

# Nectar drinking

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**Nectar drinkers must feed quickly and efficiently due to the threat of predation. While the sweetest nectar offers the greatest energetic rewards, the sharp increase of viscosity with sugar concentration makes it the most difficult to transport. We here demonstrate that the sugar concentration that optimizes energy transport depends exclusively on the drinking technique employed. We identify three nectar drinking techniques: active suction, capillary suction and viscous dipping. For each, we deduce the dependence of the volume intake rate on the nectar viscosity, and thus infer an optimal sugar concentration consistent with laboratory measurements. Our results provide the first rationale for why suction feeders typically pollinate flowers with lower sugar concentration nectar than their counterparts that use viscous dipping.**

biomechanics | nectar | optimal concentration

Many insects and birds feed primarily or opportunistically on floral nectar. There are three principal techniques employed by nectar feeders: active suction, capillary suction, and viscous dipping. Lepidopterans (e.g. butterflies and moths) employ the former, sucking nectar through their probosci, along which a pressure gradient is generated by cibarial muscles [1, 2]. Nectar-feeding birds (e.g. hummingbirds<sup>1</sup> and sunbirds) employ capillary suction, in which capillary pressure drives flow along the tongue once its tip touches the nectar [3]. Most bees (except orchid bees) and some ants ingest nectar by dipping their tongue into, then extracting it from, the viscous nectar [4, 5]. It is advantageous for creatures to ingest energy rapidly due to the threat of predation during feeding. Optimal conditions might thus be sought to maximize their energy intake rate. While the sweetest nectar offers the greatest energetic rewards, the exponential increase of viscosity with sugar concentration [2] also makes it the most difficult to transport. We here rationalize the different optimal concentrations reported for the different drinking strategies by developing a dynamic model for viscous dipping and comparing it to existing models of suction feeding. Our new viscous dipping model indicates an optimal sugar concentration of 52%, which is higher than that for suction feeding, 33%. This result suggests a rationale for the fact that the nectar concentration of flowers pollinated by viscous dippers such as bees (35%) is typically higher than that of those pollinated by suction feeders such as hummingbirds or butterflies (20-25%) [6].

The sugar concentration that maximizes energy intake rate has been evaluated for a variety of nectar feeders in previous experimental studies (Fig. 1). Careful consideration of all of these results indicates that this so-called ‘optimal concentration’ depends exclusively on feeding mechanism but not on body size, quantity of intake, or species. Roughly speaking, the optimal concentration for active or capillary suction feeders is 30-40% while that for creatures using viscous dipping is 50-60%. Optimal sugar concentrations for suction feeders have been previously rationalized by Kingsolver & Daniel, who established dynamic models for both active [1] and capillary [3] suction. Pivnick & McNeil [2] advanced the active suction model by introducing the assumption of constant power output for the suction pump, and so predicted an optimal concentration of approximately 35%, consistent with that observed. Daniel *et al.* further demonstrated how the optimal concentration of 35% emerges for active suction when muscular mechanics is considered [7]. Kingsolver & Daniel [4] also suggested a dynamic model for bees that relies on capillary imbibition into the hairs of the tongue, a physical picture expected to be valid only in the limit of small nectar uptake rates.

Owing to its reliance on capillary suction, their model implies an optimal concentration of 30-40%, identical to that for suction feeders. However, for larger uptake volumes (Fig. 1), this suction model is no longer expected to be valid, and fails to rationalize the higher optimal concentrations of 50-60% reported, for example, for bees [8, 9]. We proceed by briefly reviewing the active suction model developed by Pivnick & McNeil [2] and the capillary suction model of Kingsolver & Daniel [3], then developing a new dynamic model for viscous dipping. The result is a global physical picture that describes all nectar feeders, and indicates that the optimal concentration depends exclusively on drinking style.

## Results

Suction feeding may be simply described mathematically. The pressure-driven flow of a fluid of density  $\rho$  and viscosity  $\mu$  along a tube of radius  $a$ , with mean speed  $u$ , is described by Newton’s second law:

$$h \frac{du}{dt} = \frac{\Delta P}{\rho} - \frac{8\mu h u}{\rho a^2} - gh, \quad [1]$$

where  $g$  is the gravitational acceleration and  $\Delta P$  the pressure difference applied at the height  $h$  of the nectar. For active suction,  $\Delta P$  is mainly generated by cibarial muscles [1, 2], while for capillary suction,  $\Delta P \sim \sigma/a$  results from curvature pressure, where  $\sigma$  is the surface tension [3]. A cornerstone of biomechanics is that the force that a creature of characteristic size  $l$  can generate [10]  $F \sim l^2$ ; thus, one expects the suction pressure generated by muscles,  $\Delta P \sim F/l^2 \sim l^0$ , to be independent of scale and to be of comparable magnitude for all creatures (e.g.  $\Delta P \sim 10$  kPa for both mosquitoes [4] and humans [11]). One can thus assess the tube scale  $a \sim \sigma/\Delta P \sim 10 \mu\text{m}$  below which curvature pressure dominates the applied suction pressure  $\Delta P$ . For most suction feeders, the radius  $a$  of the proboscis is of order  $100 \mu\text{m}$  [12, 13], so the curvature pressure is less than the pressure applied in active suction. Nevertheless, capillary suction is employed by certain creatures (Fig. 1) for which active suction is precluded by virtue of geometrical and physiological constraints such as the open, passive tongue of the hummingbird [13]. We further note that most suction feeders have tubes of characteristic length  $L \sim 1$  cm [12, 13]; consequently,  $\rho g L/\Delta P < 0.1$ , and the effect of gravity on the flows is negligible. Finally, the ratio of inertial to viscous terms scales as  $\rho a^2 f/\mu < 0.1$ , where  $f \sim 10$  Hz is the typical suction frequency [14, 15], indicating negligible in-

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<sup>1</sup>Rico-Guevara and Rubega [23] recently demonstrated that the hummingbird’s tongue folds around the nectar and suggested that this trapped fluid is then transported mouthward exclusively by tongue retraction. However, our recent observations indicate a meniscus rise speed of approximately 10 cm/s, greatly in excess of the tongue retraction speed, 2 cm/s, so clearly indicate capillary suction (Movie S1).

ertial effects. Neglecting the gravitational and inertial terms in (1) yields  $8\mu hu = a^2 \Delta P$ .

In active suction, the nectar motion is described by Poiseuille flow, for which the volumetric flow rate is given by  $Q = \pi a^2 u = \pi a^4 \Delta P / 8\mu L$ . By measuring the dependence of flow rate on sugar concentration, Pivnick [2] inferred that butterflies apply constant suction power in drinking, regardless of nectar concentration. The work per unit time required to overcome the viscous friction on the wall or power output  $\dot{W}$  of the pump is given by  $\dot{W} = Q\Delta P$ . Expressing  $\Delta P$  in terms of  $Q$  then yields the dependence of volume flux on viscosity:  $Q = (\pi a^4 \dot{W} / 8\mu L)^{1/2} \propto \mu^{-1/2}$ . In capillary suction,  $\Delta P = 2\sigma \cos \theta / a$ , where  $\theta$  denotes the contact angle, and the height of the nectar is time-dependent:  $h = h(t)$  and  $u = h'(t)$  (Fig. 1). The solution of the force balance,  $4\mu h h' = a\sigma \cos \theta$ , with initial condition  $h(0) = 0$  is given by  $h(t) = (a\sigma t \cos \theta / 2\mu)^{1/2}$ . Capillary suction consists of repeated cycles of tongue insertion and retraction. The whole time for a cycle is thus the sum of the time to absorb the nectar,  $T$ , and the time to unload it,  $T_0$ . The average volumetric flow rate per cycle,  $\bar{Q}$ , is given by  $\bar{Q} = \pi a^2 h(T) / (T + T_0) \propto (T/\mu)^{1/2} / (T + T_0)$ , where  $T^{1/2} / (T + T_0)$  depends weakly on viscosity [16], and so  $\bar{Q} \propto \mu^{-1/2}$ . Thus, for all suction mechanisms, we anticipate  $Q \propto \mu^{-1/2}$ .

To test these proposed scalings against experimental data, we introduce a general relation between  $Q$  and  $\mu$ :  $Q = X\mu^n$ , where  $X$  is a geometry-dependent prefactor that we expect to be different for each species. If we plot  $Q$  as a function of  $\mu$  on a log scale,  $n$  and  $X$  represent the slope and the offset on the y-axis respectively. For each species, we calculate an average value  $\langle X \rangle = \langle Q\mu^{-n} \rangle$  based on the measured dependence of flow rate on viscosity. In Fig. 2, red and blue points, respectively, indicate the dependence of  $Q/\langle X \rangle$  on  $\mu$  for active and capillary suction. The convincing collapse of the data, plus the fact that, for each species, the slopes are close to -1/2, together support the proposed scalings.

The energy intake rate  $\dot{E}$  is given by the product of the energy content per unit mass of sugar  $c$ , the sucrose concentration  $s$ , and the volumetric flow rate  $Q$ :  $\dot{E} = Q\mu c s \propto s \cdot \mu(s)^{-1/2}$ . For the sake of simplicity, density is treated as constant since its variation with sugar concentration is much less than that of viscosity. Considering the known dependence of nectar viscosity  $\mu(s)$  on  $s$  [2], the dependence of  $\dot{E}$  on  $s$  can be computed as shown in the inset of Fig. 2 and reveals an optimal concentration of 33% as inferred by Pivnick & McNeil for butterflies [2] and Kingsolver & Daniel for hummingbirds [3]. These predicted optimal concentrations are consistent with the results from the experimental studies reported in Fig. 1.

We proceed by presenting a new model for feeding in which the nectar intake relies on viscous entrainment by the outer surface of the tongue (Fig. 3 and Movie S1). Viscous dipping is generally characterized by an extendible tongue being immersed into nectar, coated, then extracted as shown in Fig. 3C, where a honeybee (*Apis*) drinks nectar from a reservoir. One expects the volume entrained to be proportional to the area of the immersed tongue surface and the thickness  $e$  of the nectar layer. As in capillary suction, the feeding by viscous dipping consists of repeated cycles. If  $T$  and  $T_0$  represent, respectively, the time needed for tongue retraction and the interval between each cycle, then the volumetric flow rate is given by  $\bar{Q} = 2\pi a e u T / (2T + T_0)$ , where  $u$  represents the average tongue retraction speed.

Encouraged by its success in the modeling of suction feeding, we introduce the assumption that the work rate applied in viscous dipping remains constant with respect to nectar concentration. The movement of the tongue in the fluid requires the power  $P_v \sim \mu L u^2$  to overcome the viscous drag, where  $L$  is the tongue length (Fig. 3). The power required for tongue acceleration  $P_t \sim m u' u \sim \rho a^2 u^3$ , where  $m \sim \rho a^2 L$  is the tongue mass. The ratio  $P_t / P_v \sim \rho u a^2 / \mu L \ll 1$ , so the effect of  $P_t$  is negligible. Assuming constant

applied power  $P_v$  thus suggests that  $u \propto \mu^{-1/2}$ . One does not expect  $T / (2T + T_0)$  to depend strongly on viscosity since if  $T$  is shorter in less viscous nectar due to a faster retraction, the unloading time  $T_0$  would also be shorter, so that  $T \propto T_0$ . Thus, the average volumetric flow rate may be expressed as  $\bar{Q} = 2\pi a e u T / (2T + T_0) \propto e \mu^{-1/2}$ .

The thickness of the fluid layer entrained by a cylinder of radius  $a$  depends explicitly on three dimensionless groups: the Bond number  $Bo = \rho g a^2 / \sigma$  (the ratio of hydrostatic to capillary pressures), the Weber number  $We = \rho u^2 a / \sigma$  (the ratio of inertial to curvature pressures), and the Capillary number  $Ca = \mu u / \sigma$  (the ratio of viscous stresses to curvature pressures). For bees,  $We \sim 10^{-3} \ll 1$ ,  $Bo \sim 10^{-3} \ll 1$ , and  $Ca < 0.1$  for  $s < 65\%$ , so the thickness of the liquid layer on a tongue is prescribed by the Landau-Levich-Derjaguin theory [17] that predicts  $e \sim Ca^{2/3} a$ . We thus anticipate that  $\bar{Q} \propto e \mu^{-1/2} \propto \mu^{-1/6}$ . In Fig. 2, this proposed scaling is validated by the data for all creatures that employ viscous dipping. The energy intake rate,  $\dot{E} = Q\mu c s$ , thus scales as  $\dot{E} \propto s \cdot \mu(s)^{-1/6}$ . In the inset of Fig. 2, the energy intake rate is plotted as a function of the sucrose concentration, and peaks at a concentration of 52%, which is consistent with the data presented in Fig. 1. Our analysis thus provides the first rationale for the different optimal concentrations reported for creatures using suction and viscous dipping. For example, we can now rationalize the observation that orchid bees that employ active suction have optimal concentrations of 35%, while honeybees and bumblebees that use viscous dipping, 50-60% [18].

## Discussion

A relatively complete physical picture of the fluid dynamics of nectar feeding has emerged. First and foremost, the optimal nectar concentration for a given creature depends only on its drinking style, being higher for viscous dippers than suction feeders. When considered in light of the coevolution between flowers and pollinators, this deduction provides rationale for the observation that the nectar concentration of flowers pollinated by bees (35%) is generally higher than that of those pollinated by butterflies and hummingbirds (20-25%) [6]. Several caveats are in order, however. The optimal concentration in the laboratory might differ from that preferred in nature due to the limited availability of nectar in the wild. Specifically, in addition to energy intake rate, nectar feeders in the wild presumably consider competition between other individuals or colonies [9], and travel costs [19]. Moreover, the nectar concentration proffered by flowers need not correspond to the optimal value owing to the pollination strategy of flowers [20]. Indeed, it has been suggested that flowers try to keep their pollinators hungry and faithful [21] because too great an energy reward would decrease the inter-flower movement of pollinators, and too small a reward would bring about desertion of the pollinators. Hence, the optimal concentrations suggested by dynamic models still need to be carefully scrutinized in attempts to understand the cues of coevolution between flowers and nectar feeders.

## Methods

Five bumblebees were captured in Cambridge, MA in April, 2010. One of the bees was kept in a transparent 50 cm  $\times$  50 cm  $\times$  30 cm cage, with the bottom covered in paper towel soaked in a sucrose solution. Three hours after capture, we took still photographs (with a Nikon AF-S DX still camera) of the bee protruding its tongue to lap nectar from the substrate (Fig. 3a). The tongue of a deceased bee was cleaned in a methanol solution, naturally dried, and examined with an SEM (Zeiss ultra 55 FE-SEM) (Fig. 3b).

Live honeybees were purchased from a merchant in Paju, South Korea in June, 2010. One of the bees was confined to a cylindrical cage of inner diameter 15 mm. One end of the cylindrical cage had a window through which the bee could extrude its head. A feeder made with glass blocks was filled with a 40% (by mass) sucrose solution and placed sufficiently close to the window that the bee could drink

from it. We filmed the drinking process with a high speed camera (Photron APX-RS) operating at 250 frames per second with a zoom lens (Navitar 12X Zoom) (Fig. 3c and Movie S1).

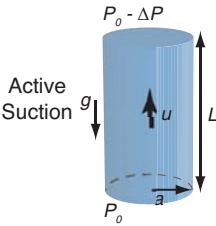
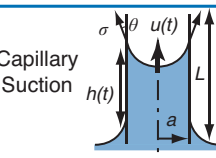
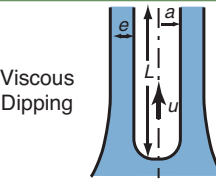
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**Fig. 1.** Optimal sugar concentrations for various nectar feeders [22]. The optimal concentration is that for which the energy intake rate is highest based on drinking rates measured at various nectar concentrations in a laboratory setting.

**Fig. 2.** The dependence of scaled volumetric flow rate  $Q/\langle X \rangle$  on nectar viscosity  $\mu$ . The red points represent data for active suction, the blue points for capillary suction, and the green points for viscous dipping. The slopes of the expected lines for suction and viscous dipping are  $-1/2$  and  $-1/6$  respectively. Inset: optimal concentrations of 33% and 52% are evident for, respectively, suction feeding and viscous dipping from the dependence of relative energy intake rate on nectar viscosity. Characteristic error bars are shown.

**Fig. 3.** Bees uptake nectar via viscous dipping. a) A bumblebee (*Bombus*) drinking. While normally protruding straight from the galeae, the tongue here bends to lap up nectar from the substrate, a paper towel soaked in a sucrose solution. b) Scanning Electron Microscope (SEM) image of the bumblebee's tongue. c) A schematic illustration of the experiment that allows us to visualize the viscous dipping of a honeybee (*Apis*) with a long-distance microscope and a high-speed camera operating at 250 frames per second. Here, the bee's tongue is dipped into a 40% sucrose solution, then withdrawn (see Movie S2).

Mechanism	Name	Genus	Optimal (%)
<p>Active Suction</p> 	Ants	<i>Atta</i> (5) <i>Camponotus</i> (5)	30 40
	Bees	<i>Euglossa</i> (18)	35
	Butterflies	<i>Agraulis</i> (12)	40
		<i>Phoebis</i> (12)	35
		<i>Speryeria</i> (24)	35
		<i>Thymelicus</i> (2)	40
		<i>Vanessa</i> (25)	40
	Moths	<i>Pseudaletia</i> (2) <i>Macroglossum</i> (26) <i>Manduca</i> (27)	40 35 30
<p>Capillary Suction</p> 	Humming-birds	<i>Selasphorus</i> (16) <i>Selasphorus</i> (28)	35-45
	Honey-eaters	<i>Anthochaera</i> (29)	50
		<i>Phylidonyris</i> (29)	40
		<i>Acanthorhynchus</i> (29)	30
	Sunbirds	<i>Cinnyris</i> (30)	30
<p>Viscous Dipping</p> 	Ants	<i>Pachycondyla</i> (5) <i>Rhytidoponera</i> (5)	50 50
	Bees	<i>Bombus</i> (8)	55
		<i>Apis</i> (9)	55
		<i>Melipona</i> (9)	60
	Bats	<i>Glossophaga</i> (31)	60

