sup>c and R<sup>2</sup>cv were never below to 0.77 and  
338 0.68, respectively. Moreover, CH<sub>4</sub> prediction did not seem to be affected (Figure 3)  
339 by the different kind of diets used during the experiments (one consisting of corn  
340 silage, another of pasture grass and other of grass silage). However, it seems that  
341 the pasture grass diet contributes mainly to the large variation in CH<sub>4</sub> emissions. This  
342 variation between animals for this diet could be due to the season and therefore a  
343 higher lignification of grass at the end of the experimentation. Also individual, e.g.  
344 genetic, differences among the animals could be the reason for these results. In  
345 order to be able to make a generalisation of the prediction, it is important to continue  
346 to collect data on different diets. Finally, The RPD, which relates the standard error of  
347 prediction to the standard deviation of the original reference data, was equal to 2.19  
348 suggesting a good robustness of the prediction. Indeed, Williams and Sobering  
349 (1993) show that a value of 2.5 and over are satisfactory for screening. Therefore,  
350 using this equation, it is currently feasible to classify the dairy cows in two groups:  
351 low and high CH<sub>4</sub> producers. More variability in the available data can improve the



352 ability of the equation to predict CH<sub>4</sub> emission. Potential use of data obtained from  
353 genetically diverse animals, fed on different diets and producing in diverse  
354 management, should allow improving the equations.

#### 355 *Correlations with Milk Components*

356 Due to the strong indirect link existing between CH<sub>4</sub> emissions and the overall  
357 milk composition, it was expected that CH<sub>4</sub> emissions would correlate strongly with  
358 milk production and milk components already predicted by MIR and whose equations  
359 are available (**Table 5**). Despite what was expected (Miettinen and Huhtanen, 1996)  
360 a low correlation was observed between lactose and CH<sub>4</sub> emissions. This is probably  
361 because the lactose concentration is less and milk yield is more responding to  
362 variation in propionate supply. Nevertheless R<sub>cv</sub> for the best equation is always  
363 superior to the correlation obtained from milk production and MIR milk components  
364 suggesting that the developed equation provides additional information.

365 A multiple correlation (**Table 6**) was built starting from the parameters  
366 appearing in **Table 5** to predict CH<sub>4</sub> production/kg of milk for the day +1.5. The  
367 square root of the R<sup>2</sup> (R = 0.87) was lower than the square root of the R<sup>2</sup><sub>c</sub> value (R<sub>c</sub>  
368 = 0.93) obtain by the equation for this parameter in **Table 4**. In addition, the results  
369 presented in **Table 6** suggest the existence of a close relationship between FA  
370 profiles and CH<sub>4</sub> emission, as previously reported by Chilliard *et al.* (2009). However,  
371 our first results showed that the use of the full spectral data gave more information. In  
372 the future, the use of more precise equations for the prediction of the FA, could help  
373 us to understand more in detail the role of each FA and infirm or confirm these  
374 observation. In fact, the precision (SECV) of our FA equations could be upgraded

375 and met at least the precision obtain by Soyeurt *et al.* (2011). This underlines the  
376 practical interest to use MIR spectra to predict individual CH<sub>4</sub> emission of dairy cows.

377 **CONCLUSION**

378 The results of our study tended to show that the prediction of CH<sub>4</sub> emissions  
379 prediction from milk spectra is feasible. This is partially explained, by the fact, that the  
380 milk spectrum reflects the milk composition which is considered linked to CH<sub>4</sub>, due  
381 the relationship of both phenomena to ruminal fermentation. Results from this study  
382 showed that CH<sub>4</sub> is better predicted, based on R<sup>2</sup>cv, with the weighted AMS built  
383 from spectral data collected at the day 1.5 compared to the moment of CH<sub>4</sub>  
384 measurement. Additional studies providing data from genetically diverse animals and  
385 different breeds, fed on different diets and producing in diverse managements are  
386 needed to improve the obtained RPD and therefore clarify and potentially confirm the  
387 initial relationship which is reported in this study.

388 The calibration results indicated that the best equation could be used for  
389 screening purposes, differentiating high and low methane producers, even if addition  
390 research is still required to make it more reliable and potentially implementable on  
391 other spectrometers. Once this being achieved, as the existing methods for CH<sub>4</sub>  
392 measurement are difficult to apply on a large scale, this alternative method has the  
393 potential to be very useful to predict CH<sub>4</sub> emission for dairy herds at individual cow  
394 level from milk MIR spectra. These required data are already taken and used for the  
395 milk recording and the milk payment. Through the large scale generation of predicted  
396 CH<sub>4</sub> emission data, the method could help improve the knowledge about the sources  
397 of CH<sub>4</sub> variation (genetic or not) and about its link to other traits of interest.

398

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Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra

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499

500 **Table 1** Estimated statistical parameters for each calibration equation that  
 501 estimated the concentrations of fatty acid in milk (g / dl milk) used in this study

Fatty acids (g/dL of milk)	N	Mean	SD	SEC	R <sup>2</sup> <sub>C</sub>	SECV	R <sup>2</sup> <sub>CV</sub>	RPD
C4:0	137	0.11	0.03	0.01	0.82	0.01	0.73	1.93
C6:0	138	0.08	0.02	0.01	0.86	0.01	0.82	2.32
C8:0	131	0.05	0.01	0.00	0.86	0.01	0.81	2.33
C10:0	137	0.11	0.04	0.01	0.9	0.02	0.84	2.52
C12:0	138	0.13	0.05	0.02	0.88	0.02	0.84	2.48
C14:0	136	0.45	0.12	0.03	0.93	0.04	0.88	2.9
C14:1	138	0.04	0.02	0.01	0.66	0.01	0.58	1.52
C16:0	139	1.19	0.35	0.09	0.93	0.11	0.89	3.06
C16:1	143	0.08	0.04	0.02	0.82	0.02	0.77	2.06
C17:0	138	0.03	0.01	0.00	0.84	0.01	0.72	1.89
C18:0	138	0.39	0.15	0.07	0.79	0.07	0.76	2.05
C18:1 <i>trans</i>	138	0.10	0.04	0.02	0.67	0.03	0.55	1.49
C18:1 <i>cis-9</i>	138	0.80	0.42	0.08	0.97	0.09	0.95	4.5
Total C18:1 <i>cis</i>	138	0.87	0.43	0.07	0.98	0.08	0.97	5.38
Total C18:2	143	0.09	0.03	0.01	0.7	0.02	0.6	1.58
C18:2 <i>cis-9,cis-12</i>	142	0.06	0.02	0.01	0.77	0.01	0.7	1.83
C18:3 <i>cis-9,cis-12,cis-15</i>	138	0.02	0.01	0.01	0.52	0.01	0.4	1.29
C18:2 <i>cis-9,trans-11</i>	139	0.03	0.02	0.01	0.5	0.01	0.36	1.24
Saturated FA	132	2.72	0.72	0.06	0.99	0.07	0.99	10.57
Monounsaturated FA	135	1.12	0.46	0.06	0.98	0.07	0.98	6.45

502 SD : Standard deviation; SEC: Standard error of calibration; R<sup>2</sup><sub>C</sub> : Calibration coefficient of determination;  
 503 SECV: Standard error of cross-validation; R<sup>2</sup><sub>CV</sub> : Cross-validation coefficient of determination;RPD : Ratio of  
 504 standard error of cross validation to standard deviation

505

506



Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra

507 **Table 2** Means and range of CH<sub>4</sub> emissions, production and composition of  
 508 the milk, saturated fatty acid composition of the milk fat for the two experiments (day  
 509 0)

Components	N	Mean	Minimum	Maximum	Std dev
CH <sub>4</sub> , g/d	77	429	218	653	128
g/kg milk	77	21.9	10.2	47.1	7.3
Milk, kg/d	77	19.7	12.5	30	5.1
Milk Composition, g/kg					
Fat	77	4.11	2.86	5.63	0.61
Protein	77	3.40	2.39	4.82	0.42
Lactose	77	4.67	3.69	5.21	0.27
Milk Fat composition, g/kg					
SAT	77	66.1	55.4	76.9	5.2

510

511

512 **Table 3** Comparison between components prediction for average milk spectra  
 513 (AMS) and spectra of average milk in a ratio 50-50 for 37 milk sample during the  
 514 second experiment

Components	N	AMS	Average milk	P<
Fat milk <sub>predicted</sub> , g/kg	37	4.33	4.37	NS <sup>2</sup>
Protein <sub>predicted</sub> , g/kg	37	3.28	3.28	NS
Lactose <sub>predicted</sub> , g/kg	37	0.037	0.034	NS
True protein <sub>predicted</sub> , g/kg	37	3.31	3.33	NS
NPN <sup>1</sup> <sub>predicted</sub> , mg/100g <sub>milk</sub>	37	21.75	20.69	NS

515

1 Non Protein Nitrogen

516

2 NS: P>0.05

Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra

517 **Table 4** Evolution of accuracy of the resulting calibration with the different kind  
 518 of average milk spectra (AMS) with a ratio 50-50 and a weighted average for the two  
 519 experiments. The number of available data depends on the chosen day

Ratio	Day	N	R <sup>2</sup> c	R <sup>2</sup> cv	SEC	SECV	RPD	
Ratio 50-50	g of CH <sub>4</sub>	Day 0	77	0.84	0.71	73.00	98.30	1.84
		Day 0.5	71	0.81	0.68	79.39	105.19	1.77
		Day 1	65	0.84	0.72	71.73	98.29	1.88
		Day 1.5	60	0.77	0.70	87.77	100.65	1.83
		Day 2	59	0.77	0.72	86.19	95.23	1.89
	g of CH <sub>4</sub> /kg of milk	Day 0	77	0.80	0.72	4.81	5.69	1.91
		Day 0.5	71	0.83	0.77	4.39	5.69	2.06
		Day 1	65	0.86	0.69	4.16	6.13	1.81
		Day 1.5	60	0.85	0.75	4.25	5.61	2.01
		Day 2	59	0.93	0.72	3.06	6.05	1.89
Ratio defined from morning and evening milk yields produced	g of CH <sub>4</sub>	Day 0	77	0.85	0.72	68.97	95.93	1.90
		Day 0.5	71	0.83	0.73	74.82	95.45	1.92
		Day 1	65	0.85	0.69	71.55	102.21	1.80
		Day 1.5	60	0.78	0.71	86.89	99.20	1.85
		Day 2	59	0.77	0.72	85.81	83.81	1.89
	g of CH <sub>4</sub> /kg of milk	Day 0	77	0.89	0.75	3.52	5.42	2.00
		Day 0.5	71	0.86	0.78	3.96	5.00	2.12
		Day 1	65	0.84	0.69	4.35	6.22	1.78
		Day 1.5	60	0.87	0.79	4.06	5.14	2.19
		Day 2	59	0.90	0.73	3.59	5.94	1.93

520 SEC: Standard error of calibration; SECV: Standard error of cross-validation; RPD: ratio of performance to  
 521 deviation

522

523

524 **Table 5** Individual correlations between different milk parameters and the  
 525 CH<sub>4</sub>/kg of milk produced for the day1.5. Rcv = square root of the R<sup>2</sup>cv value

Milk production and Milk components <sup>1</sup>	Correlation CH <sub>4</sub> /Milk
Milk production	-0.45
Protein	0.40
Lactose	0.19
Fat	0.49
C4:0	0.26
C6:0	0.35
C8:0	0.40
C10:0	0.31
C12:0	0.34
C14:0	0.48
C14:1 <i>cis</i> -9	0.62
C16:0	0.32
C16:1 <i>cis</i> -9	0.48
C17:0	0.53
C18:0	0.25
Tot C18:1 <i>trans</i>	0.11
C18:1 <i>cis</i> -9	0.44
C18:2 <i>cis</i> -9, <i>cis</i> -12	0.23
C18:3 <i>cis</i> -9; <i>cis</i> -12, <i>cis</i> -15	0.60
c18:2 <i>cis</i> -9, <i>trans</i> -11	0.32
<b>Rcv</b>	<b>0.89</b>

<sup>1</sup> Milk components were predicted by FTIR spectrometer

526

## Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra

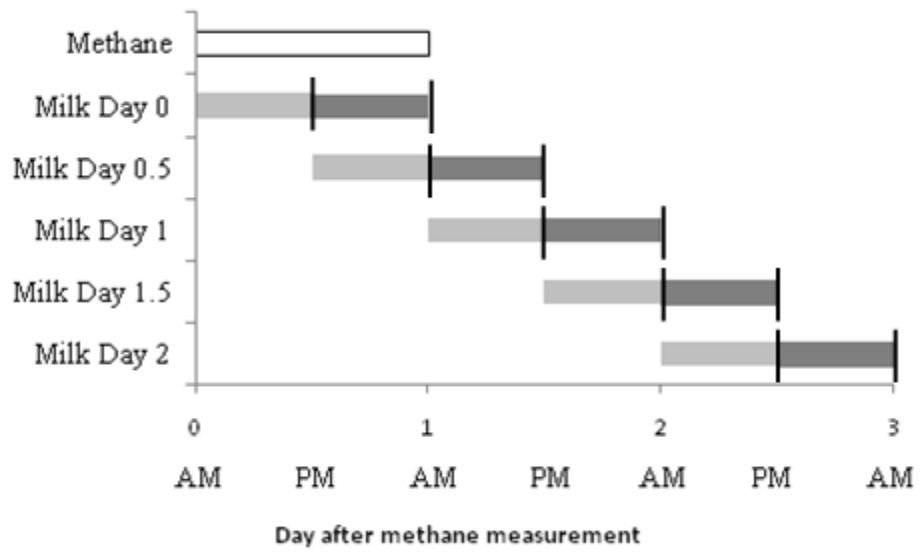
527

528 **Table 6** Multiple correlations from the parameters appearing in table 5 to  
529 predict the CH<sub>4</sub>/kg of milk produced for the day1.5. R = square root of the R<sup>2</sup> value

Correlation CH <sub>4</sub> /Milk	Name	F test
R : 0.87	1. C18:3 n-3	121.83
	2. C4:0	61.87
	3. C12:0	49.48
	4. C6:0	30.57
	5. C8:0	12.42

530

## Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra



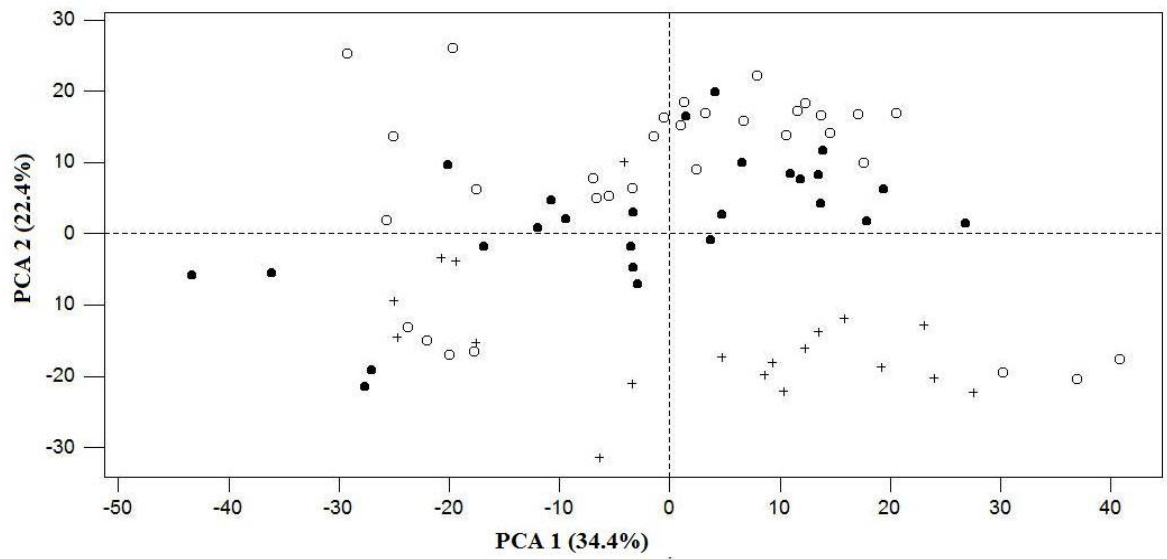
531

532 **Figure 1** Possibility to average the spectra milk in function of CH<sub>4</sub> day collect.

533 Vertical lines corresponds to milking time; surface shaded to milk production.

534

## Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra

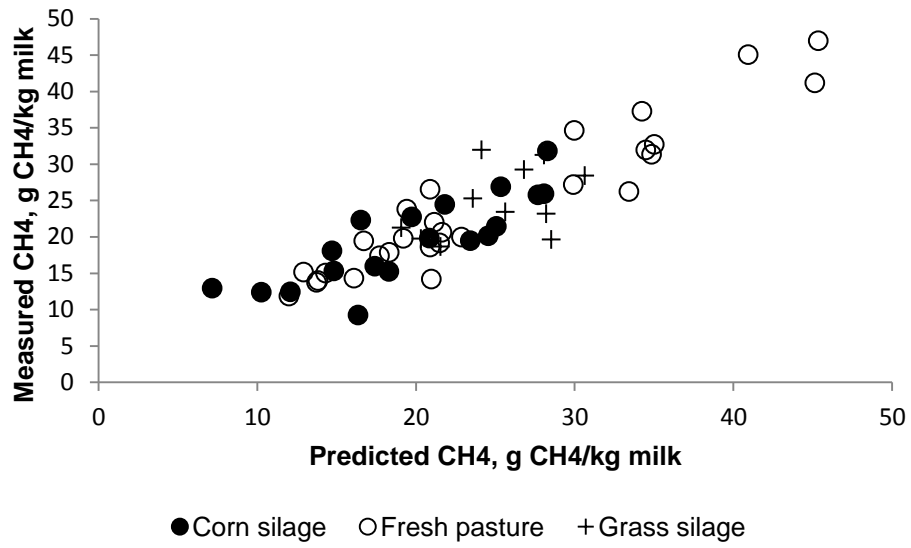


535

536 **Figure 2** Plot of the principal component analysis of milk spectra for individual cows  
537 fed three differing diets: corn silage (•), fresh pasture (o) and grass silage (+).

538

## Prediction of CH<sub>4</sub> Emission from Milk MIR Spectra



539

540 **Figure 3** Infrared CH<sub>4</sub> prediction on basis of milk spectra of the day1.5 for the  
541 different diets: corn silage (●), fresh pasture (○) and grass silage (+).